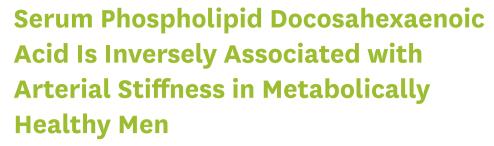


Original Article





Mi-Hyang Lee, 1,2,* Nayeon Kwon, 3,* So Ra Yoon, 4 Oh Yoen Kim^{2,4}

¹Korea Ginseng Corporation Research Institute, Korea Ginseng Corporation, Daejeon 34337, Korea ²Human Life Research Center, Dong-A University, Busan 49201, Korea

³Department of Food and Nutrition, Yonsei University, Seoul 03722, Korea

⁴Department of Food Science and Nutrition, Brain Busan 21, Dong-A University, Busan 49201, Korea



Received: Jul 13, 2016 Revised: Jul 27, 2016 Accepted: Jul 28, 2016

Correspondence to

Oh Yoen Kim

Department of Food Science and Nutrition, Dong-A University, 26 Daesingongwon-ro, Seogu, Busan 49201, Korea.

Tel: +82-51-200-7326 Fax: +82-51-200-7326 E-mail: oykim@dau.ac.kr

These two authors equally contributed to the work.

Copyright © 2016 The Korean Society of Clinical Nutrition

This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (http://creativecommons.org/licenses/by-nc/3.0/) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

ORCID

Oh Yoen Kim http://orcid.org/0000-0001-9262-3309

Funding

This study was supported by the National Research Foundation of Korea Grant funded by the Korean Government (2016R1A2B4013627) and fellowships for students were partly supported by the Brain Busan 21 project.

ABSTRACT

We hypothesized that lower proportion of serum phospholipid docosahexaenoic acid (DHA) is inversely associated with increased cardiovascular risk and vascular function in metabolically healthy men. To elucidate it, we first compared serum phospholipid free fatty acid (FA) compositions and cardiovascular risk parameters between healthy men (n = 499) and male patients with coronary artery disease (CAD, n = 111) (30-69 years) without metabolic syndrome, and then further-analyzed the association of serum phospholipid DHA composition with arterial stiffness expressed by brachial-ankle pulse wave velocity (ba-PWV) in metabolically healthy men. Basic parameters, lipid profiles, fasting glycemic status, adiponectin, high sensitivity C-reactive protein (hs-CRP) and LDL particle size, and serum phospholipid FA compositions were significantly different between the two subject groups. Serum phospholipid DHA was highly correlated with most of long-chain FAs, Metabolically healthy men were subdivided into tertile groups according to serum phospholipid DHA proportion: lower (< 2.061%), middle (2.061%-3.235%) and higher (> 3.235%). Fasting glucose, insulin resistance, hs-CRP and ba-PWVs were significantly higher and adiponectin and LDL particle size were significantly lower in the lower-DHA group than the higher-DHA group after adjusted for confounding factors. In metabolically healthy men, multiple stepwise regression analysis revealed that serum phospholipid DHA mainly contributed to arterial stiffness (β' -coefficients = -0.127, p = 0.006) together with age, systolic blood pressure, triglyceride (r = 0.548, p = 0.023). Lower proportion of serum phospholipid DHA was associated with increased cardiovascular risk and arterial stiffness in metabolically healthy men. It suggests that maintaining higher proportion of serum phospholipid DHA may be beneficial for reducing cardiovascular risk including arterial stiffness in metabolically healthy men.

Keywords: Serum phospholipid; Docosahexaenoic acid; Cardiovascular risk; Arterial stiffness; Metabolically healthy



Conflict of Interest

The authors have no potential conflicts of interest to disclose.

Author Contributions

All the authors were involved in the development of the study protocol and the experimental design. All the authors read, commented on, and contributed to the submitted manuscript. Sample collection and experiments were performed by Mi-Hyang Lee, Nayeon Kwon, So Ra Yoon and Oh Yoen Kim. Data were analyzed by Mi-Hyang Lee, Nayeon Kwon and Oh Yoen Kim. The manuscript was written by Mi-Hyang Lee, Nayeon Kwon and Oh Yoen Kim, and finalized by Oh Yoen Kim.

INTRODUCTION

Blood or dietary fatty acids (FAs) were associated with metabolic disorder such as metabolic syndrome (MS), diabetes and cardiovascular disease (CVD) [1-4]. The features of MS (i.e. insulin resistance, IR, dyslipidemia, smaller LDL particle size, decreased adiponectin and increased inflammation and oxidative stress) linked to cardiovascular risk were significantly related with serum phospholipid FA compositions and the severity of arterial stiffness [3-5]. Dietary n-3 long-chain polyunsaturated FAs (LC-PUFAs; i.e. eicosapentaenoic acid, EPA and docosahexaenoic acid, DHA) are associated with IR [6], blood pressure (BP) [7] and vascular aging [8]. Tomiyama et al. [8] reported that daily consumption of 1.8 g EPA for 12 months ameliorated arterial stiffness in patients with dyslipidemia. In our previous study, coronary artery disease (CAD) patients with MS show low proportion of PUFAs, particularly DHA in serum phospholipids [9] than the counterparts. Additional *in vitro* experiment shows that DHA treatment plays a protective role in LPS-induced vascular smooth muscle cell (VSMC) proliferation and inflammatory response by regulating ERK, p38 and NF-κB/p65 signaling pathways [9].

Numerous longitudinal and prospective studies reported the association of n-3 FAs intake with reduced CVD mortality [10]. A meta-analysis of randomized controlled trials also demonstrated that taking ≥ 3 g of n-3 LC-PUFAs/day by fish-oil supplements lowered BPs particularly in the middle aged and elderly people [11]. On the other hand, dietary intake of n-3 PUFAs ≤ 1 g/day for 12 months in healthy adults did not improve endothelial function expressed by brachial artery flow-mediated dilation (ba-FMD), arterial stiffness and BPs [12]. It was consistent with cross-sectional survey in 3,045 adults (45-84 years) [13] showing no significant relation between n-3 PUFAs intake with ba-FMD. Two more studies did not show any effects of n-3 PUFA injection (1.3-3.0 g/day for 3-12 weeks) on endothelial activation markers in young (≤ 45 years) [14] and middle-aged (50-70 years) [15] people.

However, the results are still controversial, and only a few studies have investigated the association of LC-PUFAs with cardiovascular risk parameters including vascular functions in healthy people. Therefore, this study aimed to investigate if serum phospholipid LC-PUFAs, particularly DHA are associated with the alteration of cardiovascular risk parameters and vascular function in metabolically healthy people. We hypothesized that lower proportions of LC-PUFAs particularly DHA in serum phospholipids are significantly associated with the increased risk of CVD and affect vascular function in metabolically healthy men. To elucidate it, we first compared basic and biochemical parameters related with cardiovascular risk and serum phospholipid FA compositions between healthy men and CAD men without MS, and then performed further analysis on the association of serum phospholipid DHA composition with cardiovascular risk and arterial stiffness in metabolically healthy men.

MATERIALS AND METHODS

Study population

Metabolically healthy men were recruited from the Health Service Center in the course of a routine checkup visit or by a newspaper announcement for health examinations and enrolled to the study (March 2013-March 2014). Those who were electrocardiographically normal were included. Exclusion criteria were orthopedic limitations, weight loss/gain over the previous 6 months, or any diagnosis of vascular disease, diabetes, cancer (clinically or by anamnesis),



renal disease, liver disease, thyroid disease, and acute or chronic inflammatory diseases. Subjects with MS were also excluded. MS was defined using a combination and modification of the NCEP-ATPIII guideline, Asian-Pacific guideline and American Diabetes Association guideline [16-18]. This definition requires at least three of the following components: waist circumference ≥ 90 cm (men); triglyceride ≥ 150 mg/dL; high density lipoprotein cholesterol (HDL-cholesterol) < 40 mg/dL (men); blood pressure ≥ 130 / ≥ 85 mm Hg; and fasting glucose ≥ 100 mg/dL (but fasting glucoses ≥ 126 mg/mL were considered diagnostic of diabetes). None of participants were taking any medications (antihypertensive, antidyslipidemic, antithrombotic, and antidiabetic drugs).

CAD patients were recruited from the Severance Cardiovascular Hospital, Seoul, Korea. The inclusion criteria were as follows: (1) angiographically confirmed CAD with ≥ 50% occlusion of one or more major coronary arteries, (2) myocardial infarction confirmed according to World Health Organization criteria for symptoms, enzyme elevation, or electrocardiographic changes, (3) absence of non-atherogenic occlusion, i.e. osteal stenosis and spasm, (4) neither orthopedic limitations nor any diagnosis of the diseases mentioned above (diabetes, cancer, and renal, liver, thyroid and acute or chronic inflammatory diseases). CAD patients with MS were also excluded.

Finally, 499 genetically unrelated healthy men and CAD male patients (n = 111) were included in this study (30–69 years). Participation rates in case and healthy subjects from the recruitment were 50% and 70%, respectively. Written informed consent was obtained from all participants, and the study protocol was approved by the Institutional Review Board of Yonsei University.

Anthropometric parameters and blood collection

Height, body weight and waist circumference were measured. Body mass index (BMI) was calculated as body weight/height (kg/m²). Blood pressure (BP) was obtained from the seated individuals with an automatic BP monitor (TM-2654, A&D, Tokyo, Japan) after 20 minutes of rest. Study participants were interviewed regarding their smoking and drinking behavior. After an overnight fast, venous blood specimens were collected in EDTA-treated and plain tubes. The tubes were immediately placed on ice until they arrived at the analytical laboratory (1-3 hours). Then, the blood specimens were separated into plasma or serum, and stored at -80°C until analysis.

Serum lipid profile and free fatty acids

Serum total cholesterol, triglyceride and free FAs (FFAs) were measured using commercially available kits on a Hitachi 7150 Autoanalyzer (Hitachi Ltd., Tokyo, Japan). After precipitation of serum chylomicron, LDL, and VLDL with dextran sulfatemagnesium, HDL-cholesterol left in the supernatant was measured by an enzymatic method. LDL-cholesterol was calculated indirectly using the Friedewald formula for individuals with serum triglyceride < 400 mg/dL (4.52 mol/L).

Glucose, insulin, and HOMA-IR

Fasting glucose was measured by a glucose oxidase method (Glucose Analyzer Beckman Instruments, Irvine, CA, USA). Insulin was measured by radioimmuno-assays with commercial kits (Immuno Nucleo Corporation, Stillwater, MN, USA). IR was calculated with the homeostasis model assessment using the following equation: IR = [fasting insulin (mIU/ mL) - fasting glucose (mmol/L)] / 22.5.



Liver and kidney functional markers

Serum aspartate aminotransferase (AST) and alanine aminotransferase (ALT) were measured using a modified International Federation of Clinical Chemistry (IFCC) UV method. Serum creatinine was measured with a kinetic colorimetric (Jaffe) assay. Serum blood urea nitrogen (BUN) and uric acid were measured using a kinetic UV Assay.

Plasma adiponectin and LDL particle size, and serum high sensitivity C-reactive protein

Plasma adiponectin concentrations were measured using an enzyme immunoassay (Human Adiponectin ELISA kit, B-Bridge International Inc., San Francisco, CA, USA; Mercodia, Uppsala, Sweden, respectively) and read by a Victor² (Perkin Elmer Life Sciences, Turku, Finland). Particle size distribution of LDL (density 1.019-1.063 g/mL) isolated by sequential flotation ultracentrifugation was examined using a pore-gradient lipoprotein system (CBS Scientific, San Diego, CA, USA). Serum hs-CRP was measured with an ADVIA 1650 (Bayer, Tarrytown, NY, USA) using a commercially available, high-sensitivity CRP-Latex (II) X2kit (Seiken Laboratories Ltd., Tokyo, Japan). The intra-assay and inter-assay coefficients of variance were 1.87% and 1.89%, respectively.

Fatty acid composition in serum phospholipids

Total lipids in serum were extracted with chloroform/methanol (2:1, v/v) as described by Folch et al. [19]. The phospholipid fraction was isolated from total lipids using thin-layer chromatography and then methylated during 1 hour at 95°C. FA methyl esters were measured with gas chromatography (Hewlett Packard 5890A, Palo Alto, CA, USA) using wax column (Omegawax 320, 30 m × 0.32 mm × 0.25 μm , Supelco, Bellefonte, PA, USA). Individual FAs were calculated as a relative percentage of the total of 26 FAs (set as 100%) using Chemstation software (Hewlett Packard).

Brachial-ankle pulse wave velocity

Brachial-ankle pulse wave velocity (ba-PWV) as a marker for arterial stiffness was measured using an automatic waveform analyzer (model VP-1000; Nippon Colin Ltd., Komaki, Japan). The detailed methods were described previously [20]. The average baPWV from both left and right sides were used in the analysis (correlation between the right and left baPWVs: $r^2 = 0.925$, p < 0.0001).

The assessment of dietary intake and physical activity level

Information on each participant's usual diet was obtained using both a 24-hour recall and a semi-quantitative food frequency questionnaire (SQFFQ), of which the validity had been previously tested [21]. We used the former to carry out analyses and the latter to check if the collected data was representative of the usual dietary pattern. All participants were given written and verbal instructions by a registered dietitian on completion of a 3-day (2 week days and 1 weekend) dietary record. Dietary energy values and nutrient content from the 3-day food records were calculated using the Computer Aided Nutritional Analysis Program (CAN-pro 3.0, Korean Nutrition Society, Seoul, Korea). Total energy expenditure (TEE) (kcal/day) was calculated from activity patterns (basal metabolic rate, 24 hour-physical activity, and specific dynamic action of food).

Statistical analysis

Statistical analyses were performed with Win SPSS ver. 20.0 (Statistical Package for the Social Science, SPSS Inc., Chicago, IL, USA). Differences in continuous variables among subgroups



were tested with independent t-test or one-way ANOVA followed by Bonferroni correction for multiple comparisons to reduce the rate of false positive, and non-continuous variables were tested with χ^2 test. A general linear model analysis (GLM, post-hoc multiple comparison tests) followed by Bonferroni correction was also performed to see the differences in the cardiovascular risk parameters according to major serum phospholipid FA composition with adjustment for confounding factors (i.e. age, anthropometric parameters, cigarette smoking, alcohol consumption, dietary intake etc.). Pearson correlation analysis was performed for the relationship among the variables. Multiple regression analysis was used to find the major determinants for arterial stiffness in metabolically healthy men. The skewed variables were log-transformed for statistical analysis. For descriptive purposes, the mean values are presented using untransformed values. Results are expressed as mean \pm SE or percentages. A two-tailed value of p < 0.05 was considered statistically significant.

RESULTS

General characteristics and basic biochemical parameters of study population

Table 1 presents demographic and metabolic characteristics of study population. Mean values of age, BMI, waist circumference, systolic BP, proportions of current smokers and current drinkers were significantly different between healthy and CAD patients. Regarding medications, most of CAD patients consumed anticoagulation drugs (99.1%) and antihypertensive drugs (92.7%). 70.9% of CAD patients consumed lipid-lowering drugs. None of them consumed antidiabetic drugs. Serum lipid profiles (triglyceride, HDLcholesterol, total cholesterol, and LDL-cholesterol) were significantly different between the two subject groups. CAD men show significantly higher levels of insulin and HOMA-IR and lower levels of glucose and FFAs than healthy men, but all the values were in normal range. Serum creatine levels were statistically lower in CAD men than in healthy men, but the levels in both groups are in normal range. In addition, the values of another kidney function (BUN) and liver function (AST, ALT) markers were not significantly different between the two subject groups, and all the values are in normal range. Compared with CAD men, healthy men showed higher ratio of TEE to total calorie intake (TEE/TCI) and consumed higher percent energy intake from total fats and lower percent energy intake from total carbohydrates and proteins, and similar dietary polyunsaturated FAs to saturated FAs ratio (PUFA/SFA) (Table 1).

Differences in the proportions of serum phospholipid FAs between healthy men and CAD patients

Figure 1 and **Table 2** present serum phospholipid FA composition between healthy men and CAD patients. The values were adjusted for age, waist circumference, cigarette smoking, alcohol consumption, systolic blood pressure, HDL-cholesterol, TEE/TCI, dietary carbohydrates (% of total calorie intake), dietary fats (% of total calorie intake). The proportions of linoleic acid (LA, 18:2n-6) and DHA (22:6n-3) were significantly lower and that of dihomo-γ-linolenic acid (DGLA, 20:3n-6) and the ratio of n-6 PUFAs to n-3 PUFAs were significantly higher in CAD men than in healthy men (**Figure 1**). The proportions of myristic acid (14:0), palmitic acid (PA, 16:0), stearic acid (SA, 18:0), total monounsaturated FA, oleic acid (18:1n-9), eicosadienoic acid (20:2n-6), arachidonic acid (AA, 20:4n-6), adrenic acid (22:4n-6), docosapentaenoic aicd (20:5n-6) were significantly different between healthy and CAD men (**Table 2**).



Table 1. Demographic and metabolic parameters of study population

Variables	Healthy (n = 499)	CAD (n = 111)	р
Age, yr	48.3 ± 0.43	54.2 ± 0.91	< 0.001
Body mass index, kg/m ²	23.8 ± 0.11	24.6 ± 0.26	0.003
Naist, cm	83.7 ± 0.29	89.6 ± 0.71	< 0.001
Current smokers, %	37.1	23.4	0.006
Current drinkers, %	83.6	59.1	< 0.001
Systolic BP, mmHg	120.8 ± 0.62	126.5 ± 1.38	< 0.001
Diastolic BP, mmHg	75.4 ± 0.47	74.6 ± 1.00	0.445
riglyceride, mg/dL*	118.3 ± 2.66	140.5 ± 9.14	0.006
otal cholesterol, mg/dL	190.9 ± 1.47	149.5 ± 3.60	< 0.001
IDL-cholesterol, mg/dL*	52.3 ± 0.58	45.6 ± 0.95	< 0.001
.DL-cholesterol, mg/dL	115.0 ± 1.42	76.5 ± 3.08	< 0.001
Glucose, mg/dL [*]	91.2 ± 0.46	87.1 ± 1.12	0.001
Creatine, mg/dL	0.93 ± 0.01	0.85 ± 0.02	< 0.001
BUN, mg/dL	14.5 ± 0.17	14.9 ± 0.43	0.258
AST, U/L	24.6 ± 0.37	23.8 ± 0.67	0.317
ALT, U/L	23.1 ± 0.61	19.2 ± 1.04	0.005
nsulin, μIU/mL*	8.18 ± 0.15	9.70 ± 0.70	0.009
ree fatty acid, µEq/L*	493.2 ± 10.4	379.7 ± 16.4	< 0.001
IOMA-IR*	1.85 ± 0.04	2.14 ± 0.20	0.016
s-CRP, mg/dL	1.46 ± 0.13	2.04 ± 0.37	0.004
DL particle size, nm	23.8 ± 0.03	23.5 ± 0.05	< 0.001
diponectin, μg/dL	5.31 ± 0.12	4.12 ± 0.25	< 0.001
Dietary intake and energy expenditure			
TEE, kcal	2323.6 ± 10.1	2317.2 ± 24.0	0.805
TCI, kcal	2399.0 ± 10.1	2439.6 ± 29.2	0.191
TEE/TCI, kcal	0.97 ± 0.00	0.95 ± 0.00	0.001
Carbohydrate (% of TCI)	61.8 ± 0.07	63.0 ± 0.39	0.002
Protein (% of TCI)	16.9 ± 0.06	17.3 ± 0.17	0.024
Fat (% of TCI)	21.5 ± 0.07	20.4 ± 0.21	< 0.001
PUFA/SFA, g	1.41 ± 0.03	1.49 ± 0.06	0.172
Medications, %			
Anticoagulation	-	99.1	
Antihypertensive	-	92.7	
Lipid-lowering	-	70.9	
Antidiabetic	-	-	
Mean ± S.E or %.			

ALT, alanine aminotransferase; AST, aspartate aminotransferase; BP, blood pressure; BUN, blood urea nitrogen; HOMA-IR, homeostasis model assessment of insulin resistance; hs-CRP, high sensitivity C-reactive protein; TEE, total energy expenditure; TCI, total caloric intake; PUFA, polyunsaturated fatty acid; SFA, saturated fatty acid.

Correlations among serum phospholipid FA composition

Pearson correlation analysis revealed that DHA (C22:6n-3) was highly correlated with many of LC-FAs in serum phospholipid FA composition in all the subject groups (whole subjects, healthy men and CAD patients respectively): positively correlated with α -linolenic acid (ALNA, 18:3n-3, r = 0.138, p = 0.001; r = 0.102, p = 0.041; r = 0.369, p < 0.001, respectively), EPA (20:5n-3, r = 0.525, p < 0.0001; r = 0.434, p < 0.001; r = 0.830, p < 0.001, respectively), LA (18:2n-6, r = 0.448, p < 0.0001; r = 0.479, p < 0.001; r = 0.317, p = 0.001, respectively), γ -linolenic acid (GLA, 18:3n-6, r = 0.191, p < 0.0001; r = 0.236, p < 0.001; r = 0.140, p = 0.042, respectively), DGLA (20:3n-6, r = 0.498, p < 0.0001; r = 0.537, p < 0.001; r = 0.361, p < 0.001, respectively), AA (20:4n-6, r = 0.684, p < 0.0001; r = 0.716, p < 0.001; r = 0.621, p < 0.001, respectively) and docosapentaenoic acid (DPA, 22:5n-6, r = 0.220, p < 0.0001; r = 0.214, p < 0.0000.001; r = 0.320, p = 0.001, respectively), and negatively correlated with n-6 PUFAs/n-3 PUFAs (r = -0.716, p < 0.001; r = -0.695, p < 0.001; r = -0.851, p < 0.001, respectively).

^{&#}x27;Tested after log transformed; tested by independent t-test (student t-test).



Table 2. Differences in the proportions of serum phospholipid fatty acids between healthy men and CAD patients

Variables	Healthy (n = 499)	CAD (n = 111)	р
Total SFA	54.420 ± 0.266	54.160 ± 0.578	0.689
12:0, Lauric acid	0.389 ± 0.012	0.404 ± 0.026	0.643
14:0, Myristic acid*	0.651 ± 0.039	0.432 ± 0.084	0.024
16:0, Palmitic acid	32.270 ± 0.245	33.834 ± 0.532	0.011
18:0, Stearic acid [*]	18.740 ± 0.162	17.230 ± 0.353	< 0.001
Total MUFA*	11.340 ± 0.109	11.800 ± 0.237	0.048
16:1, Palmitoleic acid	0.672 ± 0.032	0.599 ± 0.070	0.373
18:1 n-9, Oleic acid	6.811 ± 0.082	7.345 ± 0.178	0.010
18:1 n-7, Vaccenic acid	1.721 ± 0.021	1.776 ± 0.046	0.297
Total PUFA	25.140 ± 0.300	24.930 ± 0.651	0.783
Total n-6 FA	20.250 ± 0.229	20.220 ± 0.498	0.946
18:3 n-6, γ-linolenic acid [*]	0.221 ± 0.012	0.187 ± 0.025	0.246
20:2 n-6, Eicosadienoic acid'	0.861 ± 0.076	0.345 ± 0.166	0.007
20:4 n-6, Arachidonic acid	4.717 ± 0.094	5.503 ± 0.205	0.001
22:4 n-6, Adrenic acid	0.242 ± 0.013	0.092 ± 0.028	< 0.001
22:5 n-6, Docosapentaenoic acid	0.209 ± 0.010	0.100 ± 0.023	< 0.001
Total n-3 FA [*]	4.882 ± 0.110	4.713 ± 0.238	0.542
18:3 n-3, α-Linolenic acid [*]	0.157 ± 0.018	0.131 ± 0.038	0.555
20:3 n-3, Eicosatrienoic acid	0.078 ± 0.006	0.053 ± 0.012	0.076
20:5 n-3, Eicosapentaenoic acid*	1.185 ± 0.040	1.336 ± 0.086	0.132
22:5 n-3, Docosapentaenoic acid	0.538 ± 0.016	0.519 ± 0.035	0.638

Mean ± S.E.

p, p-value adjusted for age, waist circumference, cigarette smoking, alcohol consumption, systolic blood pressure, HDL-cholesterol, total energy expenditure/total calorie intake, dietary carbohydrate (% of total calorie intake), dietary fat (% of total calorie intake); FA, fatty acid; SFA, saturated fatty acid; MUFA, monounsturated fatty acid; PUFA, polyunsaturated fatty acid.

Tested after log transformed; tested by general linear model (GLM) followed by bonferroni correction with adjustment.

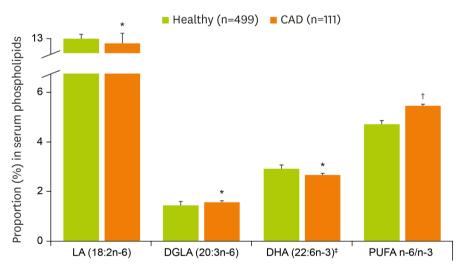


Figure 1. Proportions of serum phospholipid fatty acids between healthy men and CAD patients. Mean ± S.E.

LA, linoleic acid; DGLA, dihomo- γ -linolenic acid; DHA, docosahexaenoic acid; PUFA, polyunsaturated fatty acid. $\dot{p} < 0.05 \,^{\dagger}p < 0.001$ tested by general linear model (GLM) with adjustment for age, waist circumference, cigarette smoking, alcohol consumption, systolic blood pressure, HDL-cholesterol, total energy expenditure/total calorie intake, dietary carbohydrate (% of total calorie intake), dietary fat (% of total calorie intake); $\dot{\tau}$ Tested after log transformed.



Table 3. General characteristics and biochemical parameters according to the proportion (%) of DHA in serum phospholipids in healthy men

Variables	Lower DHA (n = 168) DHA (%) ≤ 2.061	Middle DHA (n = 163) 2.061 < DHA (%) ≤ 3.235	Higher DHA (n = 168) 3.235 < DHA (%)
DHA (%) in serum phospholipids*	1.41 ± 0.04§	$2.65 \pm 0.03^{\ddagger}$	$4.19 \pm 0.09^{\dagger}$
Age, yr	47.8 ± 0.75 ^{†,‡}	$46.9 \pm 0.80^{\ddagger}$	$49.9 \pm 0.72^{+}$
Body mass index, kg/m²	23.6 ± 0.18	23.8 ± 0.22	23.9 ± 0.19
Weight, kg	67.8 ± 0.63	68.7 ± 0.75	68.2 ± 0.67
Waist, cm	83.5 ± 0.50	83.8 ± 0.58	83.7 ± 0.49
Current smokers, %	45.1	38.1	32.9
Current drinkers, %	83.0	86.5	80.9
Systolic blood pressure, mmHg	121.3 ± 1.23	119.8 ± 1.05	120.5 ± 1.12
Diastolic blood pressure, mmHg	74.8 ± 0.87	74.3 ± 0.86	75.6 ± 0.83
Triglyceride, mg/dL	125.1 ± 5.73	114.3 ± 4.33	116.7 ± 4.32
Free fatty acid, µEq/L°	504.0 ± 18.8	499.6 ± 17.9	503.0 ± 19.3

Mean \pm S.E or %.

The association of serum phospholipid DHA proportion with cardiovascular risk related parameters and arterial stiffness in metabolically healthy men

In order to investigate the relationship between serum phospholipid DHA and cardiovascular risk parameters including ba-PWV, healthy men were subdivided into three tertile groups according to the proportion (%) of DHA in serum phospholipids [lower-DHA group: DHA (%) \leq 2.061; middle-DHA group: 2.061 < DHA (%) \leq 3.235; and higher-DHA group: 3.235 < DHA (%)]. There were no significant differences in general and basic biochemical parameters except age among the DHA tertile groups (**Table 3**). After adjusted for age, cigarette smoking, alcohol consumption, and dietary intake, fasting glycemic index (glucose and HOMA-IR) were significantly higher in the lower-DHA group than in the higher-DHA group. On the other hand, plasma adiponectin levels were significantly lower in the lower-DHA group than in the higher-DHA group. Hs-CRP and ba-PWVs were significantly higher, but LDL particle size was lower in the lower-DHA group than in the higher-DHA group (**Figure 2**).

Association of DHA with arterial stiffness by multiple regression analysis

A multiple stepwise regression analysis was performed to identify the association of serum phospholipid DHA composition with arterial stiffness in metabolically healthy men. Input factors were age, waist circumference, BPs, triglyceride, HDL-cholesterol, LDL-cholesterol, HOMA-IR, LDL particle size, hs-CRP, plasma adiponectin, serum phospholipid DHA and basic parameters such as TEE/TCI, cigarette smoking and alcohol consumption which may be closely related with MS and CAD. Among the factors, serum phospholipid DHA was found one of main contributors to arterial stiffness [adjusted β -coefficients (β ') = -0.127, p = 0.006] together with age (β ' = 0.315, p < 0.001), systolic BP (β ' = 0.394, p < 0.001), triglyceride (β ' = 0.105, p = 0.023) in metabolically healthy men (r = 0.548, p = 0.023).

DISCUSSION

This study presents that serum phospholipid DHA was significantly associated with the alterations of MS and cardiovascular risk related parameters, and arterial stiffness in metabolically healthy men. In our study, fasting glucose, HOMA-IR, hs-CRP and ba-PWVs were significantly higher and adiponectin and LDL particle size were significantly lower in the lower-DHA group than the higher-DHA group. Multiple stepwise regression analysis also revealed that serum phospholipid DHA was a protective contributor against arterial

HOMA-IR, Homeostasis model assessment of insulin resistance; DHA, docosahexaenoic acid.

^{&#}x27;Tested after log transformed; tested by one-way analysis of variance (ANOVA) with bonferroni correction; †.‡.*Sharing the same mark indicates no significant difference in each variable in the same row.



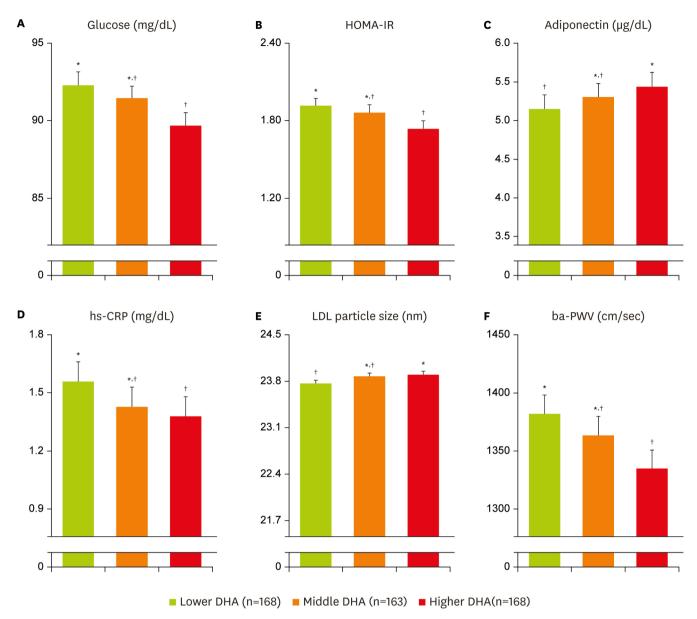


Figure 2. Association of serum phospholipid DHA proportion with cardiovascular risk related parameters and arterial stiffness in healthy men. (A) Glucose (mg/dL); (B) HOMA-IR; (C) Adiponectin (μg/dL); (D) hs-CRP (mg/dL); (E) LDL particle size (nm); (F) ba-PWV (cm/sec); (A-D, F) Tested after log transformed; Tested by general linear model (GLM) analysis followed by Bonferroni correction with adjustment for age, cigarette smoking, alcohol consumption, dietary intake.

Mean ± S.E.

DHA, docosahexaenoic acid; HOMA-IR, homeostasis model assessment of insulin resistance; hs-CRP, high sensitivity C-reactive protein; ba-PWV, brachial-ankle pulse wave velocity.

'. 'Variables marked with the same mark indicates no significant difference. Healthy men were subdivided into three tertile groups according to the proportion (%) of DHA in serum phospholipids [lower DHA group: DHA (%) ≤ 2.061; middle DHA group: 2.061 < DHA (%) ≤ 3.235; and higher DHA group: 3.235 < DHA (%)].

stiffness. These results may give evidences that serum phospholipid DHA plays a critical role in cardiometabolism and is inversely associated with arterial stiffness, a composite risk factor for early atherosclerosis [22,23] in metabolically healthy people.

Jump et al. [24] reported that over 290 clinical and experimental studies have examined the impact of LC-PUFAs on cardiovascular risk (n = 110) including MS (n = 16), diabetes and obesity (n = 12), inflammation (n = 97), dyslipidemia (n = 55), and hypertension (n = 97)



10). According to Vessby et al., [2] FA compositions in blood can be a marker for dietary fat composition and are associated with metabolic disorders: decreased LA and increased DGLA in blood phospholipids were associated with IR. In our current study, CAD patients showed lower proportions of LA, DHA and DPA, and higher proportions of DGLA and n-6/n-3 PUFAs than healthy men. The significances were maintained after the adjustment. Our previous study [3] also shows that serum phosopholipid FA compositions were significantly related to features of MS. Dietary n-3 PUFAs and fish consumption were reported to help depression of heart rate and reduction of sudden cardiac death in humans [25,26], and to induce several genes related to cardiac cells which encode proteins involved in lipid transport and the metabolism [27,28]. It may suggest that dietary n-3 PUFAs play a pathophysiologic role in the cardiometabolism including MS and myocardial dysfunctions [29].

Among the LC-PUFAs, DHA have been mainly investigated in the human and animal studies. Consumption of 0.7 g DHA/day for 3 months significantly increased DHA proportion in erythrocytes and reduced diastolic BP in middle-aged healthy subjects, but did not alter other inflammation and vascular function parameters [6]. In our previous study, DHA treatment significantly suppressed LPS-induced VSMC proliferation and inflammation by up-regulating the phosphorylation of ERK and p38, and inhibiting the translocation of NF-kB p65 from cytoplasm to nucleus in VSMCs [8]. In the spontaneously hypertensive rats, DHA supplementation significantly reduced BPs and vascular wall thicknesses of the coronary, thoracic, and abdominal aorta than control rats [30]. These results may support that DHA plays critical roles in cardiometabolism and has cardiovascular protective effect. However, most studies were performed in CAD patients or people with history and risk factor of CVD, and in the disease-based animal models. Only a few studies including cohort studies were done in healthy people.

The Atherosclerosis Risk in Communities Study performed in 1,908 middle aged healthy women reported that higher plasma phospholipid DHA can reduce the risk of heart failure [31]. Another cohort study performed in men aged 42 and more years old showed that serum DHA was associated with the risk of atrial fibrillation, a common cardiac arrhythmia [32]. Lindqvist et al. [33] showed that plasma phospholipid EPA was positively associated with HDL-cholesterol, total cholesterol, remained after adjustment for BP, and DHA was negatively related to serum endothelial markers (E-selectin, P-selectin), and positively with systolic BP. Recently, n-3 PUFA supplementation (4 g/day for 12 weeks) significantly increased EPA and DHA contents in erythrocytes and decreased blood triglyceride and carotid-femoral PWV in the healthy elderly, but the subjects numbers were so small (n = 12) [34]. In addition, fish consumption and coronary calcification had a weak inverse association in healthy women, but it did not reach the statistical significance [35]. Our current study investigated in metabolically healthy men (n = 499) showed that hs-CRP, and ba-PWV were significantly higher, and plasma adiponectin and LDL particle size were lower in the lower-DHA group than the higher-DHA group. These significances were still maintained after adjusted for confounding factors. These results may be explained that higher proportion of DHA in serum phospholipids suppressed the mRNA expression and production of several pro-inflammatory cytokines [36] which are closely associated with cardiometabolic risk parameters [4,9]. Many studies have reported the relationship between n3 LC-PUFAs, particularly DHA and glycemic status [6,37-40]. Minihane et al. [37] reported that diet containing high n6/ n3 PUFAs significantly increased fasting insulin and HOMA-IR. Plasma DHA is negatively associated with HOMA-IR [38,39] and effective on the prevention and treatment of IR in CAD patients with or without MS [38]. Recent animal study also showed that DHA consumption



significantly reduced plasma glucose level and restored insulin sensitivity in skeletal muscle [40]. It was reported that DHA together with EPA altered membrane fluidity and interacted with peroxisome proliferator-activated receptor-gamma which regulates insulin sensitivity regulator, thereby improving glycemic status [41]. In our study, metabolically healthy men with higher DHA proportion showed lower levels of fasting glucose, insulin and HOMA-IR indicating fasting glycemic status than those with lower DHA proportion. From the previous and our current results, we may suggest that higher DHA in serum phospholipids has a beneficial effect on cardiometabolism and may reduce the cardiovascular risk even in metabolically healthy people. One limitation of the current study is that participants were only men aged 30-69, so the result may not be suitable for women. Future study needs to be done in both men and women.

CONCLUSION

This study shows that lower proportion of DHA in serum phospholipids may be associated with increased cardiovascular risk and positively related with arterial stiffness in metabolically healthy men. It suggests that maintaining higher proportion of serum phospholipid DHA may be beneficial for reducing cardiovascular risk including arterial stiffness in healthy people.

ACKNOWLEDGEMENTS

We thank the research volunteers who participated in the study described in this manuscript.

REFERENCES

- Hooper L, Thompson RL, Harrison RA, Summerbell CD, Ness AR, Moore HJ, Worthington HV, Durrington PN, Higgins JP, Capps NE, Riemersma RA, Ebrahim SB, Davey Smith G. Risks and benefits of omega 3 fats for mortality, cardiovascular disease, and cancer: systematic review. BMJ 2006;332:752-60.
 PUBMED | CROSSREF
- 2. Vessby B. Dietary fat, fatty acid composition in plasma and the metabolic syndrome. Curr Opin Lipidol 2003;14:15-9.
 - PUBMED | CROSSREF
- Kim OY, Lim HH, Lee MJ, Kim JY, Lee JH. Association of fatty acid composition in serum phospholipids with metabolic syndrome and arterial stiffness. Nutr Metab Cardiovasc Dis 2013;23:366-74.
 PUBMED L CROSSREF
- Cho JS, Baek SH, Kim JY, Lee JH, Kim OY. Serum phospholipid monounsaturated fatty acid composition and Δ-9-desaturase activity are associated with early alteration of fasting glycemic status. Nutr Res 2014;34:733-41.
 - PUBMED | CROSSREF
- 5. Kim OY, Paik JK, Lee JY, Lee SH, Lee JH. Follow-ups of metabolic, inflammatory and oxidative stress markers, and brachial-ankle pulse wave velocity in middle-aged subjects without metabolic syndrome. Clin Exp Hypertens 2013;35:382-8.
 - PUBMED | CROSSREF
- Robbez Masson V, Lucas A, Gueugneau AM, Macaire JP, Paul JL, Grynberg A, Rousseau D. Long-chain (n-3) polyunsaturated fatty acids prevent metabolic and vascular disorders in fructose-fed rats. J Nutr 2008;138:1915-22.
 - PUBMED
- Theobald HE, Goodall AH, Sattar N, Talbot DC, Chowienczyk PJ, Sanders TA. Low-dose docosahexaenoic acid lowers diastolic blood pressure in middle-aged men and women. J Nutr 2007;137:973-8.
 PUBMED



8. Tomiyama H, Takazawa K, Osa S, Hirose K, Hirai A, Iketani T, Monden M, Sanoyama K, Yamashina A. Do eicosapentaenoic acid supplements attenuate age-related increases in arterial stiffness in patients with dyslipidemia?: a preliminary study. Hypertens Res 2005;28:651-5.

PUBMED | CROSSREF

9. Song J, Kwon N, Lee MH, Ko YG, Lee JH, Kim OY. Association of serum phospholipid PUFAs with cardiometabolic risk: beneficial effect of DHA on the suppression of vascular proliferation/inflammation. Clin Biochem 2014;47:361-8.

PUBMED | CROSSREF

- Harris WS, Kris-Etherton PM, Harris KA. Intakes of long-chain omega-3 fatty acid associated with reduced risk for death from coronary heart disease in healthy adults. Curr Atheroscler Rep 2008;10:503-9.

 PUBMED | CROSSREF
- 11. Geleijnse JM, Giltay EJ, Grobbee DE, Donders AR, Kok FJ. Blood pressure response to fish oil supplementation: metaregression analysis of randomized trials. J Hypertens 2002;20:1493-9.
- 12. Sanders TA, Hall WL, Maniou Z, Lewis F, Seed PT, Chowienczyk PJ. Effect of low doses of long-chain n-3 PUFAs on endothelial function and arterial stiffness: a randomized controlled trial. Am J Clin Nutr 2011;94:973-80.

PUBMED | CROSSREF

13. Anderson JS, Nettleton JA, Herrington DM, Johnson WC, Tsai MY, Siscovick D. Relation of omega-3 fatty acid and dietary fish intake with brachial artery flow-mediated vasodilation in the Multi-Ethnic Study of Atherosclerosis. Am J Clin Nutr 2010;92:1204-13.

PUBMED | CROSSREF

- Egert S, Rassoul F, Boesch-Saadatmandi C, Richter V, Rimbach G, Erbersdobler H, Somoza V, Wahrburg U. Effects of controlled diets enriched with alpha-linolenic acid, eicosapentaenoic acid or docosahexaenoic acid on soluble adhesion molecules and endothelin-1 concentrations in healthy volunteers. Curr Top Nutraceutical Res 2007;5:189-96.
- Pot GK, Brouwer IA, Enneman A, Rijkers GT, Kampman E, Geelen A. No effect of fish oil supplementation on serum inflammatory markers and their interrelationships: a randomized controlled trial in healthy, middle-aged individuals. Eur J Clin Nutr 2009;63:1353-9.
 PUBMED | CROSSREF
- 16. National Cholesterol Education Program (NCEP) Expert Panel on Detection, Evaluation, and Treatment of High Blood Cholesterol in Adults (Adult Treatment Panel III). National Cholesterol Education Program (NCEP) Expert Panel on Detection, Evaluation, and Treatment of High Blood Cholesterol in Adults (Adult Treatment Panel III). Third report of the national cholesterol education program (NCEP) expert panel on detection, evaluation, and treatment of high blood cholesterol in Adults (Adult Treatment Panel III) final report. Circulation 2002;106:3143-421.
- 17. World Health Organization Western Pacific Region; IASO International Association for the Study of Obesity; International Obesity TaskForce. The Asian Pacific perspective: redefining obesity and its treatment. Sydney: Health Communications Australia Pty Limited on behalf of the Steering Committee; 2000.
- American Diabetes Association. Standards of medical care in diabetes--2010. Diabetes Care 2010;33 Suppl 1:S11-61.
- 19. Folch J, Lees M, Sloane Stanley GH. A simple method for the isolation and purification of total lipides from animal tissues. J Biol Chem 1957;226:497-509.

PUBMED

PUBMED | CROSSREF

PUBMED

- 20. Kim OY, Yoe HY, Kim HJ, Park JY, Kim JY, Lee SH, Lee JH, Lee KP, Jang Y, Lee JH. Independent inverse relationship between serum lycopene concentration and arterial stiffness. Atherosclerosis 2010;208:581-6. PUBMED | CROSSREF
- 21. Shim JS, Oh KW, Suh I, Kim MY, Sohn CY, Lee EJ, Nam CM. A study on validity of a semi-quantitative food frequency questionnaire for Korean adults. Korean J Community Nutr 2002;7:484-94.
- Laurent S, Boutouyrie P, Asmar R, Gautier I, Laloux B, Guize L, Ducimetiere P, Benetos A. Aortic stiffness is an independent predictor of all-cause and cardiovascular mortality in hypertensive patients.
 Hypertension 2001;37:1236-41.

 PUBMED | CROSSREF
- 23. Matsuoka O, Otsuka K, Murakami S, Hotta N, Yamanaka G, Kubo Y, Yamanaka T, Shinagawa M, Nunoda S, Nishimura Y, Shibata K, Saitoh H, Nishinaga M, Ishine M, Wada T, Okumiya K, Matsubayashi K, Yano S, Ichihara K, Cornélissen G, Halberg F, Ozawa T. Arterial stiffness independently predicts cardiovascular events in an elderly community -- Longitudinal Investigation for the Longevity and Aging in Hokkaido County (LILAC) study. Biomed Pharmacother 2005;59 Suppl 1:S40-4.

PUBMED | CROSSREF



 Jump DB, Depner CM, Tripathy S. Omega-3 fatty acid supplementation and cardiovascular disease. J Lipid Res 2012;53:2525-45.

PUBMED | CROSSREF

 Mozaffarian D, Prineas RJ, Stein PK, Siscovick DS. Dietary fish and n-3 fatty acid intake and cardiac electrocardiographic parameters in humans. J Am Coll Cardiol 2006;48:478-84.

- Leaf A, Kang JX, Xiao YF, Billman GE. Clinical prevention of sudden cardiac death by n-3 polyunsaturated fatty acids and mechanism of prevention of arrhythmias by n-3 fish oils. Circulation 2003;107:2646-52.
 PUBMED | CROSSREF
- 27. Goua M, Mulgrew S, Frank J, Rees D, Sneddon AA, Wahle KW. Regulation of adhesion molecule expression in human endothelial and smooth muscle cells by omega-3 fatty acids and conjugated linoleic acids: involvement of the transcription factor NF-kappaB? Prostaglandins Leukot Essent Fatty Acids 2008;78:33-43.

PUBMED | CROSSREF

- Massaro M, Scoditti E, Carluccio MA, De Caterina R. Basic mechanisms behind the effects of n-3 fatty acids on cardiovascular disease. Prostaglandins Leukot Essent Fatty Acids 2008;79:109-15.
- Bordoni A, Astolfi A, Morandi L, Pession A, Danesi F, Di Nunzio M, Franzoni M, Biagi P, Pession A. N-3
 PUFAs modulate global gene expression profile in cultured rat cardiomyocytes. Implications in cardiac
 hypertrophy and heart failure. FEBS Lett 2007;581:923-9.
 PUBMED | CROSSREF
- Engler MM, Engler MB, Pierson DM, Molteni LB, Molteni A. Effects of docosahexaenoic acid on vascular pathology and reactivity in hypertension. Exp Biol Med (Maywood) 2003;228:299-307.
- Yamagishi K, Nettleton JA, Folsom ARARIC Study Investigators. Plasma fatty acid composition and incident heart failure in middle-aged adults: the Atherosclerosis Risk in Communities (ARIC) Study. Am Heart J 2008:156:965-74.

PUBMED I CROSSREF

- Virtanen JK, Mursu J, Voutilainen S, Tuomainen TP. Serum long-chain n-3 polyunsaturated fatty acids and risk of hospital diagnosis of atrial fibrillation in men. Circulation 2009;120:2315-21.

 PURMED L CROSSREE
- 33. Lindqvist HM, Sandberg AS, Fagerberg B, Hulthe J. Plasma phospholipid EPA and DHA in relation to atherosclerosis in 61-year-old men. Atherosclerosis 2009;205:574-8.

 PUBMED | CROSSREF
- Monahan KD, Feehan RP, Blaha C, McLaughlin DJ. Effect of omega-3 polyunsaturated fatty acid supplementation on central arterial stiffness and arterial wave reflections in young and older healthy adults. Physiol Rep 2015;3:e12438.
 PUBMED | CROSSREF
- 35. Heine-Bröring RC, Brouwer IA, Proença RV, van Rooij FJ, Hofman A, Oudkerk M, Witteman JC, Geleijnse JM. Intake of fish and marine n-3 fatty acids in relation to coronary calcification: the Rotterdam Study. Am J Clin Nutr 2010;91:1317-23.

PUBMED | CROSSREF

PUBMED | CROSSREF

PUBMED | CROSSREF

- 36. Weldon SM, Mullen AC, Loscher CE, Hurley LA, Roche HM. Docosahexaenoic acid induces an antiinflammatory profile in lipopolysaccharide-stimulated human THP-1 macrophages more effectively than eicosapentaenoic acid. J Nutr Biochem 2007;18:250-8.
- 37. Minihane AM, Brady LM, Lovegrove SS, Lesauvage SV, Williams CM, Lovegrove JA. Lack of effect of dietary n-6:n-3 PUFA ratio on plasma lipids and markers of insulin responses in Indian Asians living in the UK. Eur J Nutr 2005;44:26-32.

 PUBMED | CROSSREF
- 38. Nigam A, Frasure-Smith N, Lespérance F, Julien P. Relationship between n-3 and n-6 plasma fatty acid levels and insulin resistance in coronary patients with and without metabolic syndrome. Nutr Metab Cardiovasc Dis 2009;19:264-70.
- 39. Damsgaard CT, Eidner MB, Stark KD, Hjorth MF, Sjödin A, Andersen MR, Andersen R, Tetens I, Astrup A, Michaelsen KF, Lauritzen L. Eicosapentaenoic acid and docosahexaenoic acid in whole blood are differentially and sex-specifically associated with cardiometabolic risk markers in 8-11-year-old danish children. PLoS One 2014;9:e109368.



- Capel F, Acquaviva C, Pitois E, Laillet B, Rigaudière JP, Jouve C, Pouyet C, Gladine C, Comte B, Vianey Saban C, Morio B. DHA at nutritional doses restores insulin sensitivity in skeletal muscle by preventing lipotoxicity and inflammation. J Nutr Biochem 2015;26:949-59.
 PUBMED | CROSSREF
- 41. Li H, Ruan XZ, Powis SH, Fernando R, Mon WY, Wheeler DC, Moorhead JF, Varghese Z. EPA and DHA reduce LPS-induced inflammation responses in HK-2 cells: evidence for a PPAR-gamma-dependent mechanism. Kidney Int 2005;67:867-74.

 PUBMED | CROSSREF