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Microscale velocity-dependent unbinding generates a macroscale performance-efficiency tradeoff in actomyosin systems

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Myosin motors are fundamental biological actuators that power diverse mechanical tasks in eukaryotic cells via ATP hydrolysis. Previous work has linked myosin's velocity-dependent detachment rate to macroscopic scale muscle dynamics described by Hill's model, yet its impact on energetic flows — power consumption, output, and efficiency — remains unclear. We develop an analytical model relating myosin unbinding, quantified by a dimensionless parameter α , to energetics. Our model agrees with published in-vivo muscle data and reveals a performance-efficiency tradeoff governed by α . To experimentally validate this tradeoff, we build HillBot, a robophysical Hill muscle model that mimics nonlinearity and decouples α 's concurrent effects on performance and efficiency, demonstrating that nonlinearity sensitively drives efficiency. We analyze 136 published α measurements from in-vivo muscle samples and find a distribution centered at $\alpha^* = 3.85 \pm 2.32$. Importantly, both our analytical model and HillBot — despite operating under entirely different mechanisms — converge on the finding that this value α^* of nonlinearity observed in muscle corresponds to generalist actuators that balance power and efficiency. These insights inform a nonlinear variable-impedance protocol that directly shifts along a performance-efficiency axis, which could be implemented in robotics applications.

Myosin motors are ubiquitous, fundamental molecular machines responsible for powering many biological processes, including intracellular transport, cytokinesis, and muscle contraction. These motors operate at submicron scales, utilizing chemical energy from ATP hydrolysis to generate mechanical force, yet their force production is vital across scales from cellular functions to organismal-level activities. By coordinating in vast ensembles, myosin motors drive macroscopic muscle dynamics, enabling a diverse set of mechanical tasks such as locomotion (walking, swimming, flight), internal regulation (stability, balance, fluid transport), and athletics (endurance, high-intensity). Myosin coordination on the organismal-scale is adaptable, either on long time scales due to selection pressures^{1,2} or on short time scales due to training regimen^{3–5}. In order to power cellular and organismal dynamics, myosin motors convert a stored energy source, in the form of adenosine triphosphate (ATP), into useful mechanical work. How myosin's energy transduction mechanisms, which are ultimately microscopic (nm) in origin, manifest on energetic flows at cellular (μm) and organismal scales (m) remains a formidable research challenge.

A multitude of mechanisms determine how microscopic activity transduces chemical free-energy consumption to macroscopic mechanical

work output. Various biochemical and structural factors, such as muscle-specific protein isoforms and distinct metabolic pathways to supply ATP⁶, and microstructural features like post-stimulation transients⁷, variable thick-thin-filament overlap⁸, filament length polydispersity⁹, length-dependent calcium concentration¹⁰, advection of substrates¹¹, and sensitivity to small changes in lattice spacing¹² modulate how microscopic myosin kinetics translates into macroscopic mechanical output. These microscopic influences shape key muscle characteristics observed at larger scales¹³, including the force-length curve¹⁴, force response to stimuli¹⁵, and work loops¹⁶. However, ultimately, it is the chemomechanical activity of actomyosin cross-bridges, where myosin-mediated ATP hydrolysis couples to the relative sliding of actin, that drives actuation in biology.

A distinctive feature of the myosin power stroke, relative to other biochemical reactions (such as typical enzymatic reactions or glycolysis), is myosin's velocity-dependent off-rate¹⁷ wherein faster contraction rates cause quicker unbinding. Recent work has shown that this velocity-dependent unbinding aligns myosin kinetics with Hill's historical muscle model, connecting microscopic activity to macroscopic force-velocity characteristics through a nonlinearity parameter α ¹⁸. Here, the nonlinearity

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parameter, α , functions as a coarse-grained descriptor translating microscopic myosin activity rates into macroscopic force-velocity curvature, as characterized by Hill's model¹⁹.

Seminal work by Hill¹⁹ experimentally measured the force-velocity (FV) curve of the sartorius muscle in frogs, and found a nonlinear, hyperbolic relation. Although Hill's experiments were performed under quick-release from isometric conditions¹⁹, and Hill's model is not precise near stall^{20,21}, or under eccentric contractions²², it remains a reasonable approximation under quasistatic conditions and is frequently used in the region of maximum power for many kinds of motions^{18,23}. In normalized form, Hill's force-velocity relation is shown in Eq. (1)

$$F = \frac{1 - V}{1 + \alpha V} \quad (1)$$

where the muscle's tension f and contraction rate v are normalized by isometric tension f_m and no-load speed v_m to yield dimensionless quantities $F = f/f_m$, $V = v/v_m \in [0, 1]$ (see SI Table 1 for a full list of variable definitions) and the non-negative, dimensionless parameter $\alpha \geq 0$ governs nonlinearity (Fig. 1b).

Within the framework of Hill's model, empirical findings have demonstrated how a concave force-velocity relation shapes energetics. Studies have correlated efficiency with higher nonlinearities and have shown that heat generation in contracting muscle fibers depends on α ^{24,25}. Early studies suggested that more curved FV relations give fewer negatively strained cross-bridges²⁵, which would tend to unbind once a certain conformation was reached²⁶, with X-ray diffraction studies confirming this mechanism¹⁷.

Although empirical connections between concavity and energetics in the context of Hill's model have been made, analytical connections between the microscale velocity-dependent unbinding of myosin and macroscale energetic flows—from free-energy consumption to mechanical power output—remain elusive.

Here, we develop an analytical 2-state model of actomyosin dynamics that relates molecular myosin unbinding rates to emergent macroscale energetics. We demonstrate that our model agrees with published in-vivo muscle data (both in power outputs and energetic efficiencies). Furthermore, the model uncovers a performance-efficiency tradeoff governed by myosin's unbinding rate α . To experimentally investigate the tradeoff, we build HillBot, a robophysical model of Hill's muscle, as experimentally measuring energy consumption and muscle mechanics in-vivo is difficult and often requires estimation^{27–30}. Moreover, it is difficult to experimentally test a series of muscles with systematically varying nonlinearities α while

keeping all other properties constant — e.g., the muscle's maximum force, contraction-rate, ratio of fast- to slow- twitch fibers, training-level, fatigue, cross-sectional area, length, etc. Through HillBot, we decouple α 's concurrent effect on performance and efficiency, demonstrating that concavity in the muscle's force-velocity relation drives efficiency at the sacrifice of power output. Finally, we compile 136 published measurements of α in muscle and myoblasts to reveal a distribution centered at $\alpha^* = 3.85 \pm 2.32$. Synthesizing data from our model and HillBot, we quantitatively show that α^* coincides with a minimum of a cost function representing the performance-efficiency tradeoff. Finally, leveraging these insights, we discuss a nonlinear variable-impedance protocol to shift along a performance-efficiency axis in robotic applications.

Results

Cross-bridge dynamics control energetic flows in muscle

We build a simple 2-state model of muscle contraction following^{17,18} to study how the nonlinear, velocity-dependent unbinding rate of cross-bridges affects energy consumption and mechanical power generation.

The model starts following Ref. 18, which connects microscopic actin-myosin dynamics to macroscopic Hill dynamics. We start by assuming a simple 2-state model of actin-myosin contraction³¹ where some proportion of myosin are attached to actin A and some are detached D (Fig. 1a). The sum of detached and attached myosin proportions must be 1, $A + D = 1$. We assume the rate of myosin attachment k_a to be constant and velocity-independent. We further assume the rate of myosin detachment $k_d(V) = K_D V + k_0$ to be velocity-dependent, with a finite detachment rate k_0 at zero velocity (i.e., the isometric detachment rate)¹⁷. We write down the rate at which the fraction of attached myosin changes as

$$\dot{A} = k_a D - k_d A \quad (2)$$

Non-dimensionalizing the above equation yields

$$\dot{A} = 1 - A(1 + \alpha V + \gamma) \quad (3)$$

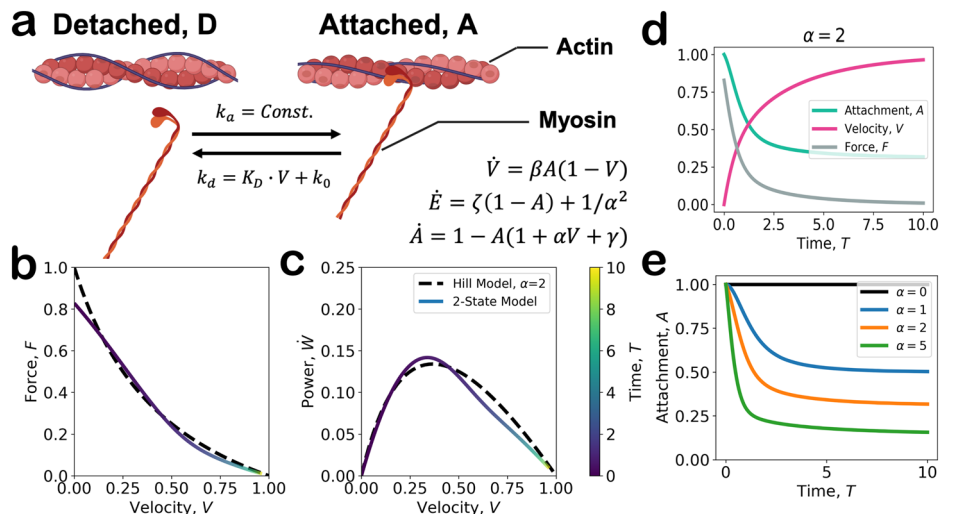
where time is rescaled by myosin's attachment rate $T = t k_a$ and the non-linearity is defined as $\alpha = K_D/k_a$ ¹⁸ and $\gamma = k_0/k_a$. Experimental data of the anterior tibialis muscle in frog¹⁷ demonstrate $\gamma \approx 1$.

Next, in order to couple myosin binding dynamics to actuation, we let this simple 2-state model of actin-myosin contract against some mass m

$$m\dot{v} = f_0 A(1 - V) \quad (4)$$

Fig. 1 | A nonlinear model accounting for 2-state myosin binding dynamics and actuation of a mass.

a Schematic of the 2-state model of myosin binding dynamics in muscle, created using BioRender⁹⁸. Myosin cycles between detached and attached states. **b, c** Time series of actuation, represented in force-velocity (**b**) and power-velocity (**c**) spaces compared with the predictions of Hill's model in Eq. (1) (black-dashed line). Color corresponds to dimensionless time T (color bar, right). **d** Time series of model attachment (teal line), velocity (magenta line), and force (gray line) for $\alpha = 2$, with $T = k_a t$ dimensionless time. **e** Time series of myosin attachment for $\alpha = 0, 1, 2, 5$ (black, blue, orange, and green lines, respectively); a larger fraction of myosin unbind with increasing nonlinearities.



where the contracting muscle's force is given by the product of the fraction of attached, force-producing myosin A , the stall force f_0 of an individual motor, and the linear force-velocity relation of a single myosin motor¹⁷. We again rescale time $T = tk_a$ and normalize acceleration \dot{v} by $v_m k_a$ to arrive at

$$\dot{V} = \beta A(1 - V) \quad (5)$$

where $\beta = f_0/mv_0k_a$ is a dimensionless quantity relating muscle actuation properties to the inertia of the attached mass. When attached to a mass $m = 10^{-1}$ kg, a rough order of magnitude estimate suggests $\beta \approx 1$ given that $k_a = 10 \text{ s}^{-1}$ ¹⁷, $f_0 = 78 \text{ mN}$, and $v_0 = 27 \text{ mm s}^{-1}$ in the soleus of a mouse³².

Although our model includes the fact that individual myosin motors have a linear FV-curve¹⁷, the velocity-dependent myosin detachment rate K_d introduces a nonlinear coupling between the equations of motion for \dot{A} and \dot{V} . The dimensionless nonlinearity parameter $\alpha = K_D/k_a$ directly relates to the curvature parameter $\alpha = f_m/a$ of Hill's original model, as demonstrated in steady state in ref. 18. Here, we confirm that the transient dynamics of our model can closely follow the hyperbolic relation originally described by Hill¹⁹ (Fig. 1b, c). Recognizing $F = \beta^{-1}\dot{V}$ and taking $\gamma \rightarrow 0$, Eq. (3) in steady state yields exactly Hill's equation (Eq. (1)). We have thus shown that our simplified system of equations is sufficient to describe how the nonlinear coupling between myosin unbinding dynamics and actuation, expressed via α , captures the kinematics in force-velocity space described by Hill's equation.

Next, we systematically assess how nonlinearity controls macroscopic energetic flows. For this, we include an additional equation that describes how α affects energy consumption in muscle, both directly by ATPase from the power stroke as well as from maintenance heat.

Myosin hydrolyzes ATP, resulting in the release of Gibbs free energy g_0 . As the total number of attaching myosin $Nk_a(1 - A)$ transition to the attached state, the total rate of Gibbs free-energy consumption \dot{g} is given by

$$\dot{g} = g_0 Nk_a(1 - A) \quad (6)$$

In addition to utilizing Gibbs free energy for contraction, muscle also produces maintenance heat for homeostatic regulation \dot{h}_m . Experiments have shown that the rate of maintenance heat production is approximately $\dot{h}_m = ab$ where a, b are constants in Hill's muscle model²⁴. Therefore, the total rate of energy liberation in the model is given as

$$\begin{aligned} \dot{e} &= \dot{g} + \dot{h}_m \\ \dot{e} &= g_0 Nk_a(1 - A) + ab \end{aligned} \quad (7)$$

Which, after rescaling by a characteristic system power $f_m v_m$, and noting that $av_m = bf_m$ for $\alpha_{\text{Hill}} = f_m/a$ in Hill's original model, we arrive at

$$\dot{E} = \zeta(1 - A) + \frac{1}{\alpha^2} \quad (8)$$

where $\zeta = Nk_a g_0 / f_m v_m$ relates the rate of Gibbs free-energy release due to myosin ATPase activity to the rate of mechanical work output. Recognizing that $f_m v_m \approx Nf_0 v_0$, with f_0 and v_0 the maximum force and velocity of an individual myosin motor, we have $\zeta \approx \frac{k_a g_0}{f_0 v_0} = 1/3$, given $g_0 = 100z$ ³³, $k_a = 10 \text{ s}^{-1}$, $f_0 = 6pN$ and $v_0 = 10 \mu\text{m s}^{-1}$ for single motors¹⁷. The rate of energy release in our 2-state model is in agreement with prior phenomenological models, which found that the heat generated during contraction is proportional to the sum of maintenance heat and the rate of shortening^{34,35}.

Equations 3, 5, and 8 represent our simplified model that describes how myosin's nonlinear unbinding affects energetics. The model parameters β, γ, ζ can be written in terms of the nonlinearity $\alpha = K_D/k_a$ such that $\beta = \frac{f_0}{mv_0 k_a} = \frac{f_0}{mv_0 K_D} \cdot \alpha$, $\gamma = \frac{k_0}{k_a} = \frac{k_0}{K_D} \cdot \alpha$, and $\zeta = \frac{k_a g_0}{f_0 v_0} = \frac{K_D g_0}{f_0 v_0} \cdot \frac{1}{\alpha}$. Each parameter, $\beta(\alpha)$, $\gamma(\alpha)$, $\zeta(\alpha)$, is still of order 1 when allowed to vary with the unbinding rate α – see SI Fig. 3 for more information on how we write $\beta(\alpha)$, $\gamma(\alpha)$, $\zeta(\alpha)$. Figure 1d shows a time series of the fraction of attached myosin

motors A and the normalized force and velocity for $\alpha = 2$. Increasing the nonlinearity decreases the fraction of attached myosin (Fig. 1e), reducing power output (Fig. 1c).

To characterize the total power of the actuator independent of actuation time or task, we introduce the power characteristic PC . The PC is a scalar that quantifies an actuator's ability to output power at all contraction rates, and we define PC as the integral of an actuator's normalized force-velocity curve. Using Hill's equation (Eq. (1)), we have:

$$PC := \int_0^1 F \cdot dV = \frac{(1 + \alpha) \cdot \ln(1 + \alpha) - \alpha}{\alpha^2} \quad (9)$$

where $PC \rightarrow 1/2$ in the limit of linear actuation $\alpha \rightarrow 0$. We show that the total mechanical work delivered by our 2-state model in a fixed amount of time increases with PC (SI Fig. 1d, Fig. 2a, solid lines) since more powerful actuators have higher work capacities.

In addition to the model's mechanics PC , we calculate the model's energetic efficiency at each time point as a ratio of instantaneous power generation to the rate of energy release, $\eta_{\text{inst}} = \frac{\dot{W}}{\dot{E}}$ (Fig. 2b). Moreover, to further validate the energetics of our model, we identify $N = 26$ measurements across 11 studies, where efficiency and α were concurrently measured over nine distinct muscle groups^{24,25,32,36–43} (Fig. 2d, Supplementary Data 1–3 file). Although some studies define efficiency as the ratio of work W to energy liberated E (or, work to the sum of work and heat)^{25,32,36,37}, here we consider the maximum instantaneous efficiency during actuation transients. Because we interpolate the data across different studies, the work done by each muscle sample is not standardized. Reporting total efficiency (work to the sum of work and heat) may be difficult to compare as the muscle samples may have spent different amounts of time in steady-state. Therefore, for the following analysis, we define $\eta = \max_{V \in [0,1]} \frac{\dot{W}}{\dot{E}}$ as this metric standardizes the efficiency data for different studies. In addition, for each of the $N = 26$ efficiency and α measurements, we calculate PC for each muscle sample by Eq. (9) (Fig. 2c). We find that, by visual inspection, our model agrees well with the PC and η results of our meta-analysis (Fig. 2c, d), demonstrating that coupling cross-bridge dynamics to transient dynamics of an accelerated object suffices to account for nonlinear energetic flows (inputs to outputs) in muscle.

For increasing nonlinearity α , we demonstrate that a tradeoff between PC and η emerges (Fig. 2c, d): as curvature in FV-space increases with α , the work capacity PC decreases yet energetic efficiency η increases. In the linear limit $\alpha \rightarrow 0$, the ratio of myosin detachment rates to attachment rates decreases, leaving the myosin mostly attached (Fig. 1e) and generating force. In this limit, we recover the highest PC (Fig. 2c). However, with relatively high myosin attachment rates, ATP consumption and heat generation increases (Fig. 2a). Increasing nonlinearity causes more cross-bridges to unbind (Fig. 1e). Although this reduces power (Fig. 2c), it increases efficiency (Fig. 2d) by reducing ATP consumption and heat generation.

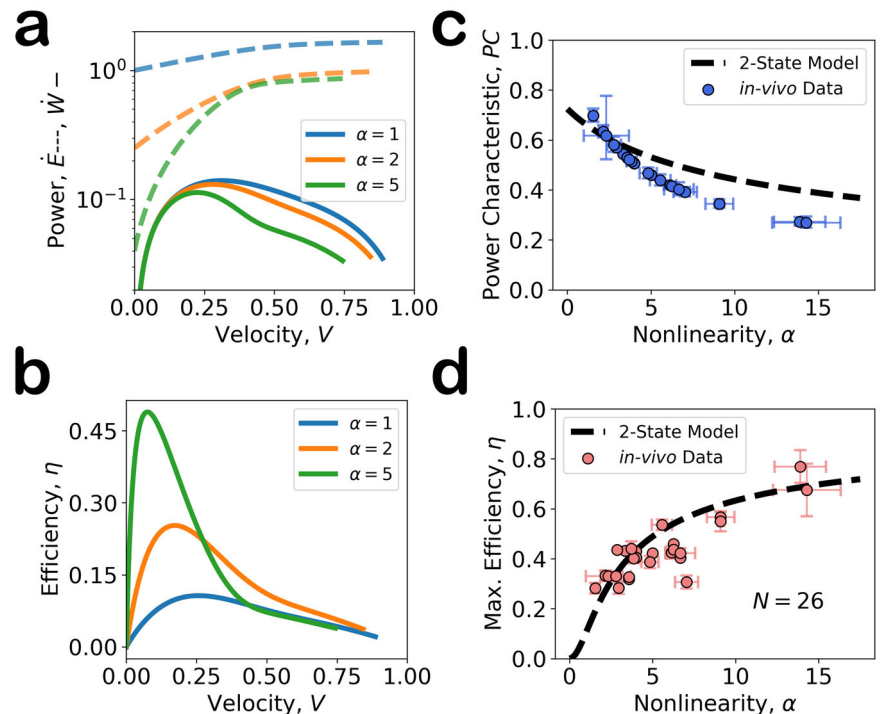
These results quantitatively elucidate the molecular basis for a tradeoff between efficiency, η , and performance, PC , in actin-myosin systems while demonstrating strong agreement with in-vivo muscle data taken from the literature. In addition to our analytical 2-state model, to experimentally assess how nonlinear unbinding $\alpha = K_D/k_a$ governs a tradeoff in performance and efficiency, keeping all other muscle parameters constant, we develop HillBot, a robophysical model of the Hill muscle.

HillBot elucidates a performance-efficiency tradeoff

In order to quantitatively assess how nonlinearity determines power and efficiency in experiment, we extend our earlier work⁴⁴. We develop a robophysical muscle model called HillBot (Fig. 3a–c) consisting of a DC motor and an Arduino microcontroller that utilizes proportional-integral-derivative (PID) feedback control (SI Fig. 1a) to mimic Hill's nonlinear force-velocity profile for a user-specified nonlinearity α (Fig. 3d–f).

We employ HillBot as it is difficult to experimentally test a series of muscles with systematically varying values of the velocity-dependence K_D

Fig. 2 | A performance-efficiency tradeoff emerges. a As the rate of unbinding (α) increases, mechanical power output (solid lines) and power consumption (dashed lines) decreases, however, because power consumption drops at a faster rate than power output, **(b)** we find increasing efficiency with the nonlinearity. **c, d** Performance-efficiency tradeoff emerges in 2-state model (black-dash lines) and in-vivo data taken from the literature (scatter points, $N = 26$ measurements with associated error bars as reported in their respective studies); as the unbinding rate α increases, the actomyosin system is less powerful (lower PC) yet it gains higher instantaneous efficiency η values.



on the myosin unbinding rate (i.e., $\alpha = K_D/k_a$) while keeping all other properties constant, e.g., the muscle's maximum force, contraction-rate, ratio of fast- to slow- twitch fibers, training-level, fatigue, cross-sectional area, length, among other muscle properties. In addition, accessing muscle state variables (the rate of energy consumption \dot{E} and power output \dot{W} , along with others) in real time is difficult to measure in living muscle and often requires estimation^{27–30}. Through HillBot, we can experimentally measure energetics in real time while keeping all muscle properties constant for a range of α , allowing us to have experimental control in monitoring α 's concurrent effects on efficiency and performance, which is otherwise difficult with in-vivo tissue.

We construct HillBot's differential equations with Newton's and Kirchhoff's second laws to describe its electro-mechanical behavior (Fig. 3b, see Methods). To couple these equations, we use the fact that the back EMF is proportional to the motor's output angular velocity by a constant k_1 and the current draw is proportional to the motor's output torque by a constant k_2 . To achieve Hill-type actuation, the PID minimizes the error function in Eq. (13) (defined as the difference between the predicted Hill muscle force in Eq. (1) and the measure output force of HillBot) by dynamically reducing the voltage in Eq. (12) (Fig. 3g), similar to the myosin unbinding from earlier.

$$\dot{\omega}(t) = \frac{1}{J} \cdot (k_2 i - b\omega) \quad (10)$$

$$\dot{i}(t) = \frac{1}{L} \cdot (\mathcal{E} - iR - k_1\omega) \quad (11)$$

$$\mathcal{E}(t) = k_p e(t) + k_i \int_0^t e(\tau) d\tau + k_d \frac{de(t)}{dt} \quad (12)$$

$$e(t) = 12 \cdot \left(\frac{1 - V}{1 + \alpha V} - F \right) \quad (13)$$

The applied voltage $\mathcal{E} \in [0, 12]$ (in Volts) is determined by the measured current draw i (in Amps) and angular velocity ω (in Rads/s) (which are proportional to the normalized force F and velocity V of the motor; see Methods). The PID gains (k_p, k_i, k_d) are set by the user beforehand and are

thus known constants. The resistance, inductance, angular velocity-back EMF constant, torque-current constant, moment of inertia, and damping constants (written as $\Pi = \{R, L, k_1, k_2, J, b\}$) are unknowns, which we determine through fitting (see Methods, SI Fig. 1c).

For a range of α , we integrate Eqs. (10–13) forward in time. Also, varying α across $N = 169$ trials, HillBot lifts a small mass m , and we record HillBot's angular displacement and current draw every ~ 10 ms until the system reaches steady state ($T = 0.68$ s). We extract the system's state, expressed in terms of the force f applied and the velocity v with which the mass rises (Fig. 3d). We verify that the model and HillBot are consistent with Hill's equation (Eq. (1); Fig. 3e, f, SI Fig. 1b), and find strong agreement, demonstrating that our system accurately mimics the mechanics of Hill's model by dynamically “unbinding” the voltage (Fig. 3g). HillBot serves as the physical instantiation of the physical mechanisms represented in the electromechanical model, confirming the the model captures the correct underlying dynamics.

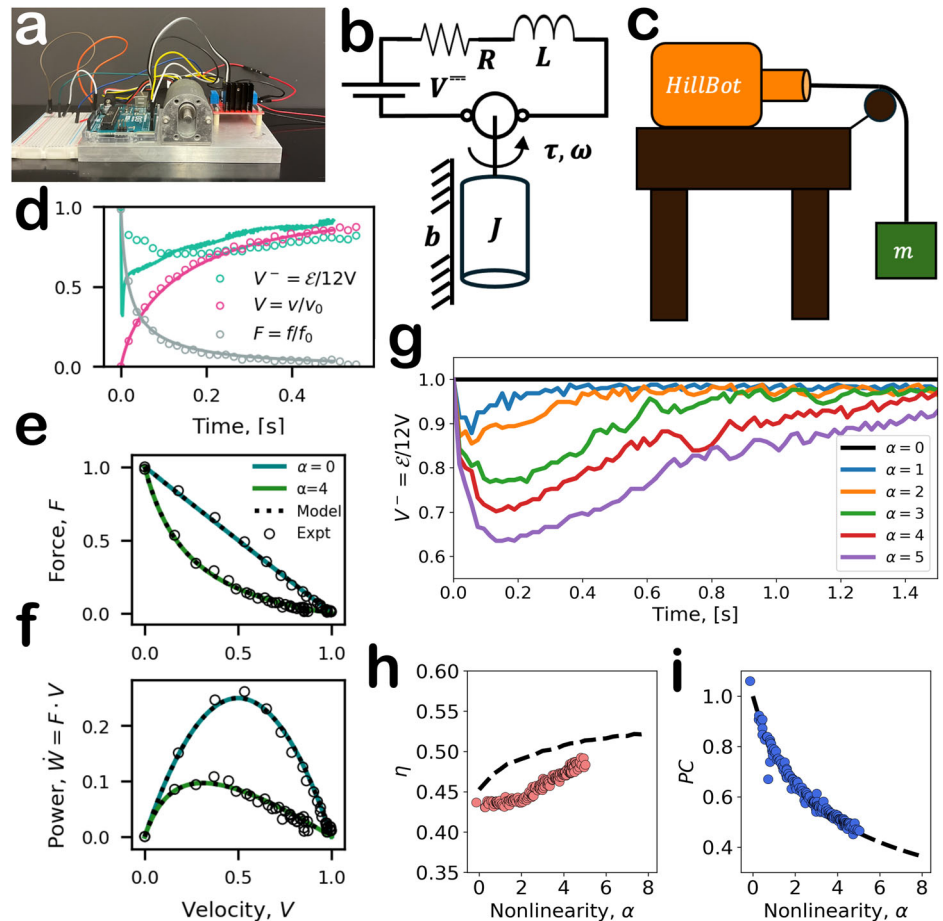
During a lifting task, we quantify HillBot's total energy consumption E by integrating a time series of electrical power consumption $\dot{E} = \mathcal{E} \cdot i$ up to steady state time T . Similarly, we integrate a time series of the actuator's mechanical power $\dot{W} = f \cdot v$ to calculate its total work output W . We then define HillBot's efficiency during actuation as the ratio of work to energy consumption, $\eta = W/E$. Furthermore, we quantify HillBot's mechanics through the power characteristic PC in Eq. (9) as before.

We observe that PC decreases as α increases (Fig. 3i). This is consistent with the observation that increasing α increases the curvature of the actuator's FV-curve (Fig. 3e) and hence suppresses power outputs (Fig. 3f). We also note that η increases with α (Fig. 3h); this result aligns with our previous findings⁴⁴ where we varied α while holding HillBot's work output constant. At the onset of each trial, the controller described in Eq. (12) applies a high initial voltage in response to a large initial error in Eq. (13). As the motor begins to rotate, to achieve a response consistent with the functional form of Hill's model (Eq. (1)), the controller subsequently decreases voltage to moderate force output – higher nonlinearities require larger “unbinding” of voltage (Fig. 3g). This voltage reduction, coupled with the motor's inertia sustaining its rotation, facilitates efficiency gains with the nonlinearity, as demonstrated in Fig. 3h.

Both HillBot and the electromechanical model dynamically “unbind” voltage (Fig. 3d, g), akin of myosin's detachment mechanism during

Fig. 3 | Introducing nonlinearity in HillBot demonstrates output and efficiency tradeoff.

a Photograph of HillBot. **b** Schematic of electro-mechanical model of HillBot. **c** Schematic of HillBot lifting a weight against gravity. **d** Time series of data collected from HillBot; normalized, measured force (gray open circles), velocity (magenta open circles), and voltage (teal open circles) as a function of time overlaid on the model's predictions (solid lines). **e** FV and **(f)** $\dot{W}V$ curves for $\alpha = 0, 4$ given by Hill's equation (blue and green lines, respectively), experimental measurements with HillBot (open circles), and electromechanical model (black dashed lines). **g** The PID controller dynamically reduces voltage for increasing α to match Hill-type outputs (similar to cross-bridge unbinding in the 2-state model). **h, i** HillBot's electromechanical model (black dashed lines) and $N = 169$ trials in HillBot (scatter points) show an exchange of power characteristic (blue circles) and efficiency (red circles) with α .



actuation (Fig. 1e). HillBot applies a voltage that drives motion on a fast timescale, while the controller measures and corrects this motion on a slower ~ 10 ms timescale. These corrections cause a characteristic voltage drop and recovery (Fig. 3g). The electromechanical model, by contrast, lacks an explicit separation of these timescales, leading to a more abrupt voltage drop and recovery (Fig. 3d). This quick, large drop in voltage with a slow recovery causes the model to over predict efficiency (Fig. 3h) as it overall draws less electrical power while producing the same mechanical outputs of HillBot (Fig. 3e, f). A longer, outer timescale is present in HillBot as measurements take ~ 10 ms to record and process, whereas in the electromechanical model, the equations are coupled and thus integrated forward in time together. This results in a single timescale in the model: the integration time step $dt \sim 0.1$ ms, where this small step size is necessary due to the nonlinear nature of the coupled equations.

Increasing η and decreasing PC as the unbinding rate α increases mirrors the main results of the 2-state model from earlier (Fig. 2c, d): through HillBot, we again find a performance-efficiency tradeoff (Fig. 3h, i) parameterized by the nonlinearity α . This result is striking: although HillBot and muscle consume energy by entirely different mechanisms, we show that dynamically “unbinding” to generate Hill-type dynamics (whether it be through voltage (Fig. 3g) or cross-bridges (Fig. 1e)) generates energetic efficiency at the sacrifice of power output. This similarity suggests that nonlinear couplings in other actuators could yield similar energetic tradeoffs and optima. Thus, HillBot serves to demonstrate the generality of the physics uncovered in our model.

So far, we have demonstrated that increasing α simultaneously decreases power but increases efficiency. But because of these simultaneous effects, the mechanism underlying efficiency gains still remains unclear. Efficiency gains could result from the fact that lower-power actuators simply draw less energy. Alternatively, efficiency gains could result directly from

nonlinearity. In order to disentangle α 's simultaneous effects on power and efficiency, we perform a corresponding control experiment with an equivalent PC actuated by a linear force-velocity profile (Fig. 4a, b), called “Series-L”. These control experiments allow us to experimentally isolate the effect of the nonlinearity and disentangle α 's concurrent effects on efficiency and performance. Furthermore, we conduct Series-L as a reference set of data to ensure that any findings through HillBot are the result of the muscle's inherent nonlinear FV-shape and not due to any unknown artifacts of our model system or due to the reduction in the actuator's PC .

In the Series-L, we do not perform full-state feedback and actuate with a constant, reduced voltage, thus preserving the inherent linear $\alpha = 0$ FV-relationship of the DC motor:

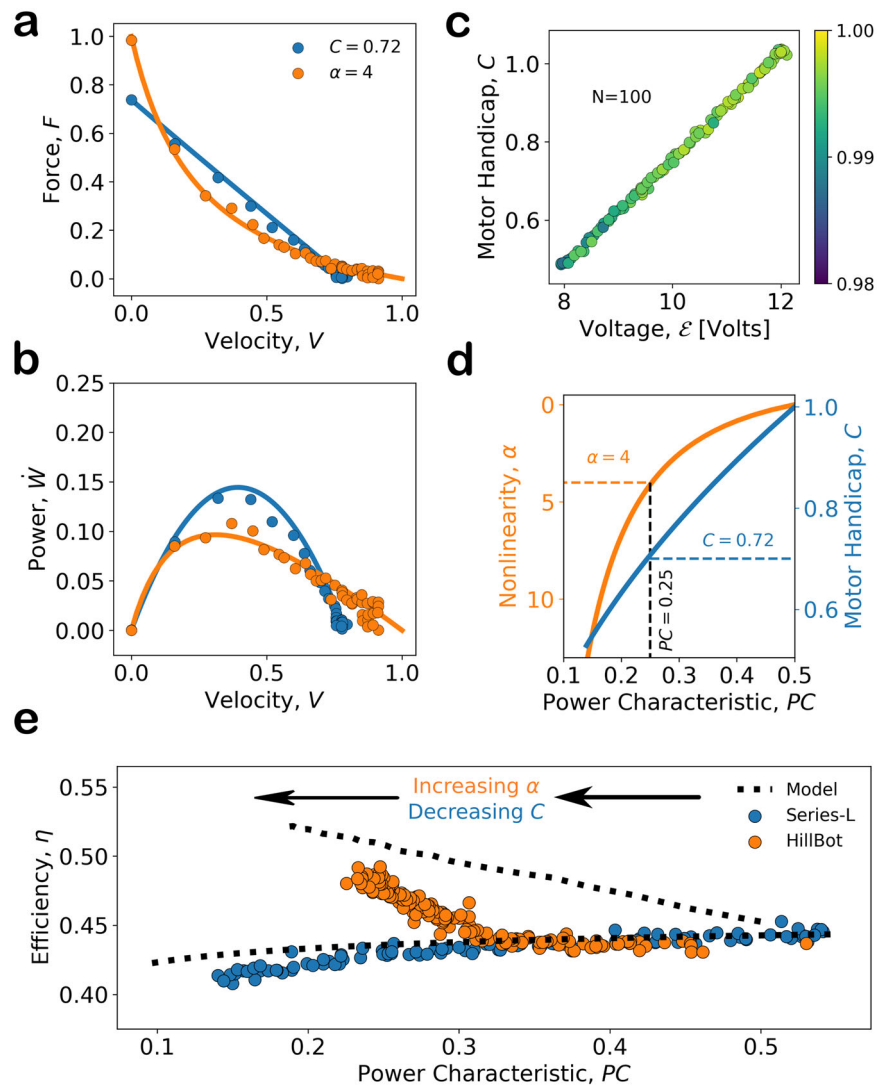
$$F = C - V \quad (14)$$

where the motor's normalized force and velocity $F = f/f_m$, $V = v/v_m \in [0, C]$ for $0 \leq C \leq 1$. To create Series-L, we reduce the constant input voltage \mathcal{E} which effectively reduces f_m and v_m to a constant C , which we call the motor handicap (Fig. 4c) and thus reduces the motor's power characteristic (Fig. 4d).

For each experiment with HillBot (with a given value α), we perform a corresponding experiment in Series-L (with a value $C(\alpha)$) to produce a series of nonlinear-linear pairs that each have an equivalent PC (Fig. 4a, b). A calculation shows $C = \sqrt{2} \cdot \sqrt{\ln(1 + \alpha)} \cdot (1 + \alpha) - \alpha/\alpha$ for any non-negative α value (see SI for derivation).

Through Series-L, we find decreasing efficiency η for decreasing PC values (Fig. 4e, blue points), whereas in HillBot, efficiency increases as the PC is reduced through the nonlinearity α (Fig. 4e, orange points). This result disentangles the effect of PC on efficiency and suggests that any efficiency gains in HillBot are emergent from the curvature of Hill's force-velocity

Fig. 4 | Nonlinear coupling generates a tradeoff in performance and efficiency not found in linear actuation. **a, b** Comparison of linear $C = 0.72$ and nonlinear $\alpha = 4$ force-power-velocity relations for $PC = 0.25$. **c** Linear relationship between applied voltage and motor handicap; experimentally, we decrease the voltage supply to decrease the motor constant to lower the PC in Series-L. **d** Motor constant C and nonlinearity α values that generate FV-relations with an equal PC ; for example, $PC(C = 0.72) = PC(\alpha = 4) = 0.25$ as in **(a, b)**. **e** Efficiency decreases with PC in Series-L, whereas efficiency increases with nonlinear actuation, disentangling PC 's effect on η .



relationship and the dynamic “unbinding” of voltage. These experiments confirm that a nonlinear coupling between actuation and the resulting dynamics controls an energy-efficiency tradeoff.

First, we started with a simple 2-state model of actin-myosin dynamics, and we were able to show that the molecular mechanism generating Hill’s nonlinear force-velocity relation is the velocity-dependent unbinding of myosin. We found that this nonlinearity sensitively parametrizes a tradeoff in efficiency (η) and performance (PC); low α -valued muscles are powerful yet inefficient, while high α -valued muscles are efficient yet weak. Next, using a robophysical model of muscle, we reproduced these results in the experiment and, moreover, through Series-L, showed that the efficiency gains of decreasing PC are the result of the muscle’s nonlinear FV-shape. This result leads us to ask: what is the physiological significance of a tradeoff in efficiency (η) and performance (PC) in muscle and non-muscle cells? Is there any utility for it? How does this tradeoff, parameterized by the unbinding rate α , manifest itself in nature?

Observation of a characteristic nonlinearity α^*

Researchers have long known that muscle has a nonlinear force-velocity curve¹⁹ (Fig. 5a, b). What values of the nonlinearity does naturally occurring muscle express? Hill initially speculated that the nonlinearity parameter α assumed a constant value of $\alpha \approx 4$ across all muscles. Since then, a broad range of α -valued muscles have been reported in the literature. To understand how α is distributed across organisms, we perform a meta-analysis

across $N = 136$ individual measurements of α in biological muscle samples from 39 different studies^{19,24,25,32,36,37,39–43,45–72}. (Fig. 5c–e, see “Methods”, Supplementary Data 1–3 file). Across evolutionarily distant species, we observe that most muscle groups in our meta-analysis exhibit values of α near a characteristic value of approximately four ($\alpha^* = 3.85 \pm 2.32$, median \pm IQR). In addition to muscle tissue, Hill’s hyperbolic force-velocity relation has also been observed in individual mouse myoblast cells⁷² where AFM experiments measured a value of $\alpha \approx 4$ (Fig. 5e, red dashed vertical line). Our meta-analysis aligns with Hill’s original estimation of $\alpha \approx 4$, a value widely accepted in the literature⁷³; however, our analysis also reveals a broader distribution of reported α values ranging from near zero to as high as 15. In order to classify muscle groups, we identify four distinct categories: muscle groups with a broad distribution of α (Fig. 5c, yellow), with a narrow distribution centered around the characteristic value $\alpha \approx 4$ (green), with lower values of α (blue), and with higher values (red). For the muscle groups that are present in humans, we depict their approximate locations in the human body (Fig. 5d). Although relating α to muscle function is not within the scope of this work, we discuss how nonlinearity and energetics could affect behavior on organismal scales (see Discussion).

The observation of a characteristic value of $\alpha^* = 3.85 \pm 2.32$ (which notably excludes linear actuation $\alpha = 0$) in muscle and myoblast cells raises the question whether this particular value confers energetic advantages. In order to provide a potential basis to explain this prevalence, we define a simple objective function $\phi_{\text{Hill}}(\alpha)$ that quantifies the performance-efficiency

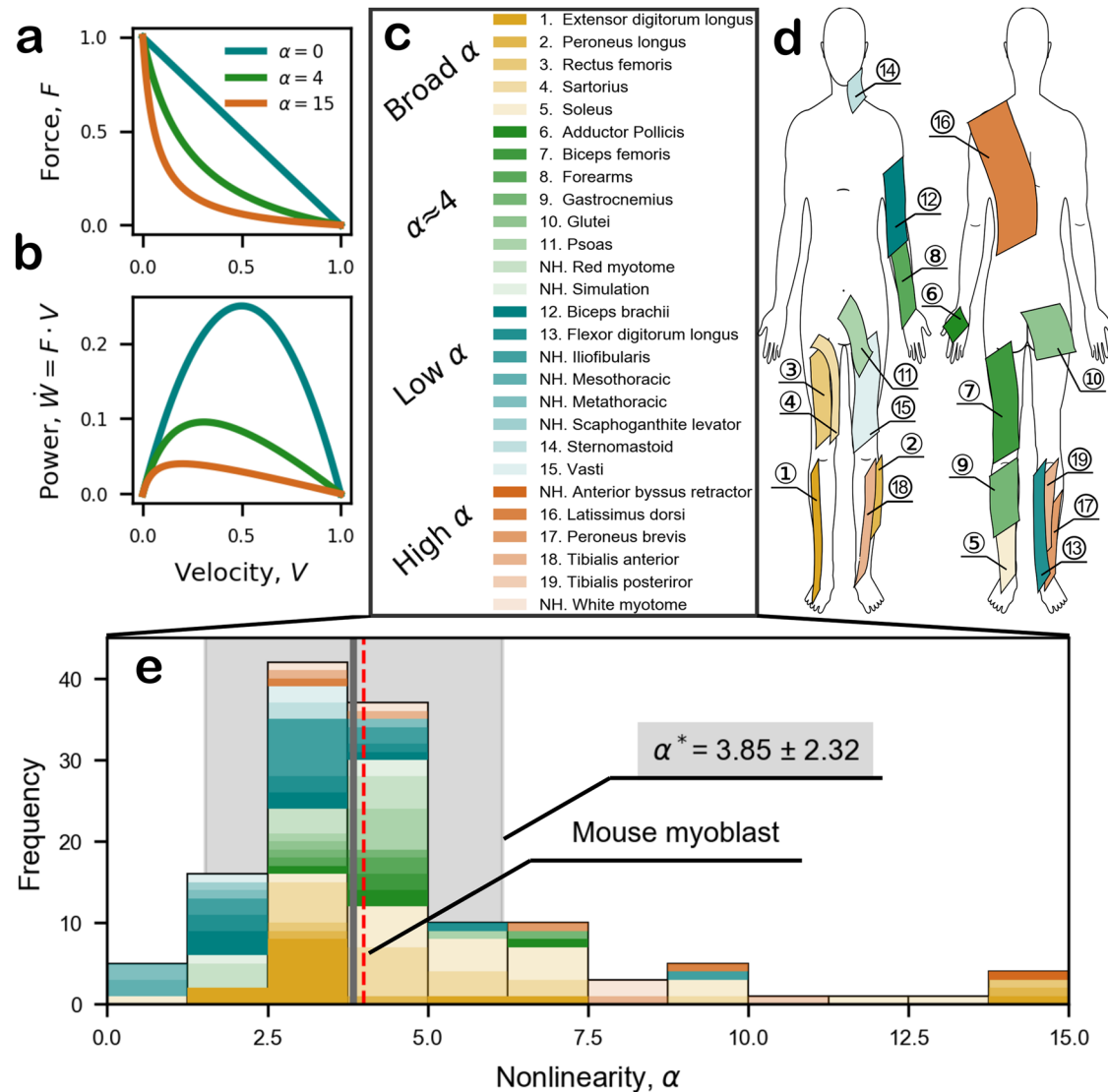


Fig. 5 | α meta-analysis results. **a** Hill's force-velocity (FV) and **(b)** mechanical power-velocity ($\dot{W}V$) relations for $\alpha = 0, 4, 15$. **c** List of muscle types found in the meta-analysis. Muscles are split into four groups: those where α has a broad range (yellow), low values (blue), high values (red), and near α^* (green). Numbered label corresponds to **(d)**; those labeled NH indicate non-human muscles. **d** Muscles found in the human body from the meta-analysis. Muscles are colored according to their

label in **(c)**. **e** Distribution of α in measured biological muscle tissue for $N = 136$ muscle samples, including those from mammals, birds, mollusks, fish, amphibians, insects, reptiles, and single mouse myoblast cells (red dashed line). The red dashed line indicates a measurement of the FV-relation from a myoblast cell. The nonlinearity α ranges between 0 and 15, centered at $\alpha^* = 3.85 \pm 2.32$ (median, gray line \pm IQR, shaded gray).

tradeoff in these actuators:

$$\phi_{\text{Hill}}(\alpha) := |\eta^*(\alpha) - PC^*(\alpha)| \quad (15)$$

where $\eta^*(\alpha)$ and $PC^*(\alpha)$ are the normalized energetic efficiency and normalized power characteristic as a function of the nonlinearity α . Energetic efficiency, by definition, is reported as a normalized quantity, $\eta^* := \eta/1$, and the power characteristic is normalized as $PC^* := PC(\alpha)/PC(\alpha=0)$. These normalization conditions ensure that maximally efficient actuators correspond to $\eta^* = 1$ and maximally powerful actuators correspond to $PC^* = 1$. The minimum of $\phi_{\text{Hill}}(\alpha)$ represents nonlinearities in FV-space that equally favor the power characteristic and efficiency.

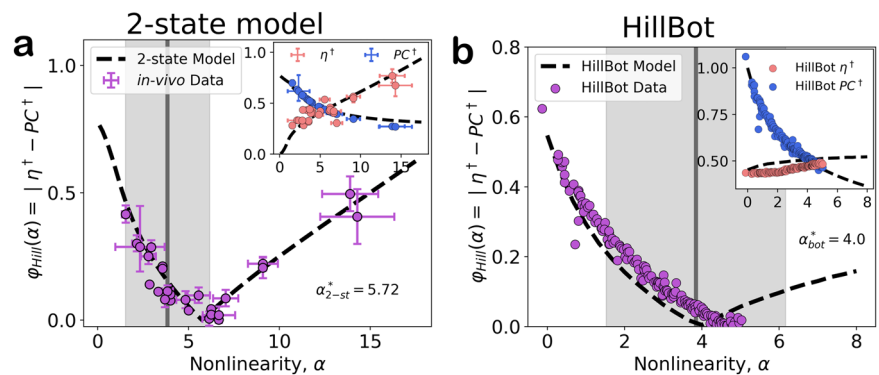
We compute $\phi_{\text{Hill}}(\alpha)$ based on the η and PC data from our 2-state model and find a functional form that, by visual inspection, agrees with the results of the in-vivo efficiency data from the literature (Fig. 6a). Our model predicts a minimum of ϕ_{Hill} at $\alpha_{\text{bot}}^* = 5.72$, agreeing with the characteristic value $\alpha^* = 3.85 \pm 2.32$ from the meta-analysis. We further compute $\phi_{\text{Hill}}(\alpha)$ using the recorded η and PC data from our experiments with

HillBot. We again observe a non-monotonic dependence of ϕ_{Hill} on α , with a minimum at $\alpha_{\text{bot}}^* = 4.0$ (Fig. 6b). This result again coincides with the characteristic nonlinearity $\alpha^* = 3.85 \pm 2.32$ from our meta-analysis (Fig. 5e, gray-shaded region). These results raise the hypothesis that the most commonly observed nonlinearity $\alpha^* \approx 4$ balances performance and efficiency in muscle and contractile cells. However, testing this hypothesis in muscle cells would be difficult as it requires simultaneous measurement of energy consumption and mechanical power, as well as systematic tuning of α (or, respectively, myosin's velocity-dependent unbinding rate) while keeping all other parameters constant.

Discussion

In summary, we have investigated how Hill's nonlinearity parameter α relates to the performance-economy tradeoff in muscle through a 2-state model of actin-myosin dynamics accompanied by published in-vivo data (Figs. 1, 2) in addition to a robophysical model of Hill-type dynamics (Figs. 3, 4). In each study, we quantified the performance-economy tradeoff through an objective function $\phi(\alpha)$ (Fig. 6) and found minima (Table 1) that

Fig. 6 | The performance-efficiency tradeoff quantified. **a** Objective function applied to in-vivo meta-analysis data (purple scatter points, $N = 26$ measurements with associated error bars as reported in their respective studies) and the simple 2-state actomyosin model (dashed black line). ϕ_{Hill} is minimized at $\alpha_{2\text{-st}}^* = 5.72$, agreeing with $\alpha^* = 3.85 \pm 2.32$ (gray shaded region). Inset: Efficiency η (red symbols) and power characteristic PC (blue symbols) as functions of the nonlinearity parameter α . **b** Objective function ϕ_{Hill} as a function of the nonlinearity parameter α in HillBot (purple circles, $N = 169$ trials) with electromechanical model (black dashed line). The gray region corresponds to the characteristic nonlinearity α^* . Sub-panel shows an exchange of power characteristic (blue circles) and efficiency (red circles) with α ; ϕ_{Hill} is minimized at $\alpha_{\text{bot}}^* = 4.0$.



agreed with the characteristic range of published nonlinearities $\alpha^* = 3.85 \pm 2.32$ (Fig. 5) from the literature. Together, these results demonstrate that energetic flows are sensitively related to α . We propose that α controls a tradeoff between mechanical power output and energetic efficiency through cross-bridge (un-)binding rate constants K_D and k_u , and that a value of $\alpha \approx 4$ optimizes this tradeoff, helping to explain the observed frequency of α^* in biological muscle tissue.

Notably, HillBot and the 2-state model both converge to nature's most commonly observed α^* (Figs. 5, 6) despite operating through entirely different mechanisms of power consumption and production (Eqs. 2–8 and Eqs. 10–13). This suggests that nonlinear couplings in other actuators could yield similar energetic tradeoffs and optima, demonstrating the generality of the physics within Hill's model.

Our results have investigated connections between myosin's velocity-dependent unbinding dynamics and energetics (free energy consumption, mechanical power output, and efficiency). In our microscopic model, $\alpha = K_D/k_u$ controlled the nonlinear coupling between myosin unbinding dynamics and actuation of a coupled mass. Meanwhile, in HillBot, we implemented α by dynamically reducing voltage to mimic the concave force-velocity relation described by Hill. We demonstrated in our microscopic model that the nonlinearity parameter $\alpha = K_D/k_u$ describing myosin's velocity-dependent unbinding rate resulted in transient dynamics resembling Hill's concave force-velocity curve (Fig. 1b, c). Furthermore, in both experiments on HillBot (Fig. 3) and our microscopic model of muscle (Fig. 2), we found similar dependencies of mechanical power output $\dot{W}(\alpha)$ and efficiency $\eta(\alpha)$ on the nonlinearity parameter α . Overall, our results demonstrate that nonlinear coupling between myosin binding dynamics and actuation impacts energetics, and that the energetics can be experimentally instantiated by mimicking Hill's relation in robotic systems.

Although it is interesting that $\dot{W}(\alpha)$ and $\eta(\alpha)$ coincided between our microscopic nonlinear model of myosin and HillBot's mimicking of the muscle's concave force-velocity curve, our study does not attempt to relate myosin binding dynamics to Hill's force-velocity curve for in-vivo systems. Although velocity-dependent unbinding does contribute to a concave force-velocity relation, other factors can affect the force-velocity curve. Coupling of muscle to the surrounding elastic connective tissue, and to elastic tendon, can

affect the force-velocity curve⁷. Values of α can vary between individuals and species depending on fitness training⁷⁴, fatigue^{5,76}, and temperature^{50,77,78}. Muscle groups are recruited cooperatively rather than in isolation⁷⁹. Furthermore, Hill's original experiments were performed in the specific case where muscle contracts a constant load after relaxation from isometric tension¹⁹, and Hill's relation is not precise near stall^{20,21}. A concave force-velocity curve can result from entirely different mechanisms as well, such as capillary forces⁸⁰. In addition, other muscle properties contribute to muscle energetics and function¹³, such as the muscle's force-length relationship¹⁴, the tendon's elastic energy storage⁸¹, coupling to multiple joints^{82–84}, Henneman's size principle⁸⁵, and differences in metabolic supply of ATP⁷³.

Future research could explore the impact of separating fast dynamics from slow control corrections, as seen in HillBot but absent in the electro-mechanical model, and investigate how this separation might apply to biological systems. Furthermore, relating microscopic-scale myosin binding dynamics to macroscopic-scale energetics and biological mechanical function would be an interesting future research direction. We encountered qualitative observations while performing our meta-analysis that could potentially lead to testable hypotheses. For muscle groups with broad distributions of α (Fig. 5c, yellow), and those with narrower distributions, close to the characteristic value of $\alpha \approx 4$ (green), we found some muscle groups that are implicated in a wide variety of tasks. The psoas ("11."), for example, performs both dynamic and posture functions⁸⁶. Meanwhile, muscle groups with high values of α (Fig. 5c, red), such as the latissimus dorsi ("16.") and the peroneus brevis ("17."), tend to sustain low levels of activity over long periods of time to assist in stabilization and posture regulation⁸⁷, activities where energetic efficiency may be vital. Finally, muscle groups such as the biceps brachii ("12.") and vasti ("15."), exhibit almost linear force-velocity curves (Fig. 5c, blue). These muscle groups are mostly implicated in explosive, ballistic motions⁸⁸ where power generation is key. Again, we caution that these observations remain qualitative. However, we anticipate that studying nonlinear unbinding dynamics and energetics across muscle groups and species could yield interesting insights into how animal-scale function is sustained by regulation of molecular-scale energetic conversion. Furthermore, it would be interesting to compare and contrast muscle groups with low and high α , and investigate whether their microscopic unbinding dynamics differ from the more common $\alpha \approx 4$ muscle groups.

Interestingly, our work shows that the power-efficiency tradeoff due to the nonlinearity α is not necessarily specific to actomyosin systems. In actomyosin, an increased fraction of myosin unbinds at higher contraction rates; similarly, HillBot reduces the applied voltage as the coupled mass increases velocity. Both responses amount to a "letting go" of actuation effort during transients. Although our analytical model of actomyosin dynamics included an expression for power consumption and heat generation, HillBot consumes and dissipates energy via entirely different

Table 1 | α that minimizes the power-efficiency tradeoff determined by different mechanisms

Distribution	HillBot	in-vivo muscle	STF-FTF
$\alpha^* = 3.85 \pm 2.32$	$\alpha_{\text{bot}}^* = 4.0$	$\alpha_{2\text{-st}}^* = 5.72$	$\alpha_{\text{fibr}}^* = 3.67$

The minima of $\phi(\alpha)$ for HillBot (α_{bot}^*) and the simple 2-state model ($\alpha_{2\text{-st}}^*$) are within the range of the most frequently observed nonlinearities from the literature α^* . Muscles that share an equal composition of slow- and fast-twitch fibers (α_{fibr}^*) are also within the bounds of α^* (see SI Fig. 2 for more information).

mechanisms. Yet, we still found a performance-efficiency tradeoff in both systems. This result highlights the generality of nonlinear actuation's control over energetic flows. We therefore propose HillBot as a novel experimental platform for investigating biological principles in a controlled, mechanistic framework and to leverage HillBot's actuation mechanism as a method of dynamically regulating energetic flows in robotics applications.

Seminal work demonstrated that dynamically regulating an actuator's end-point impedance to mimic a linear damped-spring system stabilizes contact interactions⁸⁹, and more recent work has highlighted the benefit of biologically-inspired hysteric damping^{90,91}. Here, we propose enhancing these approaches by incorporating dynamic control over the nonlinearity parameter α in variable impedance protocols. We anticipate that mimicking the nonlinearity affords high-level control over the energetic flows between an actuator and its coupled environment, dynamically biasing actuator-environment interactions along a performance-efficiency axis depending on context, energetic demands, and reserves. This method improves existing designs which mimic a static value of α using hardware components^{80,92}.

One further consideration in impedance protocols relates to human safety and comfort in human-robot interactions, which are crucial factors in designing assistive devices⁹³. Human prosthetics need to carefully regulate power to avoid injury⁹⁴. Surprisingly, reducing prosthetic power does not impact the user's metabolic cost⁹⁵. Moreover, amplifying the prosthetic's energetic efficiency allows for a lightweight, compact design⁹⁶. Under a nonlinear actuation scheme with α , the concavity of Hill's FV-relation simultaneously boosts efficiency and reduces power output (cf. Fig. 3h, i), addressing design and safety concerns without raising the user's metabolic cost. We anticipate that a variable nonlinear impedance control will result in more precise control over energetic flows, ultimately improving user experience with actuated prosthetics.

To conclude, here we have investigated the effect of the nonlinearity parameter α , which governs myosin's velocity-dependent unbinding rate, on energetic flows during actuation. We demonstrated this quantification with a simplified theoretical model coupling myosin binding dynamics to a mass. The energetic predictions of this model agreed with measurements of 26 muscle samples from the literature. Moreover, the model illustrated a performance-efficiency tradeoff parameterized by the unbinding rate α . Furthermore, we sought to mimic this velocity-dependent unbinding with HillBot, a robophysical model of Hill's hyperbolic force-velocity curve. Experiments with HillBot allowed us to systematically control α while holding all other muscle properties constant. As in our actomyosin model, we found similar dependencies of the mechanical power output $\dot{W}(\alpha)$ and the efficiency $\eta(\alpha)$ on the nonlinearity parameter α in HillBot. Although the precise microscopic mechanisms by which α regulates actuation in both systems differ greatly, our main result appears to be generalizable: a negative feedback from velocity onto actuation (i.e., "letting go" or "unbinding") reduces power generation but boosts efficiency. Lastly, we compile 136 published measurements of α in muscle and myoblasts to reveal a distribution centered around $\alpha^* = 3.85 \pm 2.32$. Synthesizing data from our model and HillBot, we quantitatively show that the performance-efficiency tradeoff may underpin the prevalence of α^* in nature. We propose to leverage this understanding in variable-impedance protocols to enable real-time control over energetic flows in robotics applications. This high level of abstraction could provide more nuanced control over robot-environment and human-robot interactions, including prosthetics and exoskeletons.

Methods

Meta-analysis

To acquire the meta-analysis data, we searched for peer-reviewed research articles that reported a measured value of α from a biological muscle tissue sample. The literature often describes force-velocity curvature as a ratio of the muscle's coefficient of shortening heat to the muscle's isometric force, using the notation a/P_0 or a/F_0 , where α is the inverse of this ratio. For each study where FV-curvature is measured, we record the animal's name and nickname, the muscle sample's group (EDL, bicep brachii, etc.), the

temperature at measurement (if available), and whether the measured α is the result of an average. Once we acquired all $N = 136$ measurements of α in muscle (in addition to the lone α measurement in mouse myoblasts⁷³), we created Fig. 5 to show the distribution of α in nature. Data from the meta-analysis is shown in the associated supplemental data file. Furthermore, to generate Figs. 2 and 6, we analyze a subset of $N = 26$ samples where energetic efficiency and FV-curvature are reported for the same muscle sample (see associated supplemental data file for raw data).

HillBot

HillBot is a physical model of the Hill muscle that uses feedback control to mimic the muscle's nonlinear force-velocity behavior in Eq. (1). Fig. 3 depicts HillBot, and SI Fig. 1 shows the control scheme implemented for biomimetic actuation. Our methods in physical-model development and data acquisition follow from our previous paper⁴⁴.

Hardware. HillBot comprises a motor (Pololu 70:1 gear motor) with a rotary encoder (4480 counts per revolution resolution). A time series of positional changes are used to calculate the motor's velocity. The measured velocity is analogous to the muscle contraction rate. A Hall effect-based current sensor (ACS714 Current Sensor ± 5 A) with low resistance (~ 1.2 m Ω) is placed in series with the motor and reads the motor's electrical current. The torque applied by a DC motor is proportional to the current and is analogous to muscle force.

Software. We program a microcontroller (Arduino UNO R3) with proportional-integral-derivative (PID) control with optimized gain coefficients to mimic the muscle's nonlinear FV-behavior (SI Fig. 1a). At each time step (loop sampling rate ~ 10 ms), we record force and velocity using the current sensor and encoder. The controller maps system state (force and velocity) to mimic Hill's relation for a given initialized α value by dynamically controlling the applied voltage \mathcal{E} .

Normalized state variables. For congruence with the normalized Hill model in Eq. (1), we normalize measured force and velocity by $V = v/v_m$ where v and v_m are the measured and maximum angular velocities of the motor. To normalize the measured force, we use $F = (i - i_s)/(i_m - i_s)$ so that force F is proportional to current i as expected. We subtract off the steady state current i_s (that is, i when the motor is at v_m) so that the actuator produces little force close to the no-load velocity.

Model. We start with Newton's and Kirchhoff's second laws to construct differential equations describing the electro-mechanical behavior of our actuator (Eq. (10–13)). The PID gains (k_p , k_i , k_d) are determined through trial and error until the model produces the expected Hill-type dynamics. The resistance, inductance, angular velocity-back EMF constant, torque-current constant, moment of inertia, and damping constants (written as $\Pi = R, L, k_1, k_2, J, b$) are unknowns. We determine these constants by supplying a constant voltage to the system and recording the system state. We sweep through a range of voltages from 12 to 8 Volts in $N = 100$ trials. For each set of time series data, we perform a nonlinear fit using LMFIT (a nonlinear least-squares minimization Python package) to determine the six parameters Π (SI Fig. 1c).

Finally, we compare the characteristics of our experimental data (scatter points) and model (black dashed line) with the predictions of Hill's equation for different α values (SI Fig. 1b). For a given value of α , we record a time series of the actuator's force-velocity data and mechanical power-velocity data. Both curves are integrated, and we extract the maximum power output of the actuator for each α . We then compare how the area under the actuator's FV and $\dot{W}V$ curves and maximum power output agree with Hill's original muscle model in Eq. (1). The results in SI Fig. 1b show strong agreement between our model, data, and Hill's muscle model for all tested values of α , demonstrating that our system accurately mimics the mechanics of Hill's model.

2-State model

The 2-state model presented in this work is based on Ref. 18. In this framework, myosin motors cycle between attached and detached states, consuming energy and generating mechanical work. The nonlinear relationship between contraction rate and force production arises from the velocity-dependent unbinding rate of the cross-bridges. Full details and derivations of the 2-state model are provided in the Results section of the main text.

Statistics and reproducibility

The study employed a metanalysis, sampling $N = 136$ measurements of the nonlinearity parameter α across the literature. These data span a wide range of experimental methods as they are interpolated from 39 different published studies. Such diverse sampling across experiments, methods, protocols, authors, and potential biases offers a holistic view of the muscle nonlinearity α . The distribution is summarized in Fig. 5 using the median and inter-quartile range (IQR), yielding $\alpha^* = 3.85 \pm 2.32$. A subset of the metanalysis data ($N = 26$) also includes corresponding measurements of energetic efficiency during contraction, as reported in Figs. 2, 6. Throughout this study, the error bars used in the metanalysis reflect those reported in the original publications. In experiments with HillBot, we conducted a large number of independent trials ($N = 169$) to support the robustness of our findings. For the Series-L experiments, we similarly performed $N = 100$ trials to maintain statistical reliability. Given that HillBot's code and the accompanying data analysis Jupyter notebooks are available in the repository, the results presented here should be fully reproducible by the reader.

Reporting summary

Further information on research design is available in the Nature Portfolio Reporting Summary linked to this article.

Data availability

All data related to this manuscript can be found on the Texas Data Repository under the name "Microscale velocity dependent unbinding generates a macroscale performance efficiency tradeoff in actomyosin systems," DOI found in references list⁹⁷. The associated Supplemental Data 1–3 file (in addition to the "alpha_metanalysis.tab" file in the repository) contains all of the metanalysis source data (α measurements with the associated efficiency and slow-twitch fiber data) used to create Figs. 2, 5, 6 and Supplementary Fig. SI2. HillBot source data used to create Figs. 3, 4 and Supplementary Fig. SI1 are found in the Excel files titled "newest_nonlin.tab" and "newest_lin.tab" on the repository. Finally, the source data related to the 2-state model used in Figs. 1, 2, 6 and Supplementary Fig. SI3 are found in the Jupyter Notebook titled "2-state-model" on the repository.

Code availability

All code related to this manuscript can be found on the Texas Data Repository under the name "Microscale velocity dependent unbinding generates a macroscale performance efficiency tradeoff in actomyosin systems," DOI found in the references list⁹⁷. Scripts for HillBot and the 2-state model can be found in the Jupyter Notebooks "generate-hillbot-model" and "2-state-model," respectively. Data analysis scripts are titled using the prefix analyze-, followed by a brief description of the analysis performed.

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Author contributions

J.M. and J.A. designed experiments. J.M. and B.K. performed research; J.M. analyzed data. J.M., C.J. and J.A. developed models. J.M. and J.A. wrote the manuscript.

Competing interests

Authors declare no competing interests.

Additional information

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