

Draft Genome Sequences of *Marinobacter* Strains Recovered from Utica Shale-Produced Fluids

Shantal Tummings,^a Jenny Panescu,^a Rebecca A. Daly,^b Kelly C. Wrighton,^b [©]Paula J. Mouser^{a*}

^aDepartment of Civil, Environmental, and Geodetic Engineering, The Ohio State University, Columbus, Ohio, USA

^bDepartment of Microbiology, The Ohio State University, Columbus, Ohio, USA

ABSTRACT The genomes of three *Marinobacter* strains, isolated from saline fluids produced from a Utica-Point Pleasant shale well, have been sequenced. These genomes provide novel information on the degradation of petroleum distillates and virulence mechanisms under microaerophilic conditions in fractured shale.

Prizontal drilling coupled to hydraulic fracturing well completion methods are the industry standard for recovering hydrocarbon from low-permeability black shales (1, 2). Microorganisms play a role in degrading shale-derived hydrocarbons (3) and altering xenobiotic organic compounds introduced during the fracturing process (4). The bacterial strains *Marinobacter persicus* UTICA-S1B3, UTICA-S1B6, and UTICA-S1B9 were isolated from saline fluids produced from a Utica-Point Pleasant shale well in Ohio. Cosmopolitan *Marinobacter* species are distributed across marine lakes, oceans, sediments, and deep mines (5–8), especially where hydrocarbons are present (8, 9), and recently were observed in fractured shale brines (4, 10–12). Here, we describe the genomic sequencing of three *Marinobacter* strains isolated from fractured shale and highlight their capacity for aromatic compound degradation and bacterial virulence.

The *Marinobacter* isolates were cultivated from fluids collected on the first day of flowback in Difco marine broth 2216 medium supplemented with 40 mM nitrate at 30°C. Cells were harvested via centrifugation, and genomic DNA was isolated using a DNA minikit (Qiagen, Hilden, Germany), with sequencing performed at the Department of Energy Joint Genome Institute (Walnut Creek, CA, USA). Assemblies were constructed from Illumina MiSeq sequence data (SPAdes version 3.6.2) and generated 94, 96, and 100 contigs for UTICA-S1B3, UTICA-S1B6, and UTICA S1B9, respectively, with a G+C content of 57.7% and 92% genome completeness. Annotation was performed in the Integrated Microbial Genomes platform (Pipeline version 4.12.1) and resulted in 3,287, 3,294, and 3,288 protein-coding genes for UTICA-S1B3, UTICA-S1B6, and UTICA S1B9, respectively. While these strains had an average nucleotide identity (ANI) of 99.9% to each other, their genomes were more distantly related to *Marinobacter persicus* IRBC-M 10445 (ANI, 83%) and *Marinobacter hydrocarbonoclasticus* ATCC 49840 (ANI, 78%).

These *Marinobacter* strains have the genomic potential to degrade toluene and benzene to (methyl)catechol using phenol 2-monooxgenases, further metabolizing catechol through *meta*-cleavage to formate, acrylate, pyruvate, or acetyl-coenzyme A (acetyl-CoA). All three strains contain genes for denitrification and alternative nitrogen source utilization (e.g., urea). Of 35 predicted cytochromes in each genome, 15 cytochromes are annotated for (per)oxidase activity, which may be important for outer membrane processes, including iron oxidation. Unlike other *Marinobacter* species that utilize a type IV secretion system (13), the *Marinobacter* strains encode a type VI secretion system that has a known role in the delivery of toxic effectors to other bacteria using a phage-like tubule (14). Specifically, the three strains can target the peptidoglycan of recipient bacterial cells using amidases and proteases or attack outer

Received 24 February 2018 Accepted 16 March 2018 Published 5 April 2018

Citation Tummings S, Panescu J, Daly RA, Wrighton KC, Mouser PJ. 2018. Draft genome sequences of *Marinobacter* strains recovered from Utica shale-produced fluids. Genome Announc 6:e00155-18. https://doi.org/10.1128/ genomeA.00155-18.

Copyright © 2018 Tummings et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Paula J. Mouser, Paula.Mouser@unh.edu.

* Present address: Paula J. Mouser, Department of Civil and Environmental Engineering, University of New Hampshire, Durham, New Hampshire, USA. membranes using phospholipases (14, 15). Amidases may also catalyze the degradation of polyacrylamides (16), which are common additives used during slick-water hydraulic fracturing (17). The isolation of *Marinobacter persicus* UTICA-S1B3, UTICA-S1B6, and UTICA S1B9 provides new insight into hydrocarbon metabolism, polymer degradation, and opportunistic survivability in the shale ecosystem.

Accession number(s). The whole-genome sequences for *M. persicus* UTICA-S1B3, UTICA-S1B6, and UTICA-S1B9 have been deposited in DDBJ/ENA/GenBank under accession numbers PTIV00000000, PTIT00000000, and PTIU00000000, respectively, and can be accessed at the JGI Integrated Microbial Genomes and Microbiome database under the IMG genome identification (ID) numbers 2700989663, 2700989662, and 2700989665, respectively.

ACKNOWLEDGMENTS

This research was performed under a sequencing award to K.C.W. (numbers 1777 and 1931) and conducted by the U.S. Department of Energy (DOE) Joint Genome Institute, a DOE Office of Science user facility, supported by the Office of Science of the U.S. Department of Energy under contract DE-AC02-05CH11231. J.P., R.A.D., K.C.W., and P.J.M. were partially supported by funding from the National Science Foundation (NSF) Dimensions of Biodiversity (award 1342701), and P.J.M. and S.T. were partially supported by NSF Chemical, Bioengineering, Environmental, and Transport Systems (award 1336326).

We are especially grateful to our industry partner for providing site access and assistance with sampling.

REFERENCES

- Arthur J, Langhus B, Alleman D. 2009. Modern shale gas development in the United States: a primer U.S. Department of Energy, Office of Fossil Energy, Washington, DC.
- Vidic RD, Brantley SL, Vandenbossche JM, Yoxtheimer D, Abad JD. 2013. Impact of shale gas development on regional water quality. Science 340:1235009. https://doi.org/10.1126/science.1235009.
- Fredrickson JK, Balkwill DL. 2006. Geomicrobial processes and biodiversity in the deep terrestrial subsurface. Geomicrobiol J 23:345–356. https://doi.org/10.1080/01490450600875571.
- Daly RA, Borton MA, Wilkins MJ, Hoyt DW, Kountz DJ, Wolfe RA, Welch SA, Marcus DN, Trexler RV, MacRae JD, Krzycki JA, Cole DR, Mouser PJ, Wrighton KC. 2016. Microbial metabolisms in a 2.5-km-deep ecosystem created by hydraulic fracturing in shales. Nat Microbiol 1:16146. https:// doi.org/10.1038/nmicrobiol.2016.146.
- Bagheri M, Amoozegar MA, Didari M, Makhdoumi-Kakhki A, Schumann P, Sproer C, Sanchez-Porro C, Ventosa A. 2013. *Marinobacter persicus* sp nov., a moderately halophilic bacterium from a saline lake in Iran. Antonie Van Leeuwenhoek 104:47–54. https://doi.org/10.1007/s10482 -013-9923-3.
- Kaye JZ, Baross JA. 2000. High incidence of halotolerant bacteria in Pacific hydrothermal-vent and pelagic environments. FEMS Microbiol Ecol 32:249–260. https://doi.org/10.1111/j.1574-6941.2000.tb00718.x.
- Bonis BM, Gralnick JA. 2015. Marinobacter subterrani, a genetically tractable neutrophilic Fe(II)-oxidizing strain isolated from the Soudan Iron Mine. Front Microbiol 6:719. https://doi.org/10.3389/fmicb.2015.00719.
- Gauthier MJ, Lafay B, Christen R, Fernandez L, Acquaviva M, Bonin P, Bertrand JC. 1992. *Marinobacter hydrocarbonoclasticus* gen. nov., sp. nov, a new, extremely halotolerant, hydrocarbon-degrading marine bacterium. Int J Syst Bacteriol 42:568–576. https://doi.org/10.1099/00207713 -42-4-568.
- Atlas RM, Stoeckel DM, Faith SA, Minard-Smith A, Thorn JR, Benotti MJ. 2015. Oil biodegradation and oil-degrading microbial populations in marsh sediments impacted by oil from the Deepwater Horizon well

blowout. Environ Sci Technol 49:8356–8366. https://doi.org/10.1021/acs .est.5b00413.

- Cluff MA, Hartsock A, MacRae J, Carter KE, Mouser PJ. 2014. Temporal changes in microbial ecology and geochemistry in produced water from hydraulically fractured marcellus shale gas wells. Environ Sci Technol 48:6508–6517. https://doi.org/10.1021/es501173p.
- Fichter J, Moore R, Braman S, Wunch K, Summer E, Holmes P. 2012. How hot is too hot for bacteria? A technical study assessing bacterial establishment in downhole drilling, fracturing, and stimulation operations Abstr NACE International Corrosion Conference & Expo, 11 to 15 March 2012, Salt Lake City, UT.
- Mohan AM, Bibby KJ, Lipus D, Hammack RW, Gregory KB. 2014. The functional potential of microbial communities in hydraulic fracturing source water and produced water from natural gas extraction characterized by metagenomic sequencing. PLoS One 9:e107682. https://doi .org/10.1371/journal.pone.0107682.
- Singer E, Webb EA, Nelson WC, Heidelberg JF, Ivanova N, Pati A, Edwards KJ. 2011. Genomic potential of *Marinobacter aquaeolei*, a biogeochemical "opportunitroph." Appl Environ Microbiol 77:2763–2771. https://doi .org/10.1128/AEM.01866-10.
- Russell AB, Peterson SB, Mougous JD. 2014. Type VI secretion system effectors: poisons with a purpose. Nat Rev Microbiol 12:137–148. https:// doi.org/10.1038/nrmicro3185.
- Sang G, Pi Y, Bao M, Li Y, Lu J. 2015. Biodegradation for hydrolyzed polyacrylamide in the anaerobic baffled reactor combined aeration tank. Ecol Eng 84:121–127. https://doi.org/10.1016/j.ecoleng.2015.07.028.
- Ma F, Wei L, Wang L, Chang C-C. 2008. Isolation and identification of the sulphate-reducing bacteria strain H1 and its function for hydrolysed polyacrylamide degradation. Int J Biotechnol 10:55–63. https://doi.org/ 10.1504/JJBT.2008.017979.
- King GE. 2010. Thirty years of gas shale fracturing: what have we learned? SPE 133456. SPE Annual Technical Conference and Exhibition, 19 to 22 September 2010, Florence, Italy.