

GOPEN ACCESS

Citation: Yan X, Wang S, Yang B, Zhang W, Cao Y, Shi Y, et al. (2020) QTL mapping for flag leafrelated traits and genetic effect of *QFLW-6A* on flag leaf width using two related introgression line populations in wheat. PLoS ONE 15(3): e0229912. https://doi.org/10.1371/journal.pone.0229912

Editor: Chengdao Li, Murdoch University, AUSTRALIA

Received: June 30, 2019

Accepted: February 17, 2020

Published: March 19, 2020

Copyright: © 2020 Yan et al. This is an open access article distributed under the terms of the <u>Creative</u> Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: All relevant data are within the manuscript and its Supporting Information files.

Funding: This work was supported by the National Transgenic Major Project of China (2018ZX0800917B), National Key R&D Program of China (2017YFD0300202), National Natural Science Foundation of China (31671607) and Key R&D Program in Shanxi (201703D211007-6, 201703D211007-4) RESEARCH ARTICLE

QTL mapping for flag leaf-related traits and genetic effect of *QFLW-6A* on flag leaf width using two related introgression line populations in wheat

Xue Yan¹, Shuguang Wang¹, Bin Yang², Wenjun Zhang¹, Yaping Cao², Yugang Shi¹, Daizhen Sun¹*, Ruilian Jing³*

1 College of Agronomy, Shanxi Agricultural University, Taigu, Shanxi, China, 2 Wheat Research Institute, Shanxi Academy of Agricultural Sciences, Linfen, Shanxi, China, 3 Chinese Academy of Agricultural Sciences, Institute of Crop Science, Beijing, China

* sdz64@126.com (DS); jingrl@caas.net.cn (RJ)

Abstract

The flag leaf is the main organ of photosynthesis during grain-filling period of wheat, and flag leaf-related traits affect plant morphology and yield potential. In this study, two BC_3F_6 introgression line (IL) populations derived from the common recipient parent Lumai 14 with Jing 411 and Shaanhan 8675, respectively, were used to map quantitative trait loci (QTL) for flag leaf length (FLL), flag leaf width (FLW), flag leaf area (FLA) and chlorophyll content (CC) at flowering stage and 15 and 20 days after anthesis (DAA) in 2016–2017 (E1) and 2017-2018 (E2) two environments. A total of 14 and 15 QTLs for flag leaf-related traits were detected in Lumai 14 / Jing 411 and Lumai 14 / Shaanhan 8675 populations, respectively. Among them, Both QFLW-6A and QFLA-6A were detected in Lumai 14 / Jing 411 population under E2 and in Lumai 14 / Shaanhan 8675 population under E1 and E2 environments, respectively. QCC_{S2}-3A from Lumai 14 / Jing 411 population and QCC_{S3}-1A, QFLL-4A and QFLL-6A from Lumai 14 / Shaanhan 8675 population were repeatedly identified under two tested environments. Moreover, eight QTL clusters controlling flag leaf-related traits were identified, which provided a genetic basis for significant correlations in phenotype among these traits. On the other hand, positive alleles of QFLW-6A for FLW detected in two populations were derived from their donors. Eighteen lines and 44 lines carried this QTL were found in Lumai 14 / Jing 411 and Lumai 14 / Shaanhan 8675 populations, respectively. The means of FLW in these lines were wider than that of the recipient parent, Lumai 14, in two environments, suggesting that QFLW-6A played an important role for increasing FLW. The IL 124 in Lumai 14 / Jing 411 population and the IL 59 and IL 127 in Lumai 14 / Shaanhan 8675 population had five, five and four donor chromosomal segments which carried no other QTL controlling FLW than QFLW-6A, respectively. And the FLWs of these lines were significantly greater than that of Lumai 14 under two environments. So these lines and their donor parent can be regarded as potential near-isogenic lines. Further, a synteny analysis found QFLW-6A was near the 574,283,851-574,283,613 bp fragment on chromosome 6A and 10 genes were in the range of 500 kb upstream and downstream of the fragment.

Competing interests: The authors have declared that no competing interests exist.

These results provide the basis for identification of candidate gene and map-based cloning and functional verification of the QTL.

Introduction

Wheat (*Triticum aestivum* L.) is a staple food crop for more than 35% of the population all over the world. The formation of grain yield in wheat is a complex physiological and biochemical process, which is related to the accumulation and assimilation of photosynthetic products during grain-filling period [1], while they are related to the function of leaves [2]. The flag leaves of wheat are considered as the main source of carbohydrates in grains, which contributed up to 50% photosynthetic activity, and about 41–43% of carbohydrates for grain filling after anthesis [3, 4]. Duwayri [5] considered that grain yield and grain number per spike decreased when flag leaves were removed. Many studies have shown that flag leaf size of wheat was positively correlated with thousand-grain weight, grain number per spike, yield per plant and other yield-related traits in cereals [6–10]. The longer the flag leaves maintain high chlorophyll content and photosynthesis, the stronger the assimilation ability of canopy, which provide more assimilation substances for grain filling, delay leaf senescence and ultimately increase grain yield [11–13]. Therefore, the flag leaf size and chlorophyll content are the main factors determining the yield potential of wheat [14–16], and optimal flag leaf size can improve photosynthesis and increase grain yield.

Flag leaf-related traits including chlorophyll content (CC), flag leaf length (FLL), flag leaf width (FLW) and flag leaf area (FLA) are all quantitative traits, and easily affected by environments. With the application of molecular marker and genetic map in crop breeding, many researchers have devoted to quantitative trait loci (QTL) mapping for flag leaf-related traits in rice [17–19], barley [20, 21], sorghum [22] and durum wheat [4, 23]. The *qFL1* for FLL and *aFLW4* and *aFSR4* for FLW have been fine-mapped, and even two genes related to FLW, *Nal1* and Nal7 have been cloned in rice [24-28]. In wheat, QTLs controlling flag leaf-related traits have been identified on almost all 21 chromosomes [16, 29-31]. For example, using recombinant inbred line (RIL) population, 12 QTLs for chlorophyll content-related traits were mapped on chromosomes 1A, 3A, 3B, 3D, 4A, 5A, 6A, 6D, 7A and 7D [32]. Using different RIL populations, Yang et al. [33] reported five additive QTLs for CC, among them, QChl-5A.1 was detected in multiple stages. Twenty-eight QTLs for CC were identified on chromosomes 1B, 2A, 2B, 2D, 3A, 3B, 4B, 4D, 5A, 6B, 6D and 7A by Shi et al. using double haploid (DH) population [34]. Using RIL population with an integrated high-density simple sequence repeat (SSR) and single-nucleotide polymorphism (SNP) genetic linkage map, 61 QTLs for flag leaf morphology trait were detected [9]. A total of 34 QTLs for flag leaf morphology trait were mapped under eight environments using RIL population, among them, two QTLs for FLW qFlw-4B.3 and qFlw-6B.2 and one QTL for FLA qFla-5B detected under more seven environments were stable QTLs [10]. Liu et al. [8] also identified 23 QTLs for FLL, FLW, FLA and flag leaf angle (FLANG) using RIL population, and four QTLs for FLL, two QTLs for FLW, four QTLs for FLA and five QTLs for FLANG were detected at least two environments. Furthermore, a major QTL for FLW, TaFLW1, was fine mapped at 0.2 centiMorgan (cM) interval in the 5AL12-0.35–0.57 deletion bin, which was closely linked with Fhb5 [35]. However, these QTLs were detected using DH or RIL populations. The results were affected by both the genetic background and environment, which was difficult to be applied to breeding program. Introgression lines (IL), also known as chromosomal segment substitution lines (CSSL), are constructed by transferring chromosomal fragment from donor parent into receptor by multiple generations of backcrossing and self-crossing coupled with molecular marker-assisted

selection [<u>36</u>]. Mapping QTL using IL population can reduce the influence of genetic background and improve the accuracy of QTL.

In this study, QTL for FLL, FLW, FLA and CC were mapped using Lumai 14 / Jing 411 and Lumai 14 / Shaanhan 8675 BC_3F_6 populations. The objectives of this study were to (1) identify stably expressed QTLs for flag leaf-related traits in two IL populations, (2) analyze the genetic effects of QTLs that were detected repeatedly in two populations under different environments, (3) detect potential near-isogenic lines controlling flag leaf-related traits. The propose is to provide a foundation for further fine mapping and map-based cloning.

Materials and methods

Plant materials and field trials

Two related BC3F6 IL populations were used in the present study, which were obtained from crossing the common receptor parent Lumai 14 with Jing 411 and Shaanhan 8675, respectively. Both IL populations contained 160 lines. Lumai 14, a variety with high grain yield potential, which was developed by the Yantai Academy of Agricultural Sciences, Shandong, China, was widely cultivated under irrigated condition [37, 38]. The donor parent, Jing 411, with strong cold resistance had been widely grown as one of the main varieties at the Northern Winter Wheat Region of China in the 1990s [39]. The other donor parent, Shaanhan 8675, was a drought-resistant and high-yield cultivar and was released in 1996 by Shaanxi Wheat Research Center, China.

Field trials were conducted at experimental station of Shanxi Agricultural University, Taigu, China (37°25′N, 112°35′E) during 2016–2017 (E1) and 2017–2018 (E2) crop seasons. All the trials were performed in randomized complete block design with three replications. The ILs together with their parents were grown in 2.5 m rows spaced 25 cm apart. Fifty seeds were sown in each row. All of the trials were irrigated before sowing. Plants only relied on natural precipitation during the whole growing period after sowing. The rainfalls in E1 and E2 growing seasons were 138.0 and 196.8 mm (http://data.cma.cn/), respectively. All field experiments were employed in accordance with standard local practices.

Phenotyping and statistical analysis

Five plants with flowering at the same day and developing normally were randomly selected and tagged from the middle of each row. Chlorophyll content (CC) of flag leaves of the tagged plants was measured with a handheld portable chlorophyll meter (SPAD-502, Konica-Minolta, Tokyo) at flowering stage, 15 and 20 days after anthesis (DAA). The flowering stage, 15 and 20 DAA were denoted S1, S2 and S3, respectively. The reading was taken from the average of the base, middle and apical of the flag leaf. The average of CC from five plants was used as phenotypic value for each line. At S1, FLL and FLW were evaluated. Trait means of the five tagged samples from each row were used in the data analysis based on three replications. FLL was measured as the distance from the base to the tip of the leaf. The FLW measurement was taken at the widest part of the flag leaf. FLA, a derived trait, was defined as FLL × FLW × 0.75 [8, 40]. Basic statistics and Pearson's correlation analysis among FLL, FLW, FLA and CC were performed using SPSS 20.0 (SPSS, Chicago, MI, USA).

QTL analysis

The linkage maps of Lumai 14 / Jing 411 and Lumai 14 / Shaanhan 8675 BC₃F₆ populations were constructed, based on 156 and 185 polymorphic simple sequence repeat (SSR) markers, respectively [36, 41]. QTL analysis for flag leaf-related traits was performed by IciMapping 4.0

software (http://www.isbreeding.net/) with the likelihood ratio test based on stepwise regression (RSTEP-LRT). The threshold LOD values were calculated using 1,000 permutations with a type 1 error of 0.05. The QTL nomenclature was according to the rule "QTL + trait + chromosome" formula [34, 42].

Results

Phenotypic variation of flag leaf-related traits

FLL and FLA of both donor parents, Jing 411 and Shaanhan 8675, were much higher than those of recipient parent, Lumai 14, in all environments, and there was significant difference for FLL between Jing 411 and Shaanhan 8675 and Lumai 14, respectively. While, FLW of Jing 411 and Shaanhan 8675 was lower than that of Lumai 14, respectively. Jing 411 and Shaanhan 8675 consistently showed higher values of CC than Lumai 14 in the whole filling grain stage (Table 1). The means of FLL and CC for both IL populations in all environments were intermediate between their parents, except for CC_{S1} in E1, respectively. In both IL populations, bidirectional transgressive segregation was observed for all tested traits, showing wide

Table 1. Phenotypic values for flag leaf-related traits in two IL wheat populations and their parents under two environments.

Trait ¹	Environment ²		Paren	t		Luma	i 14 / Jing 4	11	Lumai 14 / Shaanhan 8675									
		Lumai 14	Jing 411	Shaanhan 8675	Mean ±SD ³	Variation	Skewness	Kurtosis	CV (%) ⁴	Mean ±SD	Variation	Skewness	Kurtosis	CV (%)				
FLL/ cm	E1	15.13	17.12**	18.25**	15.45 ±1.03	13.93– 17.97	0.45	0.41	6.64	15.92 ±1.53	12.49– 18.42	-0.12	0.05	9.62				
	E2	11.83	13.93**	14.26**	11.87 ±1.17	9.69–14.21	0.19	-0.30	9.88	11.87 ±1.35	9.72–14.42	0.19	-0.10	11.38				
FLW/ cm	E1	1.47	1.42	1.31**	1.46 ±0.08	1.28–1.59	-0.14	-0.26	5.61	1.54 ±0.10	1.31–1.79	0.16	0.31	6.52				
	E2	1.31	1.27	1.18*	1.30 ±0.09	1.13–1.48	-0.09	0.01	6.56	1.33 ±0.10	1.14–1.51	-0.06	-0.20	7.17				
FLA/ cm ²	E1	16.72	18.22*	17.98	16.89 ±1.73	14.34- 20.33	0.34	0.25	10.24	18.47 ±2.62	14.48– 23.55	0.10	-0.10	14.19				
	E2	11.66	13.29**	12.62	11.62 ±1.70	8.26-15.76	0.32	0.00	14.66	11.94 ±2.00	8.85-15.89	0.10	-0.10	16.73				
CC _{S1}	E1	59.83	57.55	53.69*	60.34 ±2.02	56.15– 63.97	-0.01	-0.26	3.35	60.04 ±2.50	52.09- 64.45	-0.63	0.59	4.17				
	E2	63.48	59.07**	55.43**	62.70 ±2.78	56.93- 67.19	0.04	-0.16	4.44	63.22 ±3.06	54.11- 67.84	-0.15	-0.02	4.84				
CC_{S2}	E1	59.31	55.23	52.94**	56.81 ±4.21	45.79– 62.72	-0.95	1.39	7.41	58.97 ±2.64	51.21- 63.84	-0.87	1.19	4.47				
	E2	61.52	56.14*	54.12**	58.01 ±3.75	49.93- 63.06	-0.67	0.90	6.46	60.39 ±2.75	52.90- 64.57	-0.42	0.18	4.55				
CC _{S3}	E1	41.42	30.94*	32.75*	35.17 ±10.41	14.51– 56.70	-0.03	-0.57	29.61	38.71 ±8.92	21.65- 56.60	0.11	-0.63	23.04				
	E2	46.61	33.42**	37.14*	35.19 ±9.07	16.07– 54.07	-0.03	-0.49	25.78	39.39 ±7.61	22.80- 55.84	-0.06	-0.28	19.30				

¹: FLL, flag leaf length; FLW, flag leaf width; FLA, flag leaf area; CC_{S1} , chlorophyll content at flowering stage; CC_{S2} , chlorophyll content at 15 days after anthesis; CC_{S3} , chlorophyll content at 20 days after anthesis

²: E1: 2016–2017; E2: 2017–2018

³: SD: standard deviation

⁴: CV: coefficient of variation

* and ** indicate significant difference at the 0.05 and 0.01 probability level, respectively

https://doi.org/10.1371/journal.pone.0229912.t001

phenotypic variability with the coefficients of variation (CV) ranging from 3.35 to 29.61%. The skewness and kurtosis for all treatments were less than 1.00, with the exception of CC_{S2} for Lumai 14 / Shaanhan 8675 population in E2, indicating that they were continuous variation and quantitative genetic basis.

Correlation analysis for flag leaf-related traits

Significantly positive correlations were observed between FLL, FLW and FLA for both IL populations in all experiments. The correlation coefficients between FLL and FLA (r = 0.80 to 0.94) were higher than those between FLW and FLA (r = 0.76 to 0.83), which implied that FLL may be the main contributor to affect FLA. FLW was significantly positive correlated with CC at different stages after anthesis, respectively, except CC_{S3} for Lumai 14 / Jing 411 population and CC_{S1} and CC_{S2} for Lumai 14 / Shaanhan 8675 population in E2. FLA showed a highly significant positive correlation with CC_{S1} and CC_{S2} for both IL populations in E1 and CC_{S3} for Lumai 14 / Shaanhan 8675 population in E2. In addition, CC_{S1} had significantly positive with CC_{S2} and CC_{S3} in all environments, except CC_{S3} for Lumai 14 / Shaanhan 8675 population in E2. There was strongly significantly positive correlation between CC_{S2} and CC_{S3} (Table 2).

Additive QTL analysis for flag leaf-related traits

Fourteen additive QTLs for flag leaf-related traits were detected in Lumai 14 / Jing 411 IL population under two environments, including two QTLs for FLL, three QTLs for FLW, four QTLs for FLA, one QTL for CC_{S1} , three QTLs for CC_{S2} and one QTL for CC_{S3} . These QTLs were distributed on chromosomes 1A, 2A, 3A, 6A, 3B, 4B and 3D with individual QTL contributing 3.02–7.21% to the phenotypic variance (Table 3 and Fig 1). Among them, the favorable alleles of ten QTLs detected were contributed from the donor parent Jing 411, while the favorable alleles of the rest four QTLs mapped were derived from recipient Lumai 14. QCC_{S2} -3A was detected across two tested environments, with explaining 4.09 and 3.96% of the phenotypic variance. And the locus had a favorable allele from Lumai 14 for increasing CC. The rest QTLs were detected only in one environment. In addition, four QTL clusters for flag leaf-

Population	Trait	FLL	FLW	FLA	CC _{S1}	CC _{S2}	CC _{S3}
Lumai 14 / Jing 411	FLL	1.00	0.55**	0.94**	-0.13	0.06	0.11
	FLW	0.21**	1.00	0.80**	0.18*	0.23**	0.11
	FLA	0.80**	0.76**	1.00	-0.01	0.13	0.13
	CC _{S1}	-0.08	0.54**	0.28**	1.00	0.44**	0.27**
	CC _{S2}	-0.14	0.49**	0.21**	0.67**	1.00	0.46**
	CC _{S3}	-0.29**	0.42**	0.09	0.57**	0.64**	1.00
Lumai 14 / Shaanhan 8675	FLL	1.00	0.58**	0.93**	-0.03	0.03	0.41**
	FLW	0.45**	1.00	0.83**	0.14	0.03	0.20*
	FLA	0.89**	0.80**	1.00	0.04	0.03	0.36**
	CC _{S1}	0.11	0.14	0.17*	1.00	0.62**	0.14
	CC _{S2}	0.09	0.22**	0.19*	0.72**	1.00	0.24**
	CC _{S3}	0.10	0.17*	0.15	0.34**	0.43**	1.00

FLL, flag leaf length; FLW, flag leaf width; FLA, flag leaf area; CC_{S1} , chlorophyll content at flowering stage; CC_{S2} , chlorophyll content at 15 days after anthesis; CC_{S3} , chlorophyll content at 20 days after anthesis

Correlation coefficients at the lower and upper triangle part are for 2016–2017 (E1) and 2017–2018 (E2), respectively

* and ** indicate significant difference at the 0.05 and 0.01 level, respectively

https://doi.org/10.1371/journal.pone.0229912.t002

related traits were identified. Two QTL clusters controlling FLL (*QFLL-2A* and *QFLL-4B*) and FLA (*QFLA-2A* and *QFLA-4B*) were detected on chromosomes 2A and 4B. *QFLW-6A* and *QFLA-6A* was co-located on chromosome 6A based on marker *Xwmc201*. Moreover, *QFLW-3B* was clustered with *QFLA-3B*, QCC_{S2} -3B and QCC_{S3} -3B with marker *Xwmc754*. The favorable alleles of these QTL clusters were derived from Jing 411.

In Lumai 14 / Shaanhan 8675 IL population, a total of 15 QTLs controlling flag leaf-related traits were mapped on chromosomes 1A, 4A, 6A, 1B, 2D and 5D in all experiments, seven of which carried the favorable alleles from Lumai 14 (Table 3 and Fig 1). There were four, three, five, one and two QTLs for FLL, FLW, FLW, CC_{S1} and CC_{S3}, respectively, with the phenotypic variation ranging from 2.81 to 14.79%. Of them, five QTLs, *QFLL-4A*, *QFLL-6A*, *QFLW-6A*, *QFLA-6A* and *QCC_{S3}-1A*, were repeatedly identified in two environments. The rest of ten QTLs were detected just in one environment, accounting for phenotypic variation of 2.81 to 9.30%. Four QTL clusters with common trait for FLL were found in this study. *QFLL-4A* was co-localized with *QFLA-4A* based on marker *Xwmc757*, with the Lumai 14-derived alleles simultaneously increasing FLL and FLA. *QFLL-1A* associated with *QFLA-1A* and *QCC_{S3}-1A* were detected on chromosome 1A, and the favorable alleles were also derived from Lumai 14. The alleles of the QTL clusters formed by the remaining two QTLs for FLL (*QFLL-5D* and *QFLL-6A*) and QTLs for FLA (*QFLA-5D* and *QFLA-6A*) on chromosomes 5D and 6A, respectively.

Compared with the above results, we found two QTLs, *QFLW-6A* for FLW and *QFLA-6A* for FLA, were detected in Lumai 14 / Jing 411 IL population in E2 environment and in Lumai 14 / Shaanhan 8675 population in E1 and E2 environments, respectively, suggesting that they were stable QTLs. And the two loci were linked with marker *Xwmc201*, indicating that they may be pleiotropic or tightly linked.

Genetic effect analysis of QFLW-6A

Based on the above results, we found that the *QFLW-6A* was a stable QTL detected under E2 environment in Lumai 14 / Jing 411 IL population and under E1 and E2 environments in Lumai 14 / Shaanhan 8675 IL population. Therefore, it is necessary to analyze the genetic effect *QFLW-6A* on flag leaf width.

Genome-wide scanning and QTL mapping found that 18 lines in Lumai 14 / Jing 411 population contained *QFLW-6A* with the favorable allele originating from Jing 411. The number of chromosomal fragments from donor (Jing 411) ranged from four to twenty-one in all 18 lines. Among them, IL 70 was introgressed four donor chromosomal fragments, while IL 18, 69 and 124 contained five donor fragments, respectively. On the other hand, the mean value of FLW in all lines with the locus *QFLW-6A* was higher than that of recipient parent (Lumai 14) in two environments. The FLW of IL 12 and 110 showed significant difference from Lumai 14 in E1. The FLW of IL 4, 8 and 31 showed significant difference compared with Lumai 14 in E2 environment. In particular, The FLW of IL 124 was significantly wider than Lumai 14 in both E1 and E2. It's worth noting that IL 124 carried no other QTL controlling FLW than *QFLW-6A* (S1 Table, Fig 2A). Therefore, the IL 124 and their recurrent parent can be regarded as potential near-isogenic lines (NILs).

For Lumai 14 / Shaanhan 8675 IL population, *QFLW-6A* was repeatedly detected both in E1 and E2, with positive additive effect from Shaanhan 8675. Forty-four lines anchored this locus. Among of them, 11 lines (IL 2, 19, 59, 76, 77, 80, 104, 118, 126, 127 and 157) were substituted fragments from Shaanhan 8675 no more than five fragments. On the other hand, the mean FLW of all lines with *QFLW-6A* was wider than that of Lumai 14 in two environments.

Population	Trait ¹	QTL ²	Environment ³	Marker	LOD ⁴	Add ⁵	PVE(%) ⁶
Lumai 14 / Jing 411	FLL	QFLL-4B	E1	Xwmc47	3.90	0.33	5.12
		QFLL-2A	E2	Xwmc667	3.37	0.41	6.38
	FLW	QFLW-3B	E1	Xwmc754	5.18	0.03	8.43
		QFLW-3D	E2	Xgwm314	2.85	-0.03	3.53
		QFLW-6A	E2	Xwmc201	4.52	0.03	6.96
	FLA	QFLA-3B	E1	Xwmc754	3.00	0.47	3.66
		QFLA-4B	E1	Xwmc47	2.55	0.43	3.16
		QFLA-2A	E2	Xwmc667	3.77	0.57	5.79
		QFLA-6A	E2	Xwmc201	3.03	0.58	5.28
	CC _{S1}	QCC _{S1} -3D	E2	Xgwm161	2.83	-0.68	3.02
	CC _{S2}	QCC _{S2} -1A	E1	Xwmc24	4.31	-2.70	4.86
		QCC _{S2} -3A	E1	Xwmc11	3.66	-1.55	4.09
			E2	Xwmc11	3.01	-0.94	3.96
		QCC _{S2} -3B	E2	Xwmc754	2.78	1.04	3.63
	CC _{S3}	QCC _{S3} -3B	E2	Xwmc754	3.91	3.54	4.10
Lumai 14 / Shaanhan 8675	FLL	QFLL-4A	E1	Xwmc757	7.47	-0.67	9.29
			E2	Xwmc757	3.25	-0.37	3.70
		QFLL-6A	E1	Xwmc201	7.95	0.54	10.08
			E2	Xwmc201	4.92	0.36	5.84
		QFLL-1A	E2	Xbarc148	3.32	-0.40	3.79
		QFLL-5D	E2	Xcfd189	3.64	0.35	4.32
	FLW	QFLW-1A	E1	Xwmc312	2.92	0.03	4.81
		QFLW-6A	E1	Xwmc201	5.39	0.03	9.28
			E2	Xwmc201	4.83	0.03	7.41
		QFLW-5D	E2	Xcfd189	3.58	0.03	5.34
	FLA	QFLA-4A	E1	Xwmc757	4.08	-0.84	5.05
		QFLA-6A	E1	Xwmc201	9.73	1.07	13.32
			E2	Xwmc201	5.35	0.55	6.18
		QFLA-1A	E2	Xbarc148	3.04	-0.55	3.32
		QFLA-2D	E2	Xwmc144	2.53	-0.53	2.78
		QFLA-5D	E2	Xcfd189	5.44	0.62	6.34
	CC _{S1}	QCC _{S1} -1B	E2	Xwmc367	3.97	-2.63	5.08
	CC _{S3}	QCC _{S3} -1B	E1	Xwmc134	3.55	2.51	7.38
		QCC _{S3} -1A	E1	Xbarc148	5.76	-4.72	12.19
			E2	Xbarc148	3.32	-3.17	7.46

Table 3. QTLs for flag leaf-related traits detected in two IL wheat populations under two environments.

¹: FLL, flag leaf length; FLW, flag leaf width; FLA, flag leaf area; CC_{S1}, chlorophyll content at flowering stage; CC_{S2}, chlorophyll content at 15 days after anthesis; CC_{S3}, chlorophyll content at 20 days after anthesis

²: QTL, quantitative trait locus

³: E1, 2016–2017, E2, 2017–2018

⁴: LOD, logarithm of the odds

⁵: Add, additive effect, positive and negative values indicate that phenotypic variation are contributed by recipient parent and donor parent, respectively

⁶: PVE, phenotypic variation explained

https://doi.org/10.1371/journal.pone.0229912.t003

Compared with Lumai14, 17 lines (IL 42, 57, 58, 73, 78, 79, 86, 90, 91, 92, 94, 95, 102, 104, 125, 128 and 145) presented significantly wider FLW in E1, and three lines (IL 75, 80 and 126) did in E2. Especially, the FLW of ten lines (IL 3, 22, 24, 26, 59, 60, 74, 93, 127 and 151) was significantly wider than that of recipient parent in both E1 and E2 environments (S1 Table, Fig 2B).



Fig 1. Distribution of QTLs for flag leaf-related traits on the genetic linkage map. The map distance in cM are shown on the left. The QTLs are listed on the right. A and B indicate Lumai 14 / Jing 411 and Lumai 14 / Shaanhan 8675 populations, respectively.

https://doi.org/10.1371/journal.pone.0229912.g001

These results indicated *QFLW-6A* played an important role for increasing FLW. Because the IL 59 and 127 was introgressed five and four donor chromosomal fragments, respectively, on which no other QTL for FLW was detected, the IL 59 and 127 and their recurrent parent can be taken as potential NILs.

A	Chromosome	:			2 <i>P</i>	ł					2D									3B 4D													6A								
	Marker	X : w c 1 6 6 7	X Z g V m 0 6 4 3 (6 7	X X v b n au c c 4 2 0 1 7 2	x g m 3 5 9	X m c 1 7 7	X m c 2 9 6	X m c 6 4 4	X m c 7 9 4	X m c 5 0 3	X g m 2 6 1	X m c 1 1 2	X 2 m v c r 1 1 8	X 2 g V n (1 4 5 1 7	X 2 w g n v c n 4 5 1 3 9	X X y y n n 5 3 4 9 9	X X g c v f n d 3 1 4 6 2 1	X m c 1 6 7	X g m 3 8 2	X g m 4 9 3	X w m c 7 5 4	X g w m 5 6 6	X m c 6 7 5	X m c 7 7 7	X : w : c : 6 : 1 : 2	X 2 b 1 ar a c 0 7 7 3 1	X X ra c 7 1 7 6 4	X w m c 7 8 7	X b ar c 8 4	X m c 6 8 7	X m c 3 2 6	X g m 1 8 1	X w m 3 4 0	X m c 3 1	X m c 6 2 2	X m c 7 4	X g m 6 2 4	X g m 3 4	X : m : c 8 : 7	X X w w n n c c 2 5 0 5 1 3	K v n c b b b b
	Lumai 14	\square																																							
	IL 124	Π																																							
										_		- -											-								_					-	QI	FL	W-	6A	
B	Chromosome	vv	v	2.ª	L V V	v	v	v	3		vl	/ v	v	v	v	v	51 v l v) Iv	v	v	v	vl		lv	v	v	6A vla	Iv	v	v	v	vla	7	lv	v	//	1 v	V	v	zlv	
	Marker	w w m m c c 6 4 6 0 7 7	w m c 5 9 8	mi c 5 2 2	w w m n c c 2 6 9 4 6 4	v w n m c 8 1 9	r r c 5	g w m 3 1 4	g d m 9 9	w m c 5 2 9	w 4 m v c r 4 1	g g v w n m 7 1 1 9 0	c f d 1 8 9	c f d 6 7	c f d s	g m 1 5 8 3	g c w f m d 1 5 8 7 2	g w 2 9 2	m c 7 8 8	w m c 9 7	w m c 3 5 7	w w m n c c 7 4 5 3	v g n v c n 4 3 4 3 3 4		w m c 2 5 6	w m c 8 0 7	b v a n c 2 3 (ww mm c 5 5 3	g w m 1 6 9	w m c 5 8 0	g m 4 2 7	g V N 1 n 0 3 1 5	v b n a c r 1 c 7 1 0 8	m c 5 9 6	w m c 6 7	g w m 2 7 6	g m 2 8 2	g w m 6 3	w w w w w w w w w w w w w w w w w w w	v w n m c c 5 8 2 0 5 9	
	Lumai 14																																								
	IL59								_	_	\downarrow	_				_				_	_							_			_							Ц	-		
	IL127																																								
					-	-	-					-				_	_		_			-	-	-			1	_				_	-	-	-	-			_	-	-



https://doi.org/10.1371/journal.pone.0229912.g002

Fine mapping for QFLW-6A

Through sequence alignment with wheat reference genome sequence (IWGSC RefSeq v1.0) published by International Wheat Genome Sequence Consortium (IWGSC), the physical location of the amplified fragment of SSR marker *Xwmc201* linked with *QFLW-6A* was found to be 574,283,851–574,283,613 bp on chromosome 6A. And 10 genes were found by further analysis of the genetic information in the range of 500 kb upstream and downstream (573,783,851–574,783,851 bp) of the fragment, including genes encoding F-box family protein and protein kinase family protein, etc (http://plants.ensembl.org/index.html) (Table 4).

Discussion

Comparison with previous results

Flag leaf related-traits of wheat belong to quantitative traits, which have complex genetic basis and are greatly influenced by environment. Due to mapping QTL by different mapping population under different environments and using diversity of marker type, the results were difficult to replicate. Twenty-nine QTLs for flag leaf-related traits were identified in this study, only a few were completely consistent with previous results. For FLL, six QTLs were detected in this study. Among them, three QTLs, *QFLL-2A*, *QFLL-4A* and *QFLL-4B*, were associated

1 0	-	
Gene ID	Position (bp)	Genes Description
TraesCS6A02G341000	573,860,671-573,861,709	F-box family protein
TraesCS6A02G341200	574,220,464-574,221,965	F-box/RNI-like/FBD-like domains-containing protein
TraesCS6A02G341300	574,238,362-574,239,045	F-box domain containing protein-like
TraesCS6A02G341400	574,242,017-574,243,759	3-ketoacyl-CoA synthase
TraesCS6A02G341500	574,249,221-574,249,521	alpha-adaptin
TraesCS6A02G341600	574,378,180-574,386,049	GPI-anchored adhesin-like protein
TraesCS6A02G341700	574,472,578-574,479,877	RRP12-like protein
TraesCS6A02G341800	574,481,191-574,488,679	carboxyl-terminal peptidase, putative (DUF239)
TraesCS6A02G341900	574,489,301-574,493,373	Chaperone protein dnaJ
TraesCS6A02G342000	574,524,921-574,530,023	Protein kinase family protein

Table 4. The predicted genes on the location of QFLW-6A.

https://doi.org/10.1371/journal.pone.0229912.t004

with the SSR marker Xwmc667, Xwmc757 and Xwmc47, respectively, which were reported at similar genetic regions [10, 43–45]. And OFLL-4A were detected under two environments in this study. QFLL-6A was also identified under two environments, which was located near the marker Xwmc201 on chromosome 6A. QFLL-5D linked to the SSR marker Xcfd189 in this study accorded with the QTL for FLL reported by Fan et al. [10], which was located the flanking interval Xcfd189-Xgwm174. For FLW, six QTLs were identified in this study. Among them, two QTLs, QFLW-3B and QFLW-5D, linked to the SSR marker Xwmc754 and Xcfd189 were previously found on the chromosomes 3B and 5D [7, 10]. QFLW-6A was consistently mapped on chromosome 6A in two IL populations, with contributing 6.96-9.28% to the phenotypic variance in different environments, and the positive allele of the QTL was originated from donors. Fan et al. [10] also found a QTL controlling FLW at the similar genetic region on this chromosome. It indicated that QFLW-6A was of importance to affect the FLW. For FLA, nine QTLs were detected in this study. Of them, QFLA-1A, QFLA-2D, QFLA-4A and QFLA-6A were distributed on chromosomes 1A, 2D, 4A and 6A, which have been proved on these chromosomes [7, 10]. It was worth mentioning that QFLA-6A was simultaneously in Lumai 14 / Jing 411 population under E2 and in Lumai 14 / Shaanhan 8675 population under E1 and E2 environments, with explaining 5.28%, 13.32% and 6.18% of the phenotypic variance, respectively. It was likely a stable QTL controlling FLA. Throughout all QTLs detected in the present, QFLW-6A and QFLA-6A were identified in both IL populations, and the positive alleles of individual QTL were also derived from donors, suggesting that they could be stable QTLs. And the two loci were linked with Xwmc201 on chromosome 6A at the same time, which indicated that they may be pleiotropic or tightly linked QTL responsible for both traits. We detected eight QTLs for CC in this study. Of these QTLs, QCC_{S3}-1A associated with Xbarc148 and QCC₅₂-3A associated with Xwmc24 were repeatedly detected under two environments. QCC_{S1}-1B, QCC_{S2}-3A and QCC_{S2}-3B were identified at similar regions on chromosomes 1B, 3A and 3B by Shi et al. [34] and Zhang et al. [16].

Relationship between flag leaf-related traits and yield-related traits in wheat

The flag leaf, as the main organ for photosynthesis during the reproductive period, is responsible for the regulating final plant growth and yield formation in cereal crops [2, 46]. So, reasonablely increasing flag leaf size and decreasing the rate of chlorophyll degradation during grain-filling period can improve the photosynthetic ability and promote to increase photosynthetic products, and finally achieve to enhance yield. Numerous studies have shown that chlorophyll content and morphological traits of the flag leave were corelated with yield-related traits in

phenotype in cereals [7, 8, 24, 43, 47]. And QTLs controlling related traits were not uniformly distributed on chromosomes, but tended to be distributed in the same or adjacent regions of the same chromosome [48-50]. In barely, a QTL for FLL and a QTL for spike length were simultaneously associated with gene HvFT2. Feltus et al. [51] reported a QTL cluster for FLW and thousand-kernel weight in chromosome 3S in sorghum. Yue et al. [52] found that the QTL controlling flag leaf-related traits and QTL for yield-related traits were distributed on the same genic regions in rice. Two QTL clusters, chlorophyll content and yield and chlorophyll content, yield, heading date and flowering date, were identified on the flanking interval Xwmc718-Xwmc262 of 4B chromosome and Xbarc320-Xwmc215-Xgdm63 of 5D chromosome, respectively, using DH population in wheat [15]. In addition, QTL clusters controlling flag leaf-related traits and yield-related traits were also detected on chromosomes 1A, 1B, 2D, 4A, 4D, 5A, 5B, 6B, 6D, 7B and 7D of wheat [7-10, 53]. We found some regions that not only controlling flag leaf-related traits but also yield-related traits in the same population. For example, FLW-3B, FLA-3B, QCC₅₂-3B and QCC₅₃-3B were identified near the SSR marker Xwmc754 on chromosome 3B in Lumai 14 / Jing 411 population, where the QTgw-3B for thousand-kernel weight was also mapped using the same population [41]. And the favorable alleles of these QTLs were from Jing 411 increasing these traits. Besides, a QTL for CC, QCC_{s2} -1A, detected in this study and QPh-1A for plant height and QGwp-1A for grain weight per plant previously detected were associated with the marker Xwmc24 [41], with the alleles from Lumai 14 increasing CC₅₂ and plant height, however, decreasing grain weight per plant. Another QTL for CC, QCC_{S2}-3A, was shared the same marker Xwmc11 on chromosome 3A with a QTL controlling thousand-kernel weight previously identified [41], with the Lumai 14-derived alleles increasing CC_{S2}, but decreasing thousand-kernel weight. In Lumai 14 / Shaanhan 8675 population, QFLL-6A, QFLW-6A and QFLA-6A were located to the SSR marker Xwmc201, which has been proved to linked to QTLs for kernel morphology-related traits by Chen et al. [36]. It was worth noting that the positive alleles of these QTLs were originated from Shaanhan 8675. So this locus could be pleiotropism or closely linked OTL, which not only affected flag leaf size but also affected kernel size. As we all know, the FLA was closely related with photosynthetic production, the larger the leaf area, the more photosynthetic products accumulated, the larger the grains. Therefore, it is of great significance to further study this locus for high-yielding selection in wheat breeding. The flag leaf-related traits are one of the key factors affecting plant structure and yield, therefore, we can detect stable QTLs and develop reliable molecular markers through the further study of "active regions" with the same effect that is responsible for multiple elite traits, which not only can promote genetic improvement of plant stature and yield in cereal crops, but also may be transfer an excellent gene controlling multiple traits into a plant at a time to improve the efficiency of breeding.

Breeding of near-isogenic line with QFLW-6A

ILs are constructed through introgressing chromosomal fragments from a donor parent into a recipient parent after multiple backcross and self-cross, the genotypes of all progenies in the population are very similar to those of the recurrent parent. Phenotypic differences between lines and the recipient parent can generally be attributed to substituted fragments from donor [36, 41]. Backcross introgression has been the most commonly used method for developing NILs for QTL studies [54–58]. In the present study, the target QTLs were tracked with SSR markers. It was found that four lines in Lumai 14 / Jing 411 population and 11 lines in Lumai 14 / Shaanhan 8675 population were introgressed no more than five fragments from donor, and one of these fragments carried *QFLW-6A*, with the alleles from donors increasing FLW. In terms of FLW, only IL 124 in Lumai 14 / Jing 411 population and IL 59 and 127 showed

significant difference from recipient Lumai 14 in both E1 and E2 environment. And they were significantly wider FLW than Lumai 14. While, the FLW of the remaining lines was no significant difference or significant only in one environment compared with Lumai 14, which may be caused by interaction effect between *QFLW-6A* with other introgressed fragments. In addition, the three lines were contained no other QTL besides *QFLW-6A*. Therefore, they can be regarded as potential NILs and *QFLW-6A* can be used for further fine mapping of FLW by crossing and backcrossing with recipient Lumai 14.

Prediction of candidate gene for QFLW-6A

A synteny analysis found *QFLW-6A* was near the 574,283,851–574,283,613 bp fragment. And 10 genes were found in the range of 500 kb upstream and downstream of the fragment (Table 4 and Fig 1). These genes involved in life activities such as cell cycle regulation, cell apoptosis, signal transduction, growth and development, as well as biochemical processes such as resistance to stresses [59–62], and so on. At present, we are not sure that which one is the candidate gene of this QTL. It needs to be further analyzed and proved by biotechnology.

Conclusion

A total of six, six, nine and eight QTLs for FLL, FLW, FLA and CC were detected in two IL populations, respectively. Of them, QFLW-6A and QFLA-6A were detected under E2 in Lumai 14 / Jing 411 population and under both E1 and E2 in Lumai 14 / Shaanhan 8675 population. QCC_{S3} -1A from Lumai 14 / Jing 411 population and QCC_{S2} -3A, QFLL-4A and QFLL-6A from Lumai 14 / Shaanhan 8675 population were repeatedly identified under two environments. Besides, eight QTL clusters for flag leaf size and CC were identified in the two IL populations. On the other hand, three potential near-isogenic lines carried no other QTL controlling FLW than QFLW-6A were found. QFLW-6A had an important role for increasing the FLW, with the favorable allele stemmed from donors in both IL populations. Further, QFLW-6A was found to be near the 574,283,851–574,283,613 bp fragment on chromosome 6A. These results can lay a foundation for identification of candidate gene and map-based cloning and functional verification of the QTL.

Supporting information

S1 Table. Flag leaf width characteristics in wheat lines carrying introgressed donor chromosomal fragments at the *QFLW-6A* locus in two IL population. (XLSX)

Author Contributions

Conceptualization: Xue Yan, Wenjun Zhang, Daizhen Sun, Ruilian Jing.

Data curation: Xue Yan.

Formal analysis: Xue Yan.

Funding acquisition: Shuguang Wang, Daizhen Sun.

Investigation: Bin Yang, Yaping Cao, Yugang Shi.

Methodology: Bin Yang, Yugang Shi.

Resources: Ruilian Jing.

Supervision: Shuguang Wang, Daizhen Sun.

Writing - original draft: Xue Yan.

Writing - review & editing: Xue Yan, Wenjun Zhang, Daizhen Sun, Ruilian Jing.

References

- Cui KH, Peng SB, Xing YZ, Yu SB, Xu CG, Zhang Q. Molecular dissection of the genetic relationships of source, sink and transport tissue with yield traits in rice. Theor Appl Genet. 2003; 106(4):649–58. https://doi.org/10.1007/s00122-002-1113-z PMID: 12595994
- 2. Biswal AK, Kohli A. Cereal flag leaf adaptations for grain yield under drought: Knowledge status and gaps. Mol Breed. 2013; 31(4):749–66.
- Araus JL, Tapia L. Photosynthetic gas exchange characteristics of wheat flag leaf blades and sheaths during grain filling: The case of a spring crop grown under mediterranean climate conditions. Plant Physiol. 1987; 85(3):667–73. https://doi.org/10.1104/pp.85.3.667 PMID: 16665757
- Sharma SN, Sain RS, Sharma RK. The genetic control of flag leaf length in normal and late sown durum wheat. J Agric Sci. 2003; 141(3–4):323–31.
- 5. Duwayri M. Effect of flag leaf and awn removal on grain yield and yield components of wheat grown under dryland conditions. Field Crops Res. 1984; 8(4):307–13.
- Khaliq I, Irshad A, Ahsan M. Awns and flag leaf contribution towards grain yield in spring wheat (*Triticum aestivum* L.). Cereal Res Commun. 2008; 36(1):65–76.
- 7. Zhao C, Bao Y, Wang X, Yu H, Ding A, Guan C, et al. QTL for flag leaf size and their influence on yieldrelated traits in wheat. Euphytica. 2018; 214(11):209.
- Liu K, Xu H, Liu G, Guan P, Zhou X, Peng H, et al. QTL mapping of flag leaf-related traits in wheat (*Triticum aestivum* L.). Theor Appl Genet. 2018; 131(4):839–49. https://doi.org/10.1007/s00122-017-3040-z PMID: 29359263
- Wu Q, Chen Y, Lin F, Zhou S, Chen J, Zhao X, et al. QTL mapping of flag leaf traits in common wheat using an integrated high-density SSR and SNP genetic linkage map. Euphytica. 2016; 208(2):337–51.
- 10. Fan X, Cui F, Zhao C, Zhang W, Yang L, Zhao X, et al. QTLs for flag leaf size and their influence on yield- related traits in wheat (*Triticum aestivum* L.). Mol Breed. 2015; 35(1):24.
- Zhang CJ, Chen GX, Gao XX, Chu CJ. Photosynthetic decline in flag leaves of two field-grown spring wheat cultivars with different senescence properties. S Afr J Bot. 2006; 72(1):15–23.
- 12. Blake NK, Lanning SP, Martin JM, Sherman JD, Talbert LE. Relationship of flag leaf characteristics to economically important traits in two spring wheat crosses. Crop Sci. 2007; 47(2):491–6.
- Chen J, Liang Y, Hu X, Wang X, Tan F, Zhang H, et al. Physiological characterization of 'stay green' wheat cultivars during the grain filling stage under field growing conditions. Acta Physiol Plant. 2010; 32 (5):875–82.
- Sakamoto T, Morinaka Y, Ohnishi T, Sunohara H, Fujioka S, Ueguchi-Tanaka M, et al. Erect leaves caused by brassinosteroid deficiency increase biomass production and grain yield in rice. Nat Biotechnol. 2006; 24(1):105–9. https://doi.org/10.1038/nbt1173 PMID: 16369540
- 15. Zhang K, Zhang Y, Chen G, Tian J. Genetic analysis of grain yield and leaf chlorophyll content in common wheat. Cereal Res Commun. 2009; 37(4):499–511.
- Zhang KP, Fang ZJ, Liang Y, Tian JC. Genetic dissection of chlorophyll content at different growth stages in common wheat. J Genet. 2009; 88(2):183–9. https://doi.org/10.1007/s12041-009-0026-x PMID: 19700856
- Mei HW, Luo LJ, Ying CS, Wang YP, Yu XQ, Guo LB, et al. Gene actions of QTLs affecting several agronomic traits resolved in a recombinant inbred rice population and two testcross populations. Theor Appl Genet. 2003; 107(1):89–101. https://doi.org/10.1007/s00122-003-1192-5 PMID: 12721635
- Farooq M, Tagle AG, Santos RE, Ebron LA, Fujita D, Kobayashi N. Quantitative trait loci mapping for leaf length and leaf width in rice cv. IR64 derived lines. J Integr Plant Biol. 2010; 52(6):578–84. https://doi.org/10.1111/j.1744-7909.2010.00955.x PMID: 20590988
- Wang P, Zhou G, Cui K, Li Z, Yu S. Clustered QTL for source leaf size and yield traits in rice (*Oryza sativa* L.). Mol Breed. 2012; 29(1):99–113.
- Gyenis L, Yun SJ, Smith KP, Steffenson BJ, Bossolini E, Sanguineti MC, et al. Genetic architecture of quantitative trait loci associated with morphological and agronomic trait differences in a wild by cultivated barley cross. Genome. 2007; 50(8):714–23. https://doi.org/10.1139/g07-054 PMID: 17893731
- Liu L, Sun G, Ren X, Li C, Sun D. Identification of QTL underlying physiological and morphological traits of flag leaf in barley. BMC Genet. 2015; 16(3):29.

- Harris K, Subudhi P, Jordan D, Borrell A, Rosenow D, Nguyen H, et al. Sorghum stay-green QTL individually reduce post-flowering drought-induced leaf senescence. J Exp Bot. 2007; 58(2):327–38. https://doi.org/10.1093/jxb/erl225 PMID: 17175550
- Isidro J, Knox R, Clarke F, Singh A, DePauw R, Clarke J, et al. Quantitative genetic analysis and mapping of leaf angle in durum wheat. Planta. 2012; 236(6):1713–23. <u>https://doi.org/10.1007/s00425-012-1728-5 PMID: 22868576</u>
- Wang P, Zhou G, Yu H, Yu S. Fine mapping a major QTL for flag leaf size and yield-related traits in rice. Theor Appl Genet. 2011; 123(8):1319–30. <u>https://doi.org/10.1007/s00122-011-1669-6</u> PMID: 21830109
- Chen M, Luo J, Shao G, Wei X, Tang S, Sheng Z, et al. Fine mapping of a major QTL for flag leaf width in rice, *qFLW4*, which might be caused by alternative splicing of *NAL1*. Plant Cell Rep. 2012; 31 (5):863–72. https://doi.org/10.1007/s00299-011-1207-7 PMID: 22179305
- Ding X, Li X, Xiong L. Evaluation of near-isogenic lines for drought resistance QTL and fine mapping of a locus affecting flag leaf width, spikelet number, and root volume in rice. Theor Appl Genet. 2011; 123 (5):815–26. https://doi.org/10.1007/s00122-011-1629-1 PMID: 21681490
- Qi J, Qian Q, Bu Q, Li S, Chen Q, Sun J, et al. Mutation of the rice Narrow leaf1 gene, which encodes a novel protein, affects vein patterning and polar auxin transport. Plant Physiol. 2008; 147(4):1947–59. https://doi.org/10.1104/pp.108.118778 PMID: 18562767
- Fujino K, Matsuda Y, Ozawa K, Nishimura T, Koshiba T, Fraaije MW, et al. NARROW LEAF 7 controls leaf shape mediated by auxin in rice. Mol Genet Genomics. 2008; 279(5):499–507. https://doi.org/10. 1007/s00438-008-0328-3 PMID: 18293011
- Vijayalakshmi K, Fritz AK, Paulsen GM, Bai G, Pandravada S, Gill BS. Modeling and mapping QTL for senescence-related traits in winter wheat under high temperature. Mol Breed. 2010; 26(2):163–75.
- Yang DL, Liu Y, Cheng HB, Chang L, Chen JJ, Chai SX, et al. Genetic dissection of flag leaf morphology in wheat (*Triticum aestivum* L.) under diverse water regimes. BMC Genet. 2016; 17(1):94. <u>https://doi.org/10.1186/s12863-016-0399-9</u> PMID: 27352616
- Hussain W, Baenziger PS, Belamkar V, Guttieri MJ, Venegas JP, Easterly A, et al. Genotyping-bysequencing derived high-density linkage map and its application to QTL mapping of flag leaf traits in bread wheat. Sci Rep. 2017; 7(1):16394. https://doi.org/10.1038/s41598-017-16006-z PMID: 29180623
- 32. Barakat MN, Saleh M, Al-Doss AA, Moustafa KA, Elshafei AA, Al-Qurainy FH. Identification of new SSR markers linked to leaf chlorophyll content, flag leaf senescence and cell membrane stability traits in wheat under water stressed condition. Acta Biol Hung. 2015; 66(1):93–102. <u>https://doi.org/10.1556/ABiol.66.2015.1.8 PMID: 25740441</u>
- Yang B, Yan X, Wang H, Li X, Ma H, Wang S, et al. Dynamic QTL analysis of chlorophyll content during grain filling stage in winter wheat (*Triticum aestivum* L.). Rom Agric Res. 2016; 33:77–85.
- Shi S, Azam FI, Li H, Chang X, Li B, Jing R. Mapping QTL for stay-green and agronomic traits in wheat under diverse water regimes. Euphytica. 2017; 213(11):246.
- Xue S, Xu F, Li G, Zhou Y, Lin M, Gao Z, et al. Fine mapping *TaFLW1*, a major QTL controlling flag leaf width in bread wheat (*Triticum aestivum* L.). Theor Appl Genet. 2013; 126(8):1941–9. <u>https://doi.org/10.1007/s00122-013-2108-7</u> PMID: 23661078
- **36.** Chen W, Sun D, Yan X, Li R, Wang S, Shi Y, et al. QTL analysis of wheat kernel traits, and genetic effects of *qKW-6A* on kernel width. Euphytica. 2019; 215(2):11.
- Hao ZF, Chang XP, Guo XJ, Lian JR, Li RZ, Jia JZ. QTL mapping for drought tolerance at stages of germination and seedling in wheat (*Triticum aestivum* L.) using a DH population. Agricultural Sciences in China. 2003; 2(9):943–9.
- Wu X, Chang X, Jing R. Genetic insight into yield-associated traits of wheat grown in multiple rain-fed environments. PLoS One. 2012; 7(2):e31249. <u>https://doi.org/10.1371/journal.pone.0031249</u> PMID: 22363596
- Xu YF, Wang RF, Tong YP, Zhao HT, Xie QG, Liu DC, et al. Mapping QTLs for yield and nitrogenrelated traits in wheat: influence of nitrogen and phosphorus fertilization on QTL expression. Theor Appl Genet. 2014; 127(1):59–72. https://doi.org/10.1007/s00122-013-2201-y PMID: 24072207
- Edae EA, Byrne PF, Manmathan H, Haley SD, Moragues M, Lopes MS, et al. Association mapping and nucleotide sequence variation in five drought tolerance candidate genes in spring wheat. Plant Genome. 2013; 6(2):547–62.
- Yan X, Sun D, Li R, Wang S, Ma G, Cao Y, et al. Mapping quantitative trait loci for important agronomic traits and developing potential near-isogenic lines in wheat. Int J Agric Biol. 2019; 22(1):20–8.
- Liu G, Jia L, Lu L, Qin D, Zhang J, Guan P, et al. Mapping QTLs of yield-related traits using RIL population derived from common wheat and Tibetan semi-wild wheat. Theor Appl Genet. 2014; 127(11):2415– 32. https://doi.org/10.1007/s00122-014-2387-7 PMID: 25208643

- 43. Jia H, Wan H, Yang S, Zhang Z, Kong Z, Xue S, et al. Genetic dissection of yield-related traits in a recombinant inbred line population created using a key breeding parent in China's wheat breeding. Theor Appl Genet. 2013; 126(8):2123–39. https://doi.org/10.1007/s00122-013-2123-8 PMID: 23689745
- 44. Lu L, Yang B, Zhang T, Zhang W, Yuan K, Shi X, et al. Quantitative trait loci analysis of flag leaf size and grain relative traits in winter wheat. Acta Agriculturae Boreali-sinica. 2018; 33(5):1–8.
- Yan X, Shi Y, Liang Z, Yang B, Li X, Wang S, et al. QTL mapping for morphological traits of flag leaf in wheat. Journal of Nuclear Agricultural Sciences. 2015; 29(7):1253–9.
- **46.** Tian Y, Zhang H, Xu P, Chen X, Liao Y, Han B, et al. Genetic mapping of a QTL controlling leaf width and grain number in rice. Euphytica. 2015; 202(1):1–11.
- Mason RE, Mondal S, Beecher FW, Hays DB. Genetic loci linking improved heat tolerance in wheat (*Tri-ticum aestivum* L.) to lower leaf and spike temperatures under controlled conditions. Euphytica. 2011; 180(2):181–94.
- 48. Quarrie SA, Steed A, Calestani C, Semikhodskii A, Lebreton C, Chinoy C, et al. A high-density genetic map of hexaploid wheat (*Triticum aestivum* L.) from the cross Chinese Spring × SQ1 and its use to compare QTLs for grain yield across a range of environments. Theor Appl Genet. 2005; 110(5):865–80. https://doi.org/10.1007/s00122-004-1902-7 PMID: 15719212
- 49. Ma ZQ, Zhao DM, Zhang CQ, Zhang ZZ, Xue SL, Lin F, et al. Molecular genetic analysis of five spikerelated traits in wheat using RIL and immortalized F₂ populations. Mol Genet Genomics. 2007; 277 (1):31–42. https://doi.org/10.1007/s00438-006-0166-0 PMID: 17033810
- 50. Liu Y, Tao Y, Wang Z, Guo Q, Wu F, Yang X, et al. Identification of QTL for flag leaf length in common wheat and their pleiotropic effects. Mol Breed. 2018; 38(1):11.
- Feltus FA, Hart GE, Schertz KF, Casa AM, Kresovich S, Abraham S, et al. Alignment of genetic maps and QTLs between inter- and intra-specific sorghum populations. Theor Appl Genet. 2006; 112 (7):1295–305. https://doi.org/10.1007/s00122-006-0232-3 PMID: 16491426
- 52. Yue B, Xue WY, Luo LJ, Xing YZ. QTL analysis for flag leaf characteristics and their relationships with yield and yield traits in rice. Acta genet Sin. 2006; 33(9):824–32. <u>https://doi.org/10.1016/S0379-4172</u> (06)60116-9 PMID: 16980129
- Edae EA, Byrne PF, Haley SD, Lopes MS, Reynolds MP. Genome-wide association mapping of yield and yield components of spring wheat under contrasting moisture regimes. Theor Appl Genet. 2014; 127(4):791–807. https://doi.org/10.1007/s00122-013-2257-8 PMID: 24408378
- Dorweiler J, Stec A, Kermicle J, Doebley J. Teosinte glume architecture 1: A genetic locus controlling a key step in maize evolution. Science. 1993; 262(5131):233–5. <u>https://doi.org/10.1126/science.262</u>. 5131.233 PMID: 17841871
- 55. Mia MS, Liu H, Wang X, Yan G. Multiple near-isogenic lines targeting a QTL hotspot of drought tolerance showed contrasting performance under post-anthesis water stress. Front Plant Sci. 2019; 10:271. https://doi.org/10.3389/fpls.2019.00271 PMID: 30906308
- 56. Zhai H, Feng Z, Du X, Song Y, Liu X, Qi Z, et al. A novel allele of *TaGW2-A1* is located in a finely mapped QTL that increases grain weight but decreases grain number in wheat (*Triticum aestivum* L.). Theor Appl Genet. 2018; 131(3):539–53. https://doi.org/10.1007/s00122-017-3017-y PMID: 29150697
- 57. Farre A, Sayers L, Leverington-Waite M, Goram R, Orford S, Wingen L, et al. Application of a library of near isogenic lines to understand context dependent expression of QTL for grain yield and adaptive traits in bread wheat. BMC Plant Biol. 2016; 16(1):161. <u>https://doi.org/10.1186/s12870-016-0849-6</u> PMID: 27436187
- Guan P, Di N, Mu Q, Shen X, Wang Y, Wang X, et al. Use of near-isogenic lines to precisely map and validate a major QTL for grain weight on chromosome 4AL in bread wheat (*Triticum aestivum* L.). Theor Appl Genet. 2019.
- Somers DE, Fujiwara S. Thinking outside the F-box: novel ligands for novel receptors. Trends Plant Sci. 2009; 14(4):206–13. https://doi.org/10.1016/j.tplants.2009.01.003 PMID: 19285909
- Zhang Y, Xu W, Li Z, Deng XW, Wu W, Xue Y. F-box protein DOR functions as a novel inhibitory factor for abscisic acid-induced stomatal closure under drought stress in Arabidopsis. Plant Physiol. 2008; 148(4):2121–33. https://doi.org/10.1104/pp.108.126912 PMID: 18835996
- Yoshida R, Hobo T, Ichimura K, Mizoguchi T, Takahashi F, Aronso J, et al. ABA-activated SnRK2 protein kinase is required for dehydration stress signaling in Arabidopsis. Plant Cell Physiol. 2002; 43 (12):1473–83. https://doi.org/10.1093/pcp/pcf188 PMID: 12514244
- 62. Shao Y, Zhang X, van Nocker S, Gong X, Ma F. Overexpression of a protein kinase gene MpSnRK2.10 from Malus prunifolia confers tolerance to drought stress in transgenic Arabidopsis thaliana and apple. Gene. 2019; 692:26–34. https://doi.org/10.1016/j.gene.2018.12.070 PMID: 30641216