




Review

Forecasting Zoonotic Infectious Disease Response to Climate Change: Mosquito Vectors and a Changing Environment

Andrew W. Bartlow ¹, Carrie Manore ², Chonggang Xu ³, Kimberly A. Kaufeld ⁴, Sara Del Valle ², Amanda Ziemann ⁵, Geoffrey Fairchild ² and Jeanne M. Fair ^{1,*}

¹ Los Alamos National Laboratory, Biosecurity and Public Health, Los Alamos, NM 87545, USA; abartlow@lanl.gov

² Los Alamos National Laboratory, Information Systems and Modeling, Los Alamos, NM 87545, USA; cmanore@lanl.gov (C.M.); gfairchild@lanl.gov (G.F.); sdelvall@lanl.gov (S.D.V.)

³ Los Alamos National Laboratory, Earth Systems Observations, Los Alamos, NM 87545, USA; cxu@lanl.gov

⁴ Los Alamos National Laboratory, Statistical Sciences, Los Alamos, NM 87545, USA; kkaufeld@lanl.gov

⁵ Los Alamos National Laboratory, Space Data Science and Systems, Los Alamos, NM 87545, USA; ziemann@lanl.gov

* Correspondence: jmfair@lanl.gov; Tel.: +1-505-606-1650

Received: 9 March 2019; Accepted: 29 April 2019; Published: 6 May 2019



Abstract: Infectious diseases are changing due to the environment and altered interactions among hosts, reservoirs, vectors, and pathogens. This is particularly true for zoonotic diseases that infect humans, agricultural animals, and wildlife. Within the subset of zoonoses, vector-borne pathogens are changing more rapidly with climate change, and have a complex epidemiology, which may allow them to take advantage of a changing environment. Most mosquito-borne infectious diseases are transmitted by mosquitoes in three genera: *Aedes*, *Anopheles*, and *Culex*, and the expansion of these genera is well documented. There is an urgent need to study vector-borne diseases in response to climate change and to produce a generalizable approach capable of generating risk maps and forecasting outbreaks. Here, we provide a strategy for coupling climate and epidemiological models for zoonotic infectious diseases. We discuss the complexity and challenges of data and model fusion, baseline requirements for data, and animal and human population movement. Disease forecasting needs significant investment to build the infrastructure necessary to collect data about the environment, vectors, and hosts at all spatial and temporal resolutions. These investments can contribute to building a modeling community around the globe to support public health officials so as to reduce disease burden through forecasts with quantified uncertainty.

Keywords: infectious disease; zoonotic; mosquito; vector-borne; climate change; range expansion; epidemiology

1. Introduction

The epidemiology of infectious diseases is constantly fluctuating in response to environmental changes and changing interactions among hosts, reservoirs, vectors, and pathogens. This is particularly true for zoonotic diseases that infect humans, animals of veterinary importance, and wildlife. Within the subset of zoonotic diseases, vector-borne pathogens are changing more rapidly with climate change and potentially have a more complex epidemiology. In the past 80 years, the majority of global emerging infectious diseases have been zoonotic [1]. While most of the zoonoses arise from wildlife (72%), vector-borne emerging diseases are increasing at a more rapid rate [1]. Along with an increase in emerging zoonotic diseases, there have been range expansions of reservoir hosts, vectors, and the pathogens they harbor.

Vector-borne diseases account for more than 17% of all infectious diseases, causing more than 700,000 deaths annually [2,3]. For example, almost 4 billion people in over 128 countries are at risk of contracting dengue, with 96 million cases estimated per year [2,4]. Dengue virus has both animal hosts and reservoirs that are both impacted and help maintain virus circulating in populations [5]. Many of the vectors that transmit important zoonotic infectious diseases are bloodsucking insects that ingest disease-producing microorganisms during a blood meal from an infected host and then later inject it into a new host during their subsequent blood meal. Mosquitoes are the best-known disease vector. However, other vectors, such as ticks, black flies, sandflies, midges, fleas, and triatomine bugs are also important vectors of human pathogens. Table 1 lists the primary zoonotic arboviruses and other pathogens vectored by Dipterans, and Table 2 lists the primary animal (non-zoonotic) pathogens vectored by Dipterans. For a recent review on tick-borne pathogens, see [6]. The rest of this review focuses on modeling mosquito vectors and mosquito-borne infectious diseases in response to climate change.

Mosquitoes are heavily dependent and closely tied to the environment [7,8]. A mosquito's life is a microcosm of water, available habitats, temperature, predators, and competitors. Each aspect of a mosquito's life history is greatly influenced by even the slightest changes in the environment in unpredictable ways. For example, droughts can increase mosquito habitat by increasing stagnant water in streams, thereby increasing mosquito populations [9]. In other cases, range expansion is simply due to warmer winters in the northern latitudes [10,11].

Table 1. Primary zoonotic arboviruses and other pathogens vectored by Dipterans. The taxon groups listed for viruses are at the family level. Taxon groups for vectors are at the genus level.

Pathogen	Taxon Group	Vectors	Animal Hosts	Range
Banna virus	Reoviridae	Mosquitoes (<i>Culex</i> , <i>Anopheles</i> , <i>Aedes</i>) [12]	Pigs, cattle, humans [12]	Southeast Asia, Indonesia [12]
Chikungunya virus	Togaviridae [13]	Mosquitoes (<i>Aedes</i>) [13]	Humans [13]	Central America, North America, South America, Oceania, central/southern Africa, southern/eastern Asia, western/central Europe [14]
Dengue virus	Flaviviridae	Mosquitoes (primarily <i>Aedes aegypti</i>) [15]	Humans [15] In vitro successful infection of amphibians, mammals, & reptiles [16]	North America, Central America, South America, Oceania, Europe, southern Asia, and Africa [17]
Dirofilaria spp. (Dirofilariasis)	Nematode	Mosquitoes (<i>Culex</i> , <i>Aedes</i> , <i>Anopheles</i> , <i>Mansonia</i>) [18]	Dogs, cats, humans [19]	Southern/central/eastern Europe, Middle East, eastern/southeastern/central Asia, southeastern North America [19]
Eastern Equine Encephalitis virus	Flaviviridae	Mosquitoes (<i>Culiseta</i> , <i>Culex</i>) [20]	Birds, humans, horses [20]	United States [21]
Indiana vesiculovirus	Rhabdoviridae	Sand flies (<i>Phlebotomus</i> , <i>Lutzomyia</i>), mosquitoes (<i>Aedes</i>), black flies (family <i>Simuliidae</i>) [22]	Equids, bovids [22]	North America, South America [22]
Japanese Encephalitis virus	Flaviviridae	Mosquitoes (<i>Culex</i>) [17]	Pigs, birds, horses, humans [17]	Western Europe, Russia, southern Asia, Oceania [17]
Plasmodium spp. (Malaria)	Protozoan	Mosquitoes (<i>Anopheles</i>) [23]	Humans, birds (carriers) [23]	United States, central/southern Africa, southeast Asia, southern Europe [23,24]
Mayaro virus	Togaviridae	Mosquitoes (<i>Haemagogus</i>) [25]	Humans [25]	Northern South America *tourists infected [25]
Murray Valley Encephalitis virus	Flaviviridae	Mosquitoes (<i>Culex</i>) [26]	Humans [26]	Australia, Northern Territories [26]

Table 1. Cont.

Pathogen	Taxon Group	Vectors	Animal Hosts	Range
O'nyong'nyong virus	Togaviridae	Mosquitoes (<i>Anopheles</i>) [27]	Humans [27]	Western/central Africa [27]
Oropouche virus	Peribunyaviridae	Mosquitoes (<i>Coquillettia</i>) [28]	Humans [29]	Central/northern South America, southeastern Central America [28]
Rift Valley Fever virus	Phenuviridae	Mosquitoes (<i>Culex</i> , <i>Aedes</i> , <i>Anopheles</i> , <i>Eretmapodites</i> , <i>Mansonia</i> , <i>Culicoides</i> , <i>Coquillettia</i>) [30]	Ruminants (reservoir), humans [30], Bats [31]	Africa, southern Middle East [30]
Saint Louis Encephalitis virus	Flaviviridae	Mosquitoes (<i>Culex</i>) [32]	Birds [32], humans [33]	South America [33], North America [33]
Spondweni virus	Flaviviridae	Mosquitoes (<i>Culex</i> , <i>Aedes</i>) [34]	Humans [34]	Caribbean, southern Africa [34]
<i>Trypanosoma brucei</i> (Sleeping Sickness)	Protozoan	Tsetse fly (<i>Glossina</i>) [35]	Wild ungulates, ruminants, equids, dogs, humans [35]	Africa, Central America, South America [35]
Usutu virus	Flaviviridae	Mosquitoes (<i>Culex</i>) [36]	Birds, horses, humans [36], bats [37]	Central/southern Africa, central Europe [36]
Venezuelan Equine Encephalitis virus	Togaviridae	Mosquitoes (<i>Culex</i>) [38]	Horses, humans [38]	Southern North America, central America, northern/central South America [38]
Western Equine Encephalitis virus	Togaviridae	Mosquitoes (<i>Culex</i>) [39]	Birds, humans [39]	Western North America, South America, Cuba [39]
West Nile virus	Flaviviridae	Mosquitoes (<i>Culex</i> , <i>Aedes</i>) [40]	Birds (reservoir), equids, humans [40]	Africa, the Middle East, Oceania, North America, Central America, northwestern and southern/central South America, and Europe [40]
Yellow Fever virus	Flaviviridae	Mosquitoes (<i>Aedes aegypti</i> , <i>Haemagogus</i>) [41]	Humans [41]	Central America, South America, southern North America, Oceania, Europe, southern Asia, and Africa [17]
Zika virus	Flaviviridae	Mosquitoes (<i>Aedes</i>) [42]	Primates, humans [42]	central-western and southern Africa, Oceania, South America, North America, Central America, southern Asia, Europe [17]

Table 2. Primary animal (non-zoonotic) pathogens vectored by Dipterans. The taxon groups listed for viruses are families. Taxon groups for vectors are at the genus level.

Pathogen	Taxon Group	Vectors	Animal Hosts	Range
African Horse Sickness virus	Reoviridae	Midges (<i>Culicoides</i>) [43]	Equids [43]	Central/southern Africa [43]
Bluetongue Disease virus	Reoviridae	Midges (<i>Culicoides</i>) [44]	Ruminants [44]	North America, Central America, South America, Africa, southern Asia, northern Australia, Estonia and Russia, southern and central Europe [45]
Epizootic Haemorrhagic Disease virus	Reoviridae	Midges (<i>Culicoides</i>) [46]	Ruminants [46]	North America, Australia, Africa, Asia, and the Mediterranean [46]
Lumpy Skin Disease virus	Poxviridae	Mosquitoes (<i>Aedes Anopheles</i>), flies (<i>Musca</i> , <i>Stomoxys</i> , <i>Glossina</i>), and midges (<i>Culicoides</i>) [47]	Cattle [47]	Africa, the Middle East, South-Eastern Europe, Russia [47]

Table 2. Cont.

Pathogen	Taxon Group	Vectors	Animal Hosts	Range
Schmallenberg virus	Peribunyaviridae	Midges (<i>Culicoides</i>) [48]	Ruminants [49]	Europe [49]
<i>Trypanosoma evansi</i> (Surra)	Protozoan	Hematophagous flies (<i>Tabanus, Musca</i>) [50]	Vampire bats (reservoir), ungulates, ruminants, dogs, cats [50]	Asia, northern Africa, Central America, South America, the Middle East [50]

2. Changing Mosquito Vector Biology and Range Expansion

Emerging infectious diseases are directly influenced by changing environmental conditions [1,51]. The results of these geographic expansions and range shifts represent major health crises in many parts of the world [52,53]. Vector-borne infectious diseases make up a significant portion of zoonotic diseases, which have increased in the last few decades [1]. Mosquitoes shift and expand their ranges into new areas as environments change and become more suitable, bringing the pathogens they harbor with them. Mosquitoes—important vectors for many infectious diseases of human importance—are sensitive to environmental conditions, especially temperature and precipitation [7]. As aquatic insects, their life cycle and developmental time depends on water availability [54]. Temperature is also important for their development time and their ability to overwinter [55]. Being able to overwinter due to warmer winter temperatures may aid in the northward expansion of many species [10,11]. In addition, increased temperatures induce faster development times [7,56], which may be especially important in arctic environments [57].

Most mosquito-borne infectious diseases are transmitted by mosquitoes in three genera: *Aedes*, *Anopheles*, and *Culex*. Species in the genus *Aedes*, particularly *Ae. Albopictus* (Asian tiger mosquito) and *Ae. aegypti* are vectors for a number of zoonotic diseases, including dengue virus, chikungunya virus, Zika virus, and yellow fever. *Anopheles* mosquitoes are responsible for transmitting malaria and a few other pathogens, such as canine heartworm (*Dirofilaria immitis*) and species that cause filariasis (e.g., *Wuchereria bancrofti*). *Anopheles gambiae* and *An. arabiensis* are the most responsible vectors for malaria transmission in Africa. *Culex* mosquitoes, predominantly *C. pipiens*, *C. tarsalis*, and *C. quinquefasciatus*, carry and transmit West Nile virus and Saint Louis encephalitis virus. Mosquitoes in all three genera have experienced range expansions [58–65]. *Aedes aegypti* is the most widespread, with an almost global distribution through repeated invasions [66]. As mosquito species expand their geographic distributions, the diseases they harbor are expanding and are predicted to continue to expand [67], representing new challenges for places previously not affected. For example, chikungunya virus has spread from Africa and Southeast Asia to the subtropics and the western hemisphere [68], Zika virus has spread rapidly from Africa to the Americas [69], and West Nile virus has spread to British Columbia, Canada [70].

Two species of mosquitoes in the genus *Aedes* have experienced the most range expansion globally—*Aedes albopictus* and *Ae. aegypti* [66]. *Aedes albopictus* is the most invasive mosquito species in the world [71], spreading from southeast Asia to every continent except Antarctica [72]. Global air and sea travel [73] and the used tire trade [72,74,75] have been proposed to be major causes of the worldwide dispersal of *Ae. albopictus*. Eggs of this species are long-lived, desiccation-resistant [76], and respond to shorter, colder days by going into diapause [72,74]. *Aedes aegypti* spread from Africa to the tropics and subtropics around the world by way of human movement [77,78]. In addition to climate impacts on vector distributions, urbanization may exacerbate the invasive potential of vectors following the colonization of new areas [66]. Both *Ae. albopictus* and *Ae. aegypti* have urban and suburban habitat preferences, and the habitats of larval development include man-made containers [76].

Species of *Anopheles* and *Culex* mosquitoes have also increased their geographic range [58,59,70], but have not become as invasive as the two species of *Aedes*. The increase in *Anopheles* mosquitoes, mainly *An. arabiensis*, may be due to their ability to resist desiccation and survive severe dry seasons, resulting in perennial transmission of malaria to humans [79–81]. However, *Anopheles gambiae* has also been shown to transmit malaria continuously during the dry season [80] and expand their niches into marginal habitats [61], both of which may act to increase their geographic range in the future [82].

One of the most important malaria vectors in the neotropics (Central and South America) is *An. darlingi*, and is predicted to undergo range expansions [62,83].

The *Culex pipiens* complex, made up of *C. pipiens sensu stricto* and *C. quinquefasciatus*, is distributed worldwide. *Culex quinquefasciatus* is most prevalent in the tropics and subtropics, while *C. pipiens* is found in temperate climates [84]. Recently, a new species of concern, *C. coronator*, was discovered in the southwestern United States [85,86], and as far north as Virginia [87]. This species has expanded its range from the America subtropics and tropics [85]. Importantly, this species has been documented harboring several arboviruses of human importance [86]. Other *Culex* species are experiencing range shifts northward into Canada [64,70]. Both *Culex pipiens* and *C. quinquefasciatus* are predicted to spread further northward in Canada [64,88].

Mosquito responses to environmental conditions are variable and often stray from predictions from models that take into account climate to determine future distributions. For example, when *Ae. albopictus* invaded new continents (e.g., South America), it resulted in niche shifts, which may be due to adaptive genetic changes or result from founder effects [63]. The ability to shift niches upon invading new areas may be one reason why *Ae. albopictus* is so widespread. Additionally, several species have been shown to become more tolerant to saline and brackish waters. These species include *Anopheles sundaicus*, *An. culicifacies*, *An. stephensi*, *Aedes aegypti*, *Ae. Albopictus*, and *Culex sitiens* [89]. Salinity tolerance may be important to consider in studies predicting future mosquito distributions because brackish and saline water along coastlines is predicted to increase [90]. Hybridization is another ecological factor that may determine the extent of species distribution and influence range expansion of certain species (reviewed in [91]). For example, *An. gambiae* gained a critical gene from *An. arabiensis* allowing it to move from rainforests to drier habitats, such as savannahs [92,93]. Further research is needed to understand how hybridization may contribute to range expansion in other mosquito genera. Predicting shifts in range distributions of mosquito species is challenging and requires a comprehensive modeling approach that incorporates vegetation modeling, hydrology, epidemiological modeling, human movement behavior, mosquito behavior, and host, vector, and pathogen biology and evolution.

Mosquitoes are closely tied to and track environmental conditions because their life cycle depends on environmental conditions. Mosquito expansion will likely continue well into the future because of increased temperatures, greater probabilities of overwintering, and changes to precipitation regimes. Although climate change directly influences mosquito distributions through changing environmental conditions, climate change can also impact distributions indirectly. Humans respond to climate change by altering their surroundings. For example, drought has caused southeast Australian communities to install water storage tanks, which is predicted to increase the distribution of *Ae. aegypti* and increase the risk of emerging and re-emerging diseases in these areas [94]. Changes in human movement will also affect mosquito dispersal and impact future distributions. Predicting range expansions of mosquito vectors and mosquito-borne diseases therefore needs effective climate, human, and epidemiological modeling, which when combined, will provide more accurate forecasts for vector and disease risk.

3. Epidemiological Modeling of Vector-Borne Infectious Diseases

While yearly dengue epidemics continue [4,95,96], the chikungunya virus and Zika virus have recently emerged and caused large outbreaks in the Americas, spreading rapidly across the continent after introduction and causing long-term effects, including Zika-related birth defects. Chikungunya virus infected upwards of 30% of vulnerable populations in South and Central America. While the full effects of Zika virus are still being understood, recent sero-surveys in Brazil indicate a more than 60% attack rate [97–100]. It is not clear if chikungunya virus and Zika virus will continue to expand geographically and become endemic. In addition, climate change and globalization have increased the potential for wider spread of vector-borne diseases [101]. Thus, there is an urgent need to study these diseases in different regions and to produce a generalizable approach capable of mapping risk and forecasting outbreaks to alert vulnerable populations and inform decision support while increasing scientific understanding [102].

Prior research has primarily focused on understanding outbreaks at limited spatial and temporal scales [103–110]. This is because outbreak size, timing, and relationship to exogenous factors varies widely across time and space [105,106,111–116] and because reliable data directly related to outbreak risk (e.g., case counts, biting rates, mosquito infection rates) is hard to come by. Data coverage as well as team expertise is also incomplete. Previous studies have focused on analyzing a single or small number of satellite images over one region [104,106,110,117–120] or analyzing a subset of data streams such as weather or Internet data to study vector-borne disease dynamics [121–128]. These provide a limited picture and sometimes-conflicting results [105,110,129–132] (e.g., role of El Niño). Modeling of Infectious Disease Agents (MIDAS) groups have focused on smaller spatial-scale transmission (<200 m transmission chains [133]) for Thailand and on mechanistic stochastic models with dependence on temperature and other factors at continental or global scales [134]. Other teams have used ecological niche modeling to produce static maps of suitability (not risk per se) for Zika, dengue, and chikungunya and their associated vector species, *Ae. aegypti* and *Ae. albopictus* [95,135–138].

Ecological studies have demonstrated that certain variables such as Normalized Difference Vegetation Index (NDVI), precipitation, and temperature can predict the severity of mosquito-borne disease transmission months in advance [110,129,139]. These variables have been used to predict disease risk such as West Nile virus in the United States [140]. Mosquito-borne disease modeling has been limited to either local studies that consider selected fine-scale key processes or to larger-scale models that have a limited representation of key processes and their interactions. For example, some process-based models consider the impact of temperature [141–148] or rainfall [147,149–151] on mosquito lifespan and development rates without explicitly considering the nonlinear response of mosquito habitat to weather (i.e., formation and persistence of standing water in the landscape). Recent modeling studies (e.g., malaria [24,152,153] and Rift Valley fever [154]) have begun to consider the linkage to hydrology and climate. But they are either limited to small scales (e.g., watersheds or small regions) [153,154] or are based on a simple calculation of water balance without considering real land surface characteristics [152]. Meanwhile, statistical models built on the relationship between local meteorological/environmental factors, mosquito physiology, and/or reported disease cases have been developed for understanding the ecological niches of mosquito and mosquito-borne diseases [78,95,116,155–158]. Because they are trained solely on past observations, they generally do not capture the coupled nonlinear processes (e.g., vector range expansion and human-mosquito contact) [112,148] in conditions not yet encountered (i.e., “no-analog” and beyond the space of available data for model calibration) resulting from weather extremes, climate, and socioeconomic changes.

Laboratory and field studies have shown relationships between weather/climate and risk for mosquito-borne diseases (e.g., Rift Valley Fever) [159], tick-borne diseases [160], and other vector-borne diseases. Several research teams, including ours, have incorporated these factors into working models [161–163]. A recent systematic review by the National Exposure Research Laboratory [160] highlights the need for a “rigorous multi-system modeling approach to improve our knowledge about the important mosquito vector, *Aedes* spp. presence/abundance response to the interaction between environmental, socioeconomic, and meteorological systems”. The current state of the art has focused on particular pathogens in particular regions using a subset of the needed factors [160,164–166]. The few global or continental models that exist focus on a single pathogen [137,141,163] or on ecological niche models rather than explicit disease dynamics [137].

4. Earth System Modeling: The State of Climate Forecasting

The modeling of global climate systems dates back to 1960, and were based on weather prediction models (e.g., [167]). Most of these early models only include atmosphere without detailed representation of land (e.g., no explicit representation of vegetation) and ocean process (e.g., motionless ocean). However, there are also pioneering models that coupled atmosphere and ocean currents [168,169]. Because these models capture the circulation of both atmospheric and ocean currents, they are referred to as general circulation models (GCMs). With the advancement in computing powers and our improved

understanding of coupled physical systems, GCMs have made great progress in simulating the coupled physical climate processes (e.g., winds, clouds, land surface, oceans, and ice) and atmospheric chemistry, aerosol, and static vegetation in our Earth system from the 1960s to 2000s [170]. Since the late 2000s, Earth system models (ESMs) began to evolve from GCMs to simulate the interaction between climate and biogeochemical components (e.g., dynamic terrestrial vegetation and ocean biogeochemistry) to better simulate the feedback of biological systems to our climate [171]. Meanwhile, great progress has been made to integrate the human component into ESMs (i.e., the integrated ESM or IESM) [172]. The modern ESMs and IESMs provide the potential to simulate zoonotic relevant components including biological components (e.g., vegetation dynamics), abiotic components (e.g., inundation and water temperature), and human components (e.g., urban environment and population changes). All these progresses provide great potential for seamless coupling with disease modeling for a better prediction of disease risks under population growth, warming, and climate extremes.

5. Bridging the Gap Between Epidemiological and Earth System Modeling

Although great strides have been made in simulating and predicting global climate with large-scale ESMs, coupling these forecasts with human and living-natural (HLN) systems is critical for planning and mitigating long-term impacts of changes in the environment. HLN systems interact nonlinearly with climates, and data that account for relevant parameters and states of system dynamics in a single source are limited, posing a significant challenge in understanding and predicting the response of these systems. Mosquito-borne diseases are considered a HLN system with rich traditional (e.g., number of cases) and non-traditional (e.g., remote sensing imagery) data streams. Mosquito abundance and virus development rates show both inter- and intra-seasonal variation, and are affected by long-term changes in climate regimes leading to potential range expansion of both mosquitoes and associated diseases. Changes in temperature, climate variability, and extreme weather events are already impacting mosquito-borne diseases around the world [173–175]. A recent example is the emergence of Zika virus throughout Latin America, which was likely fueled by the hot drought during the 2015–2016 El Niño [175]. The mosquito-borne disease HLN system not only affects human health in the US and troops abroad, but is also linked to poverty and regional stability.

We face three key challenges to be able to make reliable large-scale predictions of disease risks at both short-term (seasonal/sub-seasonal) and long-term (decadal) temporal scales, which is critical for planning and improving disease mitigation strategies. First, there is currently no capability to integrate key processes at large scales including (1) hydrology and vegetation that affect mosquito habitats; (2) temperature and humidity that affect mosquito population dynamics and pathogen replication; and (3) human population density and movement that affect vector human contact and facilitates the dispersal of mosquitoes and the development of additional mosquito habitats. There are numerous baseline data requirements for both the epidemiological and climate models for zoonotic infectious diseases (Figure 1). Statistical models built on the relationship between local meteorological and environmental factors and reported disease cases have been developed for understanding the ecological niches of mosquito-borne diseases [116,155,156]. However, they do not directly incorporate the coupled nonlinear processes (e.g., human behavior or mosquito population dynamics) [112], a weakness under novel future climate conditions. Meanwhile, most current process-based models have only incorporated the impact of temperature [141–145] and rainfall [149–151] on mosquito population dynamics without explicitly considering the formation and persistence of water on the landscape, which is critical for accurately predicting the nonlinear response of mosquito habitats to precipitation. One exception is the HYDREMATS model [153], but it is limited to local watersheds. Thus, it cannot account for key processes of disease transmission such as human movement or nearby pathogen reservoirs.

Second, although recent models have started to incorporate human population and movement data [141,176,177], they have not yet considered future human populations and movement that are relevant to predicting disease risk in the future. While animal movement data is also required, it can

be either easy or impossible to obtain, based on the region. Third, although field and lab studies and analysis of diverse and often non-traditional data streams critical to developing and validating coupled HLN-climate systems are increasingly available, the research community lacks an advanced data fusion and mining system to extract critical information from different data sources for parameterization, development, and evaluation of such large-scale, complex models. The climate system, resulting environmental change, vectors, animal and human hosts, and infectious pathogens are each in its own a complex system. Data requirements, existing models, a foundational understanding of infectious diseases, statistical and data fusion, and computer science are all required for predicting how climate change may impact vector-borne diseases (Figure 2). The main hurdle to the above three challenges can be attributed to the lack of large investments that enable seamless collaboration between diverse and highly skilled subject-matter experts, availability of computational resources to scale up and integrate key processes within the complex systems, and data fusion/integration approaches needed to validate models while capturing uncertainty in model parameters, inputs, and predictions.

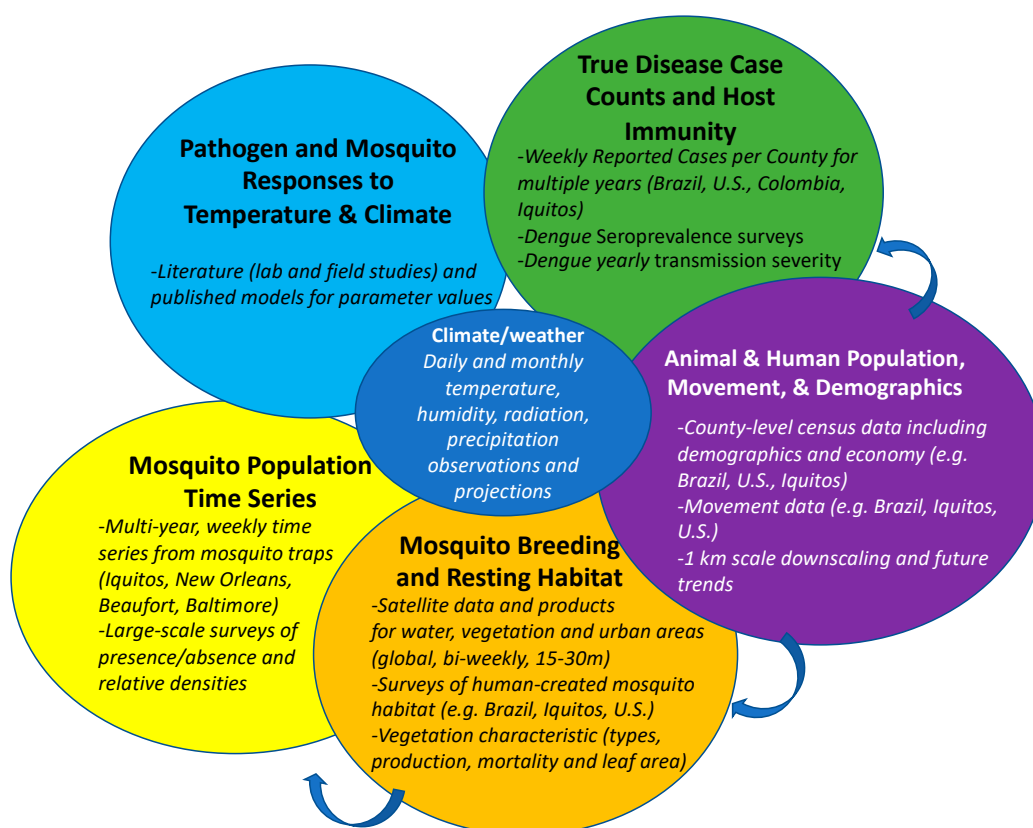


Figure 1. Data requirements to bring together climate and epidemiological models and ground truth real-time data. Arrows indicate data that can be used to inform additional data categories. Bold represents Data Categories and italics are representative subsets of Best Available Data.

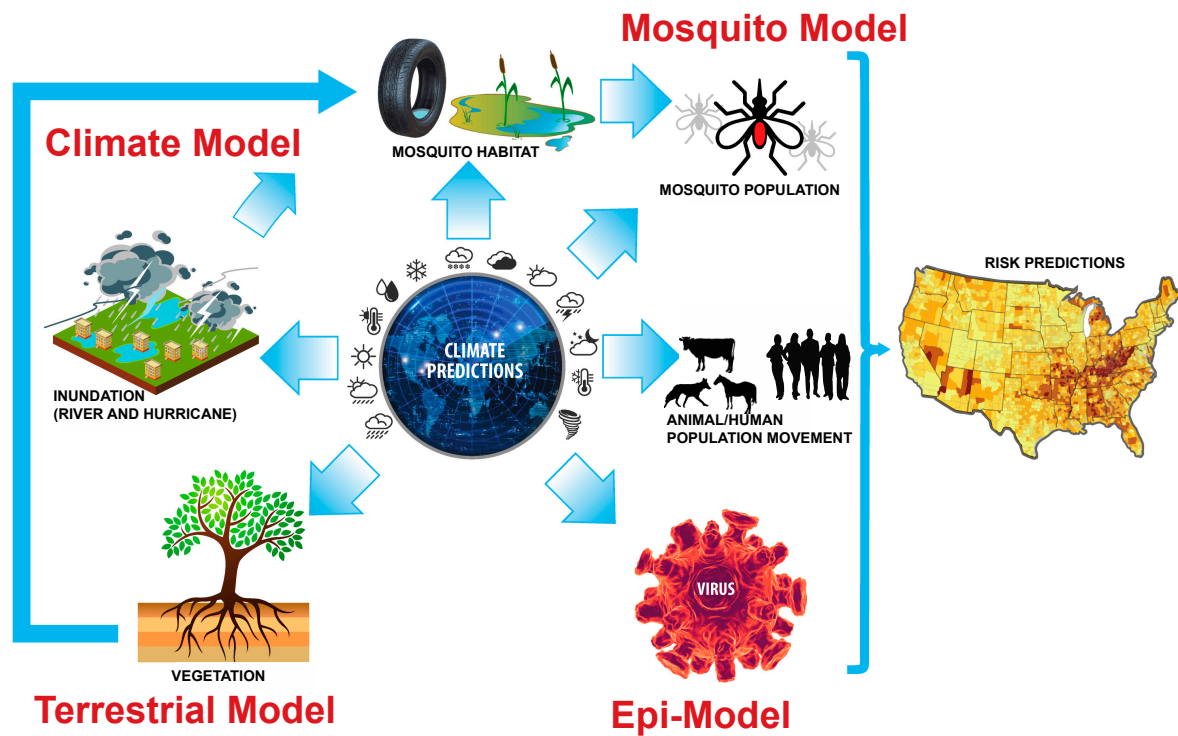


Figure 2. Disease and climate systems for mosquito borne diseases. Each system must be coupled together with validation from ground truth real-time data. Data from Figure 1 feeds into each of these systems and data fusion issues are addressed throughout the process.

6. Data Fusion and Data Requirements

Data fusion is necessary to understand the complex processes of vector-borne diseases. Fusing heterogeneous data provides a way to link the data model, the process model for the disease dynamics (including transmission rates), and R_0 , as well as account for varying levels of spatial resolution [178]. Each of these components has been well-established but have not been integrated together in a disease model. Additionally, it can be a challenge to fuse data with a mechanistic understanding [179–181]. Due to the complexity of modeling in both the process and the data, researchers generally focus on either a model-based (mathematical) or data-based (statistical) approach to understand predictions of ecological dynamics [182]. However, both the data and the mechanistic process underlying the physical process are often of interest in disease modeling. Therefore, it is not enough to fit a statistical model to observations or only use a model-based approach that ignores the data [123]. There are several statistical challenges associated with disease modeling, including modeling and estimating coupled systems, dealing with unobserved variables, and modeling the spatial-temporal dynamics [183]. Statistical inference for time-series models with incomplete data and for process models can be particularly challenging as they may be computationally expensive and oftentimes intractable. Stochastic models, on the other hand, add flexibility to the model to fit real data. This provides a framework where parameters can be identified even when they can be unidentifiable in a deterministic setting. It focuses on the distribution associated with the characteristic of a process. However, it does not incorporate all sources of data from climate, water sources, and transmission.

A particular challenge present in vector-borne disease modeling is that the underlying phenomena are generally difficult or impossible to directly measure. In an ideal situation, knowledge of mosquito population distributions through time, broken up by species and then coupled with human population distributions through time, would be used to derive risk maps. This knowledge could directly inform a mechanistic or statistical model to provide high-fidelity forecasts. However, mosquito population distributions are difficult to capture. Ball et al. [184] are developing an autonomous sensor that uses passive sugar baiting to lure mosquitoes in order to test their saliva for the presence of mosquito-borne

viral pathogens [184], but these sensors are still in their infancy, and deployment at the scale of a large city, state, or country is not yet possible. As a result, no large-scale mosquito population distribution data through time exist. This gap, along with complications coming from the structure of modern disease surveillance systems, has resulted in a revolution in the disease modeling literature in which *proxy datasets* are used to fill in the disparity in our knowledge. Such datasets indirectly relate to the phenomenon at hand. For example, internet data (e.g., [185–188]) have been used to complete the gaps in our public health data; searches for specific disease-related keywords such as cough or fever, for instance, tend to highly correlate with actual disease incidence. Using knowledge of the mosquito lifecycle (i.e., that mosquitoes require certain temperature and precipitation conditions for breeding), proxy datasets such as weather, demographics, satellite imagery, and other related datasets can be used to provide additional insight into mosquito locations. The underlying hypothesis to these data fusion approaches is that, while no individual proxy dataset will provide enough information to accurately forecast disease incidence, the fusion of *multiple weak indicators* derived from disparate data streams will provide a more complete picture.

Once this approach is considered, however, the complexity of the data fusion problem greatly increases; it may not be intuitive, for example, how internet data coming from Wikipedia and spectral signals coming from satellite imagery could be fused. Furthermore, they will relate to disease incidence on different timescales, e.g., healthy vegetation in satellite imagery (as a proxy for standing water) will be a multi-week leading indicator [189], whereas Wikipedia access logs will be closer to a real-time indicator. Another factor is that each data source has the potential to have large measurement errors, but because the data types include hard (e.g., physics-based phenomenologies) and soft (e.g., text) data sources, the mechanics of these errors are highly variable. Because each of the disparate data streams is important for forecasting and understanding disease dynamics, it is equally as important to appropriately account for leading/lagging indicators, error propagation, etc., when building the fusion models. The conditional model, for instance, is flexible as it accounts for the uncertainty in the data based upon prior knowledge of the data parameters and characterizes the relationships in the data and the inherent latent structure. The process model spatial and temporal rate variations are incorporated directly as an additional layer in the hierarchical model.

An added complexity of data fusion is that the sources for climate and mosquito data are generally inherently on different spatial and/or temporal scales, e.g., fine-scale data as in mosquito habitats, and coarse gridded data as in climate data. Oftentimes, the sources of data are spatially misaligned, meaning the data are on different grids. To account for the misalignments, a spatial interpolation method, such as kriging, can be used [190]. Additionally, if the data layers are nested or data layers overlap (non-nested), spatial modeling approaches exist to account for the misalignment in the spatial grids [191]. A hierarchical or multilevel model structure accommodates different spatial aggregations, as well as allowing for shared characteristics among the groups [192,193]. A conditional modeling framework, i.e., a Bayesian or maximum likelihood approach, can combine multiple processes and datasets in an analysis [194]. The conditional model provides a way to pool data from multiple sources and weight them based upon the content of the information.

7. Assumptions or Challenges of Disease Forecasting and Modeling in Animals and Humans

There are three major challenges in achieving the scientific and technological advances made in weather forecasting and transferring them into disease forecasting; these are: (1) data availability, (2) human behavior, and (3) funding [123]. In terms of data availability, unlike weather forecasting, there is the lack of real-time ground truth data to inform disease forecasting models. In order to accurately predict the future, mathematical models must account for the complex and interacting dynamics of the changing environment, the vectors (e.g., mosquitoes), and the host (e.g., the humans) based on past and current data. Specifically, real-time data on mosquito prevalence and diversity at various spatial resolutions is limited and often non-existent. Without these data streams, modelers use proxy information such as vegetation and hydrology to infer mosquito prevalence, however, these limitations increase the uncertainty

in the predictions. Similarly, clinical surveillance data is often biased, due to limited and costly access, and delayed, due to the bureaucratic reporting process, taking weeks or even months to be published. In order to advance our ability to forecast diseases, we need billions of sensors around the world collecting and uploading real-time information to inform forecasting models.

The second major challenge to accurately forecasting infectious diseases is human behavior. Unlike weather, which is governed by the laws of physics, humans can be unpredictable and, as a host, they play a key role in the spread of infectious diseases. Human behavior, such as vaccination, can reduce or increase the probability of infection. Thus, understanding what people do during an epidemic is crucial for modeling disease dynamics. However, there are no databases monitoring human behavior and although Internet data streams, such as social media, have been used to capture population sentiment, there are biases and uncertainty in using this information. In addition, because of the role that humans are playing in changing the climate, it will be crucial to understand how their behavior will change model predictions. Finally, infectious disease forecasting needs significant investment in building the infrastructure necessary to collect data about the environment, the vectors, and the hosts at all spatial and temporal resolutions. These investments can also contribute to building a modeling community around the globe to support public health officials and ultimately reduce disease burden through forecasts with quantified uncertainty.

8. Concluding Remarks: What is Needed Now and in the Future for Forecasting the Impacts of Climate Change on Infectious Diseases

Rapid and large-scale environmental change is occurring globally, which is already leading to dramatic changes in animal, insect, and plant populations. While some wildlife species are able to adapt and others are not, infectious pathogens and their vectors will respond and take advantage of favorable situations. Additionally, as human populations move and expand, agricultural animals will continue to replace wildlife, and new hosts and vectors will move with these migrations and expansions. Coupling cutting-edge and validated climate models with scalable epidemiological models and ground-truth, real-time data will allow the models to be validated with statistical and sensitivity analyses in order to test hypotheses and predictions of future infectious disease conditions.

A focus on the potential impacts of climate change on natural, animal, and human systems will allow for better predictions and mitigations for how these impacts will influence zoonotic infectious diseases in animals and humans. While modeling has its limitations, especially when coupling large-scale systems models, the alternative includes guesses or waiting until infectious pathogens cross into new regions. Mitigations may include animal vaccination strategies that will take time to develop, including the time it takes to stockpile vaccines in areas where they are needed most. Having more knowledge of future vaccine requirements and the impact of animal trade and movement will ensure the best use of limited resources to reduce the threat of infectious diseases globally.

Author Contributions: All authors contributed equally to this work.

Funding: This research was funded by the Laboratory Directed Research Development Program at Los Alamos National Laboratory. Los Alamos National Laboratory is operated by Triad National Security, LLC under Contract No. 89233218CNA000001 with the U.S. Department of Energy and A. Bartlow was supported by a Los Alamos Director's Postdoctoral Fellowship.

Acknowledgments: The authors are grateful to the Climate Impact Modeling Infectious Disease Team at Los Alamos National Laboratory. We also thank Tyler Wysner for help gathering background information for the manuscript.

Conflicts of Interest: The authors declare no conflict of interest.

References

1. Jones, K.E.; Patel, N.G.; Levy, M.A.; Storeygard, A.; Balk, D.; Gittleman, J.L.; Daszak, P. Global trends in emerging infectious diseases. *Nature* **2008**, *451*, 990–993. [[CrossRef](#)] [[PubMed](#)]
2. WHO. *A Global Brief on Vector-Borne Diseases*; WHO: Geneva, Switzerland, 2014.

3. Lozano, R.; Naghavi, M.; Foreman, K.; Lim, S.; Shibuya, K.; Aboyans, V.; Abraham, J.; Adair, T.; Aggarwal, R.; Ahn, S.Y.; et al. Global and regional mortality from 235 causes of death for 20 age groups in 1990 and 2010: A systematic analysis for the Global Burden of Disease Study 2010. *Lancet* **2012**, *380*, 2095–2128. [[CrossRef](#)]
4. Stanaway, J.D.; Shepard, D.S.; Undurraga, E.A.; Halasa, Y.A.; Coffeng, L.E.; Brady, O.J.; Hay, S.I.; Bedi, N.; Bensenor, I.M.; Castañeda-Orjuela, C.A.; et al. The global burden of dengue: An analysis from the Global Burden of Disease Study 2013. *Lancet. Infect. Dis.* **2016**, *16*, 712–723. [[CrossRef](#)]
5. Vasilakis, N.; Cardoso, J.; Hanley, K.A.; Holmes, E.C.; Weaver, S.C. Fever from the forest: Prospects for the continued emergence of sylvatic dengue virus and its impact on public health. *Nat. Rev. Microbiol.* **2011**, *9*, 532–541. [[CrossRef](#)]
6. Wikel, S. Ticks and Tick-Borne Infections: Complex Ecology, Agents, and Host Interactions. *Vet. Sci.* **2018**, *5*, 60. [[CrossRef](#)] [[PubMed](#)]
7. Alto, B.W.; Juliano, S.A. Precipitation and temperature effects on populations of *Aedes albopictus* (Diptera: Culicidae): Implications for range expansion. *J. Med. Entomol.* **2001**, *38*, 646–656. [[CrossRef](#)] [[PubMed](#)]
8. Afrane, Y.A.; Githeko, A.K.; Yan, G. The ecology of *Anopheles* mosquitoes under climate change: Case studies from the effects of deforestation in East African highlands. *Ann. N. Y. Acad. Sci.* **2012**, *1249*, 204–210. [[CrossRef](#)]
9. Johnson, B.J.; Sukhdeo, M.V.K. Drought-induced amplification of local and regional West Nile virus infection rates in New Jersey. *J. Med. Entomol.* **2013**, *50*, 195–204. [[CrossRef](#)] [[PubMed](#)]
10. Thomas, S.M.; Obermayr, U.; Fischer, D.; Kreyling, J.; Beierkuhnlein, C. Low-temperature threshold for egg survival of a post-diapause and non-diapause European aedine strain, *Aedes albopictus* (Diptera: Culicidae). *Parasit. Vectors* **2012**, *5*, 100. [[CrossRef](#)]
11. Rochlin, I.; Ninivaggi, D.V.; Hutchinson, M.L.; Farajollahi, A. Climate change and range expansion of the Asian tiger mosquito (*Aedes albopictus*) in Northeastern USA: Implications for public health practitioners. *PLoS ONE* **2013**, *8*, e60874. [[CrossRef](#)]
12. Liu, H.; Li, M.-H.; Zhai, Y.-G.; Meng, W.-S.; Sun, X.-H.; Cao, Y.-X.; Fu, S.-H.; Wang, H.-Y.; Xu, L.-H.; Tang, Q.; et al. Banna Virus, China, 1987–2007. *Emerg. Infect. Dis.* **2010**, *16*, 514–517. [[CrossRef](#)] [[PubMed](#)]
13. Okeoma, C.M. *Chikungunya Virus*; Okeoma, C.M., Ed.; Springer International Publishing: Cham, Switzerland, 2016; ISBN 978-3-319-42956-4.
14. Sudeep, A.B.; Parashar, D. Chikungunya: An overview. *J. Biosci.* **2008**, *33*, 443–449. [[CrossRef](#)] [[PubMed](#)]
15. Lai, C.-J.; Putnak, R. Dengue and the Dengue Viruses. *Perspec. Med. Virol.* **2006**, *16*, 269–298.
16. Barr, K.L.; Anderson, B.D.; Heil, G.L.; Friary, J.A.; Gray, G.C.; Focks, D.A. Focks Dengue serotypes 1–4 exhibit unique host specificity in vitro. *Virus Adapt. Treat.* **2012**, *4*, 65. [[CrossRef](#)]
17. Sukhralia, S.; Verma, M.; Gopirajan, S.; Dhanaraj, P.S.; Lal, R.; Mehla, N.; Kant, C.R. From dengue to Zika: The wide spread of mosquito-borne arboviruses. *Eur. J. Clin. Microbiol. Infect. Dis.* **2019**, *38*, 3–14. [[CrossRef](#)] [[PubMed](#)]
18. Kartashev, V.; Afonin, A.; González-Miguel, J.; Sepúlveda, R.; Simón, L.; Morchón, R.; Simón, F. Regional Warming and Emerging Vector-Borne Zoonotic *Dirofilariosis* in the Russian Federation, Ukraine, and Other Post-Soviet States from 1981 to 2011 and Projection by 2030. *Biomed Res. Int.* **2014**, *2014*, 1–11. [[CrossRef](#)] [[PubMed](#)]
19. Borkowski, P.K.; Rymkiewicz, G.; Golebiewska, J.; Nestoros, N.; Romejko-Jarosinska, J.; Zarnowska-Prymek, H.; Masny, A.; Palucki, J.; Cielecka, D. The first case of human autochthonous subconjunctival *dirofilariosis* in Poland and MALT lymphoma as possible consequence of this parasitosis. *Infect. Agent Cancer* **2015**, *10*, 1. [[CrossRef](#)]
20. Estep, L.K.; McClure, C.J.W.; Vander Kelen, P.; Burkett-Cadena, N.D.; Sickerman, S.; Hernandez, J.; Jinright, J.; Hunt, B.; Lusk, J.; Hoover, V.; et al. Risk of exposure to eastern equine encephalomyelitis virus increases with the density of northern cardinals. *PLoS ONE* **2013**, *8*, e57879. [[CrossRef](#)] [[PubMed](#)]
21. Silverman, M.A.; Misasi, J.; Smole, S.; Feldman, H.A.; Cohen, A.B.; Santagata, S.; Mcmanus, M.; Ahmed, A.A. Eastern equine encephalitis in children, Massachusetts and New Hampshire, USA, 1970–2010. *Emerg. Infect. Dis.* **2013**, *19*, 194–202. [[CrossRef](#)] [[PubMed](#)]
22. OIE World Organization for Animal Health. Technical Disease Cards—Vesicular Stomatitis. 2017. Available online: http://www.oie.int/fileadmin/Home/eng/Animal_Health_in_the_World/docs/pdf/Disease_cards/VESICULAR_STOMATITIS.pdf (accessed on 15 December 2018).
23. Parmet, S.; Lynn, C.; RM, G. Malaria. *JAMA* **2010**, *304*, 2084. [[CrossRef](#)]

24. Pergantas, P.; Tsatsaris, A.; Malesios, C.; Kriparakou, G.; Demiris, N.; Tselentis, Y. A spatial predictive model for malaria resurgence in central Greece integrating entomological, environmental and social data. *PLoS ONE* **2017**, *12*, e0178836. [[CrossRef](#)]
25. Hassing, R.-J.; Leparç-Goffart, I.; Blank, S.N.; Thevarayan, S.; Tolou, H.; van Doornum, G.; van Genderen, P.J. Imported Mayaro virus infection in the Netherlands. *J. Infect.* **2010**, *61*, 343–345. [[CrossRef](#)]
26. Niven, D.J.; Afra, K.; Iftinca, M.; Tellier, R.; Fonseca, K.; Kramer, A.; Safronetz, D.; Holloway, K.; Drebot, M.; Johnson, A.S. Fatal infection with murray valley encephalitis virus imported from Australia to Canada, 2011. *Emerg. Infect. Dis.* **2017**, *23*, 280–283. [[CrossRef](#)]
27. Brault, A.C.; Tesh, R.B.; Powers, A.M.; Weaver, S.C. Re-emergence of chikungunya and o'nyong-nyong viruses: Evidence for distinct geographical lineages and distant evolutionary relationships. *J. Gen. Virol.* **2000**, *81*, 471–479.
28. Travassos Da Rosa, J.F.; De Souza, W.M.; De Paula Pinheiro, F.; Figueiredo, M.L.; Cardoso, J.F.; Acrani, G.O.; Teixeira Nunes, M.R. Oropouche virus: Clinical, epidemiological, and molecular aspects of a neglected orthobunyavirus. *Am. J. Trop. Med. Hyg.* **2017**, *96*, 1019–1030. [[PubMed](#)]
29. Henry, R.; Murphy, F.A. Etymologia: Oropouche Virus. *Emerg. Infect. Dis.* **2018**, *24*, 30329. [[CrossRef](#)]
30. Berger, S. *Rift Valley Fever: Global Status*; Gideon Informatics: Los Angeles, CA, USA, 2014.
31. Kading, R.C.; Kityo, R.M.; Mossel, E.C.; Borland, E.M.; Nakayiki, T.; Nalikka, B.; Nyakarahuka, L.; Ledermann, J.P.; Panella, N.A.; Gilbert, A.T.; et al. Neutralizing antibodies against flaviviruses, Babanki virus, and Rift Valley fever virus in Ugandan bats. *Infect. Ecol. Epidemiol.* **2018**, *8*, 1439215. [[CrossRef](#)] [[PubMed](#)]
32. Ottendorfer, C.L.; Ambrose, J.H.; White, G.S.; Unnasch, T.R.; Stark, L.M. Isolation of Genotype V St. Louis Encephalitis Virus in Florida. *Emerg. Infect. Dis.* **2009**, *15*, 604–606. [[CrossRef](#)]
33. Diaz, A.; Coffey, L.L.; Burkett-Cadena, N.; Day, J.F. Reemergence of St. Louis Encephalitis Virus in the Americas. *Emerg. Infect. Dis.* **2018**, *24*, 2150. [[CrossRef](#)]
34. White, S.K.; Lednicky, J.A.; Okech, B.A.; Morris, J.G.; Dunford, J.C. Spondweni virus in field-caught culex quinquefasciatus mosquitoes, Haiti, 2016. *Emerg. Infect. Dis.* **2018**, *24*, 1765–1767. [[CrossRef](#)]
35. OIE World Organization for Animal Health. Technical Disease Cards—Trypanosomosis (Tsetse-Transmitted). 2013. Available online: www.oie.int/fileadmin/Home/eng/Animal_Health_in_the_World/docs/pdf/Disease_cards/TRYPANO_TSETSE.pdf (accessed on 15 December 2018).
36. Vazquez, A.; Jimenez-Clavero, M.; Franco, L.; Donoso-Mantke, O.; Sambri, V.; Niedrig, M.; Zeller, H.; Tenorio, A. Usutu virus: Potential risk of human disease in Europe. *Euro Surveill.* **2011**, *16*, 19935.
37. Cadar, D.; Becker, N.; de Mendonca Campos, R.; Börstler, J.; Jöst, H.; Schmidt-Chanasit, J. Usutu Virus in Bats. *Emerg. Infect. Dis.* **2014**, *20*, 2013–2015.
38. Reiss, C.S. *Neurotropic Viral Infections*; Cambridge University Press: Cambridge, UK, 2008; ISBN 9780511541728.
39. Atasheva, S.; Wang, E.; Adams, A.P.; Plante, K.S.; Ni, S.; Taylor, K.; Miller, M.E.; Frolov, I.; Weaver, S.C. Chimeric alphavirus vaccine candidates protect mice from intranasal challenge with western equine encephalitis virus. *Vaccine* **2009**, *27*, 4309–4319. [[CrossRef](#)]
40. Smith, T.L. The Emerging West Nile Virus: From the Old World to the New. *Perspect. Med. Virol.* **2006**, *16*, 133–148.
41. Monath, T.P.; Vasconcelos, P.F.C. Yellow fever. *J. Clin. Virol.* **2015**, *64*, 160–173. [[CrossRef](#)]
42. Vasudevan, J.; Skandhan, A.; Skandhan, A.K.P.; Balakrishnan, S.; Skandhand, K.P. Zika virus. *Rev. Med. Microbiol.* **2018**, *29*, 43–50. [[CrossRef](#)]
43. OIE World Organization for Animal Health. Technical Disease Cards—African Horse Sickness. 2013. Available online: http://www.oie.int/fileadmin/Home/eng/Animal_Health_in_the_World/docs/pdf/Disease_cards/AFRICAN_HORSE_SICKNESS.pdf (accessed on 15 December 2018).
44. Mellor, P.S.; Wittmann, E.J. Bluetongue Virus in the Mediterranean Basin 1998–2001. *Vet. J.* **2002**, *164*, 20–37. [[CrossRef](#)]
45. Ganter, M. Bluetongue disease—Global overview and future risks. *Small Rumin. Res.* **2014**, *118*, 79–85. [[CrossRef](#)]
46. OIE World Organization for Animal Health. Technical Disease Cards—Epizootic Haemorrhagic Disease. 2009. Available online: http://www.oie.int/fileadmin/Home/eng/Animal_Health_in_the_World/docs/pdf/Disease_cards/EPIZOOTIC_HEAMORRHAGIC_DISEASE.pdf (accessed on 15 December 2018).

47. Kara, P.D.; Mather, A.S.; Pretorius, A.; Chetty, T.; Babiuk, S.; Wallace, D.B. Characterisation of putative immunomodulatory gene knockouts of lumpy skin disease virus in cattle towards an improved vaccine. *Vaccine* **2018**, *36*, 4708–4715. [[CrossRef](#)] [[PubMed](#)]
48. Meyers, A.; Tatu, U. Schmallenberg virus. *Resonance* **2014**, *19*, 814–820. [[CrossRef](#)]
49. Almeida-Filho, R.; Miranda, F.P. Mega capture of the Rio Negro and formation of the Anavilhanas Archipelago, Central Amazônia, Brazil: Evidences in an SRTM digital elevation model. *Remote Sens. Environ.* **2007**, *110*, 387–392. [[CrossRef](#)]
50. OIE World Organization for Animal Health. Technical Disease Cards—Trypanosoma evansi Infections (Including Surra). 2013. Available online: http://www.oie.int/fileadmin/Home/eng/Animal_Health_in_the_World/docs/pdf/Disease_cards/TRYPANO_EVANSI.pdf (accessed on 15 December 2018).
51. Khasnis, A.A.; Nettleman, M.D. Global warming and infectious disease. *Arch. Med. Res.* **2005**, *36*, 689–696. [[CrossRef](#)] [[PubMed](#)]
52. Guerrant, R.L.; Blackwood, B.L. Perspective threats to global health and survival: The growing crises of tropical infectious diseases—our “unfinished agenda”. *Clin. Infect. Dis.* **1999**, *28*, 966–986. [[CrossRef](#)] [[PubMed](#)]
53. Morens, D.M.; Folkers, G.K.; Fauci, A.S. The challenge of emerging and re-emerging infectious diseases. *Nature* **2004**, *430*, 242. [[CrossRef](#)] [[PubMed](#)]
54. Yang, G.-J.; Brook, B.W.; Whelan, P.I.; Cleland, S.; Bradshaw, C.J.A. Endogenous and exogenous factors controlling temporal abundance patterns of tropical mosquitoes. *Ecol. Appl.* **2008**, *18*, 2028–2040. [[CrossRef](#)]
55. Rochlin, I.; Gaugler, R.; Williges, E.; Farajollahi, A. The rise of the invasives and decline of the natives: Insights revealed from adult populations of container-inhabiting Aedes mosquitoes (Diptera: Culicidae) in temperate North America. *Biol. Invasions* **2013**, *15*, 991–1003. [[CrossRef](#)]
56. Tun-Lin, W.; Burkot, T.R.; Kay, B.H. Effects of temperature and larval diet on development rates and survival of the dengue vector Aedes aegypti in north Queensland, Australia. *Med. Vet. Entomol.* **2000**, *14*, 31–37. [[CrossRef](#)]
57. Culler, L.E.; Ayres, M.P.; Virginia, R.A. In a warmer Arctic, mosquitoes avoid increased mortality from predators by growing faster. *Proc. R. Soc. B Biol. Sci.* **2015**, *282*, 20151549. [[CrossRef](#)]
58. Donnelly, M.J.; Licht, M.C.; Lehmann, T. Evidence for a recent population expansion in the malaria vectors Anopheles arabiensis and Anopheles gambiae. *Mol. Biol. Evol.* **2001**, *18*, 1353–1364. [[CrossRef](#)]
59. Onyabe, D.Y.; Conn, J.E. Population genetic structure of the malaria mosquito Anopheles arabiensis across Nigeria suggests range expansion. *Mol. Ecol.* **2001**, *10*, 2577–2591. [[CrossRef](#)]
60. Crowl, T.A.; Crist, T.O.; Parmenter, R.R.; Belovsky, G.; Lugo, A.E. The spread of invasive species and infectious disease as drivers of ecosystem change. *Front. Ecol. Environ.* **2008**, *6*, 238–246. [[CrossRef](#)]
61. Costantini, C.; Ayala, D.; Guelbeogo, W.M.; Pombi, M.; Some, C.Y.; Bassole, I.H.N.; Ose, K.; Fotsing, J.M.; Sagnon, N.; Fontenille, D.; et al. Living at the edge: Biogeographic patterns of habitat segregation conform to speciation by niche expansion in anopheles gambiae. *BMC Ecol.* **2009**, *9*, 1–27. [[CrossRef](#)] [[PubMed](#)]
62. Pedro, P.M.; Sallum, M.A.M. Spatial expansion and population structure of the neotropical malaria vector, Anopheles darlingi (Diptera: Culicidae). *Biol. J. Linn. Soc.* **2009**, *97*, 854–866. [[CrossRef](#)]
63. Medley, K.A. Niche shifts during the global invasion of the Asian tiger mosquito, Aedes albopictus Skuse (Culicidae), revealed by reciprocal distribution models. *Glob. Ecol. Biogeogr.* **2010**, *19*, 122–133. [[CrossRef](#)]
64. Hongoh, V.; Berrang-Ford, L.; Scott, M.E.; Lindsay, L.R. Expanding geographical distribution of the mosquito, Culex pipiens, in Canada under climate change. *Appl. Geogr.* **2012**, *33*, 53–62. [[CrossRef](#)]
65. Guagliardo, S.A.; Barboza, J.L.; Morrison, A.C.; Astete, H.; Vazquez-Prokopec, G.; Kitron, U. Patterns of geographic expansion of Aedes aegypti in the Peruvian Amazon. *PLoS Negl. Trop. Dis.* **2014**, *8*, e3033. [[CrossRef](#)]
66. Weaver, S.C. Urbanization and geographic expansion of zoonotic arboviral diseases: Mechanisms and potential strategies for prevention. *Trends Microbiol.* **2013**, *21*, 360–363. [[CrossRef](#)] [[PubMed](#)]
67. Leta, S.; Beyene, T.J.; De Clercq, E.M.; Amenu, K.; Kraemer, M.U.G.; Revie, C.W. Global risk mapping for major diseases transmitted by Aedes aegypti and Aedes albopictus. *Int. J. Infect. Dis.* **2018**, *67*, 25–35. [[CrossRef](#)] [[PubMed](#)]
68. Powers, A.M. Risks to the Americas associated with the continued expansion of chikungunya virus. *J. Gen. Virol.* **2015**, *96*, 1–5. [[CrossRef](#)]

69. Benelli, G.; Mehlhorn, H. Declining malaria, rising of dengue and Zika virus: Insights for mosquito vector control. *Parasitol. Res.* **2016**, *115*, 1747–1754. [[CrossRef](#)] [[PubMed](#)]
70. Roth, D.; Henry, B.; Mak, S.; Fraser, M.; Taylor, M.; Li, M.; Cooper, K.; Furnell, A.; Wong, Q.; Morshed, M. West Nile Virus range expansion into British Columbia. *Emerg. Infect. Dis.* **2010**, *16*, 1251–1258. [[CrossRef](#)] [[PubMed](#)]
71. Benedict, M.Q.; Levine, R.S.; Hawley, W.A.; Lounibos, L.P. Spread of the tiger: Global risk of invasion by the mosquito *Aedes albopictus*. *Vector Borne Zoonotic Dis.* **2007**, *7*, 76–85. [[CrossRef](#)] [[PubMed](#)]
72. Enserink, M. A mosquito goes global. *Science* **2008**, *320*, 864–866. [[CrossRef](#)]
73. Tatem, A.J.; Hay, S.I.; Rogers, D.J. Global traffic and disease vector dispersal. *Proc. Natl. Acad. Sci. USA* **2006**, *103*, 6242–6247. [[CrossRef](#)]
74. Hawley, W.A.; Reiter, P.; Copeland, R.S.; Pumpuni, C.B.; Craig, G.B. *Aedes albopictus* in North America: Probable introduction in used tires from Northern Asia. *Science* **1987**, *236*, 1114–1116. [[CrossRef](#)]
75. Reiter, P.; Sprenger, D. The used tire trade: A mechanism for the worldwide dispersal of container breeding mosquitoes. *J. Am. Mosq. Control Assoc.* **1987**, *3*, 494–501.
76. Juliano, S.A.; Philip Lounibos, L. Ecology of invasive mosquitoes: Effects on resident species and on human health. *Ecol. Lett.* **2005**, *8*, 558–574. [[CrossRef](#)]
77. Brown, J.E.; Evans, B.R.; Zheng, W.; Obas, V.; Barrera-Martinez, L.; Egizi, A.; Zhao, H.; Caccone, A.; Powell, J.R. Human impacts have shaped historical and recent evolution in *Aedes aegypti*, the dengue and yellow fever mosquito. *Evolution* **2014**, *68*, 514–525. [[CrossRef](#)]
78. Kraemer, M.U.G.; Sinka, M.E.; Duda, K.A.; Mylne, A.Q.N.; Shearer, F.M.; Barker, C.M.; Moore, C.G.; Carvalho, R.G.; Coelho, G.E.; Van Bortel, W.; et al. The global distribution of the arbovirus vectors *Aedes aegypti* and *Ae. Albopictus*. *eLife* **2015**, *4*, 1–18. [[CrossRef](#)]
79. Taylor, C.E.; Toure, Y.T.; Coluzzi, M.; Petrarca, V. Effective population size and persistence of *Anopheles arabiensis* during the dry season in West Africa. *Med. Vet. Entomol.* **1993**, *7*, 351–357. [[CrossRef](#)]
80. Toure, Y.T.; Traore, S.F.; Sankare, O.; Sow, M.Y.; Coulibaly, A.; Esposito, F.; Petrarca, V. Perennial transmission of malaria by the *Anopheles gambiae* complex in a North Sudan Savanna area of Mali. *Med. Vet. Entomol.* **1996**, *10*, 197–199. [[CrossRef](#)]
81. Simard, F.; Lehmann, T.; Lemasson, J.J.; Diatta, M.; Fontenille, D. Persistence of *Anopheles arabiensis* during the severe dry season conditions in Senegal: An indirect approach using microsatellite loci. *Insect Mol. Biol.* **2000**, *9*, 467–479. [[CrossRef](#)] [[PubMed](#)]
82. Tonnang, H.E.Z.; Kangalawe, R.Y.M.; Yanda, P.Z. Predicting and mapping malaria under climate change scenarios: The potential redistribution of malaria vectors in Africa. *Malar. J.* **2010**, *9*, 111. [[CrossRef](#)] [[PubMed](#)]
83. Alimi, T.O.; Fuller, D.O.; Qualls, W.A.; Herrera, S.V.; Arevalo-Herrera, M.; Quinones, M.L.; Lacerda, M.V.G.; Beier, J.C. Predicting potential ranges of primary malaria vectors and malaria in northern South America based on projected changes in climate, land cover and human population. *Parasites Vectors* **2015**, *8*, 1–16. [[CrossRef](#)]
84. Ciota, A.T.; Kramer, L.D. Vector-virus interactions and transmission dynamics of West Nile virus. *Viruses* **2013**, *5*, 3021–3047. [[CrossRef](#)] [[PubMed](#)]
85. Gray, K.M.; Burkett-Cadena, N.D.; Eubanks, M.D. Distribution expansion of *Culex coronator* in Alabama. *J. Am. Mosq. Control Assoc.* **2008**, *24*, 585–587. [[CrossRef](#)]
86. Connelly, C.R.; Alto, B.W.; O'Meara, G.F. The spread of *Culex coronator* (Diptera: Culicidae) throughout Florida. *J. Vector Ecol.* **2016**, *41*, 195–199. [[CrossRef](#)]
87. Akaratovic, K.I.; Kiser, J.P. First record of *Culex coronator* in Virginia, with notes on its rapid dispersal, trapping methods, and biology. *J. Am. Mosq. Control Assoc.* **2017**, *33*, 225–228. [[CrossRef](#)]
88. Samy, A.M.; Elaagip, A.H.; Kenawy, M.A.; Ayres, C.F.J.; Peterson, A.T.; Soliman, D.E. Climate change influences on the global potential distribution of the mosquito *Culex quinquefasciatus*, vector of West Nile virus and lymphatic filariasis. *PLoS ONE* **2016**, *11*, e0163863. [[CrossRef](#)]
89. Ramasamy, R.; Surendran, S.N. Global climate change and its potential impact on disease transmission by salinity-tolerant mosquito vectors in coastal zones. *Front. Physiol.* **2012**, *3*, 1–14. [[CrossRef](#)]
90. Ramasamy, R.; Surendran, S.N.; Jude, P.J.; Dharshini, S.; Vinobaba, M. Larval development of *Aedes aegypti* and *Aedes albopictus* in peri-urban brackish water and its implications for transmission of arboviral diseases. *PLoS Negl. Trop. Dis.* **2011**, *5*, e1369. [[CrossRef](#)] [[PubMed](#)]

91. Pfennig, K.S.; Kelly, A.L.; Pierce, A.A. Hybridization as a facilitator of species range expansion. *Proc. R. Soc. B Biol. Sci.* **2016**, *283*, 20161329. [[CrossRef](#)] [[PubMed](#)]
92. Powell, J.R.; Petrarca, V.; della Torre, A.; Caccone, A.; Coluzzi, M. Population structure, speciation, and introgression in the *Anopheles gambiae* complex. *Parassitologia* **1999**, *41*, 101–113.
93. Besansky, N.J.; Krzywinski, J.; Lehmann, T.; Simard, F.; Kern, M.; Mukabayire, O.; Fontenille, D.; Toure, Y.; Sagnon, N. Semipermeable species boundaries between *Anopheles gambiae* and *Anopheles arabiensis*: Evidence from multilocus DNA sequence variation. *Proc. Natl. Acad. Sci. USA* **2003**, *100*, 10818–10823. [[CrossRef](#)] [[PubMed](#)]
94. Beebe, N.W.; Cooper, R.D.; Mottram, P.; Sweeney, A.W. Australia's dengue risk driven by human adaptation to climate change. *PLoS Negl. Trop. Dis.* **2009**, *3*, 1–9. [[CrossRef](#)] [[PubMed](#)]
95. Bhatt, S.; Gething, P.W.; Brady, O.J.; Messina, J.P.; Farlow, A.W.; Moyes, C.L.; Drake, J.M.; Brownstein, J.S.; Hoen, A.G.; Sankoh, O. The global distribution and burden of dengue. *Nature* **2013**, *496*, 504. [[CrossRef](#)] [[PubMed](#)]
96. Imai, N.; Dorigatti, I.; Cauchemez, S.; Ferguson, N.M. Estimating dengue transmission intensity from sero-prevalence surveys in multiple countries. *PLoS Negl. Trop. Dis.* **2015**, *9*, e0003719. [[CrossRef](#)]
97. Aubry, M.; Teissier, A.; Huart, M.; Merceron, S.; Vanhomwegen, J.; Roche, C.; Vial, A.-L.; Teururai, S.; Sicard, S.; Paulous, S. Zika virus seroprevalence, French Polynesia, 2014–2015. *Emerg. Infect. Dis.* **2017**, *23*, 669. [[CrossRef](#)]
98. Netto, E.M.; Moreira-Soto, A.; Pedroso, C.; Höser, C.; Funk, S.; Kucharski, A.J.; Rockstroh, A.; Kümmerer, B.M.; Sampaio, G.S.; Luz, E.; et al. High Zika Virus Seroprevalence in Salvador, Northeastern Brazil Limits the Potential for Further Outbreaks. *mBio* **2017**, *8*, e01390-17. [[CrossRef](#)]
99. Lourenço, J.; Monteiro, M.; Tomás, T.; Monteiro Rodrigues, J.; Pybus, O.; Rodrigues Faria, N. Epidemiology of the zika virus outbreak in the Cabo Verde Islands, West Africa. *PLoS Curr.* **2018**, *1*, 5866102. [[CrossRef](#)]
100. Kucharski, A.J.; Funk, S.; Eggo, R.M.; Mallet, H.-P.; Edmunds, W.J.; Nilles, E.J. Transmission dynamics of zika virus in island populations: A modelling analysis of the 2013–14 French Polynesia Outbreak. *PLoS Negl. Trop. Dis.* **2016**, *10*, e0004726. [[CrossRef](#)] [[PubMed](#)]
101. Hales, S.; de Wet, N.; Maindonald, J.; Woodward, A. Potential effect of population and climate changes on global distribution of dengue fever: An empirical model. *Lancet* **2002**, *360*, 830–834. [[CrossRef](#)]
102. Chretien, J.-P.; Swedlow, D.; Eckstrand, I.; George, D.; Johansson, M.; Huffman, R.; Hebbeler, A. Advancing epidemic prediction and forecasting: A new US Government initiative. *Online J. Public Health Inform.* **2015**, *7*, e13. [[CrossRef](#)]
103. Stewart-Ibarra, A.M.; Lowe, R. Climate and non-climate drivers of dengue epidemics in southern coastal Ecuador. *Am. J. Trop. Med. Hyg.* **2013**, *88*, 971–981. [[CrossRef](#)] [[PubMed](#)]
104. Lowe, R.; Stewart-Ibarra, A.M.; Petrova, D.; García-Díez, M.; Borbor-Cordova, M.J.; Mejía, R.; Regato, M.; Rodó, X. Climate services for health: Predicting the evolution of the 2016 dengue season in Machala, Ecuador. *Lancet Planet. Heal.* **2017**, *1*, e142–e151. [[CrossRef](#)]
105. Liyanage, P.; Tissera, H.; Sewe, M.; Quam, M.; Amarasinghe, A.; Palihawadana, P.; Wilder-Smith, A.; Louis, V.; Tozan, Y.; Rocklöv, J. A spatial hierarchical analysis of the temporal influences of the El Niño-Southern Oscillation and weather on dengue in Kalutara District, Sri Lanka. *Int. J. Environ. Res. Public Health* **2016**, *13*, 1087. [[CrossRef](#)] [[PubMed](#)]
106. Teurlai, M.; Menkès, C.E.; Cavarero, V.; Degallier, N.; Descloux, E.; Grangeon, J.-P.; Guillaumot, L.; Libourel, T.; Lucio, P.S.; Mathieu-Daudé, F.; et al. Socio-economic and climate factors associated with dengue fever spatial heterogeneity: A worked example in New Caledonia. *PLoS Negl. Trop. Dis.* **2015**, *9*, e0004211. [[CrossRef](#)]
107. Zhang, Y.; Wang, T.; Liu, K.; Xia, Y.; Lu, Y.; Jing, Q.; Yang, Z.; Hu, W.; Lu, J. Developing a time series predictive model for dengue in Zhongshan, China based on weather and Guangzhou dengue surveillance data. *PLoS Negl. Trop. Dis.* **2016**, *10*, e0004473. [[CrossRef](#)]
108. Ramachandran, V.G.; Roy, P.; Das, S.; Mogha, N.S.; Bansal, A.K. Empirical model for calculating dengue incidence using temperature, rainfall and relative humidity: A 19-year retrospective analysis in East Delhi, India. *Epidemiol. Health* **2016**, *38*, e2016052. [[CrossRef](#)]
109. Lowe, R.; Coelho, C.A.; Barcellos, C.; Carvalho, M.S.; Catão, R.D.C.; Coelho, G.E.; Ramalho, W.M.; Bailey, T.C.; Stephenson, D.B.; Rodó, X. Evaluating probabilistic dengue risk forecasts from a prototype early warning system for Brazil. *eLife* **2016**, *5*, e11285. [[CrossRef](#)]

110. Adde, A.; Roucou, P.; Mangeas, M.; Ardillon, V.; Desenclos, J.-C.; Rousset, D.; Girod, R.; Briolant, S.; Quenel, P.; Flamand, C. Predicting dengue fever outbreaks in French Guiana using climate indicators. *PLoS Negl. Trop. Dis.* **2016**, *10*, e0004681. [[CrossRef](#)]
111. Morin, C.W.; Comrie, A.C.; Ernst, K. Climate and dengue transmission: Evidence and implications. *Environ. Health Perspect.* **2013**, *121*, 1264–1272. [[CrossRef](#)]
112. Reiner, R.C.; Perkins, T.A.; Barker, C.M.; Niu, T.; Chaves, L.F.; Ellis, A.M.; George, D.B.; Le Menach, A.; Pulliam, J.R.C.; Bisanzio, D.; et al. A systematic review of mathematical models of mosquito-borne pathogen transmission: 1970–2010. *J. R. Soc. Interface* **2013**, *10*, 20120921. [[CrossRef](#)]
113. Braga, C.; Luna, C.F.; Martelli, C.M.; de Souza, W.V.; Cordeiro, M.T.; Alexander, N.; de Albuquerque, M.F.P.M.; Júnior, J.C.S.; Marques, E.T. Seroprevalence and risk factors for dengue infection in socio-economically distinct areas of Recife, Brazil. *Acta Trop.* **2010**, *113*, 234–240. [[CrossRef](#)]
114. Akter, R.; Hu, W.; Naish, S.; Banu, S.; Tong, S. Joint effects of climate variability and socioecological factors on dengue transmission: Epidemiological evidence. *Trop. Med. Int. Heal.* **2017**, *22*, 656–669. [[CrossRef](#)]
115. Mulligan, K.; Dixon, J.; Joanna Sinn, C.-L.; Elliott, S.J. Is dengue a disease of poverty? A systematic review. *Pathog. Glob. Health* **2015**, *109*, 10–18. [[CrossRef](#)]
116. Louis, V.R.; Phalkey, R.; Horstick, O.; Ratanawong, P.; Wilder-Smith, A.; Tozan, Y.; Dambach, P. Modeling tools for dengue risk mapping—A systematic review. *Int. J. Health Geogr.* **2014**, *13*, 50. [[CrossRef](#)]
117. Buczak, A.L.; Koshute, P.T.; Babin, S.M.; Feighner, B.H.; Lewis, S.H. A data-driven epidemiological prediction method for dengue outbreaks using local and remote sensing data. *BMC Med. Inform. Decis. Mak.* **2012**, *12*, 124. [[CrossRef](#)]
118. Machault, V.; Yébakima, A.; Etienne, M.; Vignolles, C.; Palany, P.; Tourre, Y.; Guérécheau, M.; Lacaux, J.-P. Mapping entomological dengue risk levels in Martinique using high-resolution remote-sensing environmental data. *ISPRS Int. J. Geo-Inf.* **2014**, *3*, 1352–1371. [[CrossRef](#)]
119. Kalluri, S.; Gilruth, P.; Rogers, D.; Szczur, M. Surveillance of arthropod vector-borne infectious diseases using remote sensing techniques: A review. *PLoS Pathog.* **2007**, *3*, e116. [[CrossRef](#)]
120. Tian, H.; Huang, S.; Zhou, S.; Bi, P.; Yang, Z.; Li, X.; Chen, L.; Cazelles, B.; Yang, J.; Luo, L.; et al. Surface water areas significantly impacted 2014 dengue outbreaks in Guangzhou, China. *Environ. Res.* **2016**, *150*, 299–305. [[CrossRef](#)] [[PubMed](#)]
121. Teng, Y.; Bi, D.; Xie, G.; Jin, Y.; Huang, Y.; Lin, B.; An, X.; Feng, D.; Tong, Y. Dynamic forecasting of zika epidemics using Google Trends. *PLoS ONE* **2017**, *12*, e0165085. [[CrossRef](#)]
122. Gluskin, R.T.; Johansson, M.A.; Santillana, M.; Brownstein, J.S. Evaluation of internet-based dengue query data: Google Dengue Trends. *PLoS Negl. Trop. Dis.* **2014**, *8*, e2713. [[CrossRef](#)]
123. Moran, K.R.; Fairchild, G.; Generous, N.; Hickmann, K.; Osthus, D.; Priedhorsky, R.; Hyman, J.; Del Valle, S.Y. Epidemic forecasting is messier than weather forecasting: The role of human behavior and internet data streams in epidemic forecast. *J. Infect. Dis.* **2016**, *214*, S404–S408. [[CrossRef](#)]
124. Yang, S.; Kou, S.C.; Lu, F.; Brownstein, J.S.; Brooke, N.; Santillana, M. Advances in using Internet searches to track dengue. *PLOS Comput. Biol.* **2017**, *13*, e1005607. [[CrossRef](#)]
125. de Almeida Marques-Toledo, C.; Degener, C.M.; Vinhal, L.; Coelho, G.; Meira, W.; Codeço, C.T.; Teixeira, M.M. Dengue prediction by the web: Tweets are a useful tool for estimating and forecasting Dengue at country and city level. *PLoS Negl. Trop. Dis.* **2017**, *11*, e0005729. [[CrossRef](#)]
126. Rivera, R. A dynamic linear model to forecast hotel registrations in Puerto Rico using Google Trends data. *Tour. Manag.* **2016**, *57*, 12–20. [[CrossRef](#)]
127. Strauss, R.A.; Castro, J.S.; Reintjes, R.; Torres, J.R. Google dengue trends: An indicator of epidemic behavior. The Venezuelan Case. *Int. J. Med. Inform.* **2017**, *104*, 26–30. [[CrossRef](#)] [[PubMed](#)]
128. Althouse, B.M.; Ng, Y.Y.; Cummings, D.A.T. Prediction of dengue incidence using search query surveillance. *PLoS Negl. Trop. Dis.* **2011**, *5*, e1258. [[CrossRef](#)]
129. Lowe, R.; Barcellos, C.; Coelho, C.A.S.; Bailey, T.C.; Coelho, G.E.; Graham, R.; Jupp, T.; Ramalho, W.M.; Carvalho, M.S.; Stephenson, D.B.; et al. Dengue outlook for the World Cup in Brazil: An early warning model framework driven by real-time seasonal climate forecasts. *Lancet Infect. Dis.* **2014**, *14*, 619–626. [[CrossRef](#)]
130. Racloz, V.; Ramsey, R.; Tong, S.; Hu, W. Surveillance of dengue fever virus: A review of epidemiological models and early warning systems. *PLoS Negl. Trop. Dis.* **2012**, *6*, e1648. [[CrossRef](#)] [[PubMed](#)]

131. Lowe, R.; Bailey, T.C.; Stephenson, D.B.; Graham, R.J.; Coelho, C.A.S.; Sá Carvalho, M.; Barcellos, C. Spatio-temporal modelling of climate-sensitive disease risk: Towards an early warning system for dengue in Brazil. *Comput. Geosci.* **2011**, *37*, 371–381. [[CrossRef](#)]
132. Yu, H.-L.; Yang, S.-J.; Yen, H.-J.; Christakos, G. A spatio-temporal climate-based model of early dengue fever warning in southern Taiwan. *Stoch. Environ. Res. Risk Assess.* **2011**, *25*, 485–494. [[CrossRef](#)]
133. Salje, H.; Lessler, J.; Endy, T.P.; Curriero, F.C.; Gibbons, R.V.; Nisalak, A.; Nimmannitya, S.; Kalayanarooj, S.; Jarman, R.G.; Thomas, S.J.; et al. Revealing the microscale spatial signature of dengue transmission and immunity in an urban population. *Proc. Natl. Acad. Sci. USA* **2012**, *109*, 9535–9538. [[CrossRef](#)]
134. McGough, S.F.; Brownstein, J.S.; Hawkins, J.B.; Santillana, M. Forecasting zika incidence in the 2016 latin america outbreak combining traditional disease surveillance with search, social media, and news report data. *PLoS Negl. Trop. Dis.* **2017**, *11*, e0005295. [[CrossRef](#)]
135. Ashby, J.; Moreno-Madriñán, M.; Yiannoutsos, C.; Stanforth, A. Niche modeling of dengue fever using remotely sensed environmental factors and boosted regression trees. *Remote Sens.* **2017**, *9*, 328. [[CrossRef](#)]
136. Peterson, A.T.; Martínez-Campos, C.; Nakazawa, Y.; Martínez-Meyer, E. Time-specific ecological niche modeling predicts spatial dynamics of vector insects and human dengue cases. *Trans. R. Soc. Trop. Med. Hyg.* **2005**, *99*, 647–655. [[CrossRef](#)] [[PubMed](#)]
137. Messina, J.P.; Kraemer, M.U.; Brady, O.J.; Pigott, D.M.; Shearer, F.M.; Weiss, D.J.; Golding, N.; Ruktanonchai, C.W.; Gething, P.W.; Cohn, E.; et al. Mapping global environmental suitability for Zika virus. *eLife* **2016**, *5*, e15272. [[CrossRef](#)]
138. Nsoesie, E.O.; Kraemer, M.U.G.; Golding, N.; Pigott, D.M.; Brady, O.J.; Moyes, C.L.; Johansson, M.A.; Gething, P.W.; Velayudhan, R.; Khan, K.; et al. Global distribution and environmental suitability for chikungunya virus, 1952 to 2015. *Eurosurveillance* **2016**, *21*, 30234. [[CrossRef](#)]
139. Xu, L.; Stige, L.C.; Chan, K.-S.; Zhou, J.; Yang, J.; Sang, S.; Wang, M.; Yang, Z.; Yan, Z.; Jiang, T.; et al. Climate variation drives dengue dynamics. *Proc. Natl. Acad. Sci. USA* **2017**, *114*, 113–118. [[CrossRef](#)]
140. Manore, C.A.; Davis, J.K.; Christofferson, R.C.; Wesson, D.M.; Hyman, J.M.; Mores, C.N. Towards an early warning system for forecasting human West Nile virus incidence. *PLoS Curr.* **2014**, *6*, 25914857. [[CrossRef](#)]
141. Zhang, Q.; Sun, K.; Chinazzi, M.; y Piontti, A.P.; Dean, N.E.; Rojas, D.P.; Merler, S.; Mistry, D.; Poletti, P.; Rossi, L. Spread of Zika virus in the Americas. *Proc. Natl. Acad. Sci. USA* **2017**, *114*, E4334–E4343. [[CrossRef](#)]
142. Ewing, D.A.; Cobbold, C.A.; Purse, B.V.; Nunn, M.A.; White, S.M. Modelling the effect of temperature on the seasonal population dynamics of temperate mosquitoes. *J. Theor. Biol.* **2016**, *400*, 65–79. [[CrossRef](#)]
143. Lunde, T.M.; Bayoh, M.N.; Lindtjørn, B. How malaria models relate temperature to malaria transmission. *Parasit. Vectors* **2013**, *6*, 20. [[CrossRef](#)] [[PubMed](#)]
144. Magori, K.; Legros, M.; Puente, M.E.; Focks, D.A.; Scott, T.W.; Lloyd, A.L.; Gould, F. Skeeter Buster: A stochastic, spatially explicit modeling tool for studying *Aedes aegypti* population replacement and population suppression strategies. *PLoS Negl. Trop. Dis.* **2009**, *3*, e508. [[CrossRef](#)] [[PubMed](#)]
145. Xu, C.; Legros, M.; Gould, F.; Lloyd, A.L. Understanding uncertainties in model-based predictions of *Aedes aegypti* population dynamics. *PLoS Negl. Trop. Dis.* **2010**, *4*, e830. [[CrossRef](#)] [[PubMed](#)]
146. Pascual, M.; Ahumada, J.A.; Chaves, L.F.; Rodo, X.; Bouma, M. Malaria resurgence in the East African highlands: Temperature trends revisited. *Proc. Natl. Acad. Sci. USA* **2006**, *103*, 5829–5834. [[CrossRef](#)]
147. Erguler, K.; Smith-Unna, S.E.; Waldock, J.; Proestos, Y.; Christophides, G.K.; Lelieveld, J.; Parham, P.E. Large-scale modelling of the environmentally-driven population dynamics of temperate *Aedes albopictus* (Skuse). *PLoS ONE* **2016**, *11*, e0149282. [[CrossRef](#)]
148. Messina, J.P.; Brady, O.J.; Pigott, D.M.; Golding, N.; Kraemer, M.U.G.; Scott, T.W.; Wint, G.R.W.; Smith, D.L.; Hay, S.I. The many projected futures of dengue. *Nat. Rev. Microbiol.* **2015**, *13*, 230–239. [[CrossRef](#)]
149. Okuneye, K.; Gumel, A.B. Analysis of a temperature- and rainfall-dependent model for malaria transmission dynamics. *Math. Biosci.* **2017**, *287*, 72–92. [[CrossRef](#)]
150. Parham, P.E.; Michael, E. Modeling the effects of weather and climate change on Malaria transmission. *Environ. Health Perspect.* **2010**, *118*, 620–626. [[CrossRef](#)]
151. Abdelrazec, A.; Gumel, A.B. Mathematical assessment of the role of temperature and rainfall on mosquito population dynamics. *J. Math. Biol.* **2017**, *74*, 1351–1395. [[CrossRef](#)]
152. Tompkins, A.M.; Ermert, V. A regional-scale, high resolution dynamical malaria model that accounts for population density, climate and surface hydrology. *Malar. J.* **2013**, *12*, 65. [[CrossRef](#)]

153. Bomblies, A.; Duchemin, J.-B.; Eltahir, E.A.B. Hydrology of malaria: Model development and application to a Sahelian village. *Water Resour. Res.* **2008**, *44*. [[CrossRef](#)]
154. Soti, V.; Tran, A.; Degenne, P.; Chevalier, V.; Lo Seen, D.; Thiongane, Y.; Diallo, M.; Guégan, J.-F.; Fontenille, D. Combining hydrology and mosquito population models to identify the drivers of rift valley fever emergence in semi-arid regions of West Africa. *PLoS Negl. Trop. Dis.* **2012**, *6*, e1795. [[CrossRef](#)]
155. Little, E.; Bajwa, W.; Shaman, J. Local environmental and meteorological conditions influencing the invasive mosquito *Ae. albopictus* and arbovirus transmission risk in New York City. *PLoS Negl. Trop. Dis.* **2017**, *11*, e0005828. [[CrossRef](#)]
156. Colón-González, F.J.; Fezzi, C.; Lake, I.R.; Hunter, P.R. The effects of weather and climate change on dengue. *PLoS Negl. Trop. Dis.* **2013**, *7*, e2503. [[CrossRef](#)]
157. Naish, S.; Dale, P.; Mackenzie, J.S.; McBride, J.; Mengersen, K.; Tong, S.L. Climate change and dengue: A critical and systematic review of quantitative modelling approaches. *BMC Infect. Dis.* **2014**, *14*, 167. [[CrossRef](#)]
158. Johnson, L.R.; Gramacy, R.B.; Cohen, J.; Mordecai, E.; Murdock, C.; Rohr, J.; Ryan, S.J.; Stewart-Ibarra, A.M.; Weikel, D. Phenomenological forecasting of disease incidence using heteroskedastic Gaussian processes: A dengue case study. *Ann. Appl. Stat.* **2018**, *12*, 27–66. [[CrossRef](#)]
159. Martin, V.; Chevalier, V.Á.; Ceccato, P.N.; Anyamba, A.; De Simone, L.; Lubroth, J.; Domenech, J. The impact of climate change on the epidemiology and control of Rift Valley fever. *Rev. Sci. Tech.* **2008**, *27*, 413–426. [[CrossRef](#)]
160. Sallam, M.; Fizer, C.; Pilant, A.; Whung, P. Systematic review: Land cover, meteorological, and socioeconomic determinants of *Aedes* mosquito habitat for risk mapping. *Int. J. Environ. Res. Public Health* **2017**, *14*, 1230. [[CrossRef](#)] [[PubMed](#)]
161. McMahan, B.H.; Manore, C.A.; Hyman, J.M.; LaBute, M.X.; Fair, J.M. Coupling vector-host dynamics with weather geography and mitigation measures to model rift valley fever in Africa. *Math. Model. Nat. Phenom.* **2014**, *9*, 161–177. [[CrossRef](#)]
162. Manore, C.A.; Beechler, B.R. Inter-epidemic and between-season persistence of rift valley fever: Vertical transmission or cryptic cycling? *Transbound. Emerg. Dis.* **2015**, *62*, 13–23. [[CrossRef](#)] [[PubMed](#)]
163. Muñoz, Á.G.; Thomson, M.C.; Stewart-Ibarra, A.M.; Vecchi, G.A.; Chourio, X.; Nájera, P.; Moran, Z.; Yang, X. Could the recent zika epidemic have been predicted? *Front. Microbiol.* **2017**, *8*, 1291. [[CrossRef](#)]
164. Zouache, K.; Fontaine, A.; Vega-Rua, A.; Mousson, L.; Thiberge, J.-M.; Lourenco-De-Oliveira, R.; Caro, V.; Lambrechts, L.; Failloux, A.-B. Three-way interactions between mosquito population, viral strain and temperature underlying chikungunya virus transmission potential. *Proc. R. Soc. B Biol. Sci.* **2014**, *281*, 20141078. [[CrossRef](#)] [[PubMed](#)]
165. Kassa, S.M.; Ouhinou, A. The impact of self-protective measures in the optimal interventions for controlling infectious diseases of human population. *J. Math. Biol.* **2015**, *70*, 213–236. [[CrossRef](#)]
166. Agosto, F.B.; Gumel, A.B.; Parham, P.E. Qualitative assessment of the role of temperature variations on malaria transmission dynamics. *J. Biol. Syst.* **2015**, *23*, 1550030. [[CrossRef](#)]
167. Sellers, W.D. A global climatic model based on the energy balance of the Earth-atmosphere system. *J. Appl. Meteorol.* **1969**, *8*, 392–400. [[CrossRef](#)]
168. Bryan, K. Climate and the ocean circulation. *Mon. Weather Rev.* **1969**, *97*, 806–827. [[CrossRef](#)]
169. Manabe, S. Climate and the ocean circulation: I. The atmospheric circulation and the hydrology of the earth's surface. *Mon. Weather Rev.* **1969**, *97*, 739–774. [[CrossRef](#)]
170. Hayhoe, K.; Edmonds, J.; Kopp, R.E.; LeGrande, A.N.; Sanderson, B.M.; Wehner, M.F.; Wuebbles, D.J. Climate Models, Scenarios, and Projections. In *Climate Science Special Report: Fourth National Climate Assessment, Volume I*; U.S. Global Change Research Program: Washington, DC, USA, 2017.
171. Flato, G.M. Earth system models: An overview. *Wiley Interdiscip. Rev. Clim. Chang.* **2011**, *2*, 783–800. [[CrossRef](#)]
172. Calvin, K.; Bond-Lamberty, B. Integrated human-earth system modeling—State of the science and future directions. *Environ. Res. Lett.* **2018**, *13*, 063006. [[CrossRef](#)]
173. Boyce, R.; Reyes, R.; Matte, M.; Ntaro, M.; Mulogo, E.; Metlay, J.P.; Band, L.; Siedner, M.J. Severe flooding and malaria transmission in the western Ugandan Highlands: Implications for disease control in an era of global climate change. *J. Infect. Dis.* **2016**, *214*, 1403–1410. [[CrossRef](#)]

174. Paull, S.H.; Horton, D.E.; Ashfaq, M.; Rastogi, D.; Kramer, L.D.; Diffenbaugh, N.S.; Kilpatrick, A.M. Drought and immunity determine the intensity of West Nile virus epidemics and climate change impacts. *Proc. R. Soc. B Biol. Sci.* **2017**, *284*, 20162078. [[CrossRef](#)]
175. Caminade, C.; Turner, J.; Metelmann, S.; Hesson, J.C.; Blagrove, M.S.C.; Solomon, T.; Morse, A.P.; Baylis, M. Global risk model for vector-borne transmission of Zika virus reveals the role of El Niño 2015. *Proc. Natl. Acad. Sci. USA* **2017**, *114*, 119–124. [[CrossRef](#)]
176. Wesolowski, A.; Eagle, N.; Tatem, A.J.; Smith, D.L.; Noor, A.M.; Snow, R.W.; Buckee, C.O. Quantifying the impact of human mobility on malaria. *Science* **2012**, *338*, 267–270. [[CrossRef](#)] [[PubMed](#)]
177. Deville, P.; Linard, C.; Martin, S.; Gilbert, M.; Stevens, F.R.; Gaughan, A.E.; Blondel, V.D.; Tatem, A.J. Dynamic population mapping using mobile phone data. *Proc. Natl. Acad. Sci. USA* **2014**, *111*, 15888–15893. [[CrossRef](#)] [[PubMed](#)]
178. LaDeau, S.L.; Glass, G.E.; Hobbs, N.T.; Latimer, A.; Ostfeld, R.S. Data–model fusion to better understand emerging pathogens and improve infectious disease forecasting. *Ecol. Appl.* **2011**, *21*, 1443–1460. [[CrossRef](#)] [[PubMed](#)]
179. Ostfeld, R.; Glass, G.; Keesing, F. Spatial epidemiology: An emerging (or re-emerging) discipline. *Trends Ecol. Evol.* **2005**, *20*, 328–336. [[CrossRef](#)] [[PubMed](#)]
180. Kitron, U.; Clennon, J.A.; Cecere, M.C.; Gürtler, R.E.; King, C.H.; Vazquez-Prokopec, G. Upscale or downscale: Applications of fine scale remotely sensed data to Chagas disease in Argentina and schistosomiasis in Kenya. *Geospat. Health* **2006**, *1*, 49. [[CrossRef](#)]
181. Glass, G.E. Rainy with a chance of plague: Forecasting disease outbreaks from satellites. *Future Virol.* **2007**, *2*, 225–229. [[CrossRef](#)]
182. Hobbs, N.T.; Hilborn, R. Alternatives to statistical hypothesis testing in ecology: A guide to self teaching. *Ecol. Appl.* **2006**, *16*, 5–19. [[CrossRef](#)]
183. Bjørnstad, O.N.; Grenfell, B.T. Noisy clockwork: Time series analysis of population fluctuations in animals. *Science* **2001**, *293*, 638–643. [[CrossRef](#)] [[PubMed](#)]
184. Ball, C.; Priya, A.; Braswell, T.; Renzi, R.; Helm, J.; Coffey, L.; Meagher, R. Smart trap for autonomous monitoring of mosquito-borne viruses. In Proceedings of the 2017 Defense Threat Reduction Agency (DTRA) Chemical and Biological Defense Science & Technology (CBD S&T) Conference, Long Beach, CA, USA, 28–30 November 2017.
185. Polgreen, P.M.; Chen, Y.; Pennock, D.M.; Nelson, F.D. Using internet searches for influenza surveillance. *Clin. Infect. Dis.* **2008**, *47*, 1443–1448. [[CrossRef](#)]
186. Ginsberg, J.; Mohebbi, M.H.; Patel, R.S.; Brammer, L.; Smolinski, M.S.; Brilliant, L. Detecting influenza epidemics using search engine query data. *Nature* **2009**, *457*, 1012–1014. [[CrossRef](#)] [[PubMed](#)]
187. Signorini, A.; Segre, A.M.; Polgreen, P.M. The use of Twitter to track levels of disease activity and public concern in the U.S. during the Influenza A H1N1 pandemic. *PLoS ONE* **2011**, *6*, e19467. [[CrossRef](#)] [[PubMed](#)]
188. Generous, N.; Fairchild, G.; Deshpande, A.; Del Valle, S.Y.; Priedhorsky, R. Global disease monitoring and forecasting with Wikipedia. *PLoS Comput. Biol.* **2014**, *10*, e1003892. [[CrossRef](#)]
189. Ziemann, A.; Fairchild, G.; Conrad, J.; Manore, C.; Parikh, N.; Del Valle, S.; Generous, N. Predicting dengue incidence in Brazil using broad-scale spectral remote sensing imagery. In Proceedings of the IGARSS 2018—2018 IEEE International Geoscience and Remote Sensing Symposium, Valencia, Spain, 22–27 July 2018; pp. 2076–2078.
190. Stein, M.L. *Interpolation of Spatial Data*; Springer: New York, NY, USA, 1999.
191. Gelfand, A.E.; Diggle, P.; Guttorp, P.; Fuentes, M. *Handbook of Spatial Statistics*; CRC Press: Boca Raton, FL, USA, 2010.
192. Clark, J.S.; Ferraz, G.; Oguge, N.; Hays, H.; DiCostanzo, J. Hierarchical Bayes for structured, variable populations: From recapture data to life-history prediction. *Ecology* **2005**, *86*, 2232–2244. [[CrossRef](#)]
193. Qian, S.S.; Cuffney, T.F.; Alameddine, I.; McMahon, G.; Reckhow, K.H. On the application of multilevel modeling in environmental and ecological studies. *Ecology* **2010**, *91*, 355–361. [[CrossRef](#)] [[PubMed](#)]
194. Ladeau, S.L.; Clark, J.S. Elevated CO₂ and tree fecundity: The role of tree size, interannual variability, and population heterogeneity. *Glob. Chang. Biol.* **2006**, *12*, 822–833. [[CrossRef](#)]

