



## Carbon monoxide poisoning

Hiroshi Kinoshita<sup>a,\*</sup>, Hülya Türkan<sup>b</sup>, Slavica Vucinic<sup>c</sup>, Shahab Naqvi<sup>d</sup>, Rafik Bedair<sup>e</sup>,  
Ramin Rezaee<sup>f,g</sup>, Aristides Tsatsakis<sup>h</sup>

<sup>a</sup> Department of Forensic Medicine, Faculty of Medicine, Kagawa University, Kagawa, 761-0793, Japan

<sup>b</sup> Ministry of national Defense, General Directorate of Health Services, Ankara, Turkey

<sup>c</sup> National Poison Control Centre, Military Medical Academy, Medical Faculty, University of Defense, Belgrade, Serbia

<sup>d</sup> Anaesthesiology and Intensive Care, Rawal Institute of Health Sciences, Islamabad, Pakistan

<sup>e</sup> Adult Critical Care Directorate, St. George's University Hospitals, Blackshaw Road, London, SW17 9WL, United Kingdom

<sup>f</sup> Clinical Research Unit, Faculty of Medicine, Mashhad University of Medical Sciences, Mashhad, Iran

<sup>g</sup> Neurogenic Inflammation Research Center, Mashhad University of Medical Sciences, Mashhad, Iran

<sup>h</sup> Department of Toxicology & Forensic Sciences, Medical School, University of Crete, Voutes Campus, Heraklion, 71003, Greece



### ARTICLE INFO

#### Keywords:

Carbon monoxide  
Carboxyhemoglobin  
Pathophysiology  
Toxicity  
Measurement

### ABSTRACT

Carbon monoxide (CO) is the leading cause of poisoning deaths in many countries, including Japan. Annually, CO poisoning claims about 2000–5000 lives in Japan, which is over half of the total number of poisoning deaths. This paper discusses the physicochemical properties of CO and the toxicological evaluation of CO poisoning.

## 1. Introduction

Numerous poisons, from natural toxins [1,2] to synthetic chemicals existing in our environment, can produce a wide variety of deleterious effects in living organisms. This review article discusses carbon monoxide (CO) poisoning from a clinical point of view.

CO – sometimes termed a “silent killer” [3,4] is a colorless, odorless, and non-irritable gas. As the specific gravity of CO is 0.97, it is slightly lighter than air. This gas is mainly produced by incomplete combustion of organic compounds [5,6]. Vehicle exhaust, smoke from fires and improperly maintained heating systems are included as common sources.

The annual number of CO poisoning death in Japan is about 2000–5000 (Fig. 1), and it is a major cause of poisoning deaths. This is mirrored in several other countries [3,7–13]. The worldwide incidence of CO poisoning has remained stable during the last 25 years [14]. However, deaths from CO poisoning doubled in 2003 compared to that for 2001 in Japan. This may be attributable to an increase of suicides by means of CO inhalation [15]. Information from suicide-related websites may facilitate suicide by CO [16,17].

Forensic toxicologists deal with cases of fatal CO poisoning and are required to evaluate its toxicity in daily practice. This article describes the chemical properties and toxicological characteristics of CO.

## 2. Sources of CO

CO is formed by incomplete combustion of organic compounds. The main sources of CO encountered in poisoning cases are house fires (the maximum concentration of CO in air is around 5 % in the immediate vicinity of a house fire) [18], incomplete combustion of fuels (e.g., charcoal, briquette, fuel gas, petroleum) using a burner, heating or cooking equipment with insufficient ventilation or improper maintenance, exhaust gas from vehicles using internal combustion engines (the CO concentration in exhaust gas is less than a few percent) [19], and industrial accidents (such as those occurring at iron foundries or chemical plants).

Exhaust from a diesel engine contains approximately 0.01–0.06 % CO, and inhalation does not cause fatal CO poisoning [20]. Concentrations of CO in exhaust from gasoline engine have also decreased in recent years with the introduction of three-way catalytic converters [19].

CO is contained in mainstream smoke from cigarettes (3–4 %) [21], and blood carboxyhemoglobin (CO-Hb) saturation is increased approximately 10–15 % in heavy smokers [22–24].

Poisoning is occurred by inhalation of a relatively high concentration of CO gas. This is not always accidental: it is also used deliberately as a means of suicide. Charcoal-burning suicide has been increasing

\* Corresponding author at: Department of Forensic Medicine, Faculty of Medicine, Kagawa University, 1750-1, Miki, Kita, Kagawa 761-0793, Japan.  
E-mail address: [kinochin@med.kagawa-u.ac.jp](mailto:kinochin@med.kagawa-u.ac.jp) (H. Kinoshita).



Fig. 1. Statistics on annual numbers of death by CO poisoning in Japan (2000–2015).

since the late 1990's in East-Asian countries such as Hong Kong, Taiwan and Japan [10,15,25,26]. Approximately 80 % of cases under the category of “Other gases and vapors (ICD-10, X67)” in the Vital Statistics of Japan for 2007 were estimated to involve charcoal-burning suicide [15]. Cases of homicide by CO poisoning have also been reported [27].

### 3. Toxicokinetics of CO

CO is a gas at normal room temperature, and is inhaled from the lungs into the blood stream. Since the affinity of CO for hemoglobin (Hb) is 230- to 270-times greater than that of oxygen, CO-Hb is formed in erythrocytes [6,28,29]. The formation of CO-Hb in blood depends on a wide variety of factors, including the concentration of inspired CO, duration of CO exposure, pulmonary ventilation, exercise and health status [6,28]. A small amount of CO is produced by heme-protein degradation *in vivo* [28]. CO remains almost completely unoxidized following inhalation, with less than 0.1 % of inhaled CO being converted to carbon dioxide [30]. The rest is eventually discharged from the body.

CO shows a high affinity not only for hemoglobin, but also for other heme-proteins such as myoglobin and cytochrome c oxidase. CO also binds to myoglobin in myocardium and skeletal muscle [6,28,31]. As up to 15 % of the total CO in the body is taken up by tissues [32], CO can diffuse from organs into the blood as CO-Hb saturation in blood decreases [33].

The following formula has been used to estimate CO-Hb saturation in blood:

$$\text{CO-Hb (\%)} = a \times \text{CO in the inspired air (\%)} \times \text{time (min)},$$

where *a* is a constant with values of 3 at rest, 5 under light activity, 8 under light work, and 11 under heavy work [34], or

$$\text{CO-Hb (\%)} \approx 0.33 \times \text{RMV} \times \text{CO in the inspired air (\%)} \times \text{time (min)},$$

where RMV is respiratory minute volume, with standard values of 8.5 at rest, 25 under light exercise, and 50 under heavy exercise [35].

CO-Hb saturation in healthy, non-smoking subjects is less than 2 % [23,24]. It increases to 4–6 % in cases of hemolytic anemia, and can increase to nearly 10 %, depending on the state of the disease [23]. Methylene chloride, a solvent used as a paint or varnish remover, is metabolized to CO [36,37]. Severe CO poisoning with CO-Hb saturation up to 50 %, has been reported following methylene chloride exposure [38].

CO-Hb saturation in blood is easily decreased with oxygen administration [39]. The elimination half-life of CO during respiration depends on various factors, such as the concentration of inspired CO, duration of CO exposure, presence of oxygenation following rescue the oxygen concentration administered, and RMV [40,41]. This results in a half-life for a resting adult of about 4–5 h under room air ventilation at sea level, 80 min breathing 100 % oxygen at normobaric pressure, and

23 min breathing oxygen at 3 atmospheres absolute (ATA) [42,43].

### 4. Toxicity and pathophysiology of CO poisoning

Tissue hypoxia is the main toxic effect of acute CO poisoning, which is due the formation of CO-Hb. It causes decreases the oxygen transport capacity, resulting in insufficient oxygenation at the tissue level [6,28,29,44]. When CO binds to a hemoglobin subunit, other binding sites show increased affinity for the oxygen molecule. Hence, CO shifts the oxygen-hemoglobin dissociation curve to the left, inhibiting oxygen dissociation in the low-oxygen region, and potentiating tissue hypoxia [39,41,42,45–47].

Due to the much greater affinity of CO, as compared to oxygen, for hemoglobin, the bond between CO and hemoglobin is strong. However, this bond can be broken is reversible, CO is only displaced by oxygen slowly [6,28,29,47].

CO also binds to myoglobin in myocardium and skeletal muscle, causing dysfunctional tissue oxygen transport. In myocardium, this results in cardiac dysfunction [47]. It also has direct effects by inhibiting the activity of enzymes such as cytochrome c oxidase. CO poisoning may thus also be implicated in impairment of cardiac and neurological functions.

Apoptosis is a key factor in the pathogenesis of heart failure [48,49]. CO poisoning leads to apoptosis in myocardial cells. Neurotoxicity following CO exposure involves apoptosis and intracellular oxidative stress, and erythropoietin, resveratrol and hyperbaric oxygen all reduce dysfunction of the myocardium and brain by suppressing apoptosis or through other pathways [48–51].

Tissue hypoxia due to CO potentiates vascular permeability and causes increased accumulation of interstitial fluid with decreased circulating blood volume (hemoconcentration) affecting multiple organs. This includes brain edema with neurological symptoms and disorders of consciousness; pulmonary edema with respiratory failure; decreased myocardial contractility, arrhythmias and heart failure; and renal failure [46,47].

### 5. Symptoms and management of CO poisoning

The CO-Hb concentration in healthy, non-smoking subjects is less than 2 %, and less than 15 % in smokers. At CO-Hb levels below 10 %, no notable symptoms are observed. Neurological symptoms such as nausea, headache and dizziness are observed with CO-Hb levels over 10 %. Increases in respiratory and heart rates, syncope, motor paralysis and confusion are observed with CO-Hb level of 30–50 %. CO-Hb levels exceeding 50 % are considered life-threatening, and values in this range are central to the diagnosis of CO-poisoning [6,28,29,47].

Signs and symptoms such as headache, dizziness, fatigue and nausea are nonspecific [6,47]. Since behavioral disorders such as agitation, confusion and hallucination are sometimes observed, differential diagnoses such as psychosis, brain metastasis of tumor, stroke and coagulation disorders are required in clinical practice [52–55].

Motor function is depressed prior to impairment of consciousness in cases of CO poisoning [56]. This means that a victim may notice CO poisoning and try to improve the room ventilation, but may not be able to move at all.

The symptoms shown in Table 1 reportedly reflect the CO-Hb level. However, clinical symptoms of acute CO poisoning and their severity do not always correlate with concentrations of CO-Hb on admission [22,41,57,58]. This discrepancy may be due to two reasons. First, the CO-Hb saturation in blood is affected by various factors such as the concentration of inspired CO and the exposure time, oxygenation following rescue and the oxygen concentration applied [6,28], and the time elapsed between termination of CO exposure and blood sampling [57,58]. Second, as CO has a high affinity for heme proteins, CO that has diffused into tissues may not readily dissociate from them. As a result, considerable amounts of CO may be left in the body, even after

**Table 1**  
Levels of carboxyhemoglobin (CO-Hb) saturation (%) and symptoms.

CO-Hb (%)	clinical symptom
< 1	normal range (due to endogenous production)
< 10	smoker's blood (no symptom)
10–20	headache, fatigue, ear ringing
20–30	headache, weakness, nausea, vomiting
30–40	severe headache, dizziness, nausea, vomiting
40–50	syncope, confusion, increased respiration and heart rate
50k60	coma, convulsions, depressed respiration
60–70	coma, convulsions, cardiopulmonary depression, often fatal
70 <	respiratory failure, death

CO-Hb saturation has been decreased by oxygenation [31]. So, while Table 1 describe the symptoms related to increasing levels of CO-Hb following CO inhalation [41], we must also consider medical interventions and pre-hospital procedures such as oxygen administration when evaluating the toxicity of CO.

The first step of patient management is immediate evacuation from the contaminated environment. Control of the airway, intravenous access and cardiac monitoring are required for the management in the hospital. Administration of 100 % oxygen *via* facemask or endotracheal tube is required for the elimination of CO from blood as an initial treatment. Hyperbaric oxygen therapy can also be considered, where available, to accelerates the elimination of CO and reverse effects on inflammation and mitochondrial dysfunction induced by CO poisoning [6,47,59].

## 6. Autopsy findings

The cherry-red coloration of the skin is most characteristic appearance of the body surface is in CO poisoning cases. This is usually observed with CO-Hb concentrations exceeding 30 % [60]. Autopsy reveals blood, organs and muscles with similar cherry-red coloring, by the CO-Hb and carboxymyoglobin formation. Pulmonary edema and generalized organ congestion are also observed [60].

Necrosis of the globus pallidum is observed in cases of CO poisoning that occur over a prolonged period [61,62]. The underlying mechanisms are thought to involve hypoxic brain damage, as well as apoptosis [61–63].

## 7. Measurement of CO

Toxicological evaluation of CO poisoning is based on autopsy findings and CO-Hb saturation in blood. As most autopsy findings are non-specific for CO poisoning- other than the cherry-red color changes in the skin, organs and blood, the basic point of evaluation in forensic practice is CO-Hb saturation.

As a test for CO, spectrophotometric methods, gas chromatography, detection tubes and oximetry are employed [64,65], and various methods have been reported. The spectrophotometric method is the most widely used. The presence of CO-Hb can be determined by changes in the absorption spectrum [66–69]. This is simple procedure.

CO is liberated from Hb and introduced in gas chromatography. Various methods have been reported for liberation of CO from blood [70–76]. The released CO is detectable by various detectors, including a thermal conductivity detector [70–72,74,75], barrier discharge ionization detector [76], and flame ionization detector with the catalytic reduction of CO to methane (methanizer) [73]. Gas chromatography using a semiconductor detector (sensor gas chromatography) has been applied in forensic practice [77]. This system has some advantages such as portability and easy handling. As gas chromatography measures CO directly, the hemoglobin content also has to be measured for each sample, to enable calculation of the percentage CO-Hb.

Alternatively, a method using detector tube can be applied [78].

This system consists of an aspirating pump, tube for separation and tube for detection. The tube for CO separation is packed with ferricyanide coated silica gel particles [78]. The tube for detection of CO is packed with potassium palladium sulfite coated silica gel particles [79,80]. CO is released from blood following sample injection (200 µl) into the CO-separator tube, and the CO-detector tube detects the released CO gas, followed by aspiration by the pump. The detector tube can be easily used at the scene or in on-site testing.

An oximeter is widely used at the clinical laboratory [81], and also in daily forensic practice [82–90]. The instrument we use (AVOX 4000; International Technidyne Corp, Edison, NJ, USA) applies multiple wavelengths for the determination of various hemoglobin species, including CO-Hb, and requires small amounts of blood for measurement. This system allows easy handling of samples and has some advantages for on-site testing.

For details of measurement conditions and equipment, refer to the individual references [64–90].

CO-Hb is relatively stable at 4 °C for up to 24 months [91,92], and without refrigeration for up to 4 weeks [93].

## 8. Evaluation of CO toxicity

The fatal concentration for CO poisoning is a CO-Hb saturation over 50–60 % [65,94]. As mentioned above: since the CO-Hb saturation in blood is affected by multiple factors, medical interventions such as oxygen administration or cardiopulmonary resuscitation must be considered when evaluating CO toxicity. The measured value of CO-Hb at the time of death is generally found to be higher for younger casualties than for the elderly [94–96]. Elderly individuals may die at lower concentrations [6] - with a level is around 25 %, sometimes measured, with no other cause of death found. This may reflect the fact that younger individuals tend to have fewer comorbidities and are better able to tolerate the tissue hypoxia.

The brain is an organ with a very high oxygen demand, and so is especially sensitive to the effects of tissue hypoxia that results from acute CO poisoning. The heart is an organ with a high oxygen demand and is thus often affected, similar to the brain. Patients with cardiovascular disease experience reduced thresholds for angina, arrhythmias and myocardial infarction. These conditions have been observed even with CO-Hb of 5–10 % [6], with sudden death from severe arteriosclerotic heart disease reported at CO-Hb of 20–30 % [6,60]. During the last few years, several *in vivo* or *in vitro* experiments have examined various compounds such as magnesium sulfate, insulin, hesperidin, resveratrol, granulocyte colony stimulating factor and erythropoietin that can potentially combat early complications and late consequences of CO poisoning in the brain and heart [48–50,97–103].

In cases of automobile exhaust gas inhalation, the inhalation of nitrogen oxide leads to the production of methemoglobin (MetHb), and this need to be considered in addition to CO-Hb. High concentrations of MetHb have been reported in some cases, although methemoglobinemia is uncommon [104–106].

Additional consideration in cases of fire are other toxic gases (such as cyanide and phosgene) and oxygen deficiency that results from the consumption of oxygen in combustion. As cyanide is detoxicated by binding to MetHb, attention must be paid to the concentration of MetHb in the victim's blood when evaluating toxicity [107] - and so CO-Hb, cyanide and MetHb should be measured in cases of suspected fire victims [107].

Postmortem formation of CO due to putrefaction has been reported in a sample obtained from a case with a long postmortem interval. This was attributed to the degradation of heme-proteins such as hemoglobin and myoglobin. Postmortem formation of CO has also been reported in conditions and samples such as immersion in water for long periods. Values of CO-Hb over 10 % in pleural effusion are sometimes observed in cases of drowning without CO inhalation [108–112]. Since no indicators have been identified for postmortem CO formation, we should

not use body cavity fluids for measurement of CO in cases involving severe putrefaction.

## 9. Conclusion

We have discussed various issues related to CO described in previous reports. These data may be valuable for interpreting CO poisoning and may provide valuable information for forensic diagnosis. Recently, CO has been recognized as not only a toxic substance, but also a signaling gas, and research into therapeutic applications is being underway [113,114]. Further study of potential applications in daily practice are required.

## Declaration of Competing Interest

The authors declare that there are no conflicts of interest regarding the publication of this paper.

## Acknowledgements

This work was supported by JSPS KAKENHI Grant-in-aid for Scientific Research (C) Grant Number JP18K10127.

## Appendix A. Supplementary data

Supplementary material related to this article can be found in the online version, at doi:<https://doi.org/10.1016/j.toxrep.2020.01.005>.

## References

- [1] M. Tavassoli, A. Afshari, A.L. Arsene, B. Mégarbane, J. Dumanov, M.M. Bastos Paoliello, A. Tsatsakis, F. Carvalho, M. Hashemzaei, G. Karimi, R. Rezaee, Toxicological profile of *Amanita virosa* – a narrative review, *Toxicol. Rep.* 6 (2019) 143–150, <https://doi.org/10.1016/j.toxrep.2019.01.002>.eCollection 2019.
- [2] S. Govorushko, R. Rezaee, J. Dumanov, A. Tsatsakis, Poisoning associated with the use of mushrooms: a review of the global pattern and main characteristics, *Food Chem. Toxicol.* 128 (2019) 267–279, <https://doi.org/10.1016/j.fct.2019.04.016>.
- [3] G. Can, U. Sayili, Ö.A. Sayman, Ö.F. Kuyumcu, D. Yilmaz, E. Esen, E. Yurtseven, E. Erginöz, Mapping of carbon monoxide related death risk in Turkey: a ten-year analysis based on news agency records, *BMC Public Health* 19 (2019) 9, <https://doi.org/10.1186/s12889-018-6342-4>.
- [4] R.W. Byard, Carbon monoxide—the silent killer, *Forensic Sci. Med. Pathol.* 15 (2019) 1–2.
- [5] R.C. Baselt, *Disposition of Toxic Drugs and Chemicals in Man*, 11th ed., Biochemical Publications, Seal Beach, 2017.
- [6] M.J. Elenhorn, *Medical Toxicology Diagnosis and Treatment of Human Poisoning*, 2nd ed., Williams & Wilkins, Baltimore, 1997.
- [7] Ministry of Health, Labour and Welfare, *Vital Statistics of Japan, 1998–2015*.
- [8] National Research Institute of Police Science, *Annual case report of drug and toxic poisoning in Japan*, No. 44–No. 61, 2000–2019.
- [9] N. Tsunoda, Drug and toxic poisoning in recent Japan, *J. Health Sci.* 45 (1999) 356–366.
- [10] T. Ito, Y. Nakamura, Death from carbon monoxide poisoning in Japan between 1968–2007 through data from vital statistics, *J. Jpn. Soc. Emerg. Med.* 13 (2010) 275–282.
- [11] Centers for Disease Control and Prevention (CDC), Carbon monoxide –related deaths—United States, 1999–2004, *MMWR* 56 (2007) 1309–1312.
- [12] A.R. Anderson, Centers for Disease Control and Prevention (CDC), Top five chemicals resulting in injuries from acute chemical incidents – Hazardous substances emergency events surveillance, nine states, 1999–2008, *MMWR suppl.* 64 (2015) 39–46.
- [13] K. Sircar, J. Clower, M.K. Shin, C. Bailey, M. King, F. Yip, Carbon monoxide poisoning deaths in the United States, 1999 to 2012, *Am. J. Emerg. Med.* 33 (2015) 1140–1145.
- [14] C. Mattiuzzi, G. Lippi, Worldwide epidemiology of carbon monoxide poisoning, *Hum. Exp. Toxicol.* (2019), <https://doi.org/10.1177/0960327119891214>.
- [15] E. Yoshioka, S.J.B. Hanley, Y. Kawanishi, Y. Saijo, Epidemic of charcoal burning suicide in Japan, *Br. J. Psychiatry* 204 (2014) 274–282, <https://doi.org/10.1192/bjp.bp.113.135392>.
- [16] M. Hitosugi, Suicide due to carbon monoxide poisoning – trends and preventive measures, *Rinsho Byori. Suppl* 141 (2008) 40–44.
- [17] C. Ozawa-de Silva, Too lonely to die alone: internet suicide pacts and existential suffering in Japan, *Cult. Med. Psychiatry* 32 (2008) 516–551.
- [18] Y. Suzuki, M. Takeda, T. Inamura, Y. Tanaka, Analyzed gases of burning products collected at fire scenes, *Rep. Fire Sci. Lab.* 26 (1989) 45–52.
- [19] T. Kawaraya, H. Garivait, W. Laowagul, P. Sukasem, M.S. Tabucanon, M. Sakata, T. Okuno, Exhaust gases from new gasoline vehicles in Thailand, *Seikatsu. Eisei.* 41 (1997) 93–96.
- [20] H. Yamanouchi, N. Honma, R. Shigeno, A fatal case due to the exhaust of diesel engine, *Res. Pract. Forens. Med.* 28 (1985) 115–118.
- [21] M. Kano, Pharmacology of the smoking, *Diagn. Treat.* 97 (2009) 1327–1331.
- [22] J.A. Raub, M. Mathieu-Norf, N.B. Hampson, S.R. Thom, Carbon monoxide poisoning – a public perspective, *Toxicology* 145 (2000) 1–14.
- [23] E.O. Owens, Endogenous carbon monoxide production in disease, *Clin. Biochem.* 43 (2010) 1183–1188.
- [24] L.D. Prockop, R.I. Chichkova, Carbon monoxide intoxication: an updated review, *J. Neurol. Sci.* 262 (2007) 122–130.
- [25] K.Y. Liu, A. Beautrais, E. Caine, K. Chan, A. Chao, Y. Conwell, C. Law, D. Lee, P. Li, P. Yip, Charcoal burning suicides in Hong Kong and urban Taiwan: an illustration of the impact of a novel suicide method on overall regional rates, *J. Epidemiol. Community Health* 61 (2007) 248–253.
- [26] C. Yi-Han, H. Chia-Yueh, C. Qijin, C. Shu-shen, Y. Paul, The evolution of the characteristics of charcoal-burning suicide in Hong Kong, 2002–2013, *J. Affect. Disord.* 257 (2019) 390–395, <https://doi.org/10.1016/j.jad.2019.07.041>.
- [27] K. Marufuji, T. Ohnuma, M. Ohmi, T. Niwase, A homicide case camouflaged as an accidental CO poisoning, *Res. Pract. Forens. Med.* 18 (1975) 59–70.
- [28] World Health Organization, *Environmental Health Criteria 213, Carbon Monoxide*, 2nd ed., World Health Organization, Geneva, 1999.
- [29] R.H. Powers, D.E. Dean, *Pulmonary toxicology, Forensic Toxicology Mechanisms and Pathology*, CRC Press, Boca Raton, 2016, pp. 147–164.
- [30] C.A. Tobias, J.H. Lawrence, F.J.W. Roughton, W.S. Root, M.I. Gregersen, The elimination of carbon monoxide from the human body with reference to the possible conversion of CO to CO<sub>2</sub>, *Am. J. Physiol.* 145 (1945) 253–263.
- [31] I. Yamamoto, S. Inokuchi, Carbon monoxide, in: *Japanese Society for Clinical Toxicology (Ed.), Standard Clinical Practice Guide for Acute Poisoning*, Jiho, Tokyo, 2008, pp. 179–186.
- [32] R.F. Coburn, The carbon monoxide body stores, *Ann. N.Y. Acad. Sci.* 174 (1970) 11–22.
- [33] F.J.W. Roughton, W.S. Root, The fate of CO in the body during recovery from mild carbon monoxide poisoning in man, *Am. J. Physiol.* 145 (1945) 239–252.
- [34] W.H. Forbes, F. Sargent, F.J.W. Roughton, The rate of carbon monoxide uptake by normal men, *Am. J. Physiol.* 143 (1945) 594–608.
- [35] J.G. Quintiere, *Principles of fire behavior*, Delmar, New York (1997).
- [36] R.S. Ratney, D.H. Wegman, H.B. Elkins, In vivo conversion of methylene chloride to carbon monoxide, *Arch. Environ. Health* 28 (1974) 223–226.
- [37] A. Carlsson, M. Hultengren, Exposure to methylene chloride. III. Metabolism of 14C-labelled methylene chloride in rat, *Scand. J. Work Environ. Health* 1 (1975) 104–108.
- [38] J. Fagin, J. Bradley, D. Williams, Carbon monoxide poisoning secondary to inhaling methylene chloride, *Br. Med. J.* 281 (1980) 1461.
- [39] K. Taki, Hyperbaric oxygen therapy (HBOT) for CO poisoning – survey of acute CO poisoning in Japan, *J. Jpn. Assoc. Clin. Hyperbaric Oxygen Div.* 6 (2009) 7–12.
- [40] T. Shimazu, Half-life of blood carboxyhemoglobin, *Chest* 119 (2001) 661–662, <https://doi.org/10.1378/chest.119.2.661>.
- [41] T. Shimazu, Pathophysiology, myths and mysteries of acute carbon monoxide poisoning, *Chudoku Kenkyu* 19 (2006) 23–33.
- [42] H. Naito, *Poisoning of Industrial Products, Gases Pesticides, Drugs and Natural Toxins -cases, Pathogenesis and Its Treatment-*, 2nd ed., Nankodo Co., Ltd, Tokyo, 2001.
- [43] L.K. Weaver, S. Howe, R. Hopkins, K.J. Chan, Carboxyhemoglobin half-life in carbon monoxide-poisoned patients treated with 100% oxygen at atmospheric pressure, *Chest* 117 (2000) 801–808, <https://doi.org/10.1378/chest.117.3.801>.
- [44] A. Ghosh, S. Banerjee, A. Mitra, M. Muralidharan, B. Roy, R. Banerjee, A.K. Mandal, I.B. Chatterjee, Interaction of p-benzoquinone with hemoglobin in smoker's blood causes alteration of structure and loss of oxygen binding capacity, *Toxicol. Rep.* 3 (2016) 295–305.
- [45] C.G. Douglas, J.S. Haldane, J.B.S. Haldane, The laws of combination of haemoglobin with carbon monoxide and oxygen, *J. Physiol.* 44 (1912) 275–304.
- [46] Y. Okada, Carbon monoxide poisoning, *Jpn. J. Acute Med.* 3 (1979) 1114–1122.
- [47] C. Tomaszewski, et al., Carbon monoxide, in: L.S. Nelson (Ed.), *Goldfrank's Toxicologic Emergencies*, 9th ed., McGraw-Hill, New York, Chicago, 2011, pp. 1658–1670.
- [48] M.A. Rezaee, A.H. Mohammadpour, M. Imenshahidi, M. Mahmoudi, M. Sankian, K. Tsarouhas, A. Tsakalof, A.M. Tsatsakis, S.A. Moallem, Protective effect of erythropoietin on myocardial apoptosis in rats exposed to carbon monoxide, *Life Sci.* 148 (2016) 118–124, <https://doi.org/10.1016/j.lfs.2016.02.007>.
- [49] M. Hashemzaei, A.K. Barani, M. Iranshahi, R. Rezaee, K. Tsarouhas, A.M. Tsatsakis, M.F. Wilks, K. Tabrizian, Effects of resveratrol on carbon monoxide-induced cardiotoxicity in rats, *Environ. Toxicol. Pharmacol.* 46 (2016) 110–115, <https://doi.org/10.1016/j.etap.2016.07.010>.
- [50] S.A. Moallem, A.H. Mohammadpour, K. Abnous, M. Sankian, H.R. Sadeghnia, A. Tsatsakis, S. Shahsavand, Erythropoietin in the treatment of carbon monoxide neurotoxicity in rat, *Food Chem. Toxicol.* 86 (2015) 56–64, <https://doi.org/10.1016/j.fct.2015.09.015>.
- [51] D.M. Jurič, D. Šuput, M. Brvar, Hyperbaric oxygen preserves neurotrophic activity of carbon monoxide-exposed astrocytes, *Toxicol. Lett.* 253 (2016) 1–6, <https://doi.org/10.1016/j.toxlet.2016.04.019>.
- [52] V.V. Wojciechowski, D. Calina, K. Tsarouhas, A.V. Pivnik, A.A. Sergievich, V.V. Kodintsev, E.A. Filatova, E. Ozcgali, A.O. Docea, A.L. Arsene, E. Gofita, C. Tsitsimpikou, A.M. Tsatsakis, K.S. Golokhvast, A guide to acquired vitamin K coagulopathy diagnosis and treatment: the Russian perspective, *Daru* 25 (2017) 10, <https://doi.org/10.1186/s40199-017-0175-z>.
- [53] L. Nassbaum, L.M. Hoge, D. Călina, N. Andreescu, R. Grădinaru, R. Ștefănescu, M. Puiu, Modern treatment approaches in psychoses pharmacogenetic, neuro-magistic and clinical implications, *Farmacia* 65 (2017) 75–81.
- [54] A.-M. Buga, A.O. Docea, C. Albu, R.D. Malin, D.E. Branisteanu, G. Ianosi, S.L. Ianosi, A. Iordache, D. Calina, Molecular and cellular stratagem of brain metastases associated with melanoma, *Oncol. Lett.* 17 (2019) 4170–4175, <https://doi.org/10.3892/ol.2019.9933>.

- [55] A.M. Tsatsakis, A.O. Docea, D. Calina, K. Tsarouhas, L.-M. Zamfira, R. Mitrut, J. Sharifi-Rad, L. Kovatsi, V. Siokas, E. Dardiotis, N. Drakoulis, G. Lazopoulos, C. Tsitsimpikou, P. Mitsias, M. Neagu, A mechanistic and pathophysiological approach for stroke associated with drugs of abuse, *J. Clin. Med.* 8 (2019) 1295, <https://doi.org/10.3390/jcm8091295>.
- [56] O. Suzuki, Forensic toxicology, in: Y. Katsumata, O. Suzuki (Eds.), *NEW Legal Medicine and Medical Law*, Nankodo Co., Ltd, Tokyo, 2008, pp. 131–163.
- [57] J.A. Sokal, E. Kralkowska, The relationship between exposure duration, carboxyhemoglobin, blood glucose, pyruvate and lactate and the severity of intoxication in 39 cases of acute carbon monoxide poisoning in man, *Arch. Toxicol.* 57 (1985) 196–199.
- [58] N.B. Hampson, N.M. Hauff, Carboxyhemoglobin levels in carbon monoxide poisoning: do they correlate with the clinical picture? *Am. J. Emerg. Med.* 26 (2008) 665–669, <https://doi.org/10.1016/j.ajem.2007.10.005>.
- [59] J.J. Rose, L. Wang, Q. Xu, C.F. McTiernan, S. Shiva, J. Tejero, M.T. Glandwin, Carbon monoxide poisoning: pathogenesis, management, and future directions of therapy, *Am. J. Respir. Crit. Care Med.* 195 (2017) 596–606, <https://doi.org/10.1164/rccm.201606-1275CI>.
- [60] P. Saukko, B. Knight, *Knights' Forensic Pathology*, 4th ed., CRC Press, Boca Raton, FL, 2016, pp. 589–594.
- [61] M. Okada, B. Okuda, S. Okae, Bilateral necrosis of the pallidum in a case of carbon monoxide poisoning, *Neurol. Med.* 17 (1982) 304–305.
- [62] Y. Sawada, T. Sakamoto, D. Sadamitsu, J. Yokota, N. Ohashi, T. Yoshioka, S. Onishi, CT and pathological findings of globus pallidus after severe carbon monoxide poisoning, *Jpn. J. Acute Med.* 7 (1983) 1721–1724.
- [63] K. Uemura, K. Harada, D. Sadamitsu, R. Tsuruta, M. Takahashi, T. Aki, M. Yasuhara, T. Maekawa, K.-i. Yoshida, Apoptotic and necrotic brain lesions in a fatal case of carbon monoxide poisoning, *Forensic Sci. Int.* 116 (2001) 213–219.
- [64] A. Akane, Y. Fukui, A review on the development of carboxyhemoglobin analysis, *Res. Pract. Forens. Med.* 28 (1985) 185–190.
- [65] K. Sato, Carbon monoxide, in: O. Suzuki, K. Watanabe (Eds.), *Drugs and Poisons in Humans, -a Handbook of Practical Analysis*, Springer-Verlag, Berlin Heidelberg, 2005, pp. 91–99.
- [66] S. Akiya, A. Tanimura, Microdetermination of carbon monoxide-hemoglobin, *Yakugaku Zasshi* 72 (1952) 453–454.
- [67] H. Kozuka, T. Niwase, T. Taniguchi, Spectrophotometric determination of carboxyhemoglobin, *J. Hygiene Chem.* 15 (1969) 342–345.
- [68] Y. Katsumata, M. Aoki, K. Sato, O. Suzuki, M. Oya, S. Yada, A simple spectrophotometry for determination of carboxyhemoglobin in blood, *J. Forensic Sci.* 27 (1982) 928–934.
- [69] H. Ishizu, S. Ameno, Y. Yamamoto, Y. Okamura, N. Shogano, A simple spectrophotometric method for CO-Hb determination in blood and its application in legal medicine, *Acta. Crim. Japon.* 48 (1982) 187–196.
- [70] C. Miyauchi, K. Sakaki, An improved method for gas chromatographic quantification of carboxyhemoglobin in blood, *Nihon Hoigaku Zasshi* 28 (1974) 59–64.
- [71] S. Hishida, Y. Mizoi, Studies on the quantitative determination of carbon monoxide in human blood by gas chromatography, *Nihon Hoigaku Zasshi* 30 (1976) 319–326.
- [72] M. Sakata, M. Haga, Determination of carbon monoxide in blood by head space analysis, *J. Toxicol. Sci.* 5 (1980) 35–43.
- [73] J.G. Guillot, J.P. Weber, J.Y. Savoie, Quantitative determination of carbon monoxide in blood by head-space gas chromatography, *J. Anal. Toxicol.* 5 (1981) 264–266.
- [74] Y. Seto, M. Kataoka, K. Tsuge, Stability of blood carbon monoxide and hemoglobins during heating, *Forensic Sci. Int.* 121 (2001) 144–150.
- [75] R.J. Lewis, R.D. Johnson, D.V. Canfield, An accurate method for the determination of carboxyhemoglobin in postmortem blood using GC-TCD, *J. Anal. Toxicol.* 28 (2004) 59–62.
- [76] T. Ohmori, M. Otsuka, Y. Seto, Measurement of carbon monoxide in blood sample using gas-chromatography with barrier discharge ionization detection, *Rep. Natl. Res. Inst. Police Sci.* 66 (2017) 56–64.
- [77] N. Tanaka, H. Kinoshita, A. Takakura, M. Jamal, M. Kumihashi, Y. Uchiyama, K. Tsutsui, K. Ameno, Application of sensor gas chromatography for the determination of carbon monoxide in forensic medicine, *Curr. Environ. Med. Sci.* 7 (2014) 9–11.
- [78] F. Ishizawa, S. Misawa, A handy and simple apparatus for the quantitative determination of COHb in blood, *Acta. Crim. Japon.* 52 (1986) 26–32.
- [79] Komyo Rikagaku Kogyo. *Gas Detector Tube System Handbook*, 4th ed., Komyo rikagaku kogyo, Tokyo, 2003.
- [80] Pharmaceutical Society of Japan, *Standard Methods of Analysis for Hygienic Chemists -with Commentary-* (1990), Kanehara shuppan, Tokyo, 1990.
- [81] J.J. Mahoney, H.J. Vreman, D.K. Stevenson, A.L. Van Kessel, Measurement of carboxyhemoglobin and total hemoglobin by five specialized spectrophotometers (CO-oximeters) in comparison with reference methods, *Clin. Chem.* 39 (1993) 1693–1700.
- [82] M. Okada, T. Okada, K. Ide, Utilization of a CO-oximeter in the medico-legal field, *Nihon Hoigaku Zasshi.* 39 (1985) 318–325.
- [83] T. Higuchi, K. Noguchi, H. Maeda, An evaluation of analyzed data of hemoglobin derivatives by CO-oximeter in medico-legal autopsy, *Nihon Houigaku Zasshi.* 46 (1992) 416–418.
- [84] H. Maeda, K. Fukita, S. Oritani, K. Nagai, B.L. Zhu, Evaluation of post-mortem oxymetry in fire victims, *Forensic Sci. Int.* 81 (1996) 201–209.
- [85] K. Shimizu, H. Mizukami, T. Fukushima, M. Sasaki, H. Shiono, Use of a CO-oximeter for forensic diagnosis of hypothermia, *Nihon Houigaku Zasshi.* 52 (1998) 196–201.
- [86] C. Brehmer, P.X. Iten, Rapid determination of carboxyhemoglobin in blood by Oximeter, *Forensic Sci. Int.* 133 (2003) 179–181.
- [87] N. Watanabe, Medico-legal application of hemoglobin analysis using CO-oximeter, *Hokkaido Igaku Zasshi* 78 (2003) 557–566.
- [88] K.N. Olson, M.A. Hillyer, J.S. Kloss, R.J. Geiselhart, F.S. Apple, Accident or arson: is Co-oximetry reliable for carboxyhemoglobin measurement postmortem? *Clin. Chem.* 56 (2010) 515–520.
- [89] N. Tanaka, K. Ameno, M. Jamal, E. Ohkubo, M. Kumihashi, H. Kinoshita, Application of oximeter AVOX 4000 for the determination of CO-Hb in the forensic practice, *Res. Pract. Forens. Med.* 53 (2010) 39–43.
- [90] J. Fujihara, H. Kinoshita, N. Tanaka, T. Yasuda, H. Takeshita, Accuracy and usefulness of the AVOXimeter 4000 as routine analysis of carboxyhemoglobin, *J. Forensic Sci.* 58 (2013) 1047–1049.
- [91] G.W. Kunsman, C.L. Presses, P. Prodriguez, Carbon monoxide stability in stored postmortem blood samples, *J. Anal. Toxicol.* 24 (2000) 572–578.
- [92] N. Tanaka, K. Ameno, M. Jamal, M. Kumihashi, N. Miyatake, H. Kinoshita, Effects of sampling methods and storage on the value of oxyhemoglobin ratio and carboxyhemoglobin ratio, *Res. Pract. Forens. Med.* 55 (2012) 51–55.
- [93] N.B. Hampson, Stability of carboxyhemoglobin in stored and mailed blood samples, *Am. J. Emerg. Med.* 26 (2008) 191–195.
- [94] T. Mitsui, M. Ito, Concentration of carboxyhemoglobin in blood to death, *Eisei Kagaku* 36 (1990) 158–161.
- [95] B. Teige, J. Lundevall, E. Fleischer E, Carboxyhemoglobin concentrations in fire victims and in cases of fatal carbon monoxide poisoning, *Z. Rechtsmed.* 80 (1977) 17–21.
- [96] M. Yoshida, J. Adachi, T. Watabiki, Y. Tatsuno, N. Ishida, A study on house fire victims: age, carboxyhemoglobin, hydrogen cyanide and hemolysis, *Forensic Sci. Int.* 52 (1991) 13–20.
- [97] G. Bagheri, R. Rezaee, K. Tsarouhas, A.O. Docea, J. Shahraki, M. Shahriari, M.F. Wilks, H. Jahantigh, K. Tabrizian, A.A. Moghadam, S. Bagheri, D.A. Spandidos, A. Tsatsakis, M. Hashemzai, Magnesium sulfate ameliorates carbon monoxide-induced cerebral injury in male rats, *Mol. Med. Rep.* 19 (2019) 1032–1039, <https://doi.org/10.3892/mmr.2018.9771>.
- [98] K. Tabrizian, Z. Shahriari, R. Rezaee, H. Jahantigh, G. Bagheri, K. Tsarouhas, A.O. Docea, A. Tsatsakis, M. Hashemzai, Cardioprotective effects of insulin on carbon monoxide-induced toxicity in male rats, *Hum. Exp. Toxicol.* 38 (2019) 148–154, <https://doi.org/10.1177/0960327118788134>.
- [99] R. Rezaee, A. Sheidary, S. Jangjoo, S. Ekhtiary, S. Bagheri, Z. Kohkan, M. Dadres, A. Oana Docea, K. Tsarouhas, D.A. Sarigiannis, S. Karakitsios, A. Tsatsakis, L. Kovatsi, M. Hashemzai, Cardioprotective effects of hesperidin on carbon monoxide poisoned in rats, *Drug Chem. Toxicol.* 14 (2019) 1–6, <https://doi.org/10.1080/01480545.2019.1650753>.
- [100] M. Hashemzai, A.H. Mohammadpour, M. Imenshahidi, R. Rezaee, Does granulocyte colony stimulating factor have protective effects against carbon monoxide-induced apoptosis? *Biologia* 73 (2018) 1153–1157, <https://doi.org/10.2478/s11756-018-0121-7>.
- [101] K. Tabrizian, H. Khodayari, R. Rezaee, H. Jahantigh, G. Bagheri, K. Tsarouhas, M. Hashemzai, Magnesium sulfate protects the heart against carbon monoxide-induced cardiotoxicity in rats, *Res. Pharm. Sci.* 13 (2018) 65–72, <https://doi.org/10.4103/1735-5362.220969>.
- [102] K. Tabrizian, J. Shahraki, M. Bazzi, R. Rezaee, H. Jahantigh, M. Hashemzai, Neuro-protective effects of resveratrol on carbon monoxide-induced toxicity in male rats, *Phytother. Res.* 31 (2017) 1310–1315, <https://doi.org/10.1002/ptr.5855>.
- [103] S. Shahsavand, A.H. Mohammadpour, R. Rezaee, E. Behravan, R. Sakhtianchi, S.A. Moallem, Effect of erythropoietin on serum brain-derived biomarkers after carbon monoxide poisoning in rats, *Iran. J. Basic Med. Sci.* 15 (2012) 752–758.
- [104] H. Suyama, S. Morikawa, S. Noma-Tanaka, H. Adachi, Y. Kawano, K. Kaneko, S. Ishihara, Methemoglobinemia induced by automobile exhaust fumes, *J. Anesth.* 19 (2005) 333–335.
- [105] M. Vevelstad, I. Morild, Lethal methemoglobinemia and automobile exhaust inhalation, *Forensic Sci. Int.* 187 (2009) e1–e5.
- [106] Y.-M. Kuo, R.L. Nussbaum, Prolongation of chemically-induced methemoglobinemia in mice lacking  $\alpha$ -synuclein: a novel pharmacologic and toxicologic phenotype, *Toxicol. Rep.* 3 (2016) 295–305.
- [107] F. Moriya, Poisoning due to carbon monoxide and cyanide gas generated in the occurrence of fire, *Chudoku Kenkyu* 28 (2015) 339–345.
- [108] T. Kojima, M. Yashiki, I. Une, Y. Nishiyama, Post-mortem formation of carbon monoxide in a drowned body, *Nihon Hoigaku Zasshi* 34 (1980) 163–168.
- [109] T. Kojima, Y. Nishiyama, M. Yashiki, I. Une, Postmortem formation of carbon monoxide, *Forensic Sci. Int.* 19 (1982) 243–248.
- [110] T. Kojima, M. Yashiki, I. Une, Experimental study on postmortem formation of carbon monoxide, *Forensic Sci. Int.* 22 (1983) 131–135.
- [111] T. Kojima, M. Yashiki, I. Okamoto, J. Noda, I. Une, T. Miyazaki, F. Chikasue, Postmortem formation of carbon monoxide in blood and body cavity fluids of rats drowned and kept immersed in fresh water, *Hiroshima J. Med. Sci.* 33 (1984) 591–594.
- [112] T. Kojima, I. Okamoto, M. Yashiki, T. Miyazaki, F. Chikasue, K. Degawa, S. Oshida, K. Sagisaka, Production of carbon monoxide in cadavers, *Forensic Sci. Int.* 32 (1986) 67–77.
- [113] A. Nakao, J. Kotani, Application of therapeutic signaling gas for acute lung injury, *J. Jpn. Assoc. Acute Med.* 24 (2013) 59–68.
- [114] D.R. Hess, Inhaled carbon monoxide: from toxin to therapy, *Respir. Care* 62 (2017) 1333–1342.