



## Draft Genome of *Debaryomyces fabryi* CBS 789<sup>T</sup>, Isolated from a Human Interdigital Mycotic Lesion

## Hakim Tafer, Katja Sterflinger, Ksenija Lopandic

University of Natural Resources and Life Sciences Vienna, VIBT-Extremophile Center, Vienna, Austria

The yeast genus *Debaryomyces* comprises species isolated from various natural habitats, man-made environments, and clinical materials. Here, the draft genome of *D. fabryi* CBS 789<sup>T</sup>, isolated from a human interdigital mycotic lesion, is presented.

Received 16 November 2015 Accepted 9 December 2015 Published 4 February 2016

Citation Tafer H, Sterflinger K, Lopandic K. 2016. Draft genome of *Debaryomyces fabryi* CBS 789<sup>T</sup>, isolated from a human interdigital mycotic lesion. Genome Announc 4(1): e01580-15. doi:10.1128/genomeA.01580-15.

Copyright © 2016 Tafer et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 3.0 Unported license.

Address correspondence to Ksenija Lopandic, ksenija.lopandic@boku.ac.at

ebaryomyces fabryi is an ascomycetous yeast placed within the family *Debaryomycetaceae* of the order *Saccharomycetales* (1). It was considered for a long time to be a variety of the cryotolerant and halotolerant species D. hansenii (2, 3) due to ambiguity of the phenotypic and genotypic boundaries, but on the basis of different fingerprinting techniques and phylogenetic analyses of several protein encoding genes D. fabryi has been accepted as separate taxonomic entity (4-7). D. hansenii can grow in media containing up to 4 M NaCl and has frequently been isolated from see water, cheese, meat, wine, beer, fruit, and soil, while the origin of the majority of the D. fabryi isolates are human skin infections (1). In contrast to D. hansenii strains that are able to grow at 31 to 35°C, the maximum growth temperature of the D. fabryi strains is 36 to 39°C (2). Recent studies have indicated that D. fabryi CBS 789<sup>T</sup> is more resistant to oxidative stress and more sensitive to fluconazole than *D. hansenii* CBS  $767^{T}$  (8).

In order to determine differences in the genome constitution and gene regulation between two phylogenetically closely related but phenotypically different D. hansenii and D. fabryi species, the whole-genome sequence of D. fabryi CBS 789T originating from an interdigital mycotic lesion was generated. The Ion PI Hi-Q Chef Kit protocol (Life Technologies, Carlsbad, CA, USA) was used to perform emulsion PCR amplification and enrichment of the template ion sphere particles (ISPs). The enriched ISPs were loaded onto Ion PI Chip v3 and sequenced by an Ion Proton semiconductor-based sequencer. A total of 13.7 G with a median read length of 180 bp were generated and assembled with Newbler 2.9 into a 12-Mb genome containing 551 contigs ( $N_{50}$  59,311). Ninety-seven percent of the ultraconserved eukaryotic genes were recovered by CEGMA in the genome. Augustus, snap2, scipio, cegma, and glimmer were used to predict protein coding genes. Evidencemodeler summarized the predictions from the various tools into a final protein coding genes set containing 6,027 loci. TRNAscan found all tRNA-isotypes with the exception of SelCys (111 tRNAs). Furthermore, RNAse P, RNAaseMRP, 5 small nuclear RNAs (U1, U2, U4, U5, U6), and 51 snoRNAs were found. Finally two RNA cis-regulatory elements (histone 3'UTR stem loop, TPP riboswitch) were also detected.

**Nucleotide sequence accession numbers.** This whole-genome shotgun project has been deposited in DDBJ/EMBL/GenBank under the accession number LMYN00000000. The version described in this paper is the first version, LMYN01000000.

## ACKNOWLEDGMENTS

The equipment of the VIBT-Extremophile Center used in this study was financed by BOKU-Equipment GesmbH. The computational results presented have been achieved in part using the Vienna Scientific Cluster (VSC).

## REFERENCES

- 1. Suzuki M, Prasad GS, Kurtzman C. 2011. Debaryomyces Lodder & Kreger-van Rij (1952), p 361–372. *In* Kurtzman CP, Fell JW, Boekhout T (ed), The yeasts, a taxonomic study, vol 2, 5th ed. Elsevier, London, United Kingdom.
- Nakase T, Suzuki M. 1985. Taxonomic studies on *Debaryomyces hansenii* (Zopf) Lodder et Kreger-Van Rij and related species. II. Practical discrimination and nomenclature. J Gen Appl Microbiol 31:71–86. http:// dx.doi.org/10.2323/jgam.31.71.
- Kurtzman CP, Robnett CJ. 1998. Identification and phylogeny of ascomycetous yeasts from analysis of nuclear large subunit (26S) ribosomal DNA partial sequences. Antonie Van Leeuwenhoek 73:331–371. http:// dx.doi.org/10.1023/A:1001761008817.
- Prillinger H, Molnár O, Eliskases-Lechner F, Lopandic K. 1999. Phenotypic and genotypic identification of yeasts from cheese. Antonie Van Leeuwenhoek 75:267–283. http://dx.doi.org/10.1023/A:1001889917533.
- Groenewald M, Daniel HM, Robert V, Poot GA, Smith MT. 2008. Polyphasic re-examination of *Debaryomyces hansenii* strains and reinstatement of *D. Hansenii*, *D. Fabryii* and *D. Subglobosus*. Persoonia 21:17–27. http://dx.doi.org/10.3767/003158508X336576.
- Jacques N, Mallet S, Casaregola S. 2009. Delimitation of the species of the Debaryomyces hansenii complex by intron sequence analysis. Int J Syst Evol Microbiol 59:1242–1251. http://dx.doi.org/10.1099/ijs.0.004325-0.
- Nguyen HV, Gaillardin C, Neuvéglise C. 2009. Differentiation of *Debaryomyces hansenii* and *Candida famata* by rRNA gene intergenic spacer fingerprinting and reassessment of phylogenetic relationships among *D. hansenii*, *C. famata*, *D. fabryii*, *C. fareri* (=*D. subglobosus*) and *D. prosopidis*: description of *D. vietnamensis* sp.nov. closely related to *D. nepalensis*. FEMS Yeast Res 9:641–662. http://dx.doi.org/10.1111/j.1567-1364.2009.00510.x.
- Michán C, Martínez JL, Alvarez MC, Turk M, Sychrova H, Ramos J. 2013. Salt and oxidative stress tolerance in *Debaryomyces hansenii* and *Debaryomyces fabryi*. FEMS Yeast Res 13:180–188. http://dx.doi.org/10.1111/ 1567-1364.12020.