GENOME SEQUENCES





Draft Genome Sequence of *Stenotrophomonas maltophilia* CRB139-1, Isolated from Poultry Meat in Japan

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ABSTRACT Stenotrophomonas maltophilia is a nosocomial pathogen that primarily causes respiratory infection in humans. This pathogen is widely distributed in the environment, including in foods. Here, we report the draft genome sequence of *S. maltophilia* strain CRB139-1, isolated from poultry meat in Japan. The genome size was 4,619,918 bp at 90× coverage.

S tenotrophomonas maltophilia is a nosocomial pathogen that causes respiratory infection in humans (1, 2). This bacterium resides in animal hosts and in a number of different environments and food specimens (2–6). Most available genomic data were obtained from human or environmental specimens, and there is limited information available on the strains found in food; there are only two partial sequences (GenBank accession numbers KU978825 and MH450104) for the genomic data of strains from poultry meat. To address the lack of genomic data, we obtained *S. maltophilia* strain CRB139-1, a single colony isolated from violet-red bile glucose agar containing 2 $\mu g/\mu l$ meropenem, from poultry meat in Hiroshima, Japan, in 2015. This study obtained the genome sequence of CRB139-1 to explore the strain's genetic background.

Genomic DNA was extracted from a bacterial culture grown in lysogeny broth agar (Becton, Dickinson) using the Maxwell RSC blood DNA kit (Promega), and the library was prepared using the Ion Xpress Plus fragment library kit (Life Technologies). Genome sequencing was performed using the Ion Torrent Chef/GeneStudio S5 system (Life Technologies). The reads were trimmed using Qiagen CLC Genomics Workbench v.11.0 and assembled *de novo* to the contig and scaffold levels using CGE Assembler v.1.2 and CONTIGuator v.2 (7, 8) with the reference genome sequence of strain NCTC13014, as this was shown to be the closest relative of CRB139-1 (Fig. 1). The draft genome was annotated using the DDBJ Fast Annotation and Submission Tool v.1.1.4 (9). Default parameters were used for software unless otherwise specified. Identities to other strains were determined using the NCBI BLAST tool.

Sequence data comprised 461,320,092 bp from 1,895,353 reads and were assembled into 489 contigs, with an N_{50} value of 17,160 bp. The genome sequence was embedded in one scaffold with 0.854% gaps and an accumulated length of 4,619,918 bp at 90× coverage. The assembly had a G+C content of 66.3% and an N_{50} value of 4,619,918 bp. Scaffold sequences were annotated to contain 4,215 coding sequences.

S. maltophilia is known to be intrinsically resistant to carbapenems via β -lactamase production or multidrug efflux pumps (10). Strain CRB139-1 harbored the bla_{L2} gene, which is associated with decreased production of β -lactamase (11); this gene was 100% identical to that of strain FDAARGOS_507. Mutations in the bla_{L2} gene or AmpR regulator result in overproduction of β -lactamase (12, 13). AmpR, which activates the transcription of bla_{L2} (14, 15), showed four amino acid substitutions compared to that of strain KH (99.3% similarity). The start codons and promoters of these genes and a putative AmpR binding region (15) were identical to those of KH, whereas the similarity with ampR-bla_{L2} intergenic sequences was 91.6%. Previous studies indicated that

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Tree scale: 0.01 H ICU331 (accession No. CP040440)	
K279a (accession No. AM743169)	
Sm454 (accession No. CP040431)	
FDAARGOS_325 (accession No. CP022053)	
FDAARGOS_649 (accession No. CP044092)	
SVIA2 (accession No. CP033586)	
NCTC10498 (accession No. LS483406)	
13637 (accession No. CP008838)	
NCTC10257 (accession No. LT906480)	
NCTC10258 (accession No. LS483377)	
Sm53 (accession No. CP040430)	
FDAARGOS_92 (accession No. CP014014)	
FDAARGOS_507 (accession No. CP033829)	NOTOLOGIA
CRB139-1 (accession No. BKBG02000001)	NCTC13014 (accession No. LR134301)

FIG 1 Dendrogram of the genetic phylogenetic tree obtained from the whole-genome sequences of 15 representative *S. maltophilia* strains, including isolate CRB139-1. The complete genome sequences of 15 *S. maltophilia* reference strains were obtained from DDBJ/EMBL/GenBank. The phylogenetic tree was constructed using the concatenated alignment of the single nucleotide polymorphisms (SNPs) in whole-genome sequencing reads in CSI phylogeny v.1.4 with default parameters and was displayed by Interactive Tree of Life (iTOL).

*ampR-bla*_{L2} sequence variation is involved in L2 β -lactamase expression (16–18). Thus, our data suggest that a certain mutation(s) in these regions might be associated with increased meropenem resistance.

This study provides genome sequence data of *S. maltophilia* isolated from poultry meat. Further comparative genomic analysis would elucidate the molecular mechanisms underlying the growth of this bacterium in poultry meat.

Data availability. These sequences were deposited in DDBJ/ENA/GenBank under accession number BKBG02000001 and the raw reads under accession number DRA008814, with BioProject number PRJDB8492.

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REFERENCES

- Palleroni NJ, Bradbury JF. 1993. Stenotrophomonas, a new bacterial genus for Xanthomonas maltophilia (Hugh 1980) Swings et al. 1983. Int J Syst Bacteriol 43:606–609. https://doi.org/10.1099/00207713-43-3-606.
- Brooke JS. 2012. Stenotrophomonas maltophilia: an emerging global opportunistic pathogen. Clin Microbiol Rev 25:2–41. https://doi.org/10 .1128/CMR.00019-11.
- Elisha IL, Jambalang AR, Botha FS, Buys EM, McGaw LJ, Eloff JN. 2017. Potency and selectivity indices of acetone leaf extracts of nine selected South African trees against six opportunistic Enterobacteriaceae isolates from commercial chicken eggs. BMC Complement Altern Med 17:90. https://doi.org/10.1186/s12906-017-1597-3.
- Nakayama T, Ha NC, Quoc Le P, Kawahara R, Kumeda Y, Sumimura Y, Yamamoto Y. 2017. Consumption of edible ice contaminated with *Acinetobacter*, *Pseudomonas*, and *Stenotrophomonas* is a risk factor for fecal colonization with extended-spectrum β-lactamase-producing *Escherichia coli* in Vietnam. J Water Health 15:813–822. https://doi.org/10 .2166/wh.2017.054.
- Qureshi A, Mooney L, Denton M, Kerr KG. 2005. Stenotrophomonas maltophilia in salad. Emerg Infect Dis 11:1157–1158. https://doi.org/10.3201/ eid1107.040130.
- Furushita M, Okamoto A, Maeda T, Ohta M, Shiba T. 2005. Isolation of multidrug-resistant *Stenotrophomonas maltophilia* from cultured yellowtail (*Seriola quinqueradiata*) from a marine fish farm. Appl Environ Microbiol 71:5598–5600. https://doi.org/10.1128/AEM.71.9.5598-5600.2005.
- Larsen MV, Cosentino S, Rasmussen S, Friis C, Hasman H, Marvig RL, Jelsbak L, Sicheritz-Pontén T, Ussery DW, Aarestrup FM, Lund O. 2012.

Multilocus sequence typing of total-genome-sequenced bacteria. J Clin Microbiol 50:1355–1361. https://doi.org/10.1128/JCM.06094-11.

- Galardini M, Biondi EG, Bazzicalupo M, Mengoni A. 2011. CONTIGuator: a bacterial genomes finishing tool for structural insights on draft genomes. Source Code Biol Med 6:11. https://doi.org/10.1186/1751-0473-6-11.
- Tanizawa Y, Fujisawa T, Kaminuma E, Nakamura Y, Arita M. 2016. DFAST and DAGA: Web-based integrated genome annotation tools and resources. Biosci Microbiota Food Health 35:173–184. https://doi.org/10 .12938/bmfh.16-003.
- Crossman LC, Gould VC, Dow JM, Vernikos GS, Okazaki A, Sebaihia M, Saunders D, Arrowsmith C, Carver T, Peters N, Adlem E, Kerhornou A, Lord A, Murphy L, Seeger K, Squares R, Rutter S, Quail MA, Rajandream MA, Harris D, Churcher C, Bentley SD, Parkhill J, Thomson NR, Avison MB. 2008. The complete genome, comparative and functional analysis of *Stenotrophomonas maltophilia* reveals an organism heavily shielded by drug resistance determinants. Genome Biol 9:R74. https://doi.org/10 .1186/gb-2008-9-4-r74.
- 11. Liu W, Zou D, Wang X, Li X, Zhu L, Yin Z, Yang Z, Wei X, Han L, Wang Y, Shao C, Wang S, He X, Liu D, Liu F, Wang J, Huang L, Yuan J. 2012. Proteomic analysis of clinical isolate of *Stenotrophomonas maltophilia* with *bla*_{NDM-1}, *bla*_{L1} and *bla*_{L2} β-lactamase genes under imipenem treatment. J Proteome Res 11:4024 4033. https://doi.org/10.1021/pr300062v.
- Zhang L, Li XZ, Poole K. 2001. SmeDEF multidrug efflux pump contributes to intrinsic multidrug resistance in *Stenotrophomonas maltophilia*. Antimicrob Agents Chemother 45:3497–3503. https://doi.org/10.1128/ AAC.45.12.3497-3503.2001.

- Okazaki A, Avison MB. 2008. Induction of L1 and L2 β-lactamase production in *Stenotrophomonas maltophilia* is dependent on an AmpRtype regulator. Antimicrob Agents Chemother 52:1525–1528. https://doi .org/10.1128/AAC.01485-07.
- Song S, Yuan X, Liu S, Zhang N, Wang Y, Ke Y, Xu J, Huang L, Chen Z, Li Y. 2012. Genome sequence of *Stenotrophomonas maltophilia* S028, an isolate harboring the AmpR-L2 resistance module. J Bacteriol 194:6696. https://doi.org/10.1128/JB.01809-12.
- Lin CW, Huang YW, Hu RM, Chiang KH, Yang TC. 2009. The role of AmpR in regulation of L1 and L2 β-lactamases in *Stenotrophomonas maltophilia*. Res Microbiol 160:152–158. https://doi.org/10.1016/j.resmic.2008.11.001.
- 16. Kuga A, Okamoto R, Inoue M. 2000. ampR gene mutations that greatly

increase class C β -lactamase activity in *Enterobacter cloacae*. Antimicrob Agents Chemother 44:561–567. https://doi.org/10.1128/aac.44.3.561-567 .2000.

- Bagge N, Ciofu O, Hentzer M, Campbell JI, Givskov M, Høiby N. 2002. Constitutive high expression of chromosomal β-lactamase in *Pseudomonas aeruginosa* caused by a new insertion sequence (IS1669) located in *ampD*. Antimicrob Agents Chemother 46:3406–3411. https://doi.org/10.1128/aac.46.11.3406-3411.2002.
- Chang Y-C, Huang Y-W, Chiang K-H, Yang T-C, Chung T-C. 2010. Introduction of an *AmpR-L2* intergenic segment attenuates the induced β-lactamase activity of *Stenotrophomonas maltophilia*. Eur J Clin Microbiol Infect Dis 29:887–890. https://doi.org/10.1007/s10096-010-0924-0.