



Draft Genome of *Thermanaerothrix daxensis* GNS-1, a Thermophilic Facultative Anaerobe from the *Chloroflexi* Class *Anaerolineae*

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We present the draft genome of *Thermanaerothrix daxensis* GNS-1, a thermophilic member of the *Chloroflexi* phylum. This organism was initially characterized as a nonmotile, strictly anaerobic fermenter; however, genome analysis demonstrates that it encodes genes for a flagellum and multiple pathways for aerobic and anaerobic respiration.

Received 29 September 2015 Accepted 5 October 2015 Published 19 November 2015

Citation Pace LA, Hemp J, Ward LM, Fischer WW. 2015. Draft genome of *Thermanaerothrix daxensis* GNS-1, a thermophilic facultative anaerobe from the *Chloroflexi* class *Anaerolineae*. Genome Announc 3(6):e01354-15. doi:10.1128/genomeA.01354-15.

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Thermanaerothrix daxensis GNS-1 was isolated from a deep groundwater aquifer (149 m) housed within sedimentary strata of the large Mesozoic and Tertiary Aquitaine Basin in southwestern France (1). Closely related strains have been reported from a hot spring in Yellowstone National Park (2), a hot spring in southwestern Taiwan, geothermal soil, a thermophilic anaerobic digestive sludge, and a thermophilic electrochemical cell (3). *T. daxensis* is a filamentous, nonsporulating organism that can ferment a number of sugars and organic acids (1). It grows optimally at 65°C (range 50 to 73°C) and pH 7 (range pH 5.8 to 8.5) (1).

The genome of *Thermanaerothrix daxensis* GNS-1 (DSM 23592) was sequenced as part of a project to expand the phylogenetic breadth of *Chloroflexi* genomes. Genome sequencing was performed at Seqmatic using the Illumina MiSeq sequencing platform. SPAdes version 3.1.1 (4) was used to assemble the genome. The genome was screened for contaminants based on sequence coverage, GC composition, and BLAST hits of conserved singlecopy genes. Genome annotation was performed using the NCBI Prokaryotic Genome Annotation Pipeline. The draft genome is 3.06 Mb in size, assembled into 6 contigs. It encodes 2,798 genes, 2,395 coding sequences, 1 16S RNA, 47 tRNAs, and 4 CRISPR arrays. It is estimated to be ~95% complete based on conserved single-copy genes (106/111).

Genome analysis of *T. daxensis* detected the presence of aerobic and anaerobic respiration pathways, hinting at a richer physiology than previously recognized. It encodes for Complex I (NADH dehydrogenase), Complex II (succinate dehydrogenase), and an aerobic CO dehydrogenase. It also has two aerobic respiration modules; an A-family heme-copper oxygen reductase coupled to an alternative complex III (ACIII) (5), and a quinol *bd* oxidase (6). In addition, *T. daxensis* has two respiratory nitrite reductases; NirS, which reduces NO₂⁻ to NO, and NrfA that reduces NO₂⁻ to NH₄⁺. The genome provides no evidence for the presence of LPS biosynthesis genes or outer membrane proteins, suggesting that this organism has only one membrane (7). Furthermore, it encodes for a Gram-positive flagella and is likely motile under certain physiological conditions. Nucleotide sequence accession number. This whole-genome shotgun project has been deposited in DDBJ/EMBL/GenBank under the accession number LGKO00000000.

ACKNOWLEDGMENTS

Genomic DNA was obtained from the Leibniz-Institut DSMZ-Deutsche Sammlung von Mikroorganismen und Zellkulturen GmbH. Sequencing was performed at Sequatic, Fremont, CA, USA.

This work was funded in part by the Center for Environmental Microbial Interactions (CEMI) at Caltech, the Packard Foundation (W.W.F.), the Agouron Institute (J.H. and W.W.F.), and NSF GRFP (L.M.W.).

REFERENCES

- Grégoire P, Fardeau M, Joseph M, Guasco S, Hamaide F, Biasutti S, Michotey V, Bonin P, Ollivier B. 2011. Isolation and characterization of *Thermanaerothrix daxensis* gen. nov., sp. nov., a thermophilic anaerobic bacterium pertaining to the phylum "*Chloroflexi*", isolated from a deep hot aquifer in the Aquitaine basin. Syst Appl Microbiol 34:494–497. http:// dx.doi.org/10.1016/j.syapm.2011.02.004.
- 2. Hugenholtz P, Pitulle C, Hershberger KL, Pace NR. 1998. Novel division level bacterial diversity in a Yellowstone hot spring. J Bacteriol 180:366–376.
- Fu Q, Kuramochi Y, Fukushima N, Maeda H, Sato K, Kobayashi H. 2015. Bioelectrochemical analyses of the development of a thermophilic biocathode catalyzing electromethanogenesis. Environ Sci Technol 49: 1225–1232. http://dx.doi.org/10.1021/es5052233.
- Nurk S, Bankevich A, Antipov D, Gurevich AA, Korobeynikov A, Lapidus A, Prjibelski AD, Pyshkin A, Sirotkin A, Sirotkin Y, Stepanauskas R, Clingenpeel SR, Woyke T, McLean JS, Lasken R, Tesler G, Alekseyev MA, Pevzner PA. 2013. Assembling single-cell genomes and minimetagenomes from chimeric MDA products. J Comput Biol 20:714–737. http://dx.doi.org/10.1089/cmb.2013.0084.
- Refojo PN, Ribeiro MA, Calisto F, Teixeira M, Pereira MM. 2013. Structural composition of alternative complex III: variations on the same theme. Biochim Biophys Acta 1827:1378–1382. http://dx.doi.org/10.1016/ j.bbabio.2013.01.001.
- 6. Borisov VB, Gennis RB, Hemp J, Verkhovsky MI. 2011. The cytochrome bd respiratory oxygen reductases. Biochim Biophys Acta 1807:1398–1413. http://dx.doi.org/10.1016/j.bbabio.2011.06.016.
- Sutcliffe IC. 2011. Cell envelope architecture in the *Chloroflexi*: a shifting frontline in a phylogenetic turf war. Environ Microbiol 13:279–282. http:// dx.doi.org/10.1111/j.1462-2920.2010.02339.x.