ORIGINAL ARTICLE



¹⁸F-FDG PET/CT for the quantification of inflammation in large carotid artery plaques

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Background. There is currently no consensus on the methodology for quantification of ¹⁸F-FDG uptake in inflammation in atherosclerosis. In this study, we explore different methods for quantification of ¹⁸F-FDG uptake in carotid atherosclerotic plaques and correlate the uptake values to histological assessments of inflammation.

Methods and Results. Forty-four patients with atherosclerotic stenosis ≥70% of the internal carotid artery underwent 18 F-FDG PET/CT. Maximum standardized uptake values (SUV_{max}) from all plaque-containing slices were collected. SUV_{max} for the single highest and the mean of multiple slices with and without blood background correction (by subtraction (cSUV) or by division (target-to-background ratio (TBR)) were calculated. Following endarterectomy 30 plaques were assessed histologically. The length of the plaques at CT was 6-32 mm. The 18 F-FDG uptake in the plaques was 1.15-2.66 for uncorrected SUVs, 1.16-3.19 for TBRs, and 0.20-1.79 for cSUVs. There were significant correlations between the different uptake values (r = 0.57-0.99, P < 0.001). Methods with and without blood background correction showed similar, moderate correlations to the amount of inflammation assessed at histology (r = 0.44-0.59, P < 0.02).

Conclusions. In large stenotic carotid plaques, ¹⁸F-FDG uptake reflects the inflammatory status as assessed at histology. Increasing number of PET slices or background correction did not change the correlation. (J Nucl Cardiol 2019;26:883–93.)

Key Words: $^{18}\text{F-FDG}$ PET/CT $^{\bullet}$ carotid plaque $^{\bullet}$ atherosclerosis $^{\bullet}$ inflammation $^{\bullet}$ quantification method

Abbreviatio	ons	ROI	Region of interest
PET	Positron emission tomography	MDS	Most diseased segment
¹⁸ F-FDG	2-deoxy-2-(¹⁸ F)-fluoro-D-glucose	TBR	Target-to-background ratio
SUV_{max}	Maximum standardized uptake value	cSUV	Corrected SUV

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The authors of this article have provided a PowerPoint file, available for download at SpringerLink, which summarises the contents of the paper and is free for re-use at meetings and presentations. Search for the article DOI on SpringerLink.com

David Russell and Mona-Elisabeth Revheim have contributed equally to this work.

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INTRODUCTION

Inflammation is a key factor in the pathophysiology of atherosclerosis with regard to progression and destabilization of plaques.¹ Patients with unstable carotid plaques have increased risk of plaque rupture and ischemic stroke,^{2,3} and there is increasing interest in imaging carotid plaque inflammation in order to detect these unstable plaques.

Positron emission tomography (PET) imaging with 2-deoxy-2-(¹⁸F)-fluoro-D-glucose (¹⁸F-FDG) of inflammation in atherosclerotic plaques has rapidly evolved since Rudd et al first reported ¹⁸F-FDG accumulation in macrophage-rich areas of carotid artery plaques over a decade ago.4 In contrast to oncology,5 there is no consensus on methodological guidelines for ¹⁸F-FDG PET/CT in atherosclerosis imaging. A recent position paper from the Cardiovascular Committee of the European Association of Nuclear Medicine⁶ proposed optimized and standardized protocols for the imaging and interpretation of ¹⁸F-FDG PET scans in atherosclerosis. However, they admitted that many of the recommendations suffer from the absence of conclusive evidence. Compared to a solid tumor, the cells responsible for ¹⁸F-FDG uptake in carotid artery plaques are generally fewer, more dispersed, and spread around parts of the circumference of a tubular vascular structure.^{3,7} Consequently, limited spatial resolution of the PET scanner and blood background activity are of great concern. Two parallel phenomena are known to influence measured activity in a lesion:⁸ signal from the lesion lost to the surroundings (spill-out), and signal added to the lesion from the vessel lumen and adjacent anatomic structures (spill-in).^{8,9}

Different acquisition protocols and quantification methods have been suggested. They all address the same concerns but have diverging solutions. A literature search identified 53 different acquisition protocols, 51 different reconstruction protocols, and 46 different quantification methods used in 49 studies. The most

common measure is the mean of all the maximum standardized uptake values (SUVs) (mean SUV_{max}) of the regions of interest (ROIs). The ROIs include the whole plaque (in localized stenosis), or one or more vessel segments (in subclinical/generalized disease). Bural et al calculated the atherosclerotic burden of the aorta by multiplying the mean uptake values for each aorta segment with the vessel wall volume. Subanalysis looking for the most metabolically active areas of a vessel segment have been used in therapy response studies. The uptake values are either normalized to the blood background activity, 15,17,19,20 corrected for the background activity with subtraction or not corrected for background activity. The rationale for background correction has been strongly criticized. 9,23

The aim of this study was to explore different methods for the quantification of ¹⁸F-FDG uptake in large carotid artery plaques, and to correlate the uptake values to the amount of inflammation on histology.

METHODS

Study Population

Forty-four patients referred to the Department of Neurology at our institution for the evaluation of carotid artery disease were included (Table 1). Inclusion criteria were ultrasound-confirmed atherosclerosis with internal carotid artery stenosis ≥ 70% according to consensus criteria of the Society of Radiologists in Ultrasound. Exclusion criteria were prior carotid endarterectomy/angioplasty with stenting, carotid occlusion, vasculitis, malignancy, prior radiation therapy to the neck, or immunotherapy. The Regional Committee for Medical and Health Research Ethics approved the study and all patients provided informed written consent.

Of the 44 included patients, 38 underwent carotid endarterectomy due to ipsilateral ischemic events or as prophylactic treatment before heart surgery. Eight of these plaques were lost to histological assessment due to logistical reasons.

¹⁸F-FDG PET/CT Examination

Following overnight fasting blood glucose level was measured before the patient received i.v. injection of 5 MBq/

Table 1. Patient characteristics (n = 44)

Age, years; mean ± SD	66.3±8.4
Sex, male; <i>n</i> (%)	30 (68.2)
Statin therapy; n (%)	34 (77.3)
Blood glucose, mmol L^{-1} ; mean \pm SD (range)	$6.8 \pm 2.2 (4.9-14.9)$
Bodyweight, kg; mean ± SD (range)	83.5 ± 16 (55-110)
Body mass index, kg/m^2 ; mean \pm SD (range)	27.4 ± 4.5 (19.9-34.8)

kg (0.14 mCi/kg). After a mean circulation time of 100 minutes (range 68-156), a ¹⁸F-FDG PET/CT scan from the base of the skull to the aortic arch was performed with 15 minutes per bed position using a hybrid PET/CT scanner (Siemens Biograph 64, Siemens Medical Systems, Erlangen, Germany). The PET data were reconstructed to 2-mm slices with a pixel size of 2.67 mm using the ordered subset expectation-maximization 2D algorithm with four iterations (i), eight subsets (s)(4i/8s), and Gaussian post-reconstruction filter with full-width at half maximum of 3.5 mm. For attenuation correction, low-dose CT without the use of *i.v.* contrast was performed immediately before ¹⁸F-FDG PET. For patients without a recent CT angiography of the carotid arteries, this was performed after ¹⁸F-FDG PET.

¹⁸F-FDG Uptake Quantification

The Hybrid Viewer 2.0 software (Hermes Medical Solutions AB, Stockholm, Sweden) was used for image fusion and ¹⁸F-FDG uptake quantification. The CT angiography was used to guide drawing of the ROIs on the fused PET/CT slices. A plaque was defined as vessel wall thickening and a lumen contrast-filling defect on CT angiography (Figure 1). 13 An experienced nuclear medicine physician (K.J.) drew ROIs around the entire vessel wall and lumen on all plaquecontaining axial PET slices (Figure 2). ROIs were carefully placed to minimize the influence from ¹⁸F-FDG uptake in structures close to the plaque (e.g., lymph nodes, paravertebral muscles, or salivary glands). Blood pool activity was obtained from four ROIs placed in the lumen of the jugular vein away from structures with ¹⁸F-FDG uptake. Plaque localization in relation to the carotid bifurcation was recorded. The most cranial slice of the common carotid artery before the division was defined as the bifurcation. The pixel values in the PET images were converted into SUV normalized to lean body mass.5

For all plaque ROIs, SUV_{max} was obtained and the ¹⁸F-FDG uptake was quantified using the following approaches (Figure 2):

- (1) Max SUV_{max} = the single highest SUV,
- (2) Mean SUV_{max} = the mean of all plaque SUV_{max} ,
- (3) Most Diseased Segment (MDS)3 = the mean SUV_{max} of the three contiguous slices centered on the slice with the highest SUV_{max} ,
- (4) MDS5 = the mean SUV_{max} of the five contiguous slices centered on the slice with the highest SUV_{max} ,
- (5) Mean $SUV_{max}4$ = the mean SUV_{max} of the four slices with highest SUV_{max} .

For all blood pool ROIs, SUV_{mean} was obtained. Blood background-corrected values for all the SUV measurements were calculated (target-to-background ratio (TBR); SUV divided by the blood pool activity (mean SUV_{mean} in four venous regions) and corrected SUV (cSUV); blood pool-corrected SUV (subtraction of the blood pool activity (mean SUV_{mean}) from SUV).

A second independent experienced nuclear medicine physician (MER) drew ROIs on the 20 initial patients to assess inter-observer variability of the different quantification methods.

Endarterectomy and Histological Analysis

The mean time between PET/CT and endarterectomy was 6 days (range; 0-116, median; 0). The histological analysis has been described previously. ²⁵ In brief, plaques were removed *en bloc* at carotid endarterectomy, fixed in 4% formaldehyde, decalcified in ethylenediaminetetraacetic acid, and cut into 2-to 3-mm slices. After dehydration, the slices were embedded in paraffin and a 5-µm histological section from each slice was cut and stained with hematoxylin and eosin (H&E). The number of histology sections obtained from each plaque

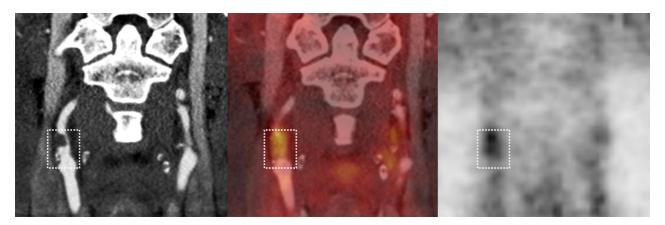


Figure 1. ¹⁸F-FDG PET/CT of stenotic plaque in the right internal carotid artery. From left to right: CT angiography, co-registered PET/CT, and ¹⁸F-FDG PET. The white box shows the plaque extension craniocaudally with lumen contrast-filling defect and vessel wall thickening on CT angiography (left image).

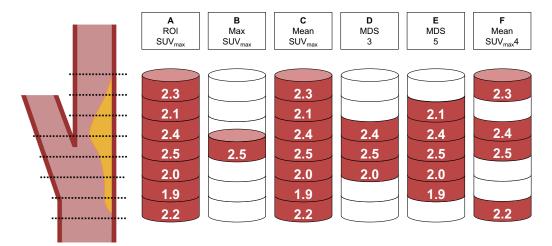


Figure 2. ¹⁸F-FDG uptake quantification was based on the SUV_{max} of all the plaque-containing axial PET slices (**A**): Max SUV_{max} = the single highest SUV_{max} (**B**), mean SUV_{max} = the mean of all plaque SUV_{max} (**C**), MDS3 = the mean SUV_{max} of the three contiguous slices centered on the slice with the highest SUV_{max} (**D**), MDS5 = the mean SUV_{max} of the five contiguous slices centered on the slice with the highest SUV_{max} (**E**), and mean SUV_{max} 4 = the mean SUV_{max} of the four slices with highest SUV_{max} within the plaque(F) (based on a figure by Tawakol et al ³²).

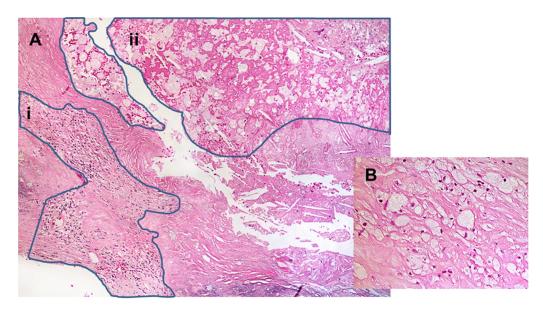


Figure 3. The histological quantification of inflammation was performed on H&E stained samples. The ocular micrometer was used to measure the total plaque area and area occupied by inflammatory cells in all the fields of view from all sections. The amount of inflammation per plaque was defined as the sum of all areas with inflammatory cells divided by the total area of all sections. **A**: ×200 magnification with inflammatory areas (marked by blue lines) containing mainly lymphocytes (*i*) and lipid macrophages (*ii*). **B**: ×400 magnification of areas with lipid macrophages.

ranged from 2 to 9 (mean 4.9, SD 2.0). The plaques were assessed by a pathologist and a research physician blinded for the clinical and the ¹⁸F-FDG PET findings. Inflammation was quantified using a modified version of the method used by Jander et al.³ The sections were evaluated with 120× magnification and the percentage area of inflammatory cells

(macrophages and leucocytes) per plaque was obtained (Figure 3): For each section, the total area and the area occupied by inflammatory cells were measured manually. The amount of inflammation per plaque was defined as the sum of all areas with inflammatory cells divided by the total area of all the sections. We performed repeated histopathological

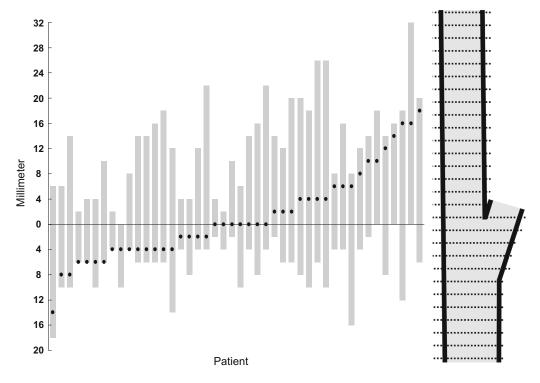


Figure 4. Plaque extension (gray bars) and max SUV_{max} location (black dots) per patient along the *x*-axis. The *y*-axis shows the distance in millimeter from the bifurcation (0) cranially in the internal carotid artery and caudally in the common carotid artery.

assessment on selected sections from this study cohort and found that the amount of inflammatory cells was in the same 5% category at both assessments for 73% of the sections (Kappa = 0.73). The amount of inflammation per plaque was 6.8% (SD 4.0; range 0.4-17.9).

Statistical Analysis

The SPSS Statistics software for Windows (IBM, version 21.0; SPSS Inc., Chicago, Ill) was used. Groups of data were compared using two-sided *t*-test and Pearson correlation for normally distributed variables. For non-normal distributions Mann-Whitney test and Spearman correlation were used. Statistical significance was set to 0.05.

RESULTS

Localization of Plaque and Highest ¹⁸F-FDG Uptake

The length of the plaques in the cranio-caudal direction was 6-32 mm (mean 19, SD 7.6) and all included the carotid artery bifurcation (Figure 4). Max SUV_{max} was located between 14 mm below to 18 mm

above the bifurcation (mean 1.2 mm below the bifurcation) (Figure 4).

Inter-observer Variability

The correlation coefficients between ¹⁸F-FDG uptakes calculated from plaques delineated by two nuclear medicine physicians independently were 0.96-0.98 for uncorrected SUVs, 0.63-0.68 for TBRs, and 0.90-0.93 for cSUVs. The correlation coefficient for background blood pool activity was 0.75.

Quantification of ¹⁸F-FDG Uptake

Mean values, SDs, and ranges for the different quantification methods are summarized in Table 2. The 18 F-FDG uptake was significantly different for the different quantification approaches (paired samples t-test, P < 0.004 for all pairs). TBR gave the highest mean values and the widest ranges, whereas the background-subtracted values (cSUV) showed the lowest mean values and the narrowest ranges.

 $^{18}\text{F-FDG}$ uptake for the different quantification methods and corresponding background values for all patients are shown in Figure 5. Mean difference between max $\mathrm{SUV}_{\mathrm{max}}$ and mean $\mathrm{SUV}_{\mathrm{max}}$ was 0.08 and ranged from 0.02 to 0.66. The effect of background correction is shown in Figure 6. Correlations between $^{18}\text{F-FDG}$ uptake within the groups were 0.93-0.99 for the uncorrected SUV, 0.94-1.0 for TBR, and 0.92-0.99 for cSUV (Table 3).

Histology and ¹⁸F-FDG Uptake

There were significant moderate (0.44-0.59) correlations between the amount of inflammation and all the different $^{18}\text{F-FDG}$ quantification methods (Table 4). The highest correlations were found for mean SUV $_{\rm max}$, and the lowest for max SUV $_{\rm max}$ independent of background correction. Figure 7 shows scatter plots of inflammation versus max and mean SUV $_{\rm max}$ with and without background correction.

Table 2. Plaque SUV for different quantification methods (n = 44)

	Uncorrected	TBR	cSUV
Max SUV _{max}	1.76 ± 0.35 (1.18-2.66)	2.07 ± 0.44 (1.34-3.19)	0.90 ± 0.33 (0.42-1.79)
Mean SUV _{max}	1.56 ± 0.28 (1.11-2.28)	1.83 ± 0.38 (1.16-2.91)	$0.69 \pm 0.28 (0.20-1.29)$
MDS3	1.70 ± 0.34 (1.17-2.51)	2.00 ± 0.43 (1.26-3.16)	$0.83 \pm 0.32 (0.33-1.64)$
MDS5	1.66 ± 0.32 (1.15-2.32)	1.95 ± 0.41 (1.22-3.14)	$0.79 \pm 0.31 (0.28 - 1.45)$
Mean $SUV_{max}4$	1.68 ± 0.33 (1.15-2.45)	1.98 ± 0.42 (1.26-3.16)	0.82 ± 0.31 (0.32-1.58)

Data are given as mean ± SD (range)

TBR, target-to-background ratio; SUV, standardized uptake value; cSUV, corrected SUV (background-subtracted SUV); MDS, most diseased segment

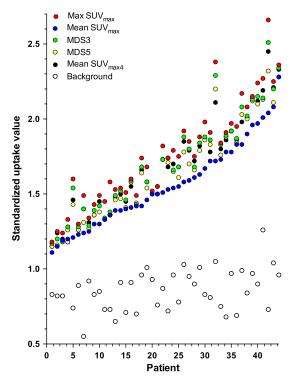


Figure 5. Uncorrected 18 F-FDG uptake values for individual patients sorted according to increasing mean SUV_{max} . Each color represents separate quantification methods. The black dots are the background values (mean SUV_{mean}).

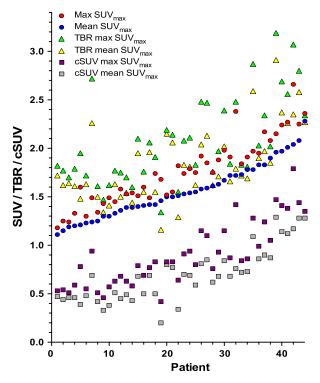


Figure 6. Uncorrected and background-corrected (TBR and cSUV) max and mean SUV $_{\rm max}$ per patient sorted according to increasing mean SUV $_{\rm max}$.

Table 3. Correlation coefficients between different methods for $^{18}\text{F-FDG}$ uptake quantification (n=44)

		Unco	Uncorrected				TBR					cSUV		
	Mean SUV _{max} MDS3 MDS5 SI	MDS3	MDS5	Mean UV _{max} 4	Max SUV _{may}	Mean SUV _{max} M	MDS3	MDS3 MDS5	Mean SUV _{max} 4	Max SUV _{may}	Mean SUV _{max} M	MDS3	MDS3 MDS5	Mean SUV _{max} 4
Uncorrected														
Max SUV _{max}	.93 *	*66:	*66:	*66:	*89:	.57*	*89.	*29.	.65	*16:	*62.	*68.	*68.	*88.
Mean SUV _{max}	_	*96	*96	*96:	*49:	.65	*69	*99:	.65*	.85	* 28.	*98.	*78.	*98.
MDS3		_	*66	*66	*69:	.59	*69:	*89:	*29.	*16.	*18:	*16.	*06:	*68.
MDS5			_	*66	*19.	*65.	*19.	*89:	.65*	*68.	*18:	*68.	*06:	*88.
Mean $SUV_{max}4$				-	*69:	.62*	*69.	*69°	*89:	*16.	*83	*06:	*06:	*06:
TBR														
Max SUV _{max}					1	* 46.	*66	*66	*66.	*16:	*68.	.92*	*16:	*26.
Mean SUV _{max}						_	*96	* 26.	*16.	.82*	.93	.85	*98.	*78.
MDS3							1	1.0*	1.0*	*16.	*06:	*26	*26	*86.
MDS5								_	*66	*06:	*16.	*16.	*26	*26.
Mean SUV _{max} 4									_	*68	*06:	*06:	*06:	* 76:
cSUV														
Max SUV _{max}										_	* 76.	*66	*66	*66.
Mean SUV _{max}											_	*46.	*36.	*96
MDS3												1	*66	*66.
MDS5													_	*66:
Mean $SUV_{max}4$														_

SUV, standardized uptake value; MDS, most diseased segment, background-corrected data; TBR, target-to-background ratio; cSUV, background-subtracted SUV *Correlation is significant at the 0.01 level

Table 4. Correlation between ¹⁸F-FDG PET uptake values and histology (n = 30)

¹⁸ F-FDG quantification method	Spearman correlation coefficient
Max SUV _{max}	0.48 (0.008)
Mean SUV _{max}	0.54 (0.002)
MDS3	0.48 (0.007)
MDS5	0.49 (0.006)
Mean SUV _{max} 4	0.52 (0.003)
TBR max SUV _{max}	0.44 (0.016)
TBR mean SUV _{max}	0.58 (0.001)
TBR MDS3	0.47 (0.009)
TBR MDS5	0.48 (0.008)
TBR mean SUV _{max} 4	0.48 (0.007)
cSUV max SUV _{max}	0.47 (0.009)
cSUV mean SUV _{max}	0.59 (0.001)
cSUV MDS3	0.52 (0.004)
cSUV MDS5	0.52 (0.003)
cSUV mean SUV _{max} 4	0.54 (0.002)

Data given as correlation coefficient (P value)

DISCUSSION

In this clinical study, we explored different methods for the quantification of ¹⁸F-FDG uptake in carotid plaques causing artery stenosis equal to or above 70%. ¹⁸F-FDG uptake was homogenously disseminated throughout the entire plaques. Although there were differences in magnitude, quantification of ¹⁸F-FDG uptake with and without background correction showed similar, moderate correlations to inflammation on histology.

Mean max SUV_{max} (mean of the max SUV_{max} for the study population) was only 13% higher than mean mean SUV_{max} (Figure 8), whereas the three other quantification methods gave values in between, increasing with decreasing number of included slices. Homogenous ¹⁸F-FDG uptake throughout the plaque contrasts findings from microPET of endarterectomized plaques showing patchy ¹⁸F-FDG uptake. ²⁶ The presence of macrophages reduces the thickness of the fibrous capsule and therefore an increasing number is likely to correlate to increasing vulnerability. As such, the highest ¹⁸F-FDG uptake within a plaque could be the most appropriate parameter for risk assessment. Currently, the most used measure for assessment of plaque inflammation has been the mean SUV_{max}. 17,21,22 No clinical studies have assessed the use of whole plaque max SUV_{max}. Max SUV_{max} is easily obtained, highly reproducible, and less influenced by partial-volume effects. $^{\!8,27}$ Although max SUV_{max} is prone to image noise,8 our findings suggest that for atherosclerotic plaque assessment in clinical PET, max SUV_{max} should be explored further.

The strong correlation between the different quantification methods suggests that for this group of patients uncorrected ¹⁸F-FDG uptake values may provide similar information as background-corrected values. As such, our findings do not support the superiority of TBR as quantification method as suggested by others. 4,10,14-17 For circulation times above one hour, the blood background is low, but highly variable, whereas carotid plaques have consistent uptake over time. 11,13 A slight variation in blood background will therefore give significant variability in TBR. From a physics perspective, Huet et al 9 have explained that there is no legitimate rationale for using TBR instead of SUV because blood contamination is an additive and not a multiplicative process. A purely additive process would require subtraction of the mean luminal blood pool activity from the SUVs (cSUVs).

The inter-observer variability analysis of ¹⁸F-FDG quantification revealed superior reproducibility of uncorrected SUV_{max} compared to blood background-corrected SUVs (TBR and cSUV). SUV measurements are highly dependent on the size, shape, and location of the drawn ROI because the ROI can either miss the voxel with the highest intensity or the ROI can inadvertently include contribution from adjacent ¹⁸F-FDG-avid organs. The use of contrast-enhanced CT to localize the plaques is likely to have contributed to the low inter-observer variability in the SUV_{max} in our study. Our correlation coefficient for blood background of 0.75

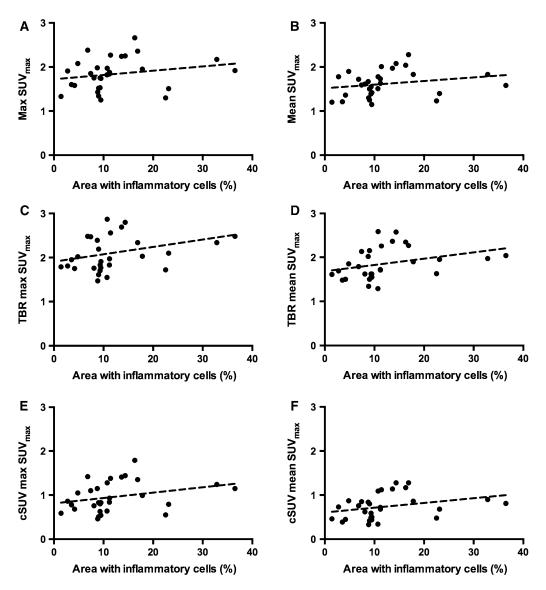


Figure 7. Scatter plots with correlation lines of total area with inflammatory cells versus uncorrected (A, B) and background-corrected (TBR: C, D) and cSUV: E, F) max and mean SUV_{max} .

inevitably increases the inter-observer variability of all background-corrected values. Other ¹⁸F-FDG uptake reproducibility studies in localized carotid artery plaques have reported moderate (background corrected)²⁸ to excellent (with and without background correction)^{22,29} inter-observer agreement. The slightly inferior inter-observer agreement for background-corrected values found in our study could be related to differences in the placement of the ROI in the jugular vein as the diameter of the jugular vein often was small, making the measurement susceptible to image noise. However, this is an inherent limitation of all quantification methods with background correction.

The optimal method should predict plaque vulnerability and clinical outcome. In the present study, all quantification methods for $^{18}\mbox{F-FDG}$ uptake showed moderate correlation to inflammation on histopathology (Table 4). The correlations were systematically slightly higher when mean SUV $_{\rm max}$ was used instead of max SUV $_{\rm max}$. This was not unexpected as total plaque inflammation score and mean SUV $_{\rm max}$ are both multislice methodologies. Total plaque inflammation score is a well-established method to correlate histology to ischemic symptoms, $^{2.3}$ or to $^{18}\mbox{F-FDG}$ uptake. 17 When comparing max SUV $_{\rm max}$ with the slice with the highest percentage inflammatory area, the correlation did not

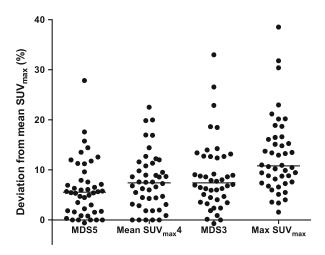


Figure 8. Deviation from the mean SUV_{max} for different quantification methods for individual patients. The black bar represents the mean values.

increase (data not shown). This is in accordance with Tawakol et al¹⁷ who found slightly higher correlations between overall plaque inflammation and histology than a slice-by-slice comparison of uptake value and histology. How to use histology as a gold standard to imaging is challenging. We know that excised plaques both shrink¹⁷ and may be partly damaged.^{17,25}

The strength of our study was the close timing between the PET examinations and the endarterectomies. Our study patients had plaques not only presumed highly inflammatory giving recent symptoms, but also plaques removed prophylactically from asymptomatic patients.

A limitation of our study is the inclusion of patients with elevated blood glucose. Four had blood glucose values > 11 mmol/L (198 mg/dL) and three 7-11 mmol/ L (126-198 mg/dL) at the time of the ¹⁸F-FDG injection. Elevated blood glucose is known to reduce the uptake of ¹⁸F-FDG into metabolic active cells in malignant diseases.³⁰ Guidelines for the clinical use of ¹⁸F-FDG in inflammation and infection³¹ also recommend the reduction of blood glucose to the lowest possible level. In studies on atherosclerosis, there is no consensus on a cutoff value. 12 Some studies do not report on blood glucose level, whereas in other studies the cut-off values have ranged from 8 mmol/L (144 mg/dL)²¹ to 11.1 mmol/L (200 mg/dL). ¹⁶ Our correlations between histology and the different ¹⁸F-FDG quantification methods were slightly increased when excluding the four patients with blood glucose level > 11 mmol/L. We have not excluded patients with high blood glucose in the correlation analysis of the different uptake parameters and thereby we do not know if this has contaminated our results. In the 44 included patients, there was no correlation between blood glucose and background SUV_{mean}. Another

limitation is the wide range of circulation times (68-156 minutes) that could have influenced the plaque ¹⁸F-FDG uptake by underestimating background-corrected values for patients with shorter circulation times and by overestimating background-corrected values for patients with longer circulation times. However, for 38 of the 44 patients the PET acquisition started between 85 and 115 minutes after ¹⁸F-FDG injection, and thus, it is unlikely that difference in circulation time would change the findings in our study.

NEW KNOWLEDGE GAINED

Our study showed that SUVs without background correction from large plaques in the carotid artery can be used as inflammatory parameter in atherosclerosis.

CONCLUSION

In conclusion, in carotid artery stenosis equal to or above 70%, ¹⁸F-FDG uptake reflects the inflammatory status as assessed on histology. Increasing number of PET slices or background correction did not improve the correlation.

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Disclosure

KJ, KS, TS, MS, DR, and MER report no conflicts of interest.

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