

Estrogen receptor beta as a prognostic factor in breast cancer patients: A systematic review and meta-analysis

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Keywords: estrogen receptor beta, breast cancer, survival, endocrine therapy, prognostic factor

Received: October 11, 2015

Accepted: January 24, 2016

Published: February 06, 2016

ABSTRACT

Background: The prognostic role of estrogen receptor beta (ER β) in early-stage breast cancer is unclear. We performed a systematic review and meta-analysis to evaluate the prognostic value of ER β in early-stage breast cancer patients.

Method: We searched Medline, Embase, and the Web of Science for studies published between 1990 and 2015 that assessed ER β status in breast cancer patients. A total of 25 studies comprising 9919 patients fitting our inclusion and exclusion criteria were included. The hazard ratios of ER β status were extracted for disease free survival (DFS) and overall survival (OS). Random or fixed-effects models were used when appropriate, and between-study heterogeneity was assessed.

Results: In the 20 studies that assessed ER β status using immunohistochemical (IHC) methods, we observed significantly improved DFS in patients positive for ER β -1 (HR=0.56, 95%CI 0.40-0.78, $P=0.0007$) and ER β -2 (HR=0.67, 95%CI 0.45-1.00, $P=0.05$). Improved OS was associated with a positive status for pan-ER β (HR=0.60, 95%CI 0.45-0.80, $P=0.0004$) and ER β -2 (HR=0.44, 95%CI 0.31-0.62, $P<0.0001$). In ER α -positive patients, ER β positivity was not associated with DFS (HR=0.77, 95%CI 0.46-1.27, $P=0.31$) or OS (HR=0.64, 95%CI 0.37-1.11, $P=0.11$). In contrast, ER β expression was significantly associated with increased DFS (HR=0.37, 95%CI 0.14-0.93, $P=0.03$) or OS (HR=0.44, 95%CI 0.30-0.65, $P<0.0001$) in ER α -negative patients. We did not observe an association between ER β mRNA levels and DFS and OS.

Conclusion: In this study, we showed that IHC ER β status, rather than mRNA levels, is a prognostic factor that is associated with DFS and OS in breast cancer patients. The prognostic value of ER β may be higher in ER α -negative patients than in ER α -positive patients.

INTRODUCTION

Estrogen receptor α (ER α) has been established as a significant predictor of the response to endocrine therapy in breast cancer patients. Immunohistochemical (IHC) examination of ER α status is the standard-of-care pathological evaluation used to guide adjuvant endocrine therapy after surgery. Anti-estrogen approaches are recommended in ER α + patients. The discovery of a second ER, ER β , has led to the re-evaluation of estrogen activity in normal mammary development, breast tumorigenesis and tumor progression. Despite over 15 years of research on ER β , its clinical significance remains unclear. Mann et

al. [1] were the first to report the significance of ER β in predicting long-term clinical outcomes (e.g., disease-free survival) in breast cancer patients, a result confirmed by other studies [2-4]. However, conflicting findings suggest that ER β status is not associated with survival [5, 6]. The aim of the present systematic review and meta-analysis was to investigate the association of ER β status (positive vs. negative) and long-term clinical outcomes (e.g. disease-free survival, overall survival) of breast cancer patients.

Table 1a: Features of included studies.

| References | Year | Patients (n) | Mean age | Methods • | ERb assessment •• | ERb status | | Median Follow up(Months) | Quality Score |
|----------------------------------|------|--------------|----------|-----------|-------------------|----------------|----------------|--------------------------|---------------|
| | | | | | | ERb+ | ERb- | | |
| Borgquist et al.[11] | 2008 | 512 | 64.2 | i | ii | 167 | 312 | 106 | ***** |
| | | 114 # | | | | 60 | 54 | NA | |
| | | 139 ## | | | | 71 | 68 | NA | |
| Chantzi et al.[20] | 2013 | 95 | 52 | i | i | b1:66 b2:65 | b1:29 b2:30 | NA | ***** |
| Gruvberger-Saal et al.[5]§, ¶¶¶¶ | 2007 | 425 | NA | i | ii | 262 | 91 | 174 | ***** |
| Guo et al. [21]¶¶¶¶¶¶ | 2014 | 490 | 49 | i | ii | 110 | 380 | 60 | ***** |
| Honma et al. [2]§ | 2008 | 442 | 56 | i | ii | 405 | 37 | 133 | ***** |
| Hopp et al.[12] | 2004 | 305 | 62 | iii | v | 141 | 164 | 65 | ***** |
| | | 186 # | | | | 89 | 97 | 74 | |
| | | 119 ## | | | | 52 | 67 | 50 | |
| Kim et al.[13] | 2012 | 139 | NA | ii | iii | 53 | 87 | 48 | ***** |
| Mahle et al.[14]§ | 2009 | 145 | 63 | i | ii | 129 | 16 | 165 | ***** |
| Mann et al.[1] | 2001 | 47 ## | NA | i | ii | 33 | 14 | 88 | **** |
| | | 118 # | NA | | ii | 78 | 40 | 49 | |
| Markey et al.[28] | 2009 | 121 | 54 | ii | iii | 50 | 71 | 38 | *** |
| Myers et al.[15] | 2004 | 150 | NA | i | i | 87 | 63 | 27 | *** |
| Nakopoulou et al.[3] | 2004 | 181 | 61 | i | ii | 128 | 50 | 76 | ***** |
| Novelli et al.[6] | 2008 | 936 | NA | i | ii | 520 | 416 | 50 | ***** |
| Omoto et al.[18] | 2002 | 57 | 60.9 | i | ii | 15 | 42 | 48 | *** |
| Omoto et al.[17] | 2001 | 88 | 54 & | i | i | 52 | 36 | NA | **** |
| O'Neill et al. ¶[16] | 2004 | 167 | NA | i | ii | 117 | 10 | NA | ***** |
| | | | NA | ii | iii | 86 | 35 | | |
| Palmieri et al.[19] | 2004 | 82 | 59 | i | i | 33 | 46 | 96 ¶¶ | **** |
| Qui et al.[22] | 2009 | 308 | 58 | i | ii | 123 | 185 | 48 | *** |
| Shaaban et al.[23] | 2008 | 880 | NA | i | i, ii | 558 | 112 | 94 | ***** |
| Sugiura et al.[24] | 2007 | 150 | 53 | i | i | 103 | 47 | 58 | *** |
| | | | | ii | iii | 52 | 98 | | |
| Vinayagam et al. [4]¶,§ | 2007 | 141 | 68 | i | i | 100 | 41 | BCS:71; BCR:79 | ***** |
| | | 100 | | ii | iii | 34 | 30 | | |

| | | | | | | | | | |
|---------------------|------|-------------------|------|-----|----|------------------|------------------|------|-------|
| Wen et al.[25] | 2002 | 116 | 53.7 | iii | v | 40 | 76 | 35.3 | ***** |
| Wimberly et al.[26] | 2014 | Yale-1:649 | NA | iv | iv | b1:228 b5:209 | b1:228 b5:209 | 95 | **** |
| | | Yale-2:398 | | | | b1:147 b5:153 | b1:148 b5:152 | 123 | **** |
| | | Toronto: 976 | | | | b1:225 b5:153 | b1:225 b5:153 | 98.2 | **** |
| | | NCI-PBCS: 1375 | | | | b5:467 | b5:468 | 116 | **** |
| Yan et al.¶¶¶,§[27] | 2011 | 147 | NA | i | ii | 90 | 20 | 64 | *** |
| Zhang et al.[29] | 2014 | 279 | 48.8 | i | ii | 40 | 109 | 92 | *** |

• i.IHC; ii, PCR; iii, Immunoblot; iv. TMA
• • i, Allred score; ii, Proportion of positive cells; iii, Ct value; iv, AQUA score; v, Band intensities
#, Tamoxifen/endocrine-treated subgroup; ##, untreated subgroup; & Median;
¶ Postmenopausal patients.
¶¶ Estimated based on the description in the text.¶¶¶ Familial breast cancer patients.
¶¶¶ Stage II patients.
¶¶¶¶ This group was reported in three publications involving the same study population. We selected the study with the longest follow-up period for analysis.
NA, Not available; ER, estrogen receptor;

RESULTS

Study characteristics

Twenty-five studies [1-25] with the full text available were identified and included in this study (Figure 1). We examined the reference list of each study and did not identify any further studies for inclusion in our analysis. We included a total of 9919 patients from these studies. All publications were full-text articles. The features of the included studies are summarized in Table 1a, 1b. The mean patient age ranged from 48 to 68 years, and the median follow-up ranged from 27 to 174 months. Nine of the included studies had a quality score ≥ 6 . None of the included studies were prospective, randomized trials. All of the studies were retrospective and did not report any information about allocation concealment or blinding methods. The matching criteria varied among the studies. Most of the studies reported the length of the follow-up period, and 12 of them exhibited a sufficiently long follow-up (defined as a median follow-up time >60 months) for the outcomes to be determined. The treatment of missing data was not sufficiently described in most of the studies.

The effect of ER β on DFS

A total of 16 studies [2-7, 11-17, 19, 25] with available DFS used IHC as the method of ER β assessment.

Pooling the data showed that a positive status for ER β -1 (HR=0.56, 95%CI 0.40-0.78, $P=0.0007$; heterogeneity: $P<0.01$, $I^2=64\%$) or ER β -2 (HR=0.67, 95%CI 0.45-1.00, $P=0.005$; heterogeneity: $P=0.10$, $I^2=45\%$) was significantly associated with improved DFS (Figure 2). Two studies [8, 21] used immunoblotting to assess pan-ER β status. Pooling the data revealed that a positive pan-ER β status was associated with an improved DFS (HR=0.51, 95%CI 0.35-0.75, $P=0.0007$; heterogeneity: $P=0.33$, $I^2=9\%$; Figure S1). Five studies [4, 9, 12, 20, 24] assessed ER β mRNA levels via PCR, and no association between total ER β mRNA levels and DFS was detected (Figure S2). Wimberly et al. [22] employed a tissue microarray (TMA) to assess the pan-ER β and ER β -1 statuses of four independent populations. However, there was no association between ER β status and DFS in these populations (Figure S3).

The effect of ER β on OS

We pooled the data from 11 studies [1-5, 10, 15, 18-20, 23] with available overall survival data and observed that improved OS was associated with a positive status for pan-ER β (HR=0.60, 95%CI 0.45-0.80, $P=0.0004$; heterogeneity: $P=0.71$, $I^2=0\%$) and ER β -2 (HR=0.44, 95%CI 0.31-0.62, $P<0.0001$; heterogeneity: $P=0.90$, $I^2=0\%$), but not ER β -1 (HR=0.55, 95%CI 0.20-1.50, $P=0.24$; heterogeneity: $P<0.01$, $I^2=88\%$; Figure 3). After excluding the study reported by Qui et al. [18], a positive ER β -1 status was shown to be associated with improved OS without significant heterogeneity (HR=0.38,

95%CI 0.25-0.57, heterogeneity: $P=1.00$, $I^2=0\%$). When the data from the two studies [8, 21] that used immunoblotting to assess pan-ER β status were pooled, we observed an association between a positive pan-ER β status and improved OS (HR=0.62, 95%CI 0.46-0.84, $P=0.002$; heterogeneity: $P=0.11$, $I^2=55\%$; Figure S4). There were 3 studies [4, 20, 24] that assessed the mRNA levels of ER β using PCR; we found no association between total ER β mRNA levels and OS (Figure S5).

ER α as an effect modifier

A total of 7 studies [2, 3, 5, 7, 10, 19, 25] reported the HR of the IHC-determined ER β status (pan-ER β /ER β -1/ER β -2) for DFS and OS in ER α -positive or negative patient subgroups. In ER α (+) patients, ER β status was not associated with DFS (HR=0.77, 95%CI 0.46-1.27, $P=0.31$; heterogeneity: $P=0.09$, $I^2=59\%$) or OS (HR=0.64, 95%CI 0.37-1.11, $P=0.11$; heterogeneity: $P=0.09$, $I^2=54\%$). In fact, Zhang [25] found that a positive ER β status was

correlated with improved DFS in univariate, but not multivariate analysis. Vinayagam [4] reported that ER β status was not correlated with DFS, but the associated HR was not available, and this study was therefore not included in the afore mentioned meta-analysis. In contrast, a positive ER β status was significantly associated with increased DFS (HR=0.37, 95%CI 0.14-0.93, $P=0.03$; heterogeneity: $P<0.01$, $I^2=77\%$) and OS (HR=0.44, 95%CI 0.30-0.65, $P<0.0001$; heterogeneity: $P=0.41$, $I^2=0\%$) in ER α (-) patients (Figures 4&5).

Sensitivity analysis and publication bias

A sensitivity analysis revealed that a positive ER β (pan ER β /ER β -1/ER β -2) status was significantly associated with improved DFS or OS in studies with a median follow-up time greater than 60 months [1-8, 10, 15, 17, 19, 20, 22, 23] (Table S1). ER β -1 was not associated with DFS or OS in studies with a sample size ≥ 200 [2, 5-8, 17-19, 25]. The funnel plots for the studies

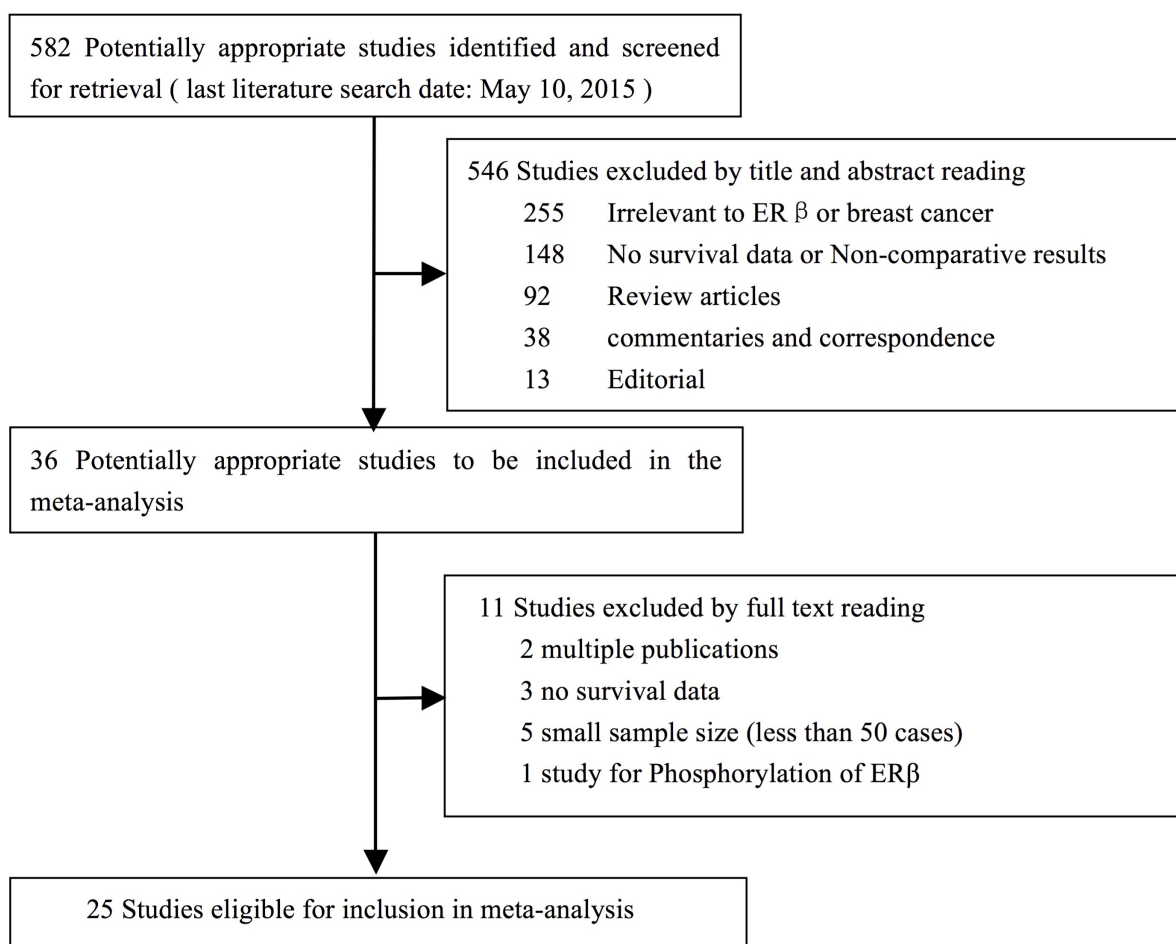


Figure 1: Flow diagram of studies identified, included, and excluded.

for DFS were symmetric, indicating no publication bias (Figure S6). However, the distribution of the OS funnel plots was not symmetric. As shown in Figure S7, the studies focusing on specific ER β (ER β 1/ER β 2) and pan-ER β reactivity were mostly located on the left and right sides of the funnel plot, respectively, indicating possible publication biases.

DISCUSSION

ER β was discovered nearly two decades ago, but its role as a prognostic or predictive factor in breast cancer remains elusive. Most studies examining ER β as a biomarker have been retrospective, and these studies have used a variety of detection methods, leading to discrepant results. IHC is the most common method employed for ER β assessment. In this meta-analysis study, we observed that a positive ER β status, as assessed via IHC, was generally associated with improved DFS and OS. Multiple ER β isoforms (ER β -1, ER β -2/cx) arise via alternative splicing of downstream coding exons or posttranslational proteolysis [26, 27]. In this study, we

noted that ER β -2 was associated with improved DFS and OS. In contrast, ER β -1 was associated with DFS, but not OS, which may be attributed to a study by Qui et al. [18], who provided the only report of an association between positive ER β -1 status and a poorer OS. After the exclusion of this study, the pooled HR(95%CI) of ER β -1 for OS changed significantly, from 0.55(95%CI: 0.20-1.50) to 0.38(95%CI: 0.25-0.57). The heterogeneity of the data synthesis was also eliminated. After careful examination, we noted in consistent results within Qui et al.'s study. In their report, they indicated that ER β -positive patients exhibit a significantly worse overall survival prognosis compared with ER β -negative patients. However, when stratified by HER2 status, the survival curves of the ER β -positive and ER β -negative patients overlapped in both strata. The authors did not attempt to explain this result. We therefore suggest that the exclusion of this study from our meta-data analysis is appropriate.

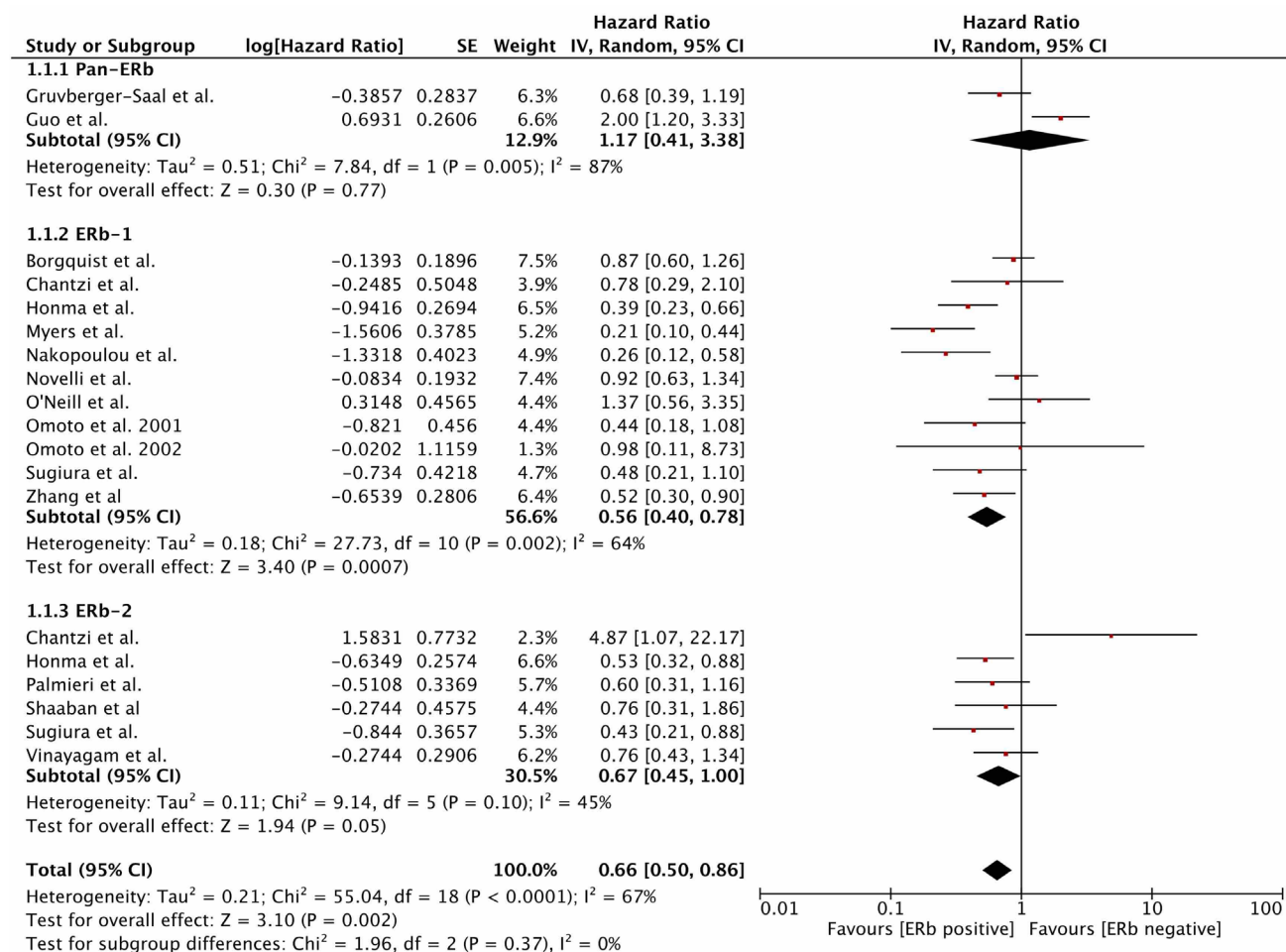


Figure 2: Prognostic role of IHC-determined ER β status for DFS. DFS, disease-free survival; IHC, immunohistochemistry; ER, estrogen receptor.

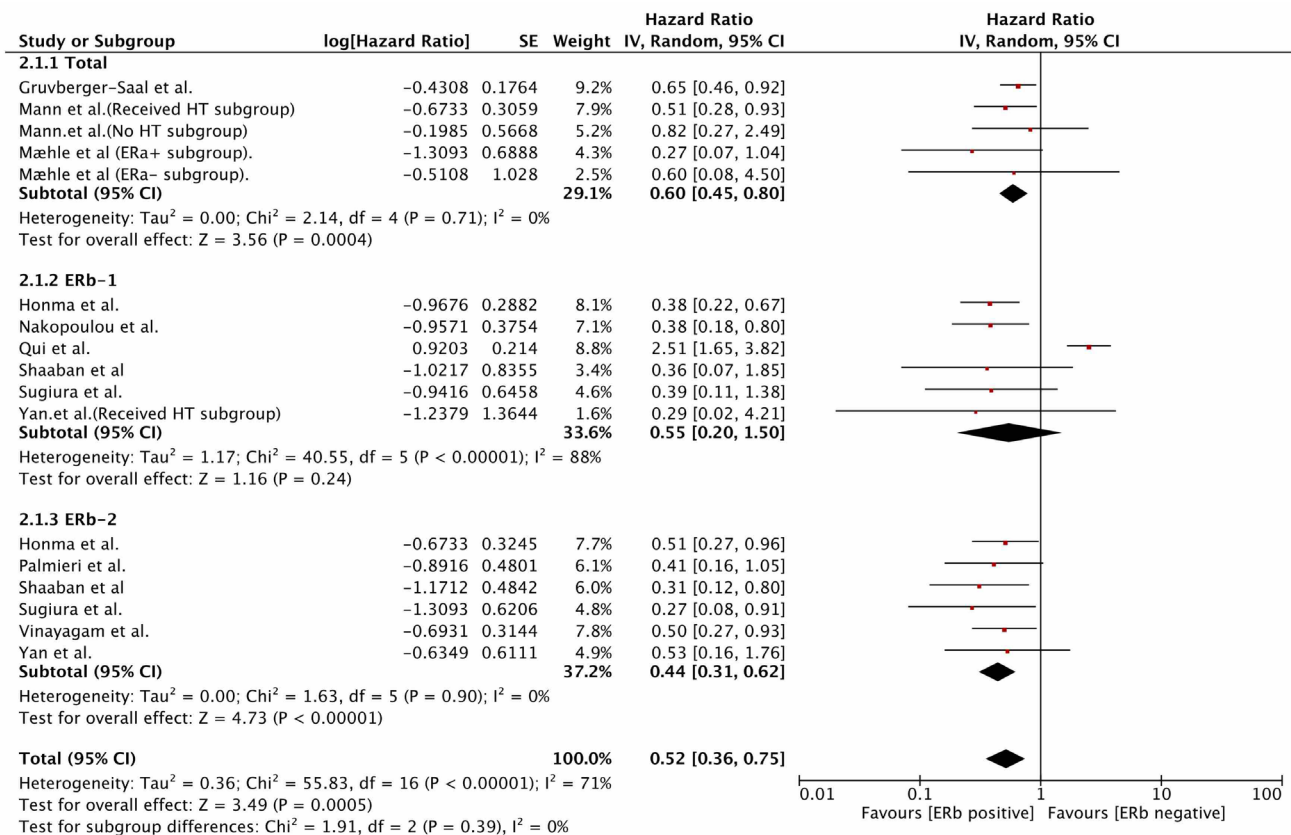


Figure 3: Prognostic role of IHC-determined ER β status for OS. OS, overall survival; IHC, immunohistochemistry; ER, estrogen receptor.

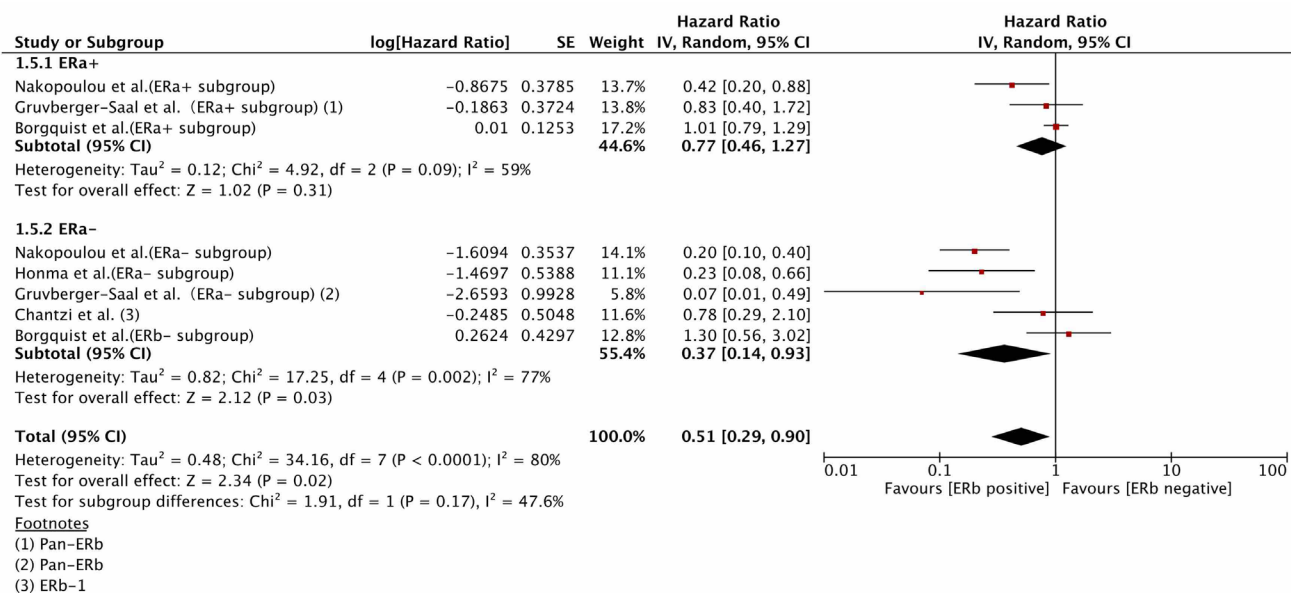


Figure 4: The prognostic role of IHC-determined ER β status for DFS varied by ER α status. DFS, disease-free survival; IHC, immunohistochemistry; ER, estrogen receptor.

Assessment method and clinical outcomes

Various methods had been used to assess ER β status. Two studies employed immunoblotting as the detection method and revealed that a positive ER β status was associated with improved DFS, similar to studies employing IHC [8, 21]. However, a study by Wimberly et al. [22] showed no association between ER β status and DFS when TMA was used to assess ER β status in four large-cohort populations. We speculate that TMA may not be an accurate method for ER β assessment. Its major limitation is that the small cores employed to construct a TMA may not accurately and comprehensively represent the whole tissue specimen. Eckel-Passow et al. [28] reported that the number of TMA cores necessary to adequately represent the whole tissue specimen is biomarker-specific. They showed that 2-3 cores appeared to be adequate for assessing the status of B7-H3, Ki-67, CAIX, and IMP3 expression in renal cancer patients, whereas as many as 10 cores were insufficient for assessing B7-H1. Thus, the association between B7-H1 determined in whole tissue sections and renal cancer-specific death is not easily revealed through TMA assessment.

Several studies found no consistent association between the mRNA and protein levels of ER β [14, 29, 30]. Furthermore, an inverse association between ER β mRNA levels and improved survival has been reported. Speirs et al. [31] noted that ER β mRNA levels were increased in tamoxifen-resistant breast cancer patients. Similarly, Kim et al. [9] reported that a higher ER β mRNA level is associated with poorer DFS in patients treated using endocrine therapy. We believe that the assessment of ER β status based on mRNA levels may be inaccurate because samples from breast tissue might contain cells from surrounding cancerous tissue. Furthermore, post-

transcriptional regulation may also compromise the prognostic value of ER β mRNA [32]. In our meta-analysis, we found no association between ER β mRNA levels and survival (DFS or OS). Hence, ER β mRNA status does not appear to be promising for clinical use.

Prognostic role of ER β varied by ER α status

As noted above, the prognostic value of ER β varies depending on a patient's ER α status. The mechanism underlying this effect may be the molecular interplay between ER α and ER β . Charn et al. [33] investigated the effects of ligand-occupied and unoccupied ER α and ER β on chromatin binding. They showed that although ER α and ER β restrict each other's binding site occupancy, ER α is dominant. The binding sites of ER α and ER β overlap substantially when they are present alone. However, when both ER α and ER β are present, only a few binding sites are shared. When both receptors are present, ER α displaces ER β and shifts ligand binding to sites that are less enriched in the estrogen response element. This finding supports our observation that in ER α + patients, the prognostic role of ER β was less significant than in ER α - patients. Because endocrine therapy is administered to ER α +, but not ER α -, patients, we suggest that endocrine therapy may play a role as an effect modifier. Unfortunately, there are insufficient data to perform a meta-analysis addressing this issue. Novelliet al. [6] reported that in patients who receive endocrine therapy, a positive ER β status is associated with increased DFS. Similar results have been reported by other investigators [1, 8]. However, Yan et al. [23] found that a positive ER β status was associated with improved OS in univariate, but not multivariate, analyses. O'Neil et al. [12] noted a trend (though not statistically significant) toward poorer DFS in patients with a positive ER β status.

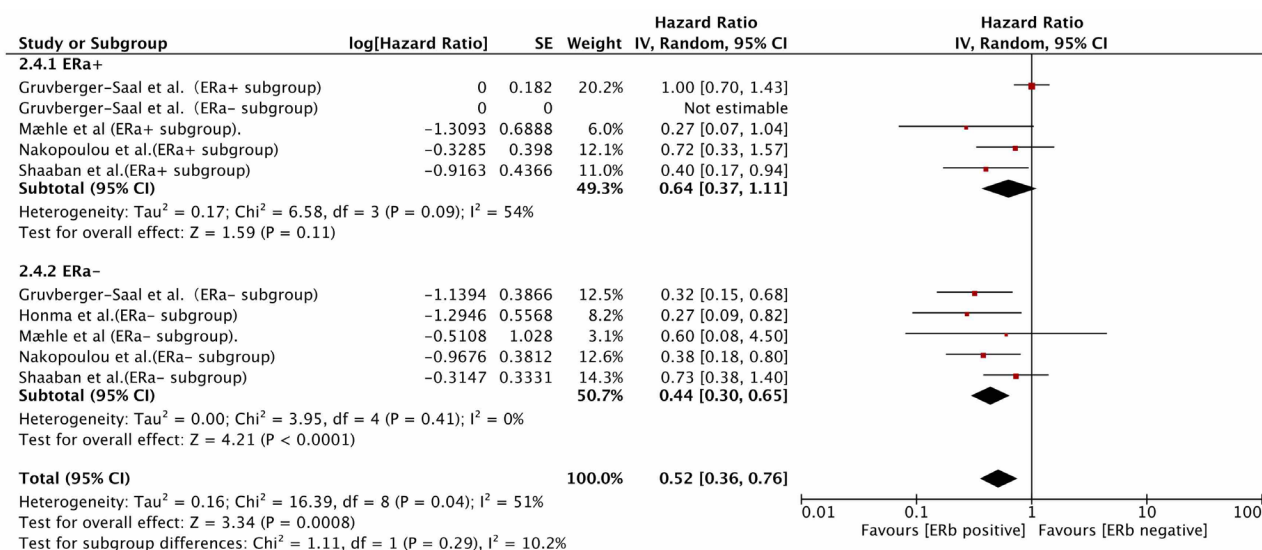


Figure 5: The prognostic role of IHC-determined ER β status for OS varied by ER α status. OS, overall survival; IHC, immunohistochemistry; ER, estrogen receptor.

Table 1b: Features of included studies.

| References | Year | Patients (n) | Antibody | ERα status | | Tumor Burden | | |
|---------------------------|------|--------------|--|------------|------|--------------|--------|-------|
| | | | | ERα+ | ERα- | T1 % | N0 % | G3 % |
| Borgquist et al.[11] | 2008 | 512 | ERβ1: anti-mouse ERβ1 monoclonal antibody (EMR02; Novocastra) | 407 | 72 | 63.1% | 63.1% | NA |
| | | 114 # | | 95 | 19 | NA | NA | NA |
| | | 139 ## | | 114 | 25 | NA | NA | NA |
| Chantzi et al.[20] | 2013 | 95 | ERβ1:anti-mouse ERβ1 monoclonal antibody (Clone PPG5/10; Serotec) ERβ2/cx: anti-human ERβ2 monoclonal antibody (Clone # 57/3; Serotec) | 0 | 95 | 44.2% | 56.8% | 47.4% |
| Gruvberger-Saal et al.[5] | 2007 | 425 | Pan-ERβ:anti-mouse ERβ monoclonal antibody (Clone 14C8; GeneTex) ERβ1:anti-mouse ERβ1 monoclonal antibody (Clone PPG5/10; Serotec) | 248 | 105 | 26.6% | 33.4% | NA |
| Guo et al. [2, 21] | 2014 | 490 | Pan-ERβ: Unclear (Fuzhou Maixin Biotechnology Development) | NA | NA | 32.8% | 51.2% | 26.1% |
| Honma et al. §[2] | 2008 | 442 | Pan-ERβ:anti-rabbit polyclonal antibody (MYEB, M.Y) ERβ1:anti-mouse ERβ1 monoclonal antibody (Clone PPG5/10; DAKO) ERβ2/cx:anti-mouse ERβ2 monoclonal antibody (Clone # 57/3; Serotec) | 364 | 78 | 39.4% | 54.8% | NA |
| Hopp et al.[12] | 2004 | 305 | Pan-ERβ:anti-mouse ERβ monoclonal antibody (Clone 14C8; GeneTex) | 272 | 33 | 23.9% | 0.0% | 43.7% |
| | | 186 # | | 176 | 10 | 26.9% | NA | 40.7% |
| | | 119 ## | | 96 | 23 | 19.5% | NA | 48.3% |
| Kim et al.[13] | 2012 | 139 | NA | 139 | 0 | 61.4% | 42.4% | 20.7% |
| Mahle et al.§[14] | 2009 | 145 | Pan-ERβ:anti-mouse ERβ monoclonal antibody (Clone 14C8; GeneTex) | 97 | 48 | 37.0% | 51.7% | 24.3% |
| Mann et al.[1] | 2001 | 47 ## | Pan-ERβ:anti-rabbit polyclonal antibody (MYEB, M.Y) | 30 | 17 | NA | NA | NA |
| | | 118 # | | 75 | 43 | NA | 100.0% | NA |
| Markey et al.[28] | 2009 | 121 | NA | 82 | 36 | 32.2% | 45.5% | 43.0% |
| Myers et al.[15] | 2004 | 150 | ERβ1:anti-mouse ERβ1 monoclonal antibody (Clone PPG5/10; Serotec) | 123 | 27 | NR | 37.3% | 49.3% |
| Nakopoulou et al.[3] | 2004 | 181 | ERβ1:anti-mouse ERβ1 monoclonal antibody (Clone PPG5/10; Serotec) | 117 | 61 | 27.1% | 38.1% | 29.3% |
| Novelli et al.[6] | 2008 | 936 | Pan-ERβ:anti-mouse ERβ monoclonal antibody (Clone 14C8; Abcam) ERβ1:anti-mouse ERβ1 monoclonal antibody (Clone PPG5/10; GeneTex) | 658 | 278 | 61.9% | 57.6% | 31.2% |
| Omoto et al.[18] | 2002 | 57 | Pan-ERβ: anti-rabbit polyclonal antibody βN; ERβ polyclonal antibody βT; ERβ1: anti-rabbit ERβ1 polyclonal antibody βC ERβ2/cx: anti-rabbit ERβ2cx polyclonal antibody | 39 | 18 | 21.1% | 62.5% | 14.0% |
| Omoto et al.[17] | 2001 | 88 | ERβ1: anti-rabbit ERβ1 polyclonal antibody βC | 62 | 26 | 22.7% | 52.3% | 4.5% |

| | | | | | | | | | |
|------------------------|----|------|----------------|---|-----|-----|----------|--------|-------|
| O'Neill et al.[16] | et | 2004 | 167 | ERβ1:anti-mouse ERβ1 monoclonal antibody (Clone PPG5/10; Serotec) | 83 | 44 | 40.6% | 53.3% | 45.5% |
| | | | | NA | 79 | 42 | | | |
| Palmieri et al.[19] | et | 2004 | 82 | Pan-ERβ:a polyclonal antibody ERβ2/cx: anti-ERβcx sheep polyclonal antibody | 46 | 33 | 25.7% | 53.2% | 40.7% |
| Qui al.[22] | et | 2009 | 308 | ERβ1:anti-rabbit ERβ polyclonal antibody(Ab-1, Oncogene research product) | 198 | 110 | 42.2% && | 37.8% | 39.6% |
| Shaaban et al.[23] | et | 2008 | 880 | ERβ1:anti-mouse ERβ1 monoclonal antibody (Clone PPG5/10; Serotec) ERβ2/cx: anti-human ERβ2 monoclonal antibody (Clone # 57/3; Serotec) | 451 | 219 | NA | NA | 45.8% |
| Sugiura et al.[24] | et | 2007 | 150 | ERβ1: anti-rabbit ERβ1 polyclonal antibody ERβ2/cx: anti-rabbit ERβ2/cx polyclonal antibody | 117 | 33 | 27.3% | 60.4% | 25.2% |
| | | | | NA | 117 | 33 | 27.3% | 60.4% | 25.2% |
| Vinayagam et al.¶,§[4] | et | 2007 | 141 | ERβ2/cx: anti-human ERβ2 monoclonal antibody (Clone # 57/3; Serotec) | 98 | 43 | 44.7% | 47.5% | 43.3% |
| | | | 100 | NA | 70 | 30 | 44.0% | 49.0% | 42.0% |
| Wen et al.[25] | et | 2002 | 116 | Pan-ERβ: anti-goat ERβ polyclonal antibody(Santa Cruz) | 73 | 43 | 12.9% | 37.1% | 38.8% |
| Wimberly et al. [26] | et | 2014 | Yale-1:649 | ERβ1: anti-mouse ERβ1 monoclonal antibody (PPG5/10; ThermoScientific) ERβ5: anti-human ERβ5 monoclonal antibody (Clone 5/25; Serotec) | 246 | 208 | 28.0% | 42.6% | NA |
| | | | Yale-2:398 | | 158 | 102 | 54.8% | 51.4% | |
| | | | Toronto: 976 | | 288 | 118 | 65.3% | 100.0% | |
| | | | NCI-PBCS: 1375 | | 656 | 271 | 52.7% | 59.0% | |
| Yan et al.¶¶¶,§[27] | et | 2011 | 147 | Pan-ERβ:anti-mouse ERβ monoclonal antibody (Clone 14C8; Abcam) ERβ1:anti-mouse ERβ1 monoclonal antibody (Clone PPG5/10; Genetex) ERβ2/cx: anti-human ERβ2 monoclonal antibody (Clone # 57/3; Serotec) | 62 | 48 | 55.3% | 69.9% | 64.4% |
| Zhang et al.[29] | et | 2014 | 279 | ERβ1:anti-mouse ERβ1 monoclonal antibody (Clone PPG5/10; Serotec) ERβ2/cx: anti-human ERβ2 monoclonal antibody (Clone # 57/3; Serotec) | 131 | 21 | 70.5% | 41.6% | 24.8% |

#, Tamoxifen/endocrine-treated subgroup; ##, untreated subgroup; & Median;
\$, Distant disease-free survival was considered to be disease-free survival in this study.
§, Breast cancer death and mortality were considered events affecting overall survival.
&& size<3 cm was considered T1-stage.

¶ Postmenopausal patients.

¶¶ Estimated based on the description in the text.

¶¶¶ Familial breast cancer patients.

¶¶¶¶ Stage II patients.

¶¶¶¶¶ This group was reported in three publications involving the same study population. We selected the study with the longest follow-up period for analysis.

NA, Not available; ER, estrogen receptor;

Hence, the predictive role of ER β for the endocrine therapy response is unclear, due to the conflicting results provided by different studies [1, 6, 7, 12, 23].

No association between ER β status and DFS/OS was observed in patients who did not receive endocrine therapy [1, 7, 23]. We believe that the sample sizes of these studies are too small to detect an association. Our group has initiated a multicenter randomized double-blind prospective clinical trial comparing the efficacy of tamoxifen as an adjuvant endocrine therapy in early-stage ER α /PR-/ER β + breast cancer patients (ClinicalTrials.gov Identifier: NCT02062489). Sun et al. has initiated a similar multicenter study, in which early stage, triple-negative breast cancer patients are randomized into a toremifene/anastrozole group or an observation group (ClinicalTrials.gov Identifier: NCT02089854).

DFS and OS as clinical endpoints

We observed heterogeneity in the synthesis of the HR of pan-ER β or ER β -1 status for DFS. However, there was no heterogeneity in the synthesis of the HR of pan-ER β , ER β -1 (with the exception of Qui's study) or ER β -2 status for OS. We suggest that this discrepancy may be due to the definition of DFS/OS. OS is a universally accepted measure of the clinical benefit of a treatment and can be precisely measured. As a result, there might be less heterogeneity for OS. In contrast, the definition of DFS varies between studies. For example, in the NSABP B-06 study [34], DFS was defined as the first recurrence of disease at a local, regional, or distant site, and the diagnosis of a second cancer and death without evidence of cancer were considered DFS events. In contrast, the guidelines from the DATECAN initiative (Definition for the Assessment of Time-to-event Endpoints in CANcer trials) [35] recommend that DFS should include death of from breast cancer as an event. Most of our included studies did not specify the definition of DFS, which may have resulted in heterogeneity in the synthesis of HR of DFS.

Publication bias

All of the included studies were retrospective and may be subject to publication bias. Insignificant HRs, especially following multivariate analysis, are less likely to be reported in retrospective studies. In the present study, we obtained asymmetric funnel plots for the synthesis of the HRs for OS. Studies reported significant HRs of ER β -1 and ER β -2 for OS tend to fall on the left side of the reference line, indicating that insignificant HRs are less likely to be reported. Several studies [4, 6, 23, 36] reported finding no association between ER β status and OS, but without an available HR and/or 95%CI. Hence, we must be cautious about the prognostic role of ER β for overall

survival.

Limitations

Several additional limitations should be addressed. First, IHC was commonly used for detecting ER β status in most studies, but different hospital used varied commercial antibodies and didn't have uniform criteria. Reported data show that many commercially available IHC stains for ER β have cross-reactivity with ER α [37]. Percentage of immunoreactive cells and allred scores were used to assess ER β status, while the cut-off values varied from 1% to 25% (Percentage of immunoreactive cells) [6, 14, 17], and from 2-4 (Allred score) [2, 11, 15] across different studies. Different cut-off values used by different studies may cause limitation to our analysis. Second, some of the HRs were not available from the full-text of the included study, and were extrapolated from survival curves. Although, this method has been demonstrated to be feasible [38, 39], we still consider this as a limitation. Additionally, HRs for synthesis in our analysis were derived from univariate and/or multivariate analysis (Table S2). This is also a major limitation, as the most standard approach should be collecting HRs derived from prospective controlled trials, with multivariate analysis adjustment.

CONCLUSION

In this meta-analysis, we showed that ER β status, determined via IHC, is generally associated with DFS/OS in breast cancer patients. Assessment of ER β mRNA levels is not recommended. As a prognostic factor, ER β may be more important in ER α (+) patients than ER α (-) patients. Based on these findings, we recommend the initiation of a prospective study to confirm the prognostic value of ER β in breast cancer patients.

MATERIALS AND METHODS

This study was waived the full IRB review of Sun Yat-sen Memorial Hospital, based on the institutional policy. This study was also performed according to the recommendations of the Cochrane Collaboration and the Quality of Reporting of Meta-analysis guidelines (MOOSE) and reported according to the PRISMA statements [40, 41].

Study selection

We searched Medline, Embase, and the Web of Science for potentially relevant studies. The following keywords were searched in the "Title" or "Abstract": "Estrogen receptor," "Beta," and "Breast cancer," without restrictions on the region and publication type. English language was required for publication. We manually

searched the retrieved articles to identify relevant studies. When multiple publications reported on the same study population, the report that was most complete or that had the longest follow-up period was used. The last date of the search was May 10th, 2015.

Inclusion and exclusion criteria

Studies were eligible if they met the following criteria: (1) the main exposure of interest was early-stage breast cancer stratified by ER β status (negative/positive or low/high expression); (2) the outcome of interest was disease-free survival or overall survival; (3) hazard ratios (HRs) with corresponding 95% confidence intervals (CIs) or survival curves for ER β were reported; and (4) over 50 patients were enrolled in the study, which did not present redundant data.

Data extraction

Two reviewers (C.G. & W.T.) independently extracted the data from the included studies. Any disagreement was resolved by the third author (E.S.). The following data were collected: first author, year of publication, clinicopathological features of the study population, methods of ER β assessment, number of included patients, and the reported outcomes. The outcomes assessed included disease-free survival (DFS) and overall survival (OS) in patients with different ER β statuses. We assessed the quality of the included studies using the Newcastle-Ottawa quality assessment tool [42]. We allocated a score of 0-9 to each included study, and those with a score ≥ 6 were considered to be of high quality.

Statistical analysis

The hazard ratio (HR) was used as a summary statistic for survival analysis, as described by Parmar and colleagues [43]. An HR of less than 1 indicated a survival benefit favoring ER β ⁺ patients. We used a random-effects model for this meta-analysis. The data were pooled and weighted using generic inverse variance. Heterogeneity between studies was assessed with the χ^2 and I² statistics. When higher values of the χ^2 and I² statistics ($>50\%$) indicated heterogeneity between studies, we applied sensitivity and subgroup analyses to further evaluate the heterogeneity. We performed a sensitivity analysis when the outcome of interest was reported in more than 3 studies. We used funnel plot analyses to analysis to determine publication bias. A two-tailed p value of less than 0.05 was considered statistically significant. Statistical analyses were performed with Review Manager Version 5.3

ACKNOWLEDGMENTS

We appreciate Yilong Education, Inc, for advices of statistical analysis.

CONFLICTS OF INTEREST

None to disclose.

GRANT SUPPORT

This work was funded by grants from Natural Science Foundation of China (81272893, 81472466, 81402201, 81172514, 81372817, 81230060, 81490750, 81442009); Program for New Century Excellent Talents in University (NCET-12-0565); National Science Foundation of Guangdong Province Grants (2014A03036003, 2014A030310070, S2012030006287, 4202037, 2011A080300002), Guangzhou Science Technology and Innovation Commission (201508020008, 201508020249), Guangdong Science and Technology Department (2015B050501004), Elite Young Scholars Program of Sun Yat-sen Memorial Hospital (Y201401). Grant KLB09001 from the Key Laboratory of Malignant Tumor Gene Regulation and Target Therapy of Guangdong Higher Education Institutes, Sun-Yat-Sen University, Grant [2013]163 from Key Laboratory of Malignant Tumor Molecular Mechanism and Translational Medicine of Guangzhou Bureau of Science and Information Technology.

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