# **INTERFACE**

## royalsocietypublishing.org/journal/rsif

# Report



**Cite this article:** Newham E, Kague E, Aggleton JA, Fernee C, Brown KR, Hammond CL. 2019 Finite element and deformation analyses predict pattern of bone failure in loaded zebrafish spines. *J. R. Soc. Interface* **16**: 20190430. http://dx.doi.org/10.1098/rsif.2019.0430

Received: 20 June 2019 Accepted: 15 October 2019

#### Subject Category:

Life Sciences-Engineering interface

#### Subject Areas:

biomechanics

#### **Keywords:**

finite element, zebrafish, spine, mechanics, deformation, geometric morphometrics

#### Author for correspondence:

Chrissy L. Hammond e-mail: chrissy.hammond@bristol.ac.uk

<sup>†</sup>These authors contributed equally to this work.

# Arrish Kate Robson Brown<sup>1</sup> and Chrissy L. Hammond<sup>2</sup> <sup>1</sup>School of Arts, Woodland Road, Bristol, UK <sup>2</sup>The School of Physiology, Pharmacology and Neuroscience, Biomedical Sciences, University of Bristol, Bristol BS8 1TD, UK

zebrafish spines

(D) EK, 0000-0002-0266-9424

The spine is the central skeletal support structure in vertebrates consisting of repeated units of bone, the vertebrae, separated by intervertebral discs (IVDs) that enable the movement of the spine. Spinal pathologies such as idiopathic back pain, vertebral compression fractures and IVD failure affect millions of people worldwide. Animal models can help us to understand the disease process, and zebrafish are increasingly used as they are highly genetically tractable, their spines are axially loaded like humans, and they show similar pathologies to humans during ageing. However, biomechanical models for the zebrafish are largely lacking. Here, we describe the results of loading intact zebrafish spinal motion segments on a material testing stage within a micro-computed tomography machine. We show that vertebrae and their arches show predictable patterns of deformation prior to their ultimate failure, in a pattern dependent on their position within the segment. We further show using geometric morphometrics which regions of the vertebra deform the most during loading, and that finite-element models of the trunk subjected reflect the real patterns of deformation and strain seen during loading and can therefore be used as a predictive model for biomechanical performance.

Finite element and deformation analyses

predict pattern of bone failure in loaded

Elis Newham<sup>1,2,†</sup>, Erika Kague<sup>1,†</sup>, Jessye A. Aggleton<sup>1,2</sup>, Christianne Fernee<sup>1</sup>,

# 1. Introduction

The spine consists of a repeated pattern of motion segments (MSs) of bony vertebrae separated by intervertebral discs (IVDs) that enable movement. Back pain and IVD degeneration affect millions of people worldwide [1,2], and vertebral compression fractures are a frequent feature of osteoporosis [3]. Biomechanical pathologies of the spine are underpinned by genetic, physiological and environmental pathways that together damage IVD, muscle and the bone, changing the mechanics of the system.

Animal models, typically rodents, are frequently used to study mechanisms of spinal pathology [4]. However, quadrupeds are disadvantageous for studying the human spine as gravitational load acts perpendicular to their axial skeleton. Zebrafish are increasingly used as a model for human disease, due to their genetic tractability. Unlike quadrupeds, but similar to humans under gravity (figure 1*a*), their spine is antero-posteriorly loaded as a result of swimming through viscous water [5]. Zebrafish are well established as models for skeletogenesis, pathology and ageing [6], and develop spinal pathologies in response to altered genetics [7] and ageing [8]. However, the biomechanics of the zebrafish spine are comparatively poorly characterized.

Finite-element analysis (FEA) has proven a pivotal tool in the study of biomechanical subjects [9], and offers a method for biomechanically characterizing the zebrafish spine, including intact MSs. This technique digitally models an object of known material properties using a series of linked nodes of known

© 2019 The Authors. Published by the Royal Society under the terms of the Creative Commons Attribution License http://creativecommons.org/licenses/by/4.0/, which permits unrestricted use, provided the original author and source are credited.



**Figure 1.** *Ex vivo* spine loading leads to MS failure in a region of high strain predicted by FEA. (*a*) Schematic of zebrafish MS dissection. (*b*) MTS schematic and X-radiograph. (*c*) Orthogonal reconstruction slices showing vertebrae and associated soft tissue. (*d*) Three-dimensional reconstruction of the FEA model with colours reflecting different materials. (*e*) Details of the nucleus-pulposus (pink) and annulus fibrosis (blue) from (*d*) showing linear measurements of IVD thickness. (*f*) Predicted compressive deformation and strain map from FEA; dashed lines indicate axes in which boundary conditions were established. (*g*) Changes to IVD width measurements (bracketed dashed line highlights IVD elastic rebound) and (*h*) changes in MS length with increasing load for the three MTS specimens; symbols correspond to those in (*d*,*e*). Values are the absolute values presented in table 1, relative to their value at 1 N. (*i*,*j*) Reconstructions of MTS specimen 1 compressed to 10 N (*i*) and 16 N (*j*) with central vertebra indicated by asterisk in each. (*k*,*l*) Antero-posterior cross-sections of the central vertebra at 10 N (*k*) and 16 N (*l*). Muscle segmented in red, and bone in grey in (*i*–*l*). Red dashed line in (*l*) denotes the angle of fracture at the vertebral centrum. (*m*,*o*) FEA strain maps at 10 N (*m*) and 16 N (*o*). Scale shown in (*n*). (Online version in colour.)

number and geometry, that can be subjected to a wide variety of forces outputting the predicted geometry, strain and deformation. Results can be validated by comparison with the results of loading experiments in which a sample is loaded *ex vivo* [10,11]. FEA has been used in zebrafish to test contributions of shape and material properties in joint morphogenesis [12,13] and to study strain patterns in a single vertebra [14].

Here, we describe a novel integrated experimental platform that brings together imaging, modelling and real-world validation to explore the biomechanics of intact zebrafish spinal MSs. We generated an FEA model of the spine, which we validated with a loading experiment using a high-precision material testing stage (MTS) under set loading regimes using micro-computed tomography ( $\mu$ CT). Three-dimensional geometric morphometrics (3D-GM) was used to explore patterns of deformation seen in each vertebra during loading. Comparison of results demonstrated that our FEA model accurately predicted the relative patterns of deformation and strain experienced by real samples loaded *ex vivo*.

# 2. Methods

## 2.1. Zebrafish samples

One-year-old, wild-type (WT) zebrafish were fixed in 4% paraformaldehyde and dehydrated to 70% EtOH. MSs were acquired by making two cuts in the trunk, between the morphologically homogeneous vertebrae 18 and 24 of a total of 33 vertebrae [5] (figure 1a-c).

#### 2.2. In vitro vertebral loading experiment

Loading experiments were conducted using a custom-built material testing stage (MTS2) in a Bruker SKYSCAN 1272 µCT system. Radiographic visualization of each MS (n=3) was performed and if required, vertebrae were trimmed to retain three complete vertebrae and associated IVDs (figure 1b-d). Samples were stabilized (anterior-up) in the MTS2 using cyanoacrylate glue. The MTS2 was programmed to perform a sequential series of seven scans at a series of increasing loads (table 1), using 60 keV X-ray energy, 50 W current, 5 µm isotropic voxel size and a 0.25 mm aluminium filter. A total of 1501 projections were collected during a 180° rotation, with 400 ms exposure time. Reconstructions were performed using NRecon (v. 1.7.1.0). Surfaces of vertebrae, muscle and IVDs in each dataset were generated using Avizo (Avizo v. 8; Vizualisation Sciences Group) (figure 1c-e and table 1) and linear measurements of IVDs and MS lengths made using the '3D Measurement' tool. Vertebrae surfaces were further processed in Meshlab (table 2).

### 2.3. Finite-element analysis

An MS surface mesh was created based on a 1-year-old WT specimen  $\mu$ CT scanned using a Nikon XTH 225ST  $\mu$ CT system as described under two conditions: (a) native state and (b) contrastenhanced following 14 day incubation in 2.5% phosphomolybdemic acid [16]. Scan (a) was used to segment vertebrae (V18–V24), and scan (b) to segment IVDs. The resulting binary labels from scans (a) and (b) were saved as 8-bit tiff stacks, manually registered in 3D space in Avizo ('Trackball' tool) and algorithmically combined ('Algebra' tool), creating a single volume of separate materials representing three vertebrae and four IVDs (figure 1*d*,*e* and table 2). A 500 µm thick cylinder was created contacting the anterior-most IVD perpendicular to the model axis, to mimic the stainless-steel compressive plate and distribution of forces applied during loading (figure 1*f*).

The complete vertebral surface mesh was imported into Simpleware ScanIP (v. 2018.12, Synopsys Inc.) to create an FE model. The model consisted of 1 054 187 linear tetrahedral elements joined at 257 392 nodes comprising four material types: vertebral bone, annulus-fibrosus, nucleus-pulposus and stainless steel (figure 1d-f, table 2). The model was analysed in Abaqus (2018) version). A custom datum coordinate system was created centred on the antero-posterior axis of the model, and a concentrated force applied to the central node of the anterior face of the compressive plate. This loading case was repeated in each of seven steps of a multi-step analysis, with load values matching the increments applied in the MTS (table 1). The model was constrained in two locations using boundary conditions, at the base of the posterior-most IVD (constrained in three axes) and at the top of the compressive plate (constrained in two axes), allowing movement along the model's antero-posterior axis (figure 1f). Deformed meshes from each step were exported as surface files and analysed using 3D-GM for quantitative comparison between relative and absolute patterns of deformation predicted by FEA and observed in MTS data.

### 2.4. Three-dimensional geometric morphometrics

Three-dimensional geometric morphometrics analysis of vertebral deformation was performed using the 'Geomorph' package for the R statistics software [17]. For each loading experiment, we used the first scan (1 N load) to create a template of 3D coordinates for 22 fixed three-dimensional landmarks (figure 2a-c) linked by 300 surface sliding semi-landmarks (using the 'buildtemplate' function). By assigning the same landmarks in each scan (using the 'digitsurface' function), we compared the first scan with subsequent scans of the same vertebra using generalized Procrustes analysis (allowing semilandmarks to 'slide' in order to remove arbitrary spacing). Resulting shape variables were subjected to principal component analysis (PCA) to identify the principal patterns of variation between scans of the same vertebra, and isolate trends in deformation with increasing compressive load.

# 3. Results and discussion

## 3.1. Vertebral motion segments fail under loading of

12–16 N at positions of maximum von Mises strain To test the range of compressive loads that the MS could resist until failure, we subjected an MS to exponentially increasing compressive forces from 1 to 100 N. This specimen failed at 16 N whereupon the central vertebra fractured midcentrum. A primary loading regime between 1 and 16 N was thus established (table 1) for the three primary specimens; occupying the elastic, plastic and failure regions of the compressive loading profile of a typical MS. Failure was considered when at least one vertebral centrum fractured across the axis (e.g. figure 1j,l). All samples failed between 12 and 16 N upon shallow angle fracture in the central vertebra, with the smallest specimen (specimen 3) failing at the lowest force (figure 1g,h). This is higher than maximum aquatic forces experienced during swim training by Fiaz et al. [5], which reached approximately 9.5 N. Minor differences in mounting orientation created differences in linear deformation between right and left sides, but specimens follow similar patterns. Prior to failure, linear measurements show an increase in IVD antero-posterior thickness (table 1, bracketed dashed line in figure 1g), suggesting the IVD acts like a coiled spring that may further contribute to the ultimate

ween	
ict bet	
conta	
ndicate	
ilues in	
ince va	
o dista	
M. Zero	
3D-G/	
using	
asured	
ta me	
ion da	
formati	
ark det	
landmä	
es of	
A scor	
and PC	
ients ä	
asuren	
ric me	
nometi	
morpl	
linear	
their	
S, and	
he MT	
using t	
lysed L	
is ana	
ecimer	
for sp.	
edule	
ng sch	
Scanni.	
le 1.	ebrae.
Tab	vert

				linear IVD p	roxy measureme	nts (µm)						3D-GM PCA scores					
		left-	right-	IVD between	top and middle	vertebra		IVD between	middle and bo	ttom vertebra		anterior vertebra		central vertebra		posterior vertebra	
		lateral motion	lateral motion	left-	right-			left-	right-								
		segment	segment	lateral	lateral	ventral	dorsal	lateral	lateral	ventral	dorsal	PC1 (antero-		PC1 (antero-		PC1 (antero-	
	compressive	length	length	central	central	central	central	central	central	central	central	posterior	PC2 (lateral	posterior	PC2 (lateral	posterior	PC2 (lateral
specimen	load (N)	(mu)	(mıl)	distance	distance	distance	distance	distance	distance	distance	distance	deformation)	deformation)	deformation)	deformation)	deformation)	deformation)
MTS 1	-	2116	2128	37.8	39.2	12.15	20.72	74.42	40.8	15.43	8.94	-0.019	0.003	-0.011	0.001	-0.021	0.008
	5	2113	2122	36.97	37.15	10.39	19.17	65.38	36.72	12.37	6.43	-0.015	0.008	-0.006	-0.009	-0.017	0.004
	8	2104	2109	34.17	31.75	7.51	19.89	55.51	31.67	0	4.26	-0.002	-0.003	0.000	0.000	-0.006	-0.002
	10	2099	2094	31.13	31.99	2.95	14.05	46.95	21.43	0	4.43	-0.001	-0.008	0.002	0.012	-0.003	-0.003
	12	2096	2092	29.42	22.42	0	12.43	42.19	17.09	0	0	0.002	-0.013	0.005	0.008	0.007	-0.010
	14	2074	2085	31.47	25.82	0	18.54	46.37	21.05	0	0	0.034	0.005	0.014	-0.004	0.013	0.009
	16	FAILURE	FAILURE	FAILURE	FAILURE	FAILURE	FAILURE	FAILURE	FAILURE	FAILURE	FAILURE	FAILURE	FAILURE	FAILURE	FAILURE	FAILURE	FAILURE
MTS 2	-	2364	2346	24.65	13.86	6.8	10.4	17.25	15.38	3.2	6.85	-0.026	0.002	-0.014	0.002	-0.015	0.001
	5	2356	2344	21.04	12.56	5.72	9.98	16.73	14.12	0	0	-0.002	-0.006	-0.005	-0.002	-0.013	0.001
	8	2339	2344	18.21	12.21	0	0	15.04	13.53	0	0	0.002	-0.008	-0.001	0.011	-0.010	0.002
	10	2336	2338	16.37	9.56	0	0	14.54	10.93	0	0	0.006	-0.008	0.001	0.005	0.005	-0.015
	12	2316	2323	17.2	12.51	9.76	7.82	19.55	17.99	4.5	7.47	0.006	0.008	0.002	0.000	0.012	-0.006
	14	2316	2326	18.94	17.53	11.92	9.4	17.52	19.32	9.5	12.34	0.011	0.004	0.004	-0.004	0.015	0.008
	16	FAILURE	FAILURE	FAILURE	FAILURE	FAILURE	FAILURE	FAILURE	FAILURE	FAILURE	FAILURE	FAILURE	FAILURE	FAILURE	FAILURE	FAILURE	FAILURE
MTS 3	-	1832	1829	15.54	11.67	0	0		15.15	0	0	-0.013	0.002	-0.012	0.007	-0.005	-0.010
	5	1827	1822	14.23	10.28	0	0	15.39	14.2	0	0	0.001	-0.010	-0.001	0.000	0.007	-0.006
	8	1820	1816	13.97	7.75	0	0	15.78	10.26	0	0	0.005	0.010	0.001	-0.012	0.013	0.005
	10	1821	1807	13.31	7.82	0	0	14.97	13.15	0	0	0.007	-0.002	0.013	0.005	0.019	0.008
	12	FAILURE	FAILURE	FAILURE	FAILURE	FAILURE	FAILURE	FAILURE	FAILURE	FAILURE	FAILURE	FAILURE	FAILURE	FAILURE	FAILURE	FAILURE	FAILURE
	14	FAILURE	FAILURE	FAILURE	FAILURE	FAILURE	FAILURE	FAILURE	FAILURE	FAILURE	FAILURE	FAILURE	Failure	Failure	FAILURE	Failure	FAILURE
	16	FAILURE	FAILURE	FAILURE	FAILURE	FAILURE	FAILURE	FAILURE	FAILURE	FAILURE	FAILURE	FAILURE	FAILURE	FAILURE	FAILURE	FAILURE	FAILURE

4

analveic	ctan.	coffware	innut filo	toolkit/function	cuhetarra cuhetarra	outmut file
MTS	1. model creation	Avizo 8.0	reconstruction 16-bit tiff stack of single	edit new label field	a. separate segmentation of individual vertebrae using magic wand	.stl surface file
			load step		tool (minimum histogram grey-scale value set at 32 000)	
				generate surface	b. generation of vertebra surface from segmentation using generate	
					surface tool (unconstrained smoothing with a value of two)	
					c. save file as .stl surface file	
	2. model simplification	Meshlab 2016	.stl surface file	quadratic edge collapse deformation	a. simplify .stl by a factor of 10; target number of faces = $10\ 000;$	.ply file
					percentage reduction = 0.1; quality threshold = $0.9$	
					b. export simplified mesh as .ply file	
	3. 3D-geometric	R; Geomorph package	.ply files	read.ply	a. import .ply file for each compressive load of a single vertebra	.csv file
	morphometrics				(from 1 N to last load before failure)	
				buildtemplate	b. build landmark template of 22 landmarks and 300 surface sliding	
					semi-landmarks along the surface of the 1 N load dataset	
				digitsurface	c. assign the same 22 landmarks for each other dataset	
				gpagen (ProcD = FALSE)	d. perform Procrustes analysis for landmark coordinates	
				plotTangentSpace	e. perform PCA on Procrustes coordinates	
				write.csv	f. save PCA scores to .csv file	
				plotRefToTarget (mag = 10)	g. plot vector map of deformation, with vectors magnified by an	1. PCA scores; 2. vector plot
					order of 10	
FEA	1. model creation	Avizo 8.0	1. native state reconstruction 16 bit tiff	edit new label field	a. find analogous vertebral region to the motion segments analysed	.stl surface files of individual motion
			stack 2. contrast-enhanced		in MTS	segment elements
			reconstruction 16 bit tiff stack		b. segment bone in the native state reconstruction	
					c. segment bone and IVD in the contrast-enhanced reconstruction	
				three-dimensional (3D) image	a. find analogous region of characteristic vertebral morphology in	
				registration using 'Algebra' tool	both label fields using the 'Orthoslice tool'	
					b. manually adjust the volumes in 3D space using the 'Volume edit'	
					tool until they overlap precisely in all three dimensions and save	
					their new position in 3D space	
					c. combine both label fields using the 'Algebra' tool	
					d. edit new label field to save individual vertebrae and IVDs as	
					separate materials	
				generate surface	e. generation of surface from each individual material using generate	
					surface tool (unconstrained smoothing with a value of two)	
					f. save each material as .stl file	
						(Continued.

Table 2. Methodology tree for image processing and analysis.

analucic	cton	cofficience	immut filo	toolbit/function	cuhetana		ut fla
cickinin	arch				Share	1000	
		ScanIP (with CAD	stl surface files of individual motion	import .stl	a. import individual .stl files	inp.	file
		plugin)	segment elements	surface to mask	b. convert all surfaces to masks		
			:	create FE model	c. incorporate all surfaces into FE model		
			:	model configuration	d. assign material properties to materials		
					material Young's modulus Pois	son's	
					(GPA) I	ratio	
					stainless steel <sup>a</sup> 200 0.3		
					vertebral bone <sup>b</sup> 20 0.3		
					anulus 4.2 0.25	- - - - - - - - - - - - - - - - - - -	
					fibrosus <sup>c</sup>		
				-	nucleus 1.72 0.45		
					pulposus <sup>d</sup>		
				full FE	generate full FE model and save as an Abaqus input file (	(inp)	
	2. compression simulation	Abaqus	.inp file	module: step	a. create seven load steps, all 'static, general', to replicate	the seven .odb	Abaqus output database
					MTS load steps, switching the 'NIgeom' option on		
				module: load	a. create a custom coordinate datum system (CSYS) so tha	it the	
					x-axis aligns with the antero-posterior axis of the motio.	n segment	
					b. create boundary conditions in all three axes of CSYS for	r elements	
					along the posterior edge of the posterior-most IVD		
					c. create boundary conditions in the $y$ and $z$ axes for eler	nents along	
					the anterior face and the rim of the stainless-steel cor	npressive	
					plate		
					d. create mechanical load of 1 N concentrated force along	the <i>x</i> -axis	
					for the central node of the anterior face of the compr	essive plate	
					e. copy this load into each subsequent step, changing the	force to	
					that of the corresponding step in the MTS analysis, de	activating	
					each load in the steps they are not required		
			:	module: job	a. create and submit job		
					b. visualize the maximum deformation		
	3. analysis	Abaqus	odb Abaqus output database.	module: visualization	a. save the image for each loading step	1. im	ages of deformation and strain
					b. repeat for von Mises strain	d	atterns under increasing loads;
					c. export deformed meshes of every loading step as .stl su	urface files 2	. stl surface files of deformed motion
						Ŋ	egment under each compressive load
		Meshlab 2016	.stl surface file	quadratic edge collapse deformation	repeat method used for MTS analysis	i ylq.	fle
		R; Geomorph package	.ply files	Geomorph package	repeat method used for MTS analysis	1. PC	A scores; 2. vector plot
<sup>a</sup> Material properti	ss from the Engineering Toolbox (ww	vw.engineeringtoolbox.com; acc	essed 18 June 2019).				
<sup>b</sup> Material properti	es from Ofer <i>et al.</i> [14].						
Material properti	es assessed using atomic force micros	scopy of joint cartilage (R. Harn	iman 2018, personal communication).				
<sup>d</sup> Material properti	es from Panzer & Cronin [15].		•				

Table 2. (Continued.)

o royalsocietypublishing.org/journal/rsif J. R. Soc. Interface 16: 20190430



**Figure 2.** Finite element and geometric morphometric analyses model deformation patterns prior to failure (a-c). Landmarks assigned for 3D-GM analysis. (d,i,n) Results of PCA of landmark deformation under increasing compressive loads for each specimen, and deformation predicted by FEA (key in (s)). Black bracketed lines indicate reduced lateral compression. (e,j,o) Three-dimensional vector plots with black line vectors representing the direction of landmark deformation and colours highlighting the extent of landmark deformation for each vertebra in specimen 1 (vector scales magnified by 10; colour scale in (t)). (g,i,q) Deformation maps predicted by FEA (scales presented in (u)). (f,h,k,m,p,r) Examples of fractures (outlined in red for clarity) occurring at compressive loads before failure; corresponding with deformation patterns predicted in FEA and seen *ex vivo*. (Online version in colour.)

strain and failure of the segment when released via smallscale bone fracture (figure 1*h*). The surrounding epaxial musculature showed no obvious deformation or damage until the entire MS failed, at which point muscle fibre organization was lost (figure 1i-l). Comparison between MTS data and FEA results demonstrated strong spatial correlation between maximum predicted strain and ultimate point of failure in the central vertebra (figure 1m-o).

## 3.2. Morphometric characterization of vertebral

compression is predicted by finite-element analysis We found characteristic patterns of deformation and strain in response to compressive loading of zebrafish vertebrae. Three-dimensional results from MTS data follow distinct trends for each vertebrae between the three specimens (figure 2d,i,n), showing consistent dorsoventral compression, and lateral compression that is reversed at higher loads potentially due to elastic rebound of the IVD and fracturing along the zygopophyses that occurs at these loads (figure 2). This relative pattern is shared between each specimen, although specimen 3 experiences this at lower loads than specimens 1-2, before failing at 12 N. Fractures are observed where the arches and zygopophyses contact the centrum, at loads that precede the failure of the segment (figure 2f,h,k,m,p,r). Comparison with FEA data (blue points in figure 2*d*,*i*,*n*) suggests that the FE model accurately predicts these patterns (figure 2*d*,*i*,*n*), and that patterns of deformation could explain the first signs of damage prior to failure. In both datasets, the anterior vertebra undergoes most deformation, particularly posterior deformation of the arches (figure 2e-h). The central vertebrae and arches show strong torsion (figure 2j-m), increasing through the loading regime leading to the failure of the segment (figure 1*l*,*o*). The posterior vertebra shows the least deformation and is most isotropic in pattern (figure 20*r*), potentially due to protection offered by the anterior IVDs.

Comparison with *ex vivo* loading of vertebral MSs validates the accuracy of our FEA model for predicting patterns of deformation and strain across these structures. This offers a step towards a digital 'sandbox' approach to modelling the effects of genetic, physiological and morphological properties on the reaction and resistance of vertebral MSs to loading. Inputting specific properties of vertebral samples into a validated FE model will allow their effects on the biomechanics of the spine to be quantitatively tested in silico, allowing the relative contributions of shape and material properties to be explored and empirically tested. This will aid comparison of mechanical performance between different model systems. As an advantage of the zebrafish system is the wealth of mutants modelling human disease genetics [18], comparisons of mechanical performance between genotype and phenotype will be possible. In the longer term, this approach may give insight into biomechanical aspects of spinal pathology, allowing identification of 'at risk sites' in the spine. This could provide a basis for more specific or earlier interventions than those commonly employed.

Data accessibility. Models are available at: http://www.bristol.ac.uk/staff/researchers/data/

Authors' contributions. E.N., E.K. and J.A.A. performed experiments, E.N., E.K., J.A.A., C.F. and C.L.H. analysed data. The project was designed by C.L.H. and K.R.B. All authors contributed to drafting the manuscript.

Competing interests. The authors report no conflicts of interest.

Funding. E.N., E.K., C.L.H. and K.R.B. were funded by STFC grant no. ST/T000678/1 and C.L.H. and E.K. by Versus Arthritis Fellowship 21937 and project grant no. 21211.

Acknowledgements. We would like to thank Rob Harniman for the atomic force microscopy values for zebrafish bone and cartilage (acquired for another project).

# References

- Laporte S, Van den Abbeele M, Rohan PY, Adam C, Rouch P, Skalli W. 2017 Spine. In *Biomechanics of living organs: hyperelastic constitutive laws for finite element modeling* (eds Y Payan, J Ohayon). Amsterdam, The Netherlands: Elsevier.
- Oxland TR. 2016 Fundamental biomechanics of the spine—what we have learned in the past 25 years and future directions. *J. Biomech.* 49, 817–832. (doi:10.1016/j.jbiomech.2015.10.035)
- Bliuc D, Nguyen ND, Milch VE, Nguyen TV, Eisman JA, Center JR. 2009 Mortality risk associated with low-trauma osteoporotic fracture and subsequent fracture in men and women. J. Am. Med. Assoc. 301, 513–521. (doi:10.1001/jama.2009.50)
- Lambers FM, Kuhn G, Weigt C, Koch KM, Schulte FA, Müller R. 2015 Bone adaptation to cyclic loading in murine caudal vertebrae is maintained with age and directly correlated to the local micromechanical environment. *J. Biomech.* 48, 1179–1187. (doi:10.1016/j.jbiomech.2014.11.020)
- Fiaz AW, Leon-Kloosterziel KM, Gort G, Schulte-Merker S, van Leeuwen JL, Kranenbarg S. 2012 Swim-training changes the spatio-temporal dynamics of skeletogenesis in zebrafish larvae (*Danio rerio*). *PLoS ONE* 7, e34072. (doi:10.1371/ journal.pone.0034072)
- 6. Askary A, Smeeton J, Paul S, Schindler S, Braasch I, Ellis NA, Postlethwait J, Miller CT, Crump JG. 2016

Ancient origin of lubricated joints in bony vertebrates. *eLife* **5**, e16415. (doi:10.7554/elife. 16415)

- Jonas A, Buranova V, Scholz S, Fetter E, Novakova K, Kohoutek J, Hilscherova K. 2014 Retinoid-like activity and teratogenic effects of cyanobacterial exudates. *Aquat. Toxicol.* **155**, 283–290. (doi:10. 1016/j.aquatox.2014.06.022)
- Hayes AJ, Reynolds S, Nowell MA, Meakin LB, Habicher J, Ledin J, Bashford A, Caterson B, Hammond CL. 2013 Spinal deformity in aged zebrafish is accompanied by degenerative changes to their vertebrae that resemble osteoarthritis. *PLoS ONE* 8, e75787. (doi:10.1371/journal.pone.0075787)
- Cole C, Wolfla C, Pintar FA, Yoganandan N. 2012 Spine biomechanics. In *Orthopaedic biomechanics* (ed. BA Winkelstein). Boca Raton, FL: CRC Press.
- Bright JA, Rayfield EJ. 2011 Sensitivity and *ex vivo* validation of finite element models of the domestic pig cranium. *J. Anat.* **219**, 456–471. (doi:10.1111/j. 1469-7580.2011.01408.x)
- Stayton CT. 2018 Warped finite element models predict whole shell failure in turtle shells. *J. Anat.* 233, 666–678. (doi:10.1111/joa.12871)
- Brunt LH, Norton JL, Bright JA, Rayfield EJ, Hammond CL. 2015 Finite element modelling predicts changes in joint shape and cell behaviour due to loss of muscle strain in jaw development.

*J. Biomech.* **48**, 3112–3122. (doi:10.1016/j. jbiomech.2015.07.017)

- Lawrence EA, Kague E, Aggleton JA, Harniman RL, Roddy KA, Hammond CL. 2018 The mechanical impact of col11a2 loss on joints; col11a2 mutant zebrafish show changes to joint development and function, which leads to early-onset osteoarthritis. *Phil. Trans. R. Soc. B* **373**, 20170335. (doi:10.1098/rstb.2017.0335)
- Ofer L *et al.* 2019 A novel nonosteocytic regulatory mechanism of bone modeling. *PLoS Biol.* 17, e3000140. (doi:10.1371/journal.pbio.3000140)
- Panzer MB, Cronin DS. 2009 C4–C5 segment finite element model development, validation, and loadsharing investigation. *J. Biomech.* 42, 480–490. (doi:10.1016/j.jbiomech.2008.11.036)
- Kague E, Hughes SM, A. Lawrence E, Cross S, Martin-Silverstone E, Hammond CL, Hinits Y. 2019 Scleraxis genes are required for normal musculoskeletal development and for rib growth and mineralization in zebrafish. *FASEB J.* 33, 9116–9130. (doi:10.1096/fj.201802654RR)
- 17. Adams D, Collyer M, Kaliontzopoulou A. 2018 Package 'geomorph': geometric morphometric analyses of 2D/3D landmark data. CRAN.
- Witten PE, Harris MP, Huysseune A, Winkler C. 2017 Small teleost fish provide new insights into human skeletal diseases. *Methods Cell Biol.* **138**, 321–346. (doi:10.1016/bs.mcb.2016.09.001)