

Research

Open Access

Enhancement of Tissue Expansion by Calcium Channel Blocker: A preliminary study

Eray Copcu*¹, Nazan Sivrioglu¹, Nejdet Sisman², Alper Aktas² and Yucel Oztan²

Address: ¹Department of the Plastic and Reconstructive Surgery, Medical Faculty, Adnan Menderes University, Aydin, Turkey and ²Department of the Plastic and Reconstructive Surgery, Atatürk Training Hospital, Izmir, Turkey

Email: Eray Copcu* - copcu@lycos.com; Nazan Sivrioglu - nsivrioglu@adu.edu.tr; Nejdet Sisman - nsisman@yahoo.com; Alper Aktas - aaktas@msn.com; Yucel Oztan - yuceloztan@yahoo.com

* Corresponding author

Published: 09 October 2003

Received: 09 July 2003

World Journal of Surgical Oncology 2003, 1:19

Accepted: 09 October 2003

This article is available from: <http://www.wjso.com/content/1/1/19>

© 2003 Copcu et al; licensee BioMed Central Ltd. This is an Open Access article: verbatim copying and redistribution of this article are permitted in all media for any purpose, provided this notice is preserved along with the article's original URL.

Abstract

Background: Reconstruction of the defects after surgical resection of tumors is one of the important issues in surgical oncology. It is essential that the defect should be covered with a tissue quite similar to the original one and is best achieved by harvesting tissue from an area adjacent to the defect. Tissue expansion is one of the most frequently used reconstructive techniques. A number of studies evaluated blood circulation, capsule formation, tissue tolerance, histomorphological changes and complications of expander placement. However, only a few attempted to enhance tissue expansion. This study we aimed to evaluate verapamil, a calcium channel blocker, to enhance tissue expansion.

Material and method: Twelve New Zealand rabbits weighing between 900 gm and 1200 gm were assigned into study and control groups. High volume expanders (100, 200 or 300 cc) were placed into the subcutaneous tissue. Rabbits in the study group received verapamil. Expanders in the control group were inflated every three days to achieve same pressure as the study group. The size of the flaps was assessed by applying pressure on tip of the flap to demonstrate the contraction. Histopathological examinations were performed.

Results: By administering liquid earlier and more quickly less flap retraction was observed in the study group. In the control group expanders were exposed in two rabbits while no complication occurred in the study group. Following extraction of the expanders, the flaps were elevated and less retraction was observed in the study group compared to controls.

Conclusion: Verapamil is safe when used topically and provides less retracted flaps. It can be suggested that verapamil acts on the myofibroblasts in the capsule around tissue expanders and thus increases efficiency of the expanders.

Introduction

Covering of the defect after surgical resection of the primary or metastatic neoplasm is essential and the best tissue for such coverage is the one, which is similar to the original resected tissue. This is best achieved by harvesting a tissue from an area adjacent to the defect. However, harvesting a large local flap from an adjacent tissue will lead to formation of an additional defect at times. In addition, the adjacent area may not provide an appropriate size tissue. The use of tissue expanders may allow an original-like tissue for coverage. Tissue expansion is a mechanical procedure, which expands the surface of an available local area [1]. All living tissues respond to mechanical strength and the mitosis in these tissues increases as the tissues gradually expand. Clinical application of tissue expanders is widespread and the expanders are indispensable for reconstruction of large defects resulting from surgical treatment of tumors, many congenital anomalies and acquired defects. Expanders are most frequently used in reconstruction of breast and scalp defects [2,3]. Schmidt *et al*, summarized the advantages of expanders as: a) best tissue match with defect areas, b) lack of a wound at the donor site, and c) maintenance of vascular supply [4]. It has been demonstrated that a fibrous capsule was formed around tissue expanders and examination of this capsule and the surrounding dermis on electron microscopy has shown an increase in the number of contractile fibroblasts [5]. These fibroblasts contain actin and myosin, which account for not only wound healing but also for formation of a fibrous capsule around the expanders [6]. Verapamil, a safe and widely used calcium channel blocker, was used in the present study as it has a topical effect on the collagen tissue fibroblasts with an aim to determine whether it can decrease pressure around the expanders and thus increasing efficiency of tissue expanders.

Material and method

The study was performed in Experimental Laboratory of Plastic and Reconstructive Surgery Clinic in Izmir Atatürk Teaching Hospital, Turkey, using twelve New Zealand rabbits weighing between 900 gm and 1200 gm. There were six rabbits in verapamil group and six rabbits in control group. Within each group, 100, 200 and 300 cc expanders were used in two animals each. Approval was obtained from the ethical committee of the hospital and the animals were kept in separate cages in temperature, light and airflow-regulated rooms. Experiments were conducted under aseptic conditions. The rabbits were anesthetized with 1 ml ketamine hydrochloride 50 mg/ml and 1 ml of 2% xylazine hydrochloride.

High volume rectangular expanders (McGhan®, Medical Corporation, USA) were used. The rabbits were assigned into a study group to receive verapamil and into a control group and each group was divided into three subgroups

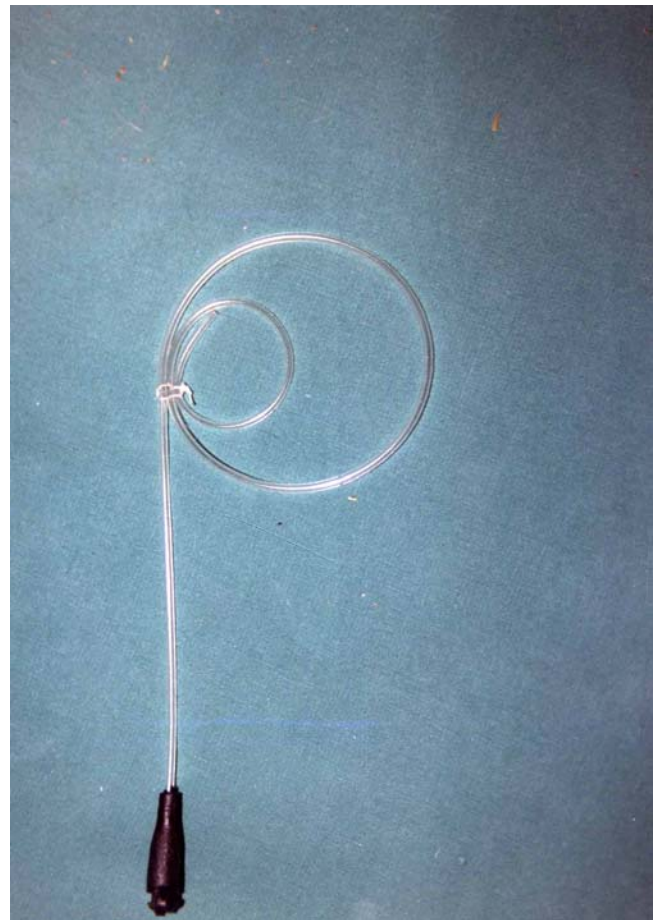


Figure 1
Apparatus for administration of verapamil

based on the volume of the expanders; Group I: 100 cc, Group II: 200 cc and Group III: 300 cc. A reservoir was connected to each expander with a 3 cm silicon tube.

Apparatus for administration of verapamil

Verapamil was administered using a special cannula: Thirty holes were drilled on an 2 mm diameter 30 cm long intravenous catheter and one end of the catheter was blocked (Figure 1). Thus, the drug was spilled only through the holes and spreads on the base where the expanders were placed. The cannula was twisted to form a spiral and sutured with 5/0 silk. This apparatus was sterilized in a gas autoclave.

The back of the rabbits were shaved and cleaned with povidon iodide and a 3 cm horizontal incision was made 5 cm away from the tail. After dividing the cutaneous and subcutaneous tissue a diverticulum was created on the panniculus carnosus. At this level, a dissection of the same



Figure 2
Photograph showing application of the apparatus

size as the base of the expander was made followed by placement of the cannula and the expander into the diverticulum (Figure 2). The reservoir of the expander was placed 2 cm away from the expander in a postero-lateral position. One end of the cannula was left outside the tissue and the skin was sutured with 3/0 prolene. Penicillin was administered once to all animals for prophylaxis.

Measurement of expansion

After the expanders were placed, four spots of crystal violet were created with a caliper on two separate planes on the shaved skin of the rabbits (Figure 3). The length and width of the expanders were labeled as AB and XY respectively. Following each expansion, the distances between the spots was measured.



Figure 3
Photograph showing AB and XY points of the tissue expansion in rabbit.

Administration of verapamil

Immediately after the expanders were placed and every three days thereafter, 2.5 ml verapamil hydrochloride (5 mg/2ml Isoptin®, Knoll Inc.) mixed with 1 cc NaCl was administered in the study group, and only 1 cc NaCl was given in the control group.

Expansion of expanders and measurement of intra-expander pressure

The expanders were not inflated immediately after they were placed, but inflation was started three days later with 15–20% inflation every three days and all expansions took place at the same time postoperatively. At each expansion, first intra-expander pressure was measured in the control group and then the expanders in the verapamil group were inflated with saline equal to this pressure. When the expansions were completed, pressure in each

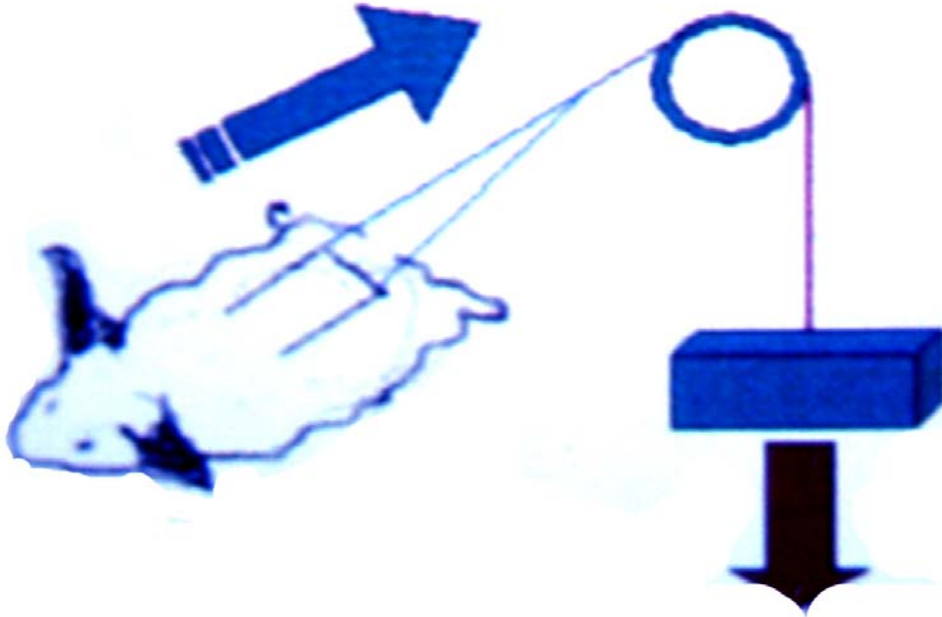


Figure 4
Schematic view of the application of 2.46 Newton force to the tip of flaps with pulleys.

expander, amount of expansion and changes in the flaps were recorded.

Determination of retraction ratios in flaps

The length of the flaps was measured twice to determine retraction ratios. The first measurement was made after the procedure of expansion was completed and the second measurement was made after the flaps were elevated. Following the elevation of the flaps, using a system of pulleys, a force of 2.46 Newton was applied on the tips of the flaps and the length of the flaps under the pressure was measured (Figure 4). Thus, the difference between the two values provided retraction ratios.

Evaluation

The differences between AB and XY lengths obtained after the completion of the expansion and those obtained after the elevation of the flaps were determined. In addition, skin specimens were collected from the study and control groups following the completion of the expansions. The specimens were stained with hematoxylin and eosin and were examined. Radiological examinations were performed to determine the apparatus function and its placement (figure 5).

Results

None of the rabbits died during the experiments. Verapamil injections were well tolerated by the rabbits and it did not cause any systemic changes or systemic or local intoxication in the rabbits. However, in the control group, one rabbit with an expander of 200 cc and one rabbit with an expander of 300 cc experienced exposition of their expanders on day 18 and 15 respectively (figure 6). No infection was detected during the procedures of expansion. However, one rabbit was observed to have immobility in his hindquarters. This may have been due to compression on the nerves.

Details of the expansions are shown in Table 1,2,3. The expansion was completed earlier in all sizes of expanders in the study group than in the control group. The extents (ABs and XYs) in all verapamil groups were longer than those in the control groups. After the expansion was completed, the expanders were removed and superior pedicled expanded flaps 50 × 30 mm in size were harvested. The sizes of the flaps measured after the application of an equal pressure on the tips of the flaps are presented in Table 4. The retraction ratios were lower in the study group than in the control group.

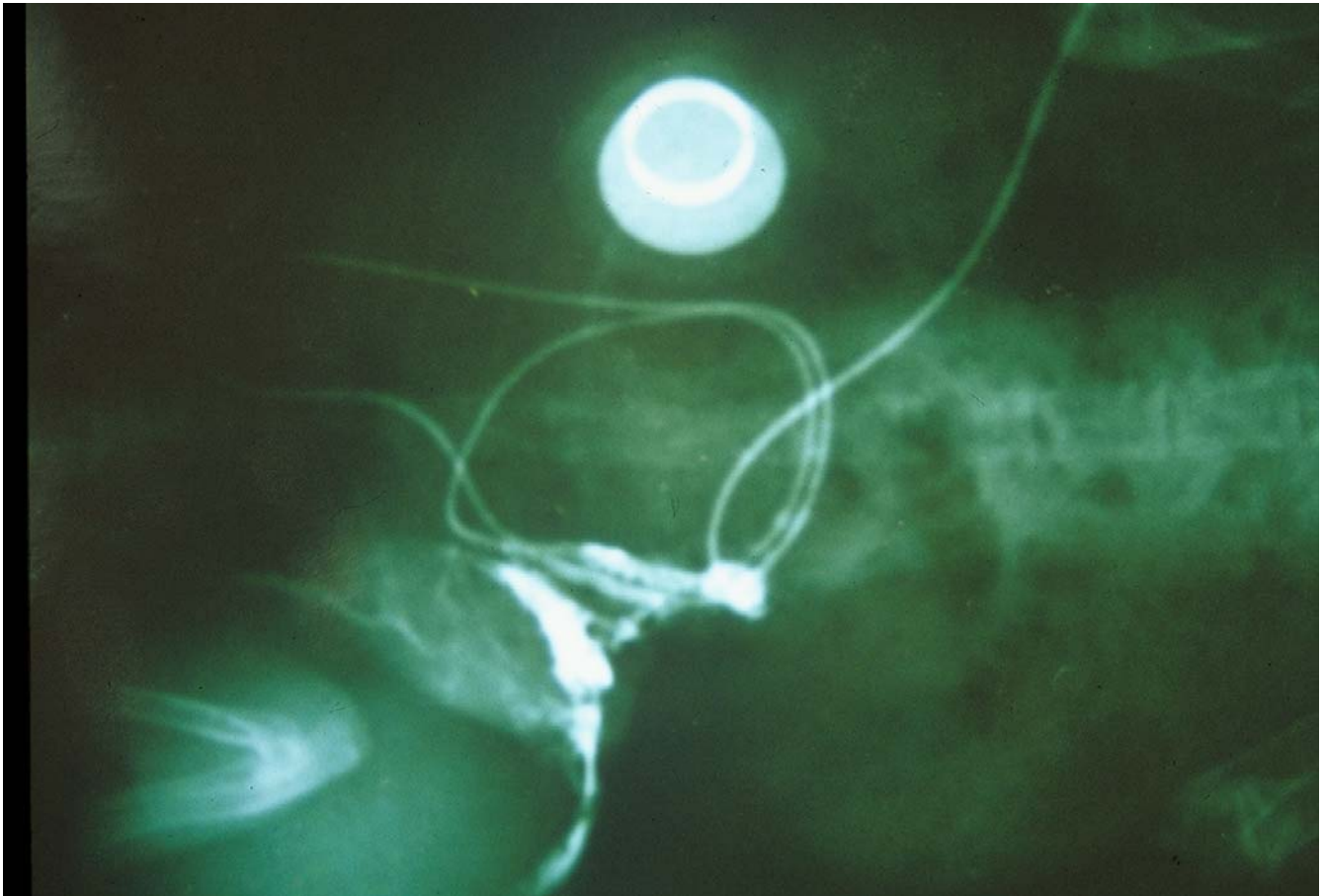


Figure 5
X-Ray of the apparatus.

Table 4: Rate of retractions of the flaps in each group after elevation.

Group	Expander volume	Retraction of AB length	Retraction of XY length
Control	100 cc	13%	13.30%
Verapamil	100 cc	8%	8.33%
Control	200 cc	18%	20%
Verapamil	200 cc	11%	15%
Control	300 cc	22%	33%
Verapamil	300 cc	13%	20%

There were two rabbits in each group, two rabbits were excluded: one rabbit in control group of the 200 cc expander, one rabbit in control group of 300 cc expander.

The microscopic examination of the specimens taken from each flap revealed that in the study group there was no thickening in the epithelial tissue, the capsule around the expander was thinner, the amount of collagen was less and that cellularity was less and scattered (Figure 7 and 8).

Discussion

Tissue expansion is a widely used technique in surgical oncology. Many reports have been published in the literature about reconstruction of the breast, head and neck, face, abdominal wall and trunk with tissue expanders after

Table 1: Details of expansions in subgroup I (100 ml expanders group)

Details	group	D0	D3	D6	D9	D12	D15	D18
Pressure (mm Hg)		0	5	8	10	20	27	30
Inflated volume (percent)	Saline	0	15	31	45	60	81	100
	Verapamil	0	21	47.5	74.5	103.5		
AB length (mm)	Saline	50	52	55.5	58	58	59.5	60.5
	Verapamil	50	53	58.5	64.5	66.5		
XY length (mm)	Saline	30	32.5	34.5	37	37.5	37.5	38
	Verapamil	30	35	41	42.5	46		

D-days.

Table 2: Details of expansions in subgroup II: 200 ml expanders group

Details	group	D0	D3	D6	D9	D12	D15	D18
Pressure (mm Hg)		0	3	6	8	30	32	39
Inflated volume (percent)	Saline	0	15.5	31	45.5	62	80	99
	Verapamil	0	22	42	65	89	106.5	
AB length (mm)	saline	50	52.5	55	59.5	62.5	66	67
	verapamil	50	54.5	61	67	69	75	
XY length (mm)	saline	40	41.5	44.5	48	49.5	51	53
	verapamil	40	47	53	57	59	61.5	

There were two rabbits in each group at the beginning of the experiment but one rabbit in saline group was excluded in day 18. D-days

Table 3: Details of expansions in subgroup III: 300 ml expanders group

Details	group	D0	D3	D6	D9	D12	D15	D18	D0
Pressure (mm Hg)		0	8	10	18	22	25	38	45
Inflated volume (percent)	saline	0	15	29	44	60	77	87	100
Inflated volume (percent)	verapamil	0	18	38	59	78	95	108	
AB length (mm)	saline	70	73.5	77	80.5	84	88	90	94
	verapamil	70	77.5	82.5	89.5	103	106	108	
XY length (mm)	saline	50	52	54	56	59	60	62	65
	verapamil	50	58.5	63.5	65	70	73	76	

There were two rabbits in each group but one rabbit in saline group was excluded in day 15. D-days

cancer treatment [7–10]. Neumann first reported the concept of tissue expansion in 1957 [11]. He described advantages of this technique as: high quality of tissue match, avoidance of the need for a donor site, and reduction in the number of procedures necessary for reconstruction. In 1976, Radovan used tissue expanders for reconstruction of the breast after mastectomy [12]. Subsequently, the use of tissue expansion has been popularized among plastic surgeons and has become the treat-

ment of choice for many congenital and acquired defects in children and adults [13–16].

Tissue expansion has two disadvantages; one it is time-consuming and second it causes complications. Exposition of the expanders is frequently encountered [17,18]. In practice, the most frequently encountered complication is capsule formation around the expanders. This capsule not only causes difficulties in the procedure of expansion but also decreases the size of flaps obtained



Figure 6
Complication in Group III: Exposed tissue expander.

after the expansion. Scar formation around the capsule may result in reduced skin elasticity and impeded flap movement, and expanded skin has a tendency to postoperative retraction [19]. Tension wounds have been shown to demonstrate increased tensile strength as a result of enhanced collagen alignment [20]. The response of the collagen to the expansion has been studied in detail by Timmenga [21]. Accelerated organization of the wound collagen associated with increased tensile strength can be considered as early maturation. The use of drugs affecting collagen synthesis and cross linkage, both in the capsule and in the surrounding dermis, might be of some value in decreasing mechanical resistance of expanded tissues, while administration of topical vasodilator drugs might increase the filling safety margin, thus reducing the risk of ischemia during the expansion [22]. Verapamil is found to be effective on anti-intimal hyperplasia in a number of animal models and decreased the myofibroblast cell proliferation in the intima [23]. Verapamil is widely used topically in the treatment of Peyronie's disease and keloid because of its effects on the collagen metabolism [24–27]. Calcium antagonists increase the extra-cellular matrix collagenase activity as well as decreasing the collagen, and fibronectin synthesis and secretion, altering fibroblastic metabolism. Lee *et al*, speculated that membrane calcium channel blockers could potentially trigger extracellular matrix degradation in dermal scars, resulting in scar volume reduction [28].

Timmenga *et al* concluded in their study that an expander placed between the skin and panniculus carnosus in the rabbit causes considerable skin stretch and affects cutaneous blood supply and therefore that it can be improved for tissue expansion in humans [21]. To date, investiga-

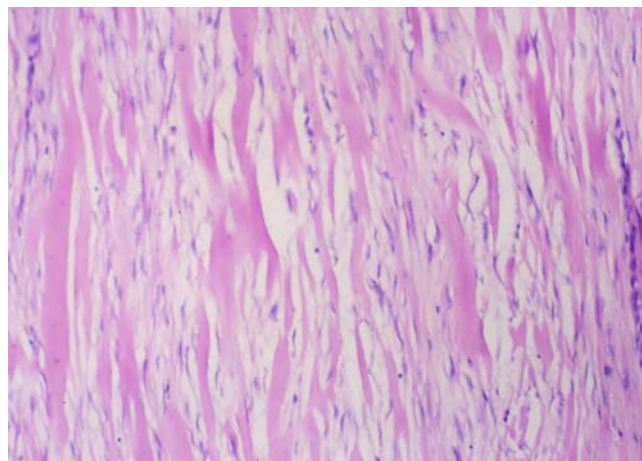


Figure 7
Photomicrograph of the subcutaneous tissue from verapamil group showing lesser and scattered cellularity on the subcutaneous tissue. There are no inflammatory cells. (Hematoxylin eosin staining × 100 magnification)

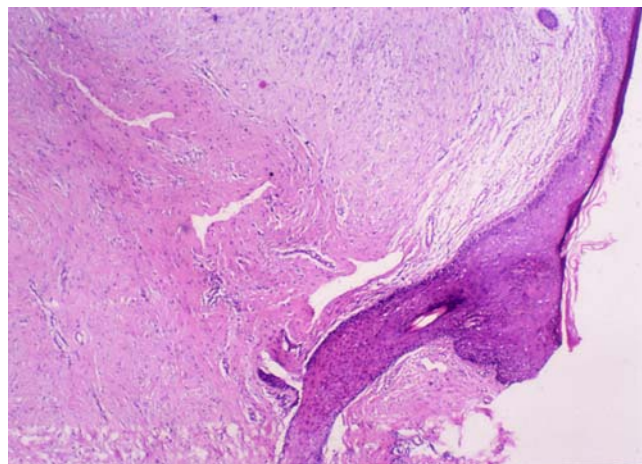


Figure 8
Photomicrograph of the subcutaneous tissue from control group showing some inflammatory cells and slightly thickened epidermis with 4 to 10 mitoses. More cellularity was seen than the verapamil group. (Hematoxylin eosin staining × 40 magnification)

tors have used low volume expanders (30–100 cc), while we preferred high volume expanders, which allowed us to evaluate frequently encountered complications in tissue expansion. In the control group in which we used the expanders of 200 cc and 300 cc, two expanders were exposed. We hypothesized that the exposition might be

associated with the pressure to which the skin is exposed and we concluded that verapamil caused relaxation in the tissues around the expander, which provided a safer expansion. Moreover, verapamil allowed the use of high volume expanders in the rabbits. There have been a number of studies on blood circulation to the tissues where expanders are placed [28], formation of the capsule, tolerance of the tissues, histomorphological changes [29] and the complications [18] since 1976. However, there have been only a few studies on the increased efficiency of the expanders. Brobmann and Huber thoroughly investigated the relation between the shape of the expanders and the outcome [30]. Their results showed that less time, pressure, and volume are needed in a larger implant to gain the same surface area as would be attained in several smaller implants. Lee *et al* investigated effects of chemical substances on the efficiency in 1985 [31] using papaverine and cytochalasine D. Other compounds like teophiline [32], dimethyl sulfoxide (DMSO) [33,34] and papaverine too have been used in humans intraoperatively [35]. The agents used to increase the efficiency of the expansion so far, especially cytochalasine D, have been reported to have toxic effects [31]. Lee in his study explained the effects of anti-contractile agents and noted that contractile fibroblasts appeared after application of tension on the dermis for only four days [36]. If the contractile actions of these cells can be diminished, then presumably the dermis will be more easily stretched and epidermal proliferation will maintain an adequate surface layer over the expanding dermis.

We made use of an ordinary catheter and invented an apparatus to administer verapamil. We also demonstrated on radiological examination that the apparatus functioned well. The mean intra-expander pressure was equal in the study and control groups. However, the tissue expansion was greater in the study group. It may be that verapamil, a muscle relaxant, may have prevented the resistance against the expansion.

As mentioned above, we measured the distances between the spots on the skin made with crystal violet, which is a standard method used to evaluate the expansion. However, there is no formula to determine efficiency of expanders precisely. The efficiency depends on the volume and the location of expanders and etiological factors, and it is claimed that an expansion of 30% can be achieved [37]. In this study, we obtained an expansion ranging from 21% to 34% in the control group and an expansion ranging from 33% to 54% in the study group.

Although there has been no study investigating the retraction ratio in the literature, it has been reported that the body responds quickly to the removal of the mechanical effect exerted by the expander. In practice, it is commonly

encountered that a little retraction of the flaps is possible after the expander is extracted and until the flaps are transferred into the recipient area. In this study, the retraction ratios determined after the elevation of the flaps were lower in the rabbits administered verapamil.

Conclusions

The topical usage of verapamil increases the efficiency of expanders, frequently used in reconstructive surgery, and decreases the complications. Thus, it provides less retracted flaps in larger volumes and in a very short time. Verapamil is safer and more potent than the other anti-contractile agents, which increases its use in practice.

References

1. Argenta LC: **Controlled tissue expansion in reconstructive surgery.** *Br J Plast Surg* 1984, **37**:520-529.
2. Manders EK, Au VK and Wong RK: **Scalp expansion for male pattern baldness.** *Clin Plast Surg* 1987, **14**:469-475.
3. Radovan C: **Breast reconstruction after mastectomy using the temporary expander.** *Plast Reconstr Surg* 1982, **69**:195-208.
4. Schmidt SC, Logan SE, Hayden JM, Ahn ST and Mustoe TA: **Continuous versus conventional tissue expansion: experimental verification of a new technique.** *Plast Reconstr Surg* 1991, **87**:10-15.
5. Pasyk KA, Austad ED, McClatchey KD and Cherry GW: **Electron microscopic evaluation of guinea pig skin and soft tissues "expanded" with a self-inflating silicone implant.** *Plast Reconstr Surg* 1982, **70**:37-45.
6. Ryan GB, Cliff WJ and Gabbiani G: **Myofibroblasts in human granulation tissue.** *Hum Pathol* 1974, **5**:55-67.
7. Kawashima T, Yamada A, Ueda K, Asato H and Harii K: **Tissue expansion in facial reconstruction.** *Plast Reconstr Surg* 1994, **94**:944-950.
8. Bauer BS, Few JW, Chavez CD and Galiano RD: **The role of tissue expansion in the management of large congenital pigmented nevi of the forehead in the pediatric patient.** *Plast Reconstr Surg* 2001, **107**:668-675.
9. Castello JR, Garro L, Najera A, Mirelis E, Sanchez-Olaso A and Barros J: **Immediate breast reconstruction in two stages using anastomotic tissue expansion.** *Scand J Plast Reconstr Surg Hand Surg* 2000, **34**:167-671.
10. Argenta LC, Watanabe MJ and Grabb WC: **The use of tissue expansion in head and neck reconstruction.** *Ann Plast Surg* 1983, **11**:31-37.
11. Neumann CG: **The expansion of an area of skin by progressive distention of a subcutaneous balloon.** *Plast Reconstr Surg* 1957, **19**:124-126.
12. Radovan C: **Breast reconstruction after mastectomy using the temporary expander.** *Plast Reconstr Surg* 1982, **69**:195-208.
13. Pusic AL and Cordeiro PG: **An accelerated approach to tissue expansion for breast reconstruction: experience with intraoperative and rapid postoperative expansion in 370 reconstructions.** *Plast Reconstr Surg* 2003, **111**:1871-1875.
14. Hata Y: **Do not forget the fundamental merits of microtia repair using a tissue expander.** *Plast Reconstr Surg* 2002, **109**:819-822.
15. Tran NV, Petty PM, Bite U, Clay RP, Johnson CH and Arnold PG: **Tissue expansion-assisted closure of massive ventral hernias.** *J Am Coll Surg* 2003, **196**:484-488.
16. Alfaro A, Garcia SS and Arenas D: **Intraoperative expansion of skin around large congenital naevi with foley catheter balloons: II new cases.** *Scand J Plast Reconstr Surg Hand Surg* 2002, **36**:273-278.
17. Cunha MS, Nakamoto HA, Herson MR, Faes JC, Gemperli R and Ferreira MC: **Tissue expander complications in plastic surgery: a 10-year experience.** *Rev Hosp Clin Fac Med Sao Paulo* 2002, **57**:93-97.
18. Friedman RM, Ingram AE Jr and Rohrich RJ: **Risk factors for complications in pediatric tissue expansion.** *Plast Reconstr Surg* 1996, **98**:1242-1246.

19. Chun JT and Rohrich RJ: **Versatility of tissue expansion in head and neck burn reconstruction.** *Ann Plast Surg* 1998, **41**:11-16.
20. van Royen BJ, O'Driscoll SW, Dhert WJ and Salter RB: **A comparison of the effects of immobilization and continuous passive motion on surgical wound healing in mature rabbits.** *Plast Reconstr Surg* 1986, **78**:360-368.
21. Timmenga EJ, Andreassen TT, Houthoff HJ and Klopper PJ: **The effect of mechanical stress on healing skin wounds: an experimental study in rabbits using tissue expansion.** *Br J Plast Surg* 1991, **44**:514-519.
22. Raposio E and Santi PL: **Topical application of DMSO as an adjunct to tissue expansion for breast reconstruction.** *Br J Plast Surg* 1999, **52**:194-197.
23. Huang P, Hawthorne WJ, Peng A, Angeli GL, Medbury HJ and Fletcher JP: **Calcium channel antagonist verapamil inhibits neointimal formation and enhances apoptosis in a vascular graft model.** *Am J Surg* 2001, **181**:492-498.
24. Lee RC, Doong H and Jellema AF: **The response of burn scars to intralesional verapamil. Report of five cases.** *Arch Surg* 1994, **129**:107-111.
25. Rehman J, Benet A and Melman A: **Use of intralesional verapamil to dissolve Peyronie's disease plaque: a long-term single-blind study.** *Urology* 1998, **51**:620-626.
26. Cavallini G, Biagiotti G, Koverech A and Vitali G: **Oral propionyl-L-carnitine and intraplaque verapamil in the therapy of advanced and resistant Peyronie's disease.** *BJU Int* 2002, **89**:895-900.
27. Levine LA: **Treatment of Peyronie's disease with intralesional verapamil injection.** *J Urol* 1997, **158**:1395-1399.
28. Pietila JP: **Tissue expansion and skin circulation. Simultaneous monitoring by laser Doppler flowmetry and transcutaneous oximetry.** *Scand J Plast Reconstr Surg Hand Surg* 1990, **24**:135-140.
29. Barone FE, Perry L, Keller T and Maxwell GP: **The biomechanical and histopathologic effects of surface texturing with silicone and polyurethane in tissue implantation and expansion.** *Plast Reconstr Surg* 1992, **90**:77-86.
30. Brobmann GF and Huber J: **Effects of different-shaped tissue expanders on transmural pressure, oxygen tension, histopathologic changes, and skin expansion in pigs.** *Plast Reconstr Surg* 1985, **76**:731-736.
31. Lee P, Squier CA and Bardach J: **Enhancement of tissue expansion by anticontractile agents.** *Plast Reconstr Surg* 1985, **76**:604-610.
32. Matt BH, Squier CA, Kelly KM and Bardach J: **Enhancement of expansion of guinea pig skin by local delivery of an anticontractile agent using a new bilumen expander.** *Ann Plast Surg* 1990, **24**:335-341.
33. Vinnik CA and Jacob SW: **Dimethylsulfoxide (DMSO) for human single-stage intraoperative tissue expansion and circulatory enhancement.** *Aesthetic Plast Surg* 1991, **15**:327-337.
34. Raposio E and Santi PL: **Topical application of DMSO as an adjunct to tissue expansion for breast reconstruction.** *Br J Plast Surg* 1999, **52**:194-197.
35. Canady JW, Squier CA, Kelly KM and Bardach J: **Blood flow in expanded tissue treated with an anticontractile agent.** *Ann Otol Rhinol Laryngol* 1991, **100**:962-965.
36. Squier CA: **The effect of stretching on formation of myofibroblasts in mouse skin.** *Cell Tissue Res* 1981, **220**:325-335.
37. Vander Kolk CA, McCann JJ, Knight KR and O'Brien BM: **Some further characteristics of expanded tissue.** *Clin Plast Surg* 1987, **14**:447-453.

Publish with **BioMed Central** and every scientist can read your work free of charge

"BioMed Central will be the most significant development for disseminating the results of biomedical research in our lifetime."

Sir Paul Nurse, Cancer Research UK

Your research papers will be:

- available free of charge to the entire biomedical community
- peer reviewed and published immediately upon acceptance
- cited in PubMed and archived on PubMed Central
- yours — you keep the copyright

Submit your manuscript here:
http://www.biomedcentral.com/info/publishing_adv.asp

