

REVIEW

Open Access

# The potential of anti-malarial compounds derived from African medicinal plants, part II: a pharmacological evaluation of non-alkaloids and non-terpenoids

Fidele Ntie-Kang<sup>1,2†</sup>, Pascal Amoa Onguéné<sup>3†</sup>, Lydia L Lifongo<sup>1</sup>, Jean Claude Ndom<sup>3</sup>, Wolfgang Sippl<sup>2</sup> and Luc Meva'a Mbaze<sup>3\*</sup>

## Abstract

Malaria is currently a public health concern in many countries in the world due to various factors which are not yet under check. Drug discovery projects targeting malaria often resort to natural sources in the search for lead compounds. A survey of the literature has led to a summary of the major findings regarding plant-derived compounds from African flora, which have shown anti-malarial/antiplasmodial activities, tested by *in vitro* and *in vivo* assays. Considerations have been given to compounds with activities ranging from "very active" to "weakly active", leading to >500 chemical structures, mainly alkaloids, terpenoids, flavonoids, coumarins, phenolics, polyacetyles, xanthenes, quinones, steroids and lignans. However, only the compounds that showed anti-malarial activity, from "very active" to "moderately active", are discussed in this review.

**Keywords:** Africa, Malaria, Medicinal plants, Natural products, Traditional medicine

## Background

Malaria is caused by protozoans of the genus *Plasmodium* (*Plasmodium falciparum*, *Plasmodium malariae*, *Plasmodium ovale*, and *Plasmodium vivax*) [1,2]. According to the World Health Organization (WHO), about half of the world's population is at risk of malaria and one to two million annual deaths (mostly among African children) can be attributed to malaria alone [3,4]. The causative agent is transmitted by the female *Anopheles* mosquito species, which has also developed resistance against insecticides, such as dichlorodiphenyltrichloroethane (DDT), and chemoprophylaxis has not often yielded the expected results [2]. Additionally, the disease-causing protozoans have developed resistance against most of the drugs currently used to treat malaria. There is an urgent need to discover new active compounds.

Nature, and particularly plants are a potential source of new anti-malarial drugs, since they contain a quantity of metabolites with a great variety of structures and pharmacological activities. Traditional preparations (with the use of macerations, extracts, steam baths, concoc-tions, and decoctions from plant materials) have been the main source of treatment of malaria in Africa [5] and other continents where the disease is endemic [6,7]. Thus, with failing treatment regimens, many research groups in Africa (African indigenous research groups and their foreign collaborators) have resorted to plant sources in the quest to expand the anti-malarial chemo-therapeutic arsenal [1,8,9]. This effort has been motivated by the use of these plant materials in the treatment of malaria and fevers in African traditional medicine (ATM). The results from Africa and other continents have been quite promising and hence there has been a general call for the use of natural products as drugs or as sources of inspiration for the development of novel anti-malarials, in order to possibly avoid problems related to drug resistance [10-12].

\* Correspondence: lmbazze@yahoo.fr

†Equal contributors

<sup>3</sup>Department of Chemistry, Faculty of Science, University of Douala, PO Box 24157, Douala, Cameroon

Full list of author information is available at the end of the article

It is believed that the next generation anti-malarials or the scaffolds necessary for their synthesis may be found in the plants currently used in ATM [13,14]. However, the last review on anti-malarial compounds from African flora dates back about ten years [13], with other reviews focusing on plant-screening campaigns in particular regions and/or countries in Africa [15-35] or on active compounds obtained by bioassay-guided fractionation efforts from given countries and/or regions, not covering an entire continent [36-41]. Even though natural products that are active against *P. falciparum* have been discussed in a number of review papers [1,42-48], the goal has been to provide an coverage of the most promising anti-malarials from the entire African continent, by giving an overview of the most pertinent *in vitro* and *in vivo* screening results reported in the literature. The most successful anti-malarials in use to date have been derived from natural product sources (quinolones/artemisinins). It is indeed a glaring omission that the African continent, despite its rich ethno-pharmacological heritage, is yet to yield a significant contribution in this respect. Clearly, as a first step, a systematic review of the many traditional therapeutic options is needed and this review addresses an important issue in this aspect. In part I, the most promising alkaloids and terpenoids were presented [49], while in this part the most interesting findings for flavonoids, coumarins, phenolics, polyacetylates, xanthenes, quinones, steroids, and lignans are shown. The last part of the work is essentially focused on the cheminformatic analysis of >500 natural products (NPs), derived from African medicinal plants, which have demonstrated from weak to very good *in vitro* anti-malarial activities, with a focus on molecular descriptors related to “drug-likeness”, drug metabolism and pharmacokinetics (DMPK). The predicted properties of plant-derived anti-malarials are those related to drug absorption, distribution, metabolism, elimination, and toxicity (ADMET) based on *in silico* computed molecular descriptors.

#### Promising anti-malarial agents derived from African flora

By convention, activities were categorised into “very potent”, “good”, “good to moderate”, “weak”, “very weak” and “inactive”. Following criteria used by Mahmoudi *et al.* [50] and Wilcox *et al.* [51], a pure compound was considered highly active if  $IC_{50} < 0.06 \mu M$ , being active with  $0.06 \mu M \leq IC_{50} \leq 5 \mu M$ , weakly active when  $5 \mu M \leq IC_{50} \leq 10 \mu M$  and compounds with  $IC_{50} > 10 \mu M$  were considered inactive. The following inhibition percentages were proposed for *in vivo* activity of antimalarial extracts at a fixed dose of  $250 \text{ mg kg}^{-1} \text{ day}^{-1}$ : 100-90% (very good activity); 90-50% (good to moderate); 50-10% (moderate to weak); 0% (inactive) [52].

#### Flavonoids

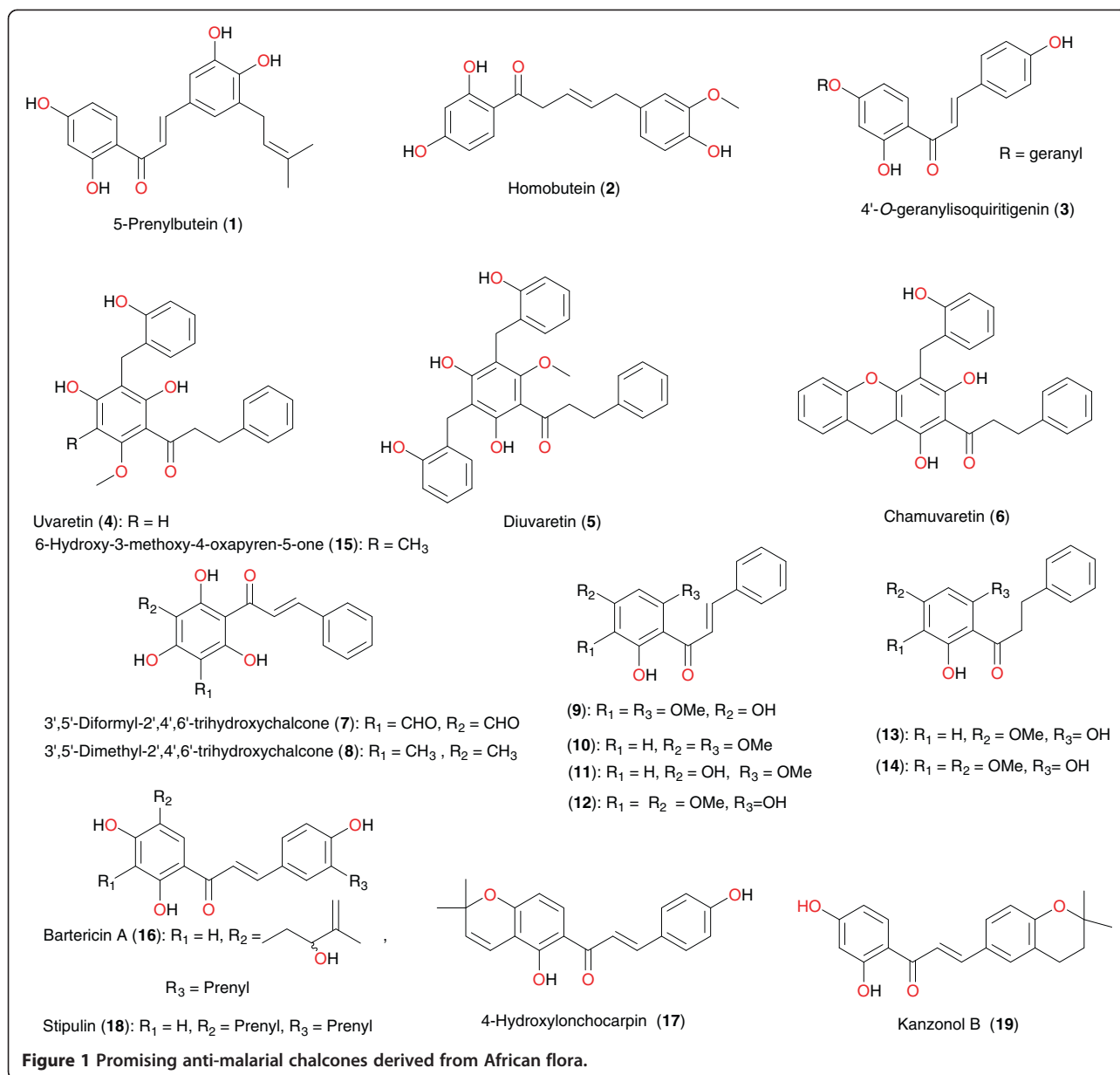
Several bioactive flavonoids have been derived from medicinal plants growing in Africa. Even though the molecular mechanism of action of anti-malarial activities of flavonoids is not fully elucidated, it is believed that flavonoids act by inhibiting the fatty acid biosynthesis (FAS II) of the parasite [53,54]. Some flavonoids have also been shown to inhibit the influx of *L*-glutamine and myoinositol into infected erythrocytes [55]. The active anti-malarial flavonoids are summarized in Table 1, while the chemical structures are shown in Figures 1, 2 and 3.

#### Chalcones

Several anti-malarial flavonoids have been isolated from the stem bark of *Erythrina abyssinica* by Yenesew *et al.* [56,57]. These include chalcones, prenylated and non-prenylated isoflavones and flavones, pterocarpenes, and flavanones. All compounds exhibited moderate anti-malarial activity against the D6 and W2 strains of *P. falciparum*. The ethyl acetate extract of the stem bark of this plant showed anti-plasmodial activity against the chloroquine-sensitive (D6) and chloroquine-resistant (W2) strains of *P. falciparum* with  $IC_{50}$  values of 7.9 and 5.3  $\mu g \text{ mL}^{-1}$ , respectively. From this extract, a new chalcone, 2',3,4,4'-tetrahydroxy-5-prenylchalcone or 5-prenylbutein (1), a new flavanone, 4',7-dihydroxy-3'-methoxy-5'-prenylflavanone (trivial name, 5-deoxyabyssinin II) and homobutein (2), along with known flavonoids have been isolated as the antiplasmodial principles. Yenesew *et al.* also investigated the stem bark of *Milletia usaramensis* ssp. *usaramensis* (Leguminosae) from Kenya [58]. The chalcone 4'-*O*-geranylisoquiritigenin (3) was isolated. This compound exhibited moderate to weak antiplasmodial activity against the D6 and W2 strains of *P. falciparum*. Nkunya *et al.* investigated several Tanzanian species of the genus *Uvaria* [59]. Petroleum ether, dichloromethane and methanol extracts of leaves, stem, and root bark of nine *Uvaria* species: *Uvaria dependens*, *Uvaria faulknerae*, *Uvaria kirkii*, *Uvaria leptocladon*, *Uvaria lucida* ssp. *lucida*, *Uvaria* sp. (Pande), *Uvaria scheffleri*, and *Uvaria tanzaniae* were tested for their *in vitro* activity against the multidrug-resistant K-1 strain of *P. falciparum*. The  $IC_{50}$  values of the extracts varied between 5 and 500  $\mu g \text{ mL}^{-1}$ . The most active extracts were obtained from the stem and root bark of *U. lucida* ssp. *lucida* and *Uvaria* sp. (Pande) and the root bark of *U. scheffleri*, all of which had  $IC_{50}$  values between 5 and 9  $\mu g \text{ mL}^{-1}$ . The investigations of these authors yielded five important chalcones, uvaretin (4), diuvaretin (5), triuvaretin, isotriuvaretin and chamuvaretin (6). These compounds showed moderate to high antiplasmodial activity against the multidrug-resistant K-1 strain of *P. falciparum*, with respective  $IC_{50}$  values of 3.49, 4.20, 46.02, 20.85 and 5.32  $\mu g \text{ mL}^{-1}$ . Joseph *et al.* also isolated two

**Table 1 Summary of promising anti-malarial flavonoids derived from African flora**

Compound subclass	Isolated metabolites	Plant species (Family)	Part of plant studied	Place of harvest (locality, country)	Author, reference	
Chalcones	<b>1</b> and <b>2</b>	<i>Erythrina abyssinica</i> (Leguminosae)	Stem bark	Thika town, Kenya	Yenesew et al. [56,57]	
	<b>3</b>	<i>Milletia usaramensis</i> ssp. <i>usaramensis</i> (Leguminosae)	Stem bark	Jadini Forest, Kenya	Yenesew et al. [58]	
	<b>4, 5</b> and <b>6</b>	<i>Uvaria</i> sp. (Annonaceae)	Leaves, stem and root bark	Tanzania	Nkunya et al. [59]	
	<b>7</b> and <b>8</b>	<i>Friesodielsia obovata</i> (Annonaceae)	Root bark	Tabora district, Tanzania	Joseph et al. [60]	
	<b>9, 10, 11, 12, 13,</b> and <b>14</b>	<i>Polygonum senegalense</i> (Polygonaceae)	Aerial parts	Nairobi, Kenya	Midowo et al. [61]	
	<b>4</b> and <b>15</b>	<i>Uvaria puguensis</i> (Annonaceae)	Stem bark	Pugu Forest Reserve, Tanzania	Makangara et al. [62]	
	<b>16, 17, 18,</b> and <b>19</b>	<i>Dorstenia barteri</i> (Moraceae)	Twigs	Tombel, Cameroon	Ngameni et al. [63]	
	Flavanones	<b>20, 21, 22, 23, 24,</b> and <b>25</b>	<i>Erythrina abyssinica</i> (Leguminosae)	Stem bark	Thika town, Kenya	Yenesew et al. [56,57]
		<b>26</b> and <b>27</b>	<i>Derris trifoliata</i> (Leguminosae)	Seed pods	Coast Province, Kenya	Yenesew et al. [64]
<b>28</b> and <b>29</b>		<i>Polygonum senegalense</i> (Polygonaceae)	Aerial parts	Nairobi, Kenya	Midowo et al. [61]	
<b>30</b>		<i>Erythrina abyssinica</i> (Leguminosae)	Stem bark	Thika town, Kenya	Yenesew et al. [57]	
<b>31</b> and <b>32</b>		<i>Morus mesozygia</i> (Moraceae)	Stem bark	Centre Province, Cameroon	Zeleafack et al. [65]	
Isoflavones	<b>33</b>	<i>Ficus mucuso</i> (Moraceae)	Figs	Tongolo-Yaoundé, Cameroon	Bankeu et al. [66]	
	<b>34</b>	<i>Erythrina saclexii</i> (Leguminosae)	Root bark	Kenya	Andayi et al. [67]	
Retonoids	<b>35, 36, 37</b> and <b>38</b>	<i>Milletia usaramensis</i> ssp. <i>usaramensis</i> (Leguminosae)	Stem bark	Jadini Forest, Kenya	Yenesew et al. [58]	

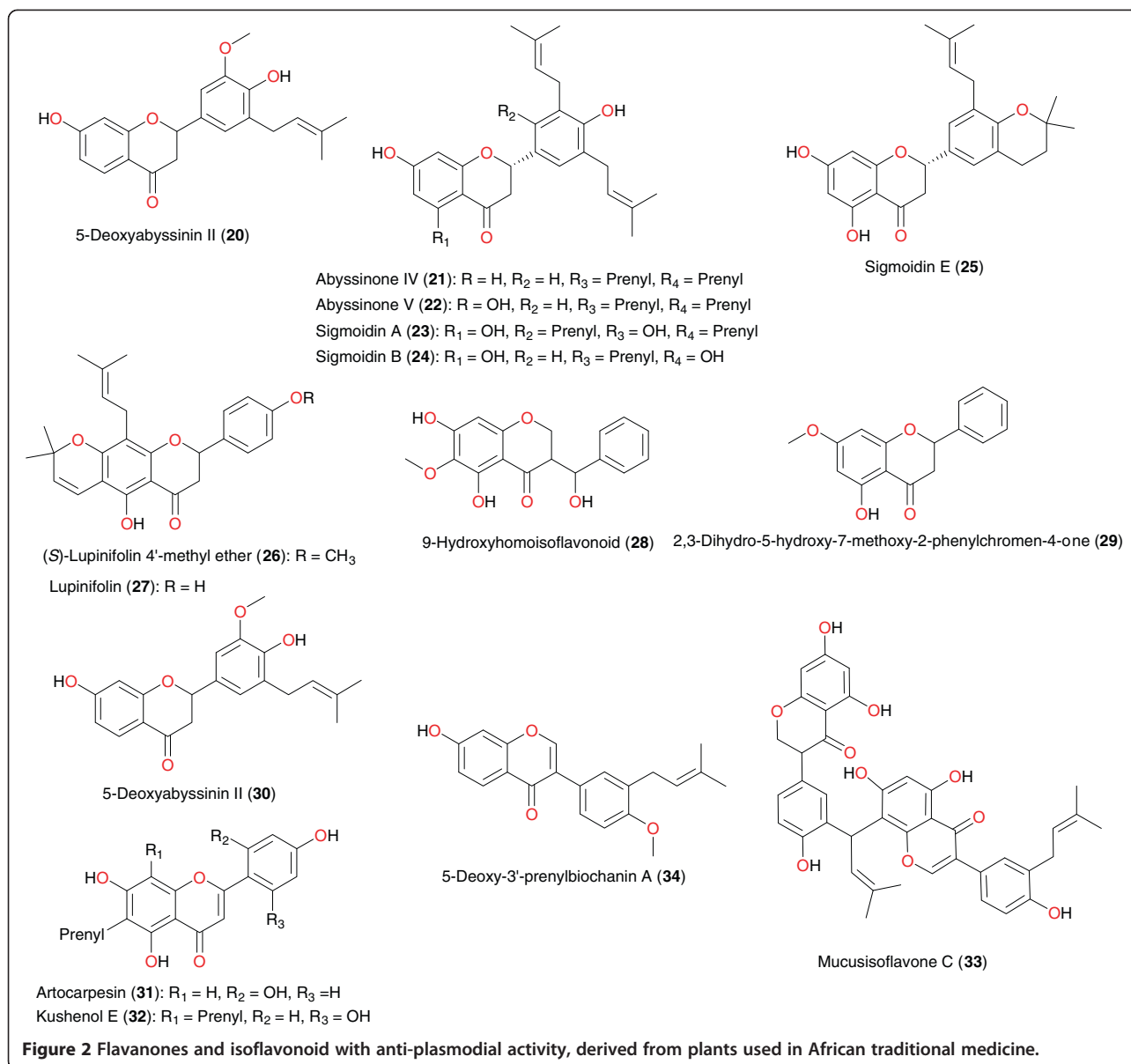


bioactive chalcones; 3',5'-diformyl-2',4',6'-trihydroxychalcone (7) and 3',5'-dimethyl-2',4',6'-trihydroxychalcone (8) from the root bark of *Friesodielsia obovata* [60]. These two compounds exhibited moderate antiplasmodial activity against the K-1 strain of *P. falciparum*, with respective IC<sub>50</sub> values of 23 and 9.7 µg mL<sup>-1</sup>. The same trend of activity was observed against the NF54 strain, against which the compounds had IC<sub>50</sub> values of 29 and 8.5 µg mL<sup>-1</sup> respectively.

The new homoisoflavonoid, 5,7-dihydroxy-3-(hydroxyphenyl-methyl)-6-methoxy-chroman-4-one or polygohomoisoflavanone (9) was isolated from the aerial exudates of *Polygonum senegalense*, along with the known chalcones 10 to 14, by Midowo *et al.* [61]. The new

compound, along with other components of the aerial exudate showed good antiplasmodial activities towards D6 and W2 strains of *P. falciparum*. Mukaranga *et al.* investigated the stem bark of *Uvaria puguensis* (Annonaceae) from Tanzania [62]. Repeated chromatography of the petroleum ether and chloroform extracts yielded uvaretin (4) and the new phenanthrenoid 6-hydroxy-3-methoxy-4-oxapyren-5-one (15), which has been named puguenolide.

The chalcones bartericin A (16), and 4-hydroxylonchocarpin (17), stipulin (18) and kanzonol B (19) were isolated from the twigs of *Dorstenia barteri* (Moraceae) from Cameroon by Ngameni *et al.* [63]. These compounds were evaluated in culture against the W2 strain

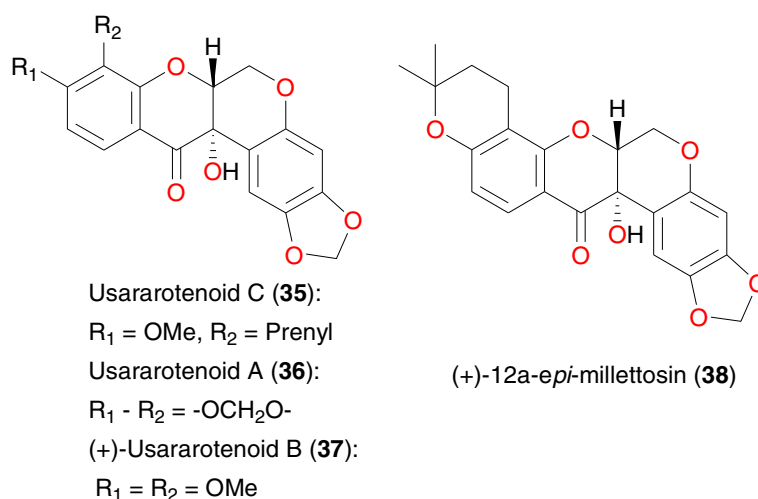


of *P. falciparum*. The evaluated compounds were found to be active *in vitro* against *P. falciparum*, **16**, **17** and **18**, demonstrating particular potencies with relatively low IC<sub>50</sub> values (2.15 μM, 3.36 μM and 5.13 μM respectively). The observed activities confirmed the chalcones as potential leads for the development of anti-malarials.

### Flavanones

The flavanones 5-deoxyabyssinin II (**20**), abyssinone IV (**21**), abyssinone V (**22**), sigmoidins A (**23**), B (**24**) and E (**25**), as well as 5-deoxyabyssinin II (**30**) were isolated from the stem bark of *Erythrina abyssinica* (Leguminosae), harvested in Kenya [57]. The investigations of Yenesew *et al.* demonstrated that these compounds

exhibited anti-malarial properties against the W2 and D6 strains of *P. falciparum* with IC<sub>50</sub> values varying from 4.9 to 13.6 μM against the latter strain and from 5.9 to 13.3 μM against the former strain [56,57]. The same authors investigated the seed pods of *Derris trifoliata* (Leguminosae) [64]. From the dichloromethane-methanol (1:1) extract, a new flavanone derivative (*S*)-lupinifolin 4'-methyl ether (**26**) was isolated, in addition to the known flavonoids lupinifolin (**27**) and rotenone. Lupinifolin only showed moderate *in vitro* antiplasmodial activity against the D6 and W2 strains of *P. falciparum*. The different parts of this plant showed larvicidal activities against *Aedes aegypti* and rotenoids were identified as the active principles [64]. Midowo *et al.* examined the aerial exudates of *Polygonum senegalense*



**Figure 3** Promising anti-malarial rotenoids from African medicinal plants.

and reported the isolation, characterization and antiplasmodial activities of the first 9-hydroxyhomoisoflavonoid (28), 2,3-dihydro-5-hydroxy-7-methoxy-2-phenylchromen-4-one (29), along with the antiplasmodial activities of some of chalconoids and a flavanone isolated along with it from the surface exudate of *Polygonum senegalense* [61].

The antiplasmodial and cytotoxic activities of flavonoids and arylbenzofuran derivatives from *Morus mesozygia* were investigated by Zelefake *et al.* [65]. This plant is used in treating many diseases, including malaria and fever. Fractionation of the methanolic extract of its stem bark led to the isolation and identification of two flavonoids: artocarpesin (31) and kushenol E (32), among other compounds (mulberrofuran F, bartericin A and 4-hydroxyonchocarpin). The methanolic extract and the isolated compounds were tested for antiplasmodial activity against the chloroquine-resistant FcB1 *P. falciparum* strain and cytotoxicity on MCF-7 human breast cancer cells. It was found that all compounds were active against the FcB1 strain of *Plasmodium*, with compounds 31, 32 and mulberrofuran F exhibiting particular potency (with the median inhibitory concentrations IC<sub>50</sub> = 2.5- 2.6 μg mL<sup>-1</sup>).

#### Isoflavones

The isoflavone dimer, mucusisoflavone C (33), derived from the figs of *Ficus mucoso*, harvested near Yaoundé in Cameroon, exhibited a weak inhibitory activity against the validated drug target *P. falciparum* enoyl-ACP reductase (PfENR), with an IC<sub>50</sub> value of 7.69 μM [66].

The acetone extracts of the root bark and stem bark of *Erythrina saculeuxii* showed antiplasmodial activities against the D6 and W2 strains of *P. falciparum*. Further chromatographic separation of the acetone extract of the root bark

by Andayi *et al.* afforded a new isoflavone, 7-hydroxy-4'-methoxy-3'-prenylisoflavone, named 5-deoxy-3'-prenylbiochanin A (34) along with known isoflavonoids as the antiplasmodial principles [67]. Flavonoids and isoflavonoids isolated from the stem bark of *E. saculeuxii* were also tested and showed antiplasmodial activities.

#### Rotenoids

*Milletia usaramensis* ssp. *usaramensis* is a plant growing in East Africa, which is reported to contain anti-malarial flavonoids, particularly rotenoids [58]. Seven rotenoids have been reported from this species, including usararotenoid C (35), usararotenoid A (36), (+)-usararotenoid B (37), and (+)-12a-*epi*-millettosin (38). These compounds exhibited moderate to weak antiplasmodial activity against the D6 and W2 strains of *P. falciparum*. Yenesew *et al.* further established some structure-activity relationships. It was observed that rotenoids containing a prenyl unit or a 2,2-dimethylpyrano substituent were more potent than the non-prenylated rotenoid, e.g., usararotenoid A. It was also reported that there is no significant activity for usararotenoid B, suggesting the importance of the carbonyl function at C-12 in usararotenoid A for the weak antiplasmodial activity observed.

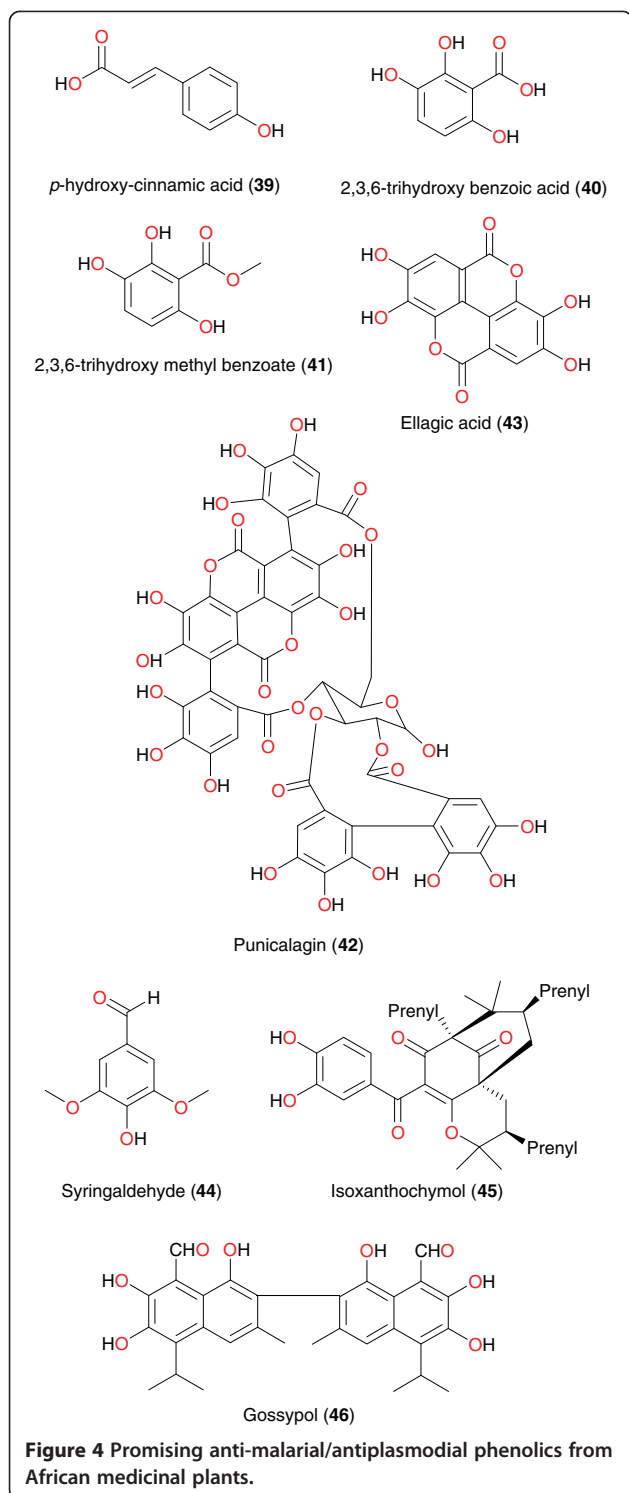
#### Phenolics

Zofou *et al.* have isolated *p*-hydroxy-cinnamic acid (39), along with other compounds, atranorin, specicoside, 2β,3β,19α-trihydroxy-urs-12-20-en-28-oic acid, from the stem bark of *Kigelia africana* (Bignoniaceae), harvested from Cameroon and performed the drug interactions of the isolated compounds among themselves, as well as their combination effects with quinine and artemether [68]. The antiplasmodial activity and drug interactions were evaluated against the multidrug-resistant W2mef



**Table 2 Summary of promising anti-malarial phenolics, polyacetylenes and quinones derived from African flora**

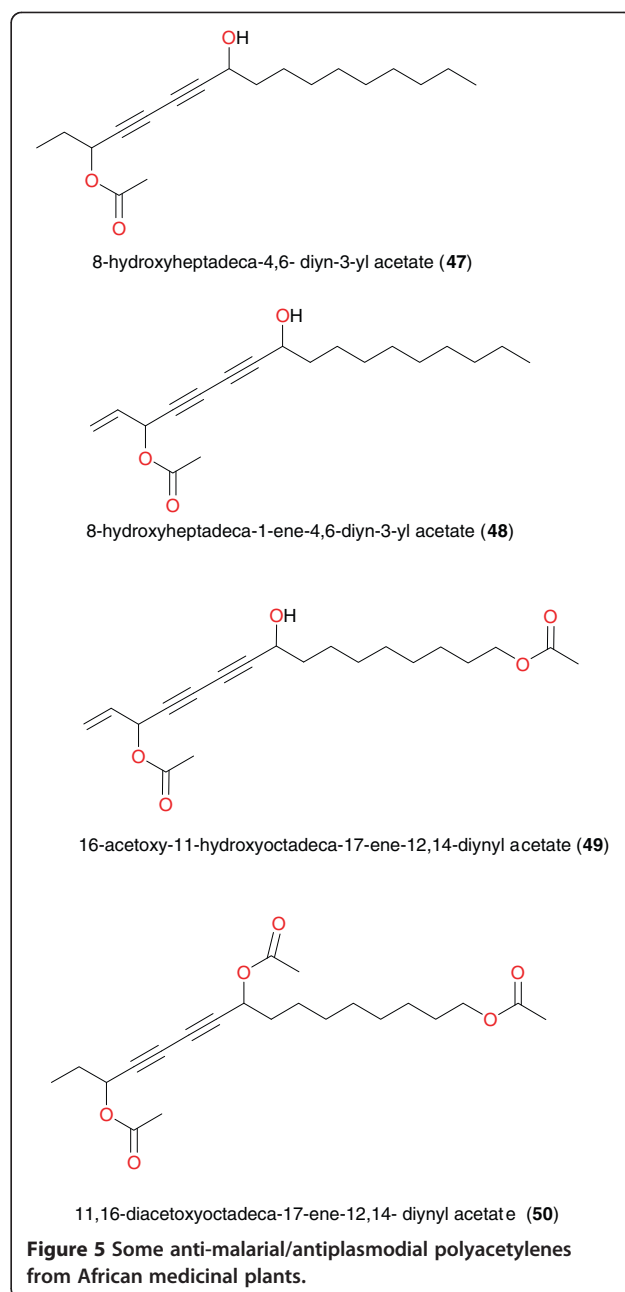
Compound class	Isolated metabolites	Plant species (Family)	Part of plant studied	Place of harvest (locality, country)	Author, reference
Phenolics	<b>39</b>	<i>Kigelia africana</i> (Bignoniaceae)	Stem bark	Bandjoun, Cameroon	Zofou et al. [68,69]
	<b>40 and 41</b>	<i>Sorindeia juglandifolia</i> (Anacardiaceae)	Fruits	Mt. Kalla, Yaoundé, Cameroon	Boyom et al. [70] Kamkumo et al. [71]
	<b>42</b>	<i>Combretum molle</i> (Combretaceae)	Stem bark	Tigray region, Northern Ethiopia	Asres et al. [72]
	<b>43</b>	<i>Alchornea cordifolia</i> (Euphorbiaceae)	Leaves	Ivory Coast	Banzouzi et al. [73]
	<b>44</b>	<i>Vepris uguenensis</i> (Rutaceae)	Roots	Baringo District, Kenya	Cheplogoi et al. [74]
	<b>45</b>	<i>Garcinia polyantha</i> (Guttiferae)	Roots	Mt Kalla, Yaoundé, Cameroon	Lannang et al. [75]
	<b>46</b>	<i>Gossypium</i> sp. (Malvaceae)	Seeds	Diverse regions from the continent	Deck et al. [76]
Polyacetylenes	<b>47, 48, 49 and 50</b>	<i>Cussonia zimmermanii</i> (Araliaceae)	Root bark	Pugu Forest, Tanzania	Senn et al. [77]
Quinones	<b>51, 52, 53 and 54</b>	<i>Hoslundia opposita</i> (Lamiaceae)	Root bark	Tanzania	Achenbach et al. [78]
	<b>55</b>	<i>Cassia siamea</i> (Fabaceae)	Stem bark	Otu (Oyo State), Nigeria	Ajaiyeoba et al. [79]
	<b>56, 57, 58, 59, 60 and 61</b>	<i>Psorospermum glaberrimum</i> (Hypericaceae)	Root bark	Ekombité, Cameroon	Lenta et al. [80]
	<b>62, 63, 64, 65 and 66</b>	<i>Harungana madagascariensis</i> (Hypericaceae)	Root bark	Bazou, Cameroon	Lenta et al. [81]
	<b>67</b>	<i>Spathodea campanulata</i> (Bignoniaceae)	Stem bark	Ibadan, Nigeria	Makinde et al. [82]
	<b>68 and 69</b>	<i>Kniphophia foliosa</i> (Asphodelaceae)	Roots		Dagne et al. [83] Bringmann et al. [84]
	<b>70 and 71</b>	<i>Kigelia pinnata</i> (Bignoniaceae)	Root bark		Weiss et al. [85]



strain of *P. falciparum*. The results equally showed a slight synergistic effect between atranorin and 2 $\beta$ ,3 $\beta$ ,19 $\alpha$ -trihydroxy-urs-12-20-en-28-oic acid (combination index, CI of 0.82) whereas the interaction between specioside and *p*-hydroxycinnamic acid was instead antagonistic (CI of 2.67). All three compounds were shown to

significantly act in synergy with some first line malaria drugs like artemether (CI of 0.42 to 0.71). More excitingly, none of these four compounds showed any significant sign of toxicity against the monkey kidney cell strains LLC-MK2 (selectivity index below 10). Compound **39** exhibited antiplasmodial activity against the W2mef strain with an IC<sub>50</sub> value of 2.11  $\mu\text{g mL}^{-1}$  [69]. The origins of the isolated anti-malarial/antiplasmodial phenolics are shown in Table 2, while the chemical structures are shown in Figure 4.

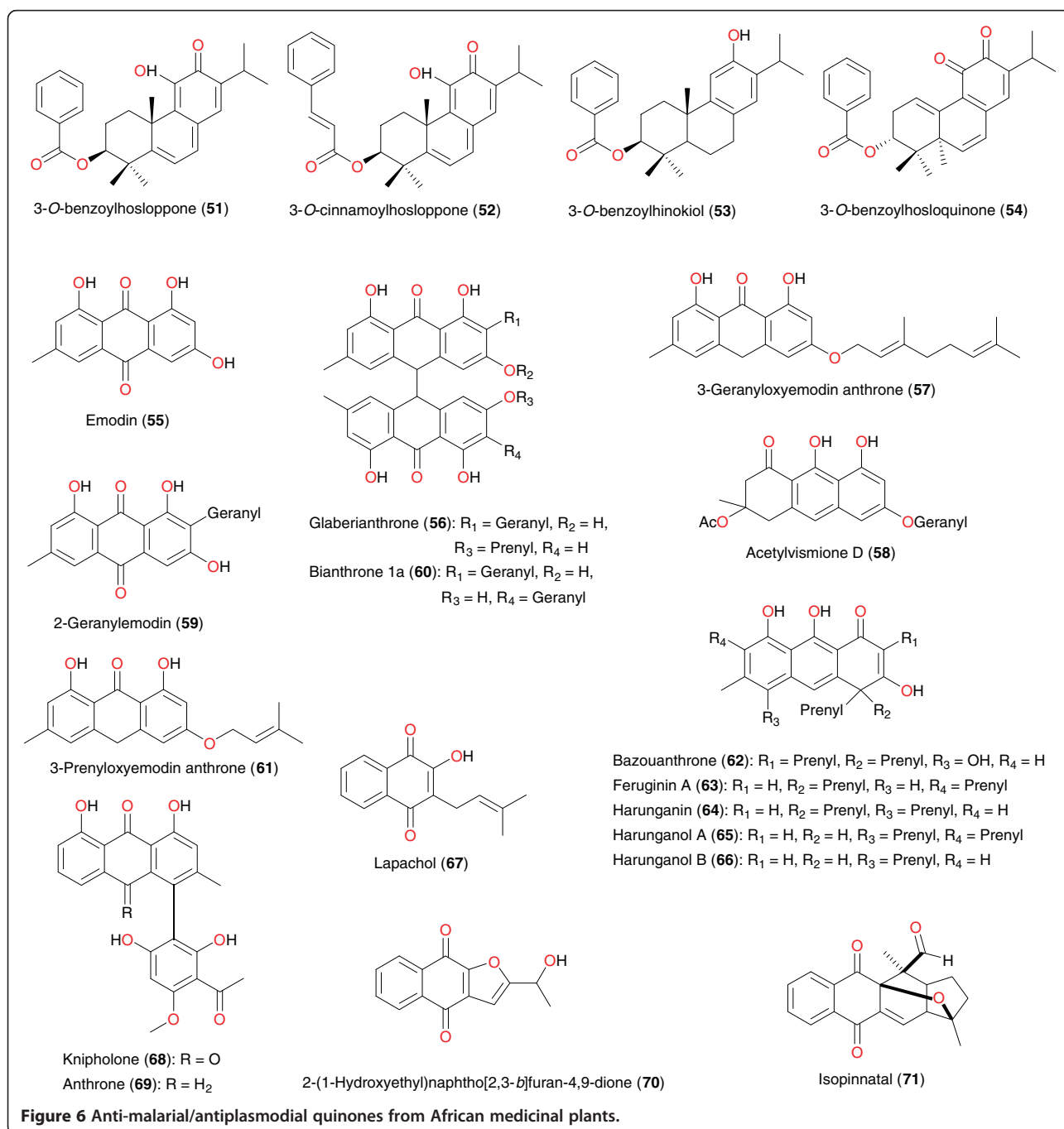
In an effort to identify a lead compound for anti-malarial drug discovery, Kamkumo *et al.* investigated the





fruits of *Sorindeia juglandifolia* (Anacardiaceae) from Mt Kalla in Cameroon and tested the isolated compounds *in vitro* against the *P. falciparum* W2, against field isolates of *P. falciparum*, and against the *P. falciparum* recombinant cysteine protease falcipain-2 [70,71]. The main end products of the activity-guided fractionation were 2,3,6-trihydroxy benzoic acid (**40**) and 2,3,6-trihydroxy methyl benzoate (**41**). Overall, nine fractions tested against *P. falciparum* W2 and falcipain-2 were active, with IC<sub>50</sub> values of varying from 2.3 to 11.6 μg mL<sup>-1</sup>

for W2, and 1.1-21.9 μg mL<sup>-1</sup> for falcipain-2. Purified compounds (**40** and **41**) also showed inhibitory effects against *P. falciparum* W2 (IC<sub>50</sub>s 16.5 μM and 13.0 μM) and falcipain-2 (IC<sub>50</sub>s 35.4 and 6.1 μM). In studies of *P. falciparum* isolates from Cameroon, the plant fractions demonstrated IC<sub>50</sub> values of 0.14-19.4 μg mL<sup>-1</sup> and compounds (**40** and **41**) values of 6.3 and 36.1 μM. *In vivo* assessment of compound **40** showed activity against *Plasmodium berghei* strain B, with mean parasitaemia suppressive dose and curative dose of 44.9 mg kg<sup>-1</sup> and



**Figure 6** Anti-malarial/antiplasmodial quinones from African medicinal plants.

**Table 3 Summary of promising anti-malarial coumarins and xanthone derived from African flora**

Compound class	Isolated metabolites	Plant species (Family)	Part of plant studied	Place of harvest (City, Country)	Author, Reference
Coumarins	<b>72</b> and <b>73</b>	<i>Vernonia brachycalyx</i> (Asteraceae)	Roots		Cubukcu <i>et al.</i> [86]
	<b>74</b>	<i>Toddalia asiatica</i> (Rutaceae)	Roots	Rachuonyo District, Kenya	Oketch-Rabah <i>et al.</i> [88]
	<b>75</b>	<i>Schefflera umbellifera</i> (Araliaceae)	Leaves	Limpopo, South Africa	Mthembu <i>et al.</i> [89]
Xanthones	<b>76</b> and <b>77</b>	<i>Hypericum lanceolatum</i> (Hypericaceae)	Stem bark	Mt. Bamboutos, Cameroon	Zofou <i>et al.</i> [90]
	<b>78, 79, 80, 81, 82</b> and <b>83</b>	<i>Allanblackia monticola</i> (Guttiferae)	Stem bark	Bagangté, Cameroon	Azebaze <i>et al.</i> [91]
	<b>84, 85</b> and <b>86</b>	<i>Symphonia globulifera</i> (Clusiaceae)	Seeds	Fundong, Cameroon	Ngouela <i>et al.</i> [92]
	<b>87, 88, 89</b> and <b>90</b>	<i>Pentadesma butyracea</i> (Guttiferae)	Fruit pericarp	Bazou, Cameroon	Lenta <i>et al.</i> [93]

42.2 mg kg<sup>-1</sup>, respectively. Active fractions were found to be safe in mice after oral administration of 7 g kg<sup>-1</sup> body weight. These results suggest that further investigation of the anti-malarial activities of natural products from *S. juglandifolia* will be appropriate.

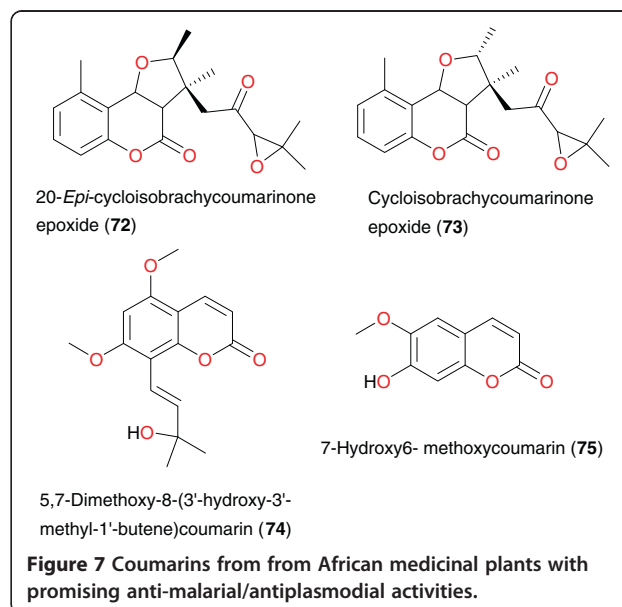
The Ethiopian medicinal plant *Combretum molle* (Combretaceae), reported to possess genuine anti-malarial activity, was investigated by Asres *et al.* [72]. The fractionation of the stem bark extract yielded punicalagin (**42**) as the active compound. This compound exhibited *in vitro* activity against the 3D7 strain of *P. falciparum* with IC<sub>50</sub> values of 2.19 µg mL<sup>-1</sup>. Ellagic acid (**43**), derived from the leaves of *Alchornea cordifolia* (Euphorbiaceae), showed good activity against *P. berghei* in mice with an ED<sub>50</sub> in the range of 0.2-0.151 µg mL<sup>-1</sup> [73]. Cheplogoi *et al.* investigated the roots of *Vepris uguenensis* (Rutaceae), harvested from the Baringo District, Kenya [74]. Syringaldehyde (**44**) was identified as an active compound, exhibiting moderate antiplasmodial activity against two strains of *P. falciparum*, with IC<sub>50</sub> values of 13.0 µg mL<sup>-1</sup> (chloroquine-susceptible 3D7 strain) and 21.4 µg mL<sup>-1</sup> (chloroquine-resistant FCM29 strain), respectively.

From the methanol extract of roots of *Garcinia polyantha*, Lannang *et al.* isolated Isoxanthochymol (**45**), which exhibited *in vitro* anti-malarial activity against *P. falciparum* and showed strong chemosuppression of parasitic growth [75]. The compound exhibited anti-malarial activity with an IC<sub>50</sub> of 2.21 µM. This was lower than the IC<sub>50</sub> of the other five co-occurring compounds (garcinane, smeathxanthones A and B, chefouxanthone, isoxanthochymol, magnificentol, and β-sitosterol and garcinianaxanthone I), which ranged from 2.5 to 4.1 µM. The compounds were administered over a period of four days to the culture and the number of parasites was determined daily. Control experiments were performed either without treatment or with administration of 0.032 µM chloroquine in the same solvent. Gossypol (**46**), derived from the seeds of cotton plant (*Gossypium* sp., Malvaceae), exhibits a variety of biological activities, including antispermatogenic,

anti-cancer, antiparasitic and antiviral activity. Deck *et al.* demonstrated that this compound also showed anti-malarial activity against both chloroquine-sensitive and chloroquine-resistant strains of *P. falciparum*, with IC<sub>50</sub> values in the order of 10 µM [76]. The presence of aldehyde functional groups renders gossypol toxic and in the light of this fact, authors further investigated synthetic analogues of compound **46** for biological activity. It was found that the synthetic analogues lost toxicity while retaining antiplasmodial activity.

#### Polyacetylenes

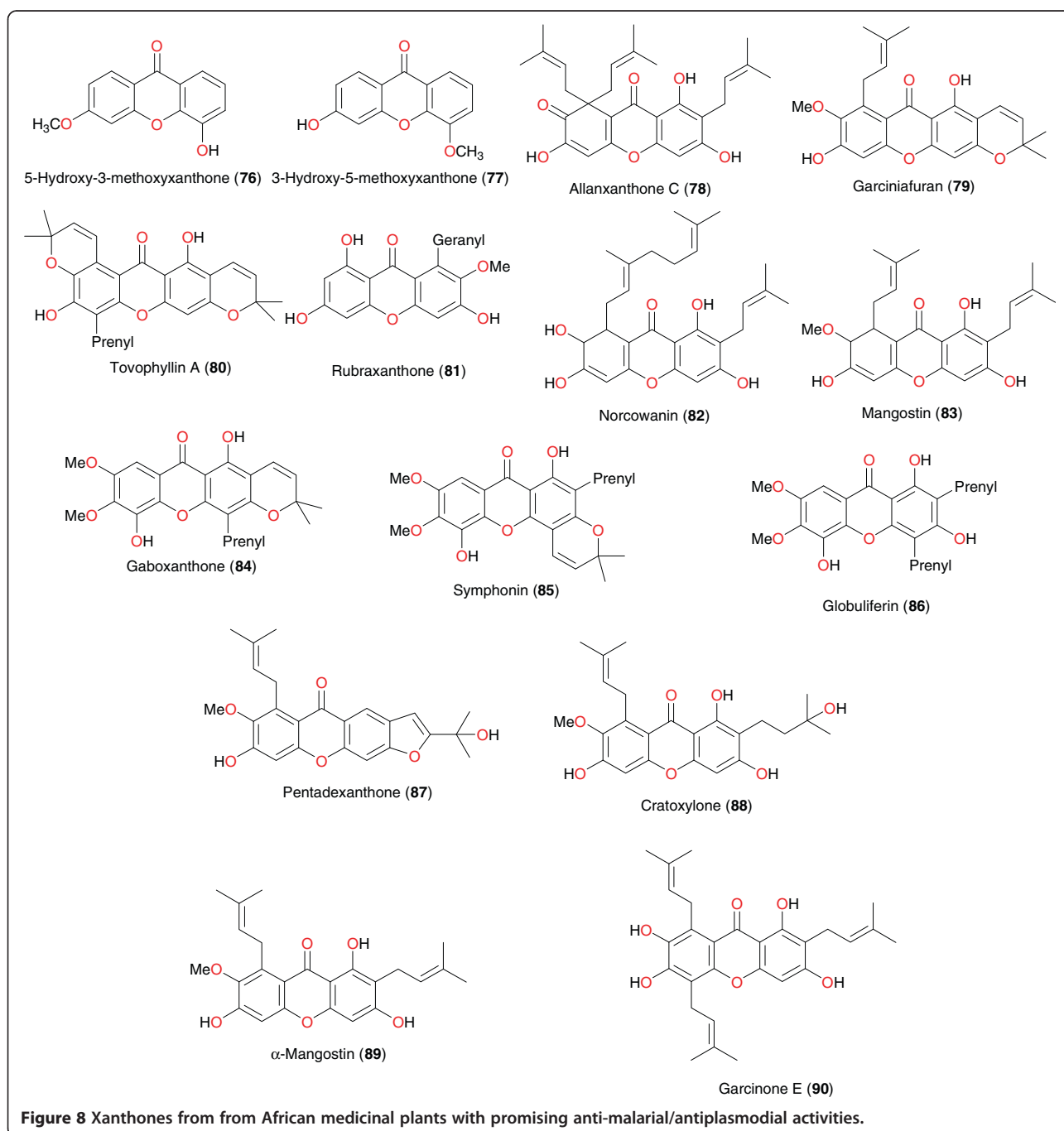
Polyacetylenes have unique chemical structures, which make them rare and often unstable and very reactive. They thus have a wide variety of biochemical and pharmacological uses. Senn *et al.* investigated the root bark extract of *Cussonia zimmermanii* (Araliaceae) from the Pugu Forest in Tanzania, a plant commonly used to



treat malaria, fever and epilepsy [77]. Four polyacetylenes were isolated, namely: 8-hydroxyheptadeca-4,6-dien-3-yl acetate (47), 8-hydroxyheptadeca-1-ene-4,6-dien-3-yl acetate (48), 16-acetoxy-11-hydroxyoctadeca-17-ene-12,14-dienyl acetate (49) and 11,16-diacetoxyoctadeca-17-ene-12,14-dienyl acetate (50), Figure 5. Compounds 47 to 49 showed high anti-malarial activity against *P. falciparum*, with  $IC_{50}$  values of 5.9, 0.44 and 0.84  $\mu$ M respectively.

### Quinones

Quinones also exhibit diverse pharmacological properties, including anti-malarial activity. Four quinones have been isolated from the root bark of *Hoslundia opposita* by Achenbach *et al.* [78], including 3-*O*-benzoylhosloppone (51), 3-*O*-cinnamoylhosloppone (52), 3-*O*-benzoylhinokiol (53), and 3-*O*-benzoylhosloppone (54), Figure 6. The antiplasmodial activities of compound 51 have helped to validate the ethnobotanical use of the plant in the



**Figure 8** Xanthones from from African medicinal plants with promising anti-malarial/antiplasmodial activities.

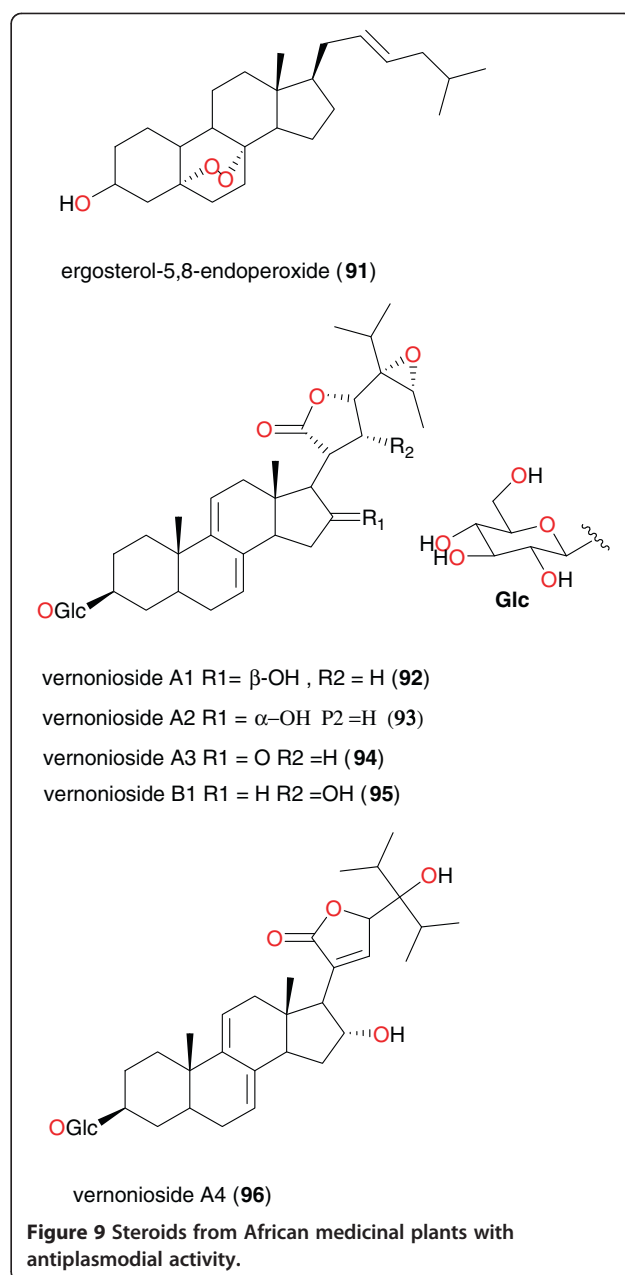
treatment of malaria [78]. The isolation of these compounds was carried out as a result of an ethnomedical use of *H. opposita* in the treatment of malaria. The *n*-hexane extract root bark gave an  $IC_{50}$  of  $5.6 \mu\text{g mL}^{-1}$  and also exhibited a 26% inhibition of growth of *P. berghei* in mice, at a daily dose of  $190 \text{ mg kg}^{-1}$  body weight, for four days [78]. Only compound **51** was tested and showed significant *in vitro* activity against the multidrug-resistant K-1 strain and the chloroquine-sensitive NF54 strain of *P. falciparum*, with  $IC_{50}$  values of 0.4 and  $0.22 \mu\text{g mL}^{-1}$ , respectively. The other metabolites were not screened due to the limited amount available [78].

*Cassia siamea* (Fabaceae) was identified from southwest Nigerian ethnobotany as a remedy for febrile illness. This led to the bioassay-guided fractionation of stem bark of the plant extract, for assessing the *in vitro* anti-malarial activity. Emodin (**55**) and lupeol were isolated from the ethyl acetate fraction. Both compounds were found to be the active principles responsible for the antiplasmodial property with  $IC_{50}$  values of  $5 \mu\text{g mL}^{-1}$  respectively [79].

Six quinones were derived from the root bark extract of *Psorospermum glaberrimum* (Hypericaceae) from Cameroon by Lenta *et al.* [80]. These include glaberianthrone (**56**), 3-geranyloxyemodin anthrone (**57**), acetylvismione D (**58**), 2-geranylemodin (**59**), bianthrone 1a (**60**), and 3-prenyloxyemodin anthrone (**61**). The *n*-hexane extracts and the isolated compounds were tested *in vitro* for their antiplasmodial activity against *P. falciparum* (W2). The *n*-hexane extract showed good antiplasmodial activity, with  $IC_{50}$  of  $0.87 \mu\text{g mL}^{-1}$ , meanwhile 3-geranyloxyemodin anthrone and acetylvismione D showed the best potencies against *P. falciparum* W2 strain with  $IC_{50}$  of  $1.68 \mu\text{M}$  and  $0.12 \mu\text{M}$ , ( $0.66 \mu\text{g mL}^{-1}$  and  $0.054 \mu\text{g mL}^{-1}$ ), respectively. The same authors investigated the root bark of *Harungana madagascariensis* (Hypericaceae), a plant whose roots and bark are used by traditional healers to treat malaria in West Province of Cameroon [81]. These authors isolated bazouanthrone (**62**), a new anthrone derivative, along with the known compounds, feruginin A (**63**), harunganin (**64**), harunganol A (**65**), and harunganol B (**66**). In order to validate its ethnobotanical use, the antiplasmodial activity of the isolated compounds were evaluated in culture against W2 strain of *P. falciparum*. All the compounds were found to be active against the *Plasmodium* parasites with bazouanthrone (**62**) showing particular potency ( $IC_{50} = 1.80 \mu\text{M}$ ).

Makinde *et al.* investigated the action of extracts of the stem bark of *Spathodea campanulata* (Bignoniaceae) from Nigeria on *Plasmodium berghei berghei* in mice [82]. The blood schizontocidal activity of the extracts was studied in early and established infections using chloroquine as the reference drug. The prophylactic

action of the extracts was also investigated with pyrimethamine as the standard drug. The hexane and chloroform extracts of the stem bark showed blood schizontocidal action in both the four-day test and Rane test. The chloroform extract demonstrated some prophylactic properties while the aqueous extract did not show any significant anti-malarial property. In addition, these authors were able to identify the active anti-malarial ingredient to be lapachol (**67**). The other anti-malarial quinones identified were knipholone (**68**) and anthrone (**69**) from *Kniphophia foliosa* (Asphodelaceae) [83,84], as well as 2-(1-hydroxyethyl)naphtho[2,3-*b*]furan-4,9-dione (**70**) and isopinnatal (**71**) from *Kigelia pinnata*



**Table 4 Summary of promising anti-malarial steroids, lignans, other antiplasmodial compounds derived from African flora**

Compound sub class	Isolated metabolites	Plant species (Family)	Part of plant studied	Place of harvest (City, Country)	Author, Reference
Steroids	<b>91</b>	<i>Ajuga remota</i> (Lamiaceae)	Aerial parts	Nairobi, Kenya	Kuria et al. [94]
	<b>92, 93, 94, 95 and 96</b>	<i>Vernonia amygdalina</i> (Asteraceae)	Young pith of trees	Mahale Mt. National Park, Tanzania	Ohigashi et al. [95]
Lignanes	<b>97, 98, 99, 100, 101, 102 and 103</b>	<i>Pycnanthus angolensis</i> (Myristicaceae)	Stem bark	São Tomé and Príncipe islands	Ramalhete et al. [96]
	<b>104</b>	<i>Asparagus africanus</i> (Asparagaceae)	Roots	Kenya	Oketch-Rabah et al. [97]
Others	<b>105</b>	<i>Lippia javanica</i> (Verbenaceae)	Leaves and stalks	Limpopo, South Africa	Ludere et al. [98]
	<b>106</b>	<i>Helichrysum cymosum</i> (Asteraceae)	Whole plant	South Africa	Jakupovic et al. [99]
	<b>107, 108, 109 and 110</b>	<i>Vernonia staehelinoides</i> (Asteraceae)	Leaves	Magaliesburg, South Africa	Vuuren et al. [100]
	<b>111</b>	<i>Hypericum lanceolatum</i> (Hypericaceae)	Stem bark	Mt. Bamboutos, Cameroon	Pillay et al. [101]
	<b>112</b>	<i>Symphonia globulifera</i> (Clusiaceae)	Seeds	Fundong, Cameroon	Zofou et al. [90]
	<b>113</b>	<i>Morus mesozygia</i> (Moraceae)	Stem bark	Centre Province, Cameroon	Ngouela et al. [92]
	<b>114 and 115</b>	<i>Kigelia africana</i> (Bignoniaceae)	Stem bark	Bandjoun, Cameroon	Zeleafack et al. [61]
	<b>116 and 117</b>	<i>Glossocalyx brevipes</i> (Monimiaceae)	Leaves	Kumba, Cameroon	Zofou et al. [64,65]
	<b>118</b>	<i>Asparagus africanus</i> (Asparagaceae)	Roots	Kenya	Mbah et al. [102]
	<b>119</b>	<i>Dracaena mannii and Dracaena arborea</i> (Dracaenaceae)	Seed pulp	Kenya	Oketch-Rabah et al. [97]
				Nigeria	Okunji et al. [103]

(Bignoniaceae) [85]. These compounds were tested against chloroquine-sensitive (T9 – 96) and -resistant (K-1) *P. falciparum* strains and for cytotoxicity using KB cells. Compound **70** possessed good activity against both strains [IC<sub>50</sub> values 627 nM (K1), 718 nM (T9 – 96)]. Isopinnatal (**71**) and the co-occurring kigelinol and isokigelinol exhibited lower activity against both strains. Bringmann *et al.* also reported that knipholone (**68**) and three of its natural derivatives from the same plant, as well as seven structurally related but simplified compounds, have been examined for their antiplasmodial activity against asexual erythrocytic stages of two strains of *P. falciparum* *in vitro* (K1/chloroquine-resistant and NF 54/chloroquine-sensitive) [84]. All the phenylanthraquinones showed considerable activity, with only little cytotoxicity, while their anthraquinone and phenyl moieties were completely inactive.

### Coumarins

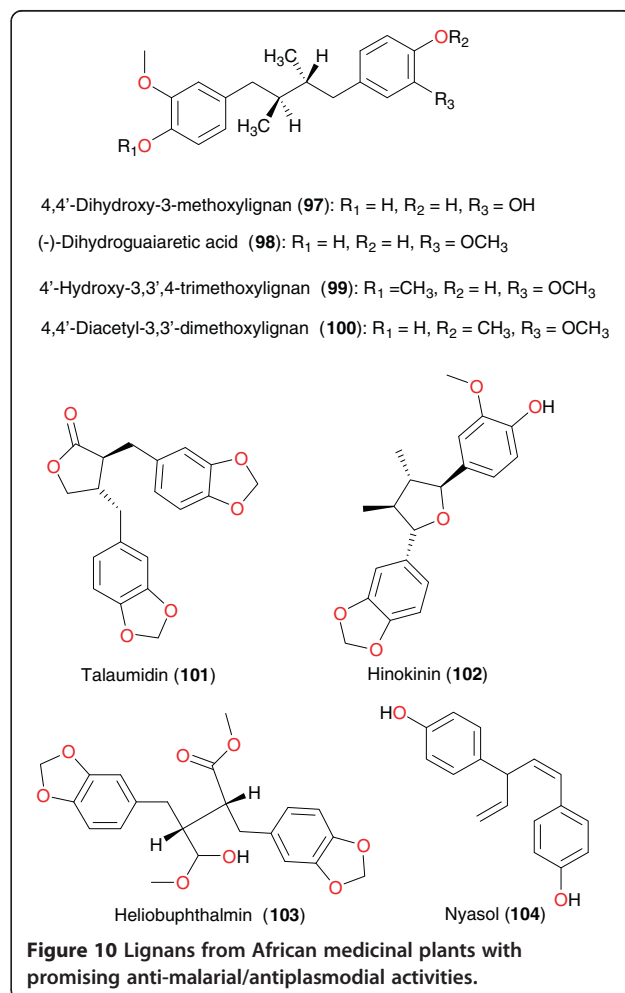
Anti-malarial coumarins have been identified by Cubukcu *et al.* [86] and by Noster *et al.* [87] from *Vernonia brachycalyx* (Asteraceae) and *Toddalia asiatica* (Rutaceae), respectively. Cubukcu *et al.* identified two isomeric 5-methylcoumarins from the roots of *V. brachycalyx*; 20-*epi*-cycloisobrachycoumarinone epoxide (**72**) and cycloisobrachycoumarinone epoxide (**73**), Table 3 and Figure 7. The results of the antiplasmodial assays against the chloroquine-susceptible 3D7 and chloroquine-resistant Dd2 strains of *P. falciparum*, showed that compound **72** was weakly active, with IC<sub>50</sub> values of 160 μM and 54 μM, while for compound **73**, the IC<sub>50</sub> values were 111 μM and 54 μM, respectively. Noster *et al.* also isolated compound **73** from the ether extract of *Exostema caribaeum* but however only moderate activity [87]. In addition, Oketch-Rabah *et al.* isolated a new anti-malarial coumarin, 5,7-dimethoxy-8-(30-hydroxy-30-methyl-10-butene) coumarin (**74**), from the roots of *Toddalia asiatica* [87]. This compound showed moderate activity against the chloroquine-sensitive K39 and chloroquine-resistant V1/S strains of *P. falciparum* strains, with IC<sub>50</sub> values of 16.2 μg mL<sup>-1</sup> and 8.8 μg mL<sup>-1</sup>, respectively.

The anti-malarial coumarin 7-hydroxy-6-methoxycoumarin or scopoletin (**75**) was isolated from the dichloromethane leaf extract of *Schefflera umbellifera* (Araliaceae), harvested from Limpopo, South Africa by Mthembu *et al.* [89]. This compound was evaluated *in vitro* against both the chloroquine-susceptible (D10) and chloroquine-resistant (K-1) strains of *P. falciparum* for anti-malarial activity, with an IC<sub>50</sub> value of 28.2 μg mL<sup>-1</sup>.

### Xanthenes

The anti-malarial xanthenes; 5-hydroxy-3-methoxyxanthone (**76**) and 3-hydroxy-5-methoxyxanthone (**77**) were isolated from stem bark of *Hypericum lanceolatum*

(Hypericaceae) from Cameroon by Zofou *et al.* [90], with IC<sub>50</sub> values of 13.56 μg mL<sup>-1</sup>, and 8.28 μg mL<sup>-1</sup>, respectively, on the multidrug-resistant W2mef strain of *P. falciparum*. Six other anti-malarial xanthenes were isolated from the methanol extract of the stem bark of *Allanblackia monticola* (Guttiferae) from Cameroon, by Azebaze *et al.* [91]. These included allanxanthone C (**78**), garciniafuran (**79**), tophophyllin A (**80**), rubraxanthone (**81**), norcowanin (**82**) and mangostin (**83**), Figure 8. Allanxanthone C exhibited an IC<sub>50</sub> of 1.3 μM on FcM29 and an IC<sub>50</sub> of 6.9 μM on F32. The molecules with interesting activities are known to be norcowanin (IC<sub>50</sub> of 6.3 μM on F32) and mangostin (IC<sub>50</sub> of 4.1 μM on FcM29 and IC<sub>50</sub> of 7.8 μM on F32 [91]. More interestingly, these molecules showed no significant toxicity against the human melanoma cell A375 cell-line. The antiplasmodial activities of xanthenes isolated from the seed shells of *Symphonia globulifera* were reported against the W2 *Plasmodium* sp. with respective IC<sub>50</sub> values of 3.53, 1.29 and 3.17 μM, for gaboxanthone (**84**), symphonin (**85**) and globuliferin (**86**) [92]. Bioassay-guided fractionation of the fruit pericarp of *Pentadesma butyracea*, using the





antiplasmodial test, led to the isolation of a new bioactive xanthone, named pentadexanthone (**87**) ( $IC_{50} = 3 \mu M$  against W2), together with three known compounds: cratoxylone (**88**) ( $IC_{50} = 2.89 \mu M$ ),  $\alpha$ -mangostin (**89**) ( $IC_{50} = 2.77 \mu M$ ), and garcinone E (**90**) ( $IC_{50} = 0.41 \mu M$ ) [93].

### Steroids

The steroid, ergosterol-5,8-endoperoxide (**91**), isolated from the aerial parts of *Ajuga remota*, exhibited high antiplasmodial activity against the chloroquine-sensitive FCA 20/GHA strain of *P. falciparum*, with an  $IC_{50}$  value of  $8.2 \mu M$  [94]. Steroidal saponins with anti-malarial activity have also been isolated from the leaves of *Vernonia amygdalina* [95]. Ohigashi et al. reported the isolation of vernonioside A1 (**92**), A2 (**93**), A3 (**94**), A4 (**95**) and B1 (**96**), Figure 9 and Table 4. These saponins had weak antiplasmodial activity against the multidrug-resistant K-1 strain of *P. falciparum*, with  $IC_{50}$  values of 139.7, 94.1, 245.1, 81.8 and  $46.1 \mu g mL^{-1}$ , respectively [95]. These saponins are also reported to be the bitter compounds in the leaves of *V. amygdalina*.

### Lignans

*Pycnanthus angolensis* (Myristicaceae) is a plant used in traditional medicine against several diseases. Its bark has been used to treat fever and malaria in São Tomé and Príncipe islands. Ramalhete et al. submitted the dichloromethane extract of the bark to anti-malarial screening and observed an activity against 3D7 *P. falciparum* strain ( $IC_{50} = 1.6 \mu g mL^{-1}$ ) [96]. This was further subjected to chromatographic bioguided fractionation, yielding the lignans 4,4'-dihydroxy-3-methoxylignan (**97**), (-)-dihydroguaiaretic acid (**98**), 4'-hydroxy-3,3',4-trimethoxylignan (**99**), 4,4'-diacetyl-3,3'-dimethoxylignan (**100**), talaumidin (**101**), hinokinin (**102**), and heliobuphthalmin (**103**), Figure 10, along with the labdane diterpene ozic acid and the steroids stigmast-4-en-6 $\beta$ -ol-3-one, stigmasterol and  $\beta$ -sitosterol. Furthermore, other compounds were obtained by derivatization. The *in vitro* anti-malarial activity of the compounds was evaluated against 3D7 and Dd2 *P. falciparum* strains. The best *in vitro* activities were exhibited by compound **97** against the 3D7 strain ( $IC_{50} = 31.0 \mu g mL^{-1}$ ) and by compound **101** against the Dd2 strain ( $IC_{50} = 20.7 \mu g mL^{-1}$ ).

*Asparagus africanus* (Asparagaceae) is used by the Akamba tribe in Kenya to treat malaria. A bioassay-guided fractionation of the root extract led to the isolation of the lignan nyasol (**104**), along with the sapogenin muzanzagenin (**119**), Figure 11, as the bioactive compounds responsible for the anti-malarial activity of this plant [97]. Nyasol moderately inhibited *P. falciparum* schizonts with the  $IC_{50}$  of  $49 \mu M$ , while muzanzagenin showed a moderate *in vitro* activity against four different

malaria schizont strains the  $IC_{50}$  values were 16, 163, 23, and  $16 \mu M$ , respectively.

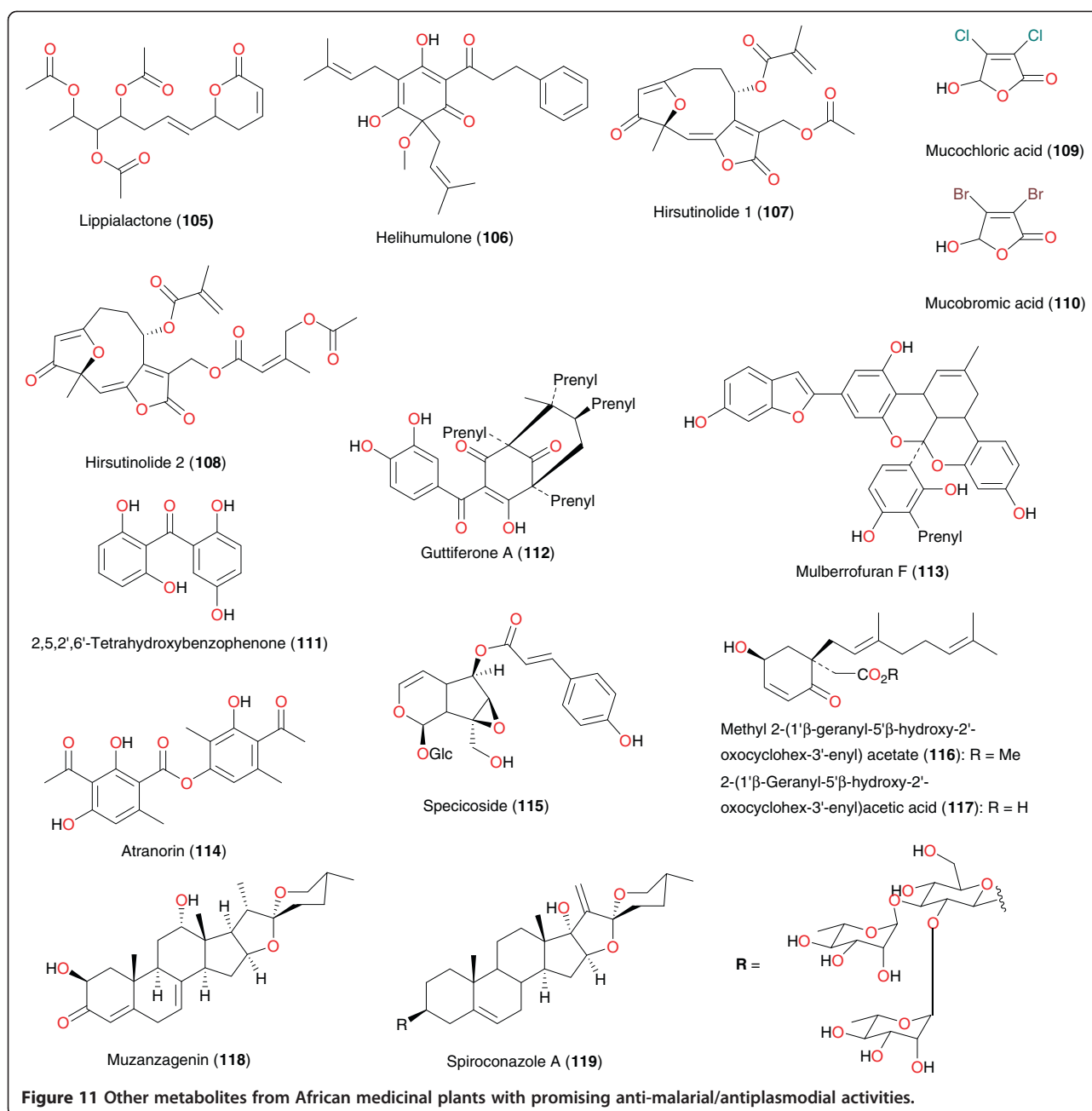
### Others

Lippialactone (**105**), derived from the ethyl acetate extract of aerial parts of *Lippia javanicais*, harvested from South Africa, was shown to be active against the chloroquine-sensitive D10 strain of *P. falciparum* with an  $IC_{50}$  value of  $9.1 \mu g mL^{-1}$ , and is also mildly cytotoxic [98]. Helihumulonone (**106**) was derived from extracts of the whole plant of *Helichrysum cymosum* (Asteraceae) from South Africa by Jakupovic et al. [99] and Vuuren et al. [100].

The dichloromethane extract of the leaves of *Vernonia staehelinoides* (Asteraceae) showed *in vitro* activity ( $IC_{50} \sim 3 \mu g mL^{-1}$ ) against the chloroquine-sensitive D10 and the chloroquine-resistant (K-1) strains of *P. falciparum* [101]. Pillay et al. further investigated the extract by bioassay-guided fractionation and two structurally related hirsutinolides displaying *in vitro* antiplasmodial activity ( $IC_{50} \sim 0.2 \mu g mL^{-1}$  against D10) were isolated. These were 8 $\alpha$ -(2-methylacryloyloxy)-3-oxo-1-desoxy-1,2-dehydrohirsutinolide-13-*O*-acetate (**107**), and 8 $\alpha$ -(5'-acetoxyseneciolyloxy)-3-oxo-1-desoxy-1,2-dehydrohirsutinolide-13-*O*-acetate (**108**). These were found to be cytotoxic to mammalian Chinese hamster ovarian (CHO) cells at similar concentrations, but proved to be attractive scaffolds for structure-activity relationship studies. Two main privileged substructures, a 2(5*H*)-furanone unit and a dihydrofuran-4-one unit, were identified as potential pharmacophores, which may be responsible for the observed biological activity. Mucochloric and mucobromic acids were selected as appropriate 2(5*H*)-furanone substructures and these were shown to have comparable activity against the D10 and superior activity against the K1 strains relative to the hirsutinolide natural product. Mucochloric and mucobromic acids (**109** and **110**) also showed selective cytotoxicity to the malaria parasites compared to mammalian (CHO) cells *in vitro*. The antiplasmodial data obtained with respect to these two acids suggest that the 2(5*H*)-furanone substructure is a key pharmacophore in the observed antiplasmodial activity. The identification of antiplasmodial hirsutinolides from *V. staehelinoides* suggests that they may play a role in the medicinal properties of the plant, but their potential for the development of anti-malarial drugs is limited due to inherent cytotoxicity and lack of selectivity. The results did however lead to the identification of potential pharmacophores, a 2(5*H*)-furanone unit and a dihydrofuran-4-one unit.

The benzophenone 2,5,2',6'-tetrahydroxybenzophenone (**111**), from the stem bark of *Hypericum lanceolatum* (Hypericaceae), exhibited an interesting activity against the multidrug-resistant strain W2mef, with an  $IC_{50}$  of  $13.56 \mu g mL^{-1}$  [90]. Ngouela et al. isolated guttiferone A





(**112**) from the seeds of *Symphonia globulifera* (Clusiaceae) [92]. This compound exhibited activity against the W2 *Plasmodium* sp. with  $IC_{50}$  of 3.17  $\mu$ M. Mulberrofuran F (**113**), from the stem bark of *Morus mesozygia* (Moraceae), was active against FcB1-Columbia strain, considered to be resistant against chloroquine, with  $IC_{50}$  of 2.6  $\mu$ g  $mL^{-1}$  [65]. Atranorin (**114**) and specicoside (**115**), derived from the stem bark extract of *Kigelia africana* (Bignoniaceae) [68,69], were both active against the multidrug-resistant W2mef strain of *P. falciparum* with respective  $IC_{50}$  values of 0.67  $\mu$ g  $mL^{-1}$  and 0.52  $\mu$ g  $mL^{-1}$ .

The homogentisic acid derivatives methyl 2-(1'β-geranyl-5'β-hydroxy-2'-oxocyclohex-3'-enyl) acetate (**116**) and 2-(1'β-geranyl-5'β-hydroxy-2'-oxocyclohex-3'-enyl) acetic acid (**117**) were isolated by Mbah *et al.* from the leaves of *Glossocalyx brevipes* (Monimiaceae) [102]. Compounds **116** and **117** exhibited both anti-malarial [102] and antisalmonellal activities [104]. The sapogenin muzanzagenin (**118**) and the saponin spiroconazole A (**119**), respectively isolated from *Asparagus africanus* (Asparagaceae) [97] and from the West Africa 'soap tree' *Dracaena* sp. (Dracaenaceae) [103], also demonstrated significant anti-malarial activities. Spiroconazole A is

reported to exhibit pronounced antileishmanial, anti-malarial and molluscicidal activities.

## Conclusions

In this review, an attempt has been made to summarise the main finding of several research groups engaged in the search for naturally occurring active principles from African medicinal plants against *P. falciparum*. With multiple resistance developed by the malaria parasite, the cry has been towards obtaining new effective drugs. Attempts to develop 'green pharmacies' for improved phytomedicines against malaria are being encouraged by some NGOs and governments as part of their efforts to control malaria [105]. Additionally, modern hit/lead discovery efforts for specific anti-malarial drug targets are being encouraged. The trend has been towards accelerating this process by employing computer-based methods such as docking, virtual screening, pharmacophore modelling and binding-free energy calculations for hit/lead identification and combinatorial design of novel inhibitors against known anti-malarial drug targets. The practice of virtual screening is beginning to occupy the centre of drug discovery efforts [106] and it has been verified that developing NP libraries containing readily available compounds for screening virtual hits could be highly useful [107]. The authors of this paper have been developing NP databases containing three-dimensional structures of compounds derived from plants used in ATM [108-110] and using computed molecular descriptors to attempt to predict the pharmacokinetic profiles of NPs [110-112]. Since the role of NPs in drug discovery cannot be overemphasized [111-116], efforts are aimed at providing tools for research groups engaged in anti-malarial drug discovery, beginning with NPs derived from African medicinal plants. This is aimed at cutting down the cost of drug discovery when computational and 'wet lab' approaches are combined [117,118]. The intention is to make the current collection of three-dimensional structures of naturally occurring anti-malarials from African medicinal plants available for virtual screening. This shall be the scope of part III of this series.

## Abbreviations

ADMET: Absorption, distribution, metabolism, excretion and toxicity; ATM: African traditional medicine; DMPK: Drug metabolism and pharmacokinetics; FAS II: Fatty acid system II; NP: Natural product; WHO: World Health Organization; WM: Western medicine.

## Competing interests

The authors declare that they have no competing interests.

## Authors' contributions

FNK, LLL, JCN, and LMM conceived the idea. FNK, LLL and PAO participated in data collection. FNK and PAO contributed to data analysis, discussion of results and the conception of the paper under the supervision of LMM, WS, LLL, and JCN. FNK and PAO wrote the first draft of the paper and all authors agreed on the final version before submission.

## Acknowledgements

Financial support is acknowledged from Lhasa Ltd, Leeds, UK through the Chemical and Bioactivity Information Centre (CBIC), University of Buea, Cameroon.

## Author details

<sup>1</sup>Chemical and Bioactivity Information Centre, Department of Chemistry, Faculty of Science, University of Buea, PO Box 63, Buea, Cameroon.

<sup>2</sup>Department of Pharmaceutical Sciences, Martin-Luther University of Halle-Wittenberg, Wolfgang-Langenbeck Str. 4, 06120 Halle, Saale, Germany.

<sup>3</sup>Department of Chemistry, Faculty of Science, University of Douala, PO Box 24157, Douala, Cameroon.

Received: 31 October 2013 Accepted: 25 February 2014

Published: 6 March 2014

## References

1. Nogueira CR, Lopes LMX: **Antiplasmodial natural products.** *Molecules* 2011, **16**:2146-2190.
2. White NJ: **Antimalarial drug resistance.** *J Clin Invest* 2004, **113**:1084-1092.
3. WHO: **World Malaria Report 2012.** Geneva: World Health Organization; 2012. Available from [http://who.int/malaria/publications/world\\_malaria\\_report\\_2012/wmr2012\\_no\\_profiles.pdf](http://who.int/malaria/publications/world_malaria_report_2012/wmr2012_no_profiles.pdf) (accessed on August 02, 2013).
4. Vogel G: **Infectious disease - new map illustrates risk from the 'other' malaria.** *Science* 2010, **329**:618-618.
5. Addae-Mensah I, Fakorede F, Holtel A, Nwaka S: **Traditional medicines as a mechanism for driving research innovation in Africa.** *Malar J* 2011, **10**(Suppl 1):S9.
6. Guantai E, Chibale K: **How can natural products serve as a viable source of lead compounds for the development of new/novel anti-malarials?** *Malar J* 2011, **10**(Suppl 1):S2.
7. Cruz LR, Spangenberg T, Lacerda MVG, Wells TNC: **Malaria in South America: a drug discovery perspective.** *Malar J* 2013, **12**:168.
8. Chin YW, Balunas MJ, Chai HB, Kinghorn AD: **Drug discovery from natural sources.** *AAPS J* 2006, **8**:E239-E253.
9. Fabricant DS, Farnsworth NR: **The value of plants used in traditional medicine for drug discovery.** *Environ Health Perspect* 2001, **109**:69-75.
10. Ginsburg H, Deharo E: **A call for using natural compounds in the development of new antimalarial treatments—an introduction.** *Malar J* 2011, **10**(Suppl 1):S1.
11. Wells TNC: **Natural products as starting points for future anti-malarial therapies: going back to our roots?** *Malar J* 2011, **10**(Suppl 1):S3.
12. Anthony MP, Burrows JN, Duparc S, Moehrle J, Wells TNC: **The global pipeline of new medicines for the control and elimination of malaria.** *Malar J* 2012, **11**:316.
13. Hostettmann K, Marston A, Ndjoko K, Wolfender JL: **The potential of African plants as a source of drugs.** *Curr Org Chem* 2000, **4**:973-1010.
14. Efange SMN: **Natural products: a continuing source of inspiration for the medicinal chemist.** In *Advances in Phytomedicine*. Edited by Iwu MM, Wootton JC. Amsterdam: Elsevier Science; 2002:61-69.
15. Tona L, Ngimbi NP, Tsakala M, Mesia K, Cimanga K, Apers S, De Bruyne T, Pieters L, Totté J, Vlietinck AJ: **Antimalarial activity of 20 crude extracts from nine African medicinal plants used in Kinshasa, Congo.** *J Ethnopharmacol* 1999, **68**:193-203.
16. Tona L, Cimanga RK, Mesia K, Musuamba CT, De Bruyne Apers TS, Hernans N, Van Miert S, Pieters L, Totté J, Vlietinck AJ: **In vitro antiplasmodial activity of extracts and fractions from seven medicinal plants used in the Democratic Republic of Congo.** *J Ethnopharmacol* 2004, **93**:27-32.
17. Dike IP, Obembe OO, Adebisi FE: **Ethnobotanical survey for potential anti-malarial plants in south-western Nigeria.** *J Ethnopharmacol* 2012, **144**:618-626.
18. Gbadamosi IT, Moody JO, Lawal AM: **Phytochemical screening and proximate analysis of eight ethnobotanicals used as antimalaria remedies in Ibadan, Nigeria.** *J Appl Biosci* 2011, **44**:2967-2971.
19. Adebayo JO, Kretli AU: **Potential antimalarials from Nigerian plants: a review.** *J Ethnopharmacol* 2011, **133**:289-302.
20. De Silva A JR, De Ramos SA, Machado M, De Moura DF, Neto Z, Canto-Cavaleiro MM, Figueiredo P, Do Rosário VE, Amaral ACF, Lopes D: **A review of antimalarial plants used in traditional medicine in communities in Portuguese-Speaking**

- countries: Brazil, Mozambique, Cape Verde, Guinea-Bissau, São Tomé and Príncipe and Angola. *Mem Inst Oswaldo Cruz* 2011, **106**(Suppl. 1):142–158.
21. Ancolio C, Azas N, Mahiou V, Ollivier E, Di Giorgio C, Keita A, Timon-David P, Balansard G: **Antimalarial activity of extracts and alkaloids isolated from six plants used in traditional medicine in Mali and Sao Tome.** *Phytother Res* 2002, **16**:646–649.
  22. Puri M, Masum H, Heys J, Singer PA: **Harnessing biodiversity: the Malagasy Institute of Applied Research (IMRA).** *BMC Int Health Hum Rights* 2010, **10**(Suppl 1):S9.
  23. Rasoanaivo P, Ratsimamanga-Urverg S, Ramanitrahasimbola D, Rafatro H, Rakoto-Ratsimamanga A: **Criblage d'extraits de plantes de Madagascar pour recherche d'activité antipaludique et d'effet potentialisateur de la chloroquine.** *J Ethnopharmacol* 1999, **64**:117–126.
  24. Mbatshi SF, Mbatshi B, Banzouzi JT, Bansimba T, Nsonde Ntandou GF, Ouamba JM, Berry A, Benoit-Vical F: **In vitro antiplasmodial activity of 18 plants used in Congo Brazzaville traditional medicine.** *J Ethnopharmacol* 2006, **104**:168–174.
  25. Randrianarivelosia M, Rasidimanana VT, Rabarison H, Cheplogoi PK, Ratsimbason M, Mulholland DA, Maucière P: **Plants traditionally prescribed to treat tazo (malaria) in the eastern region of Madagascar.** *Malar J* 2003, **2**:25.
  26. Kumlungui BS, Ondo-Azi AS, Mintsa NA, Fumoux F, Traore A: **In vitro antiplasmodial activity of seven plants commonly used against malaria in Burkina Faso.** *J Med Plant Res* 2012, **6**:2284–2288.
  27. Bero J, Ganfon H, Jonville MC, Frédéric M, Gbaguidi F, DeMol P, Moudachirou M, Quetin-Leclercq J: **In vitro antiplasmodial activity of plants used in Benin in traditional medicine to treat malaria.** *J Ethnopharmacol* 2009, **122**:439–444.
  28. Pillay P, Maharaj VJ, Smith PJ: **Investigation South African plants as a source of antimalarial drugs.** *J Ethnopharmacol* 2008, **129**:438–454.
  29. Soh PN, Benoit-Vical F: **Are West African plants a source of future antimalarial drugs?** *J Ethnopharmacol* 2007, **114**:130–140.
  30. Muthaura CN, Rukunga GM, Chhabra SC, Mungai GM, Njagi ENM: **Traditional antimalarial phytotherapy remedies used by the Kvale community of the Kenyan Coast.** *J Ethnopharmacol* 2007, **114**:377–386.
  31. Vonthron-Sénécheau C, Weniger B, Ouattara M, Bi FT, Kamenan A, Lobstein A, Brun R, Anton R: **In vitro antiplasmodial activity and cytotoxicity of ethnobotanically selected Ivorian plants.** *J Ethnopharmacol* 2003, **87**:221–225.
  32. Waako PJ, Katuura E, Smith P, Folb P: **East African medicinal plants as a source of lead compounds for the development of new antimalarial drugs.** *Afr J Ecol* 2007, **45**(Suppl. 1):102–106.
  33. Maregesi S, Van Miert S, Pannecouque C, Haddad MHF, Hermans N, Wright CW, Vlietinck AJ, Apers S, Pieters L: **Screening of Tanzanian medicinal plants against *Plasmodium falciparum* and human immunodeficiency virus.** *Planta Med* 2010, **76**:195–201.
  34. Ayuko TA, Njau RN, Cornelius W, Leah N, Ndiege IO: **In vitro antiplasmodial activity and toxicity assessment of plant extracts used in traditional malaria therapy in the Lake Victoria Region.** *Mem Inst Oswaldo Cruz* 2009, **104**:689–694.
  35. Weenen H, Nkunya MHH, Bray DH, Mwasumbi LB, Kinabo LS, Kilimali VAEB: **Antimalarial activity of Tanzanian medicinal plants.** *Planta Med* 1990, **56**:368–370.
  36. Kuete V, Efferth T: **Cameroonian medicinal plants: pharmacology and derived natural products.** *Front Pharmacol* 2010, **1**:123.
  37. Ntie-Kang F, Lifongo LL, Mbaze LM, Ekwele N, Owono Owono LC, Megnassan E, Judson PN, Sippl W, Efange SMN: **Cameroonian medicinal plants: a bioactivity versus ethnobotanical survey and chemotaxonomic classification.** *BMC Complement Altern Med* 2013, **13**:147.
  38. Titanji VPK, Zofou D, Ngemenya MN: **The antimalarial potential of medicinal plants used for the treatment of malaria in Cameroonian folk medicine.** *Afr J Trad CAM* 2008, **5**:302–321.
  39. Magadula JJ, Erasto P: **Bioactive natural products derived from the East African flora.** *Nat Prod Rep* 2009, **26**:1535–1554.
  40. Zofou D, Ntie-Kang F, Sippl W, Efange SMN: **Bioactive natural products derived from the Central African flora against neglected tropical diseases and HIV.** *Nat Prod Rep* 2013, **30**:1098–1120.
  41. Bero J, Quetin-Leclercq J: **Natural products published in 2009 from plants traditionally used to treat malaria.** *Planta Med* 2011, **77**:631–640.
  42. Schwikkard S, van Heerden FR: **Antimalarial activity of plant metabolites.** *Nat Prod Rep* 2002, **19**:675–692.
  43. Bhatnagara S, Dasa P: **Antimalarial activity in tropical plants: a review.** *J Herbs Spices Med Plants* 2007, **13**:103–132. doi:10.1300/J044v13n01\_09.
  44. Oliveira AB, Dolabela MF, Braga FC, Jacome RLRP, Varotti FP, Povoa MM: **Plant-derived antimalarial agents: new leads and efficient phytomedicines: part I: alkaloids.** *Anias da Academia Brasileira de Ciencias* 2009, **81**:715–740.
  45. Batista R, Junior AJS, Oliveira AB: **Plant-derived antimalarial agents: new leads and efficient phytomedicines: part II: non-alkaloidal natural products.** *Molecules* 2009, **14**:3037–3072.
  46. Saxena S, Pant N, Jain DC, Bhakuni RS: **Antimalarial agents from plant sources.** *Curr Sci* 2003, **85**:1314–1329.
  47. Bero J, Frédéric M, Quetin-Leclercq J: **Antimalarial compounds isolated from plants used in traditional medicine.** *J Pharm Pharmacol* 2009, **61**:1401–1433.
  48. Frédéric M, Tits M, Angenot L: **Potential antimalarial activity of indole alkaloids.** *Trans R Soc Trop Med Hyg* 2008, **102**:11–19.
  49. Amoa Onguéné P, Ntie-Kang F, Lifongo LL, Ndom JC, Sippl W, Mbaze LM: **The potential of anti-malarial compounds derived from African medicinal plants, part I: a pharmacological evaluation of alkaloids and terpenoids.** *Malar J* 2013, **12**:449.
  50. Mahmoudi N, de Julian-Ortiz JV, Cicerone L, Galvez J, Mazier D, Danism M, Derouin F, Garcia-Domenech R: **Identification of new antimalarial drugs by linear discriminant analysis and topological virtual screening.** *J Antimicrob Chemother* 2006, **57**:489–497.
  51. Willcox M, Bodeker G, Rasanaivo P: *Traditional Medicinal Plants and Malaria.* Paris: CRC Press; 2004.
  52. Rasoanaivo P, Oketch-Rabah H: *Preclinical considerations on anti-malarial phytomedicines: Part II, Efficacy evaluation.* Antananarivo: Inst., Malgache de Recherches Appliquées; 1998.
  53. Freundlich JS, Anderson JW, Sarantakis D, Shieh HM, Yu M, Valderramos JC, Lucumi E, Kuo M, Jacobs WR Jr, Fidock DA, Schiehsler GA, Jacobus DP, Sacchettiini JC: **Synthesis, biological activity, and X-ray crystal structural analysis of diaryl ether inhibitors of malarial enoyl acyl carrier protein reductase: part 1: 4'-substituted triclosan derivatives.** *Bioorg Med Chem Lett* 2005, **15**:5247–5252.
  54. Perozzo R, Kuo M, Sidhu ABS, Valiyaveetil JT, Bittman R, Jacobs WR Jr, Fidock DA, Sacchettiini JC: **Structural elucidation of the specificity of the antibacterial agent triclosan for malarial enoyl acyl carrier protein reductase.** *Biol Chem* 2002, **277**:13106–13114.
  55. Elford BC: **L-Glutamine influx in malaria-infected erythrocytes: a target for antimalarials?** *Parasitol Today* 1986, **2**:309–312.
  56. Yenesew A, Derese S, Irungu B, Midiwo JO, Waters NC, Liyala P, Akala H, Heydenreich M, Peter MG: **Flavonoids and isoflavonoids with antiplasmodial activities from the root bark of *Erythrina abyssinica*.** *Planta Med* 2003, **69**:658–661.
  57. Yenesew A, Induli M, Derese S, Midiwo JO, Heydenreich M, Peter MG, Akala H, Wangui J, Liyala P, Waters NC: **Anti-plasmodial flavonoids from the stem bark of *Erythrina abyssinica*.** *Phytochemistry* 2004, **65**:3029–3032.
  58. Yenesew A, Derese S, Midiwo JO, Oketch-Rabah HA, Lisgarten J, Palmer R, Heydenreich M, Peter MG, Akala H, Wangui J, Liyala P, Waters NC: **Anti-plasmodial activities and X-ray crystal structures of rotenoids from *Milletia usaramensis* subspecies *usaramensis*.** *Phytochemistry* 2003, **64**:773–779.
  59. Nkunya MH, Weenen H, Bray DH, Mгани QA, Mwasumbi LB: **Antimalarial activity of Tanzanian plants and their active constituents: the genus *Uvaria*1.** *Planta Med* 1991, **57**:341–343.
  60. Joseph CC, Magadula JJ, Nkunya MHH: **A novel antiplasmodial 3',5'-diformylchalcone and other constituents of *Friesodielsia obovata*.** *Nat Prod Res* 2007, **21**:1009–1015.
  61. Midiwo JO, Omoto FM, Yenesew A, Akala HM, Wangui J, Liyala P, Wasunna C, Waters NC: **The first 9-hydroxyhomoisoflavanone, and antiplasmodial chalcones, from the aerial exudates of *Polygonum senegalense*.** *Arkivoc* 2007, **9**:21–27.
  62. Makangara J, Jonker S, Nkunya M: **A novel phenanthrenolide and C-benzyl dihydrochalcones from *Uvaria puguensis*.** *Nat Prod Lett* 2010, **16**:267–272.
  63. Ngameni B, Watchueng J, Boyom FF, Keumedjio F, Ngadjui BT, Gut J, Abegaz BM, Rosenthal PJ: **Antimalarial prenylated chalcones from the twigs of *Dorstenia barteri* var. *subtriangularis*.** *Arkivoc* 2007, **13**:116–123.
  64. Yenesew A, Twinomuhwezi H, Kabaru JM, Akala HM, Kiremire BT, Heydenreich M, Peter MG, Eyase FL, Waters NC, Walsh DS: **Antiplasmodial and larvicidal flavonoids from *Derris trifoliata*.** *Bull Chem Soc Ethiop* 2009, **23**:409–414.

65. Zelefacq F, Guilet D, Valentin A, Fongang RCS, Kom B, Chevalley S, Ngouela SA, Tsamo E, Fabre N, Dijoux-Franca MG: **Antiplasmodial and cytotoxic activities of flavonoids and arylbenzofuran derivatives from *Morus mesozygia***. *Greener J Biol Sci* 2012, **2**:020–024.
66. Bankeu JJK, Khayala R, Lenta BN, Nongoué DT, Ngouela SA, Mustafa SA, Asaad K, Choudhary MI, Prigge ST, Hasanov R, Nkengfack AE, Tsamo E, Ali MS: **Isoflavone dimers and other bioactive constituents from the figs of *Ficus mucoso***. *J Nat Prod* 2011, **74**:1370–1378.
67. Andayi AW, Yenesew A, Derese S, Midiwo JO, Gitu PM, Jondiko OJ, Akala H, Liyala P, Wangui J, Waters NC, Heydenreich M, Peter MG: **Antiplasmodial flavonoids from *Erythrina saculeuxii***. *Planta Med* 2006, **72**:187–189.
68. Zofou D, Kengne AB, Tene M, Ngemenya MN, Tane P, Titanji VPK: **In vitro antiplasmodial activity and cytotoxicity of crude extracts and compounds from the stem bark of *Kigelia africana* (Lam.) Benth (Bignoniaceae)**. *Parasitol Res* 2011, **108**:1383–1390.
69. Zofou D, Tene M, Tane P, Titanji VPK: **Antimalarial drug interactions of compounds isolated from *Kigelia africana* (Bignoniaceae) and their synergism with artemether, against the multidrug-resistant W2mf *Plasmodium falciparum* strain**. *Parasitol Res* 2012, **110**:539–544.
70. Boyom FF, Madiesse EK, Bankeu JJ, Tsouh VP, Lenta BN, Mbacham WF, Tsamo E, Zollo PHA, Gut J, Rosenthal PJ: **Falcipain 2 inhibitors and antiplasmodial compounds from a bio-guided fractionation of the fruits of *Sorindeia juglandifolia* A. Rich. (Anacardiaceae) growing in Cameroon**. *Malar J* 2010, **9**(Suppl 2):P6.
71. Kamkumo RG, Ngoutane AM, Tchokouaha LRY, Fokou PVT, Madiesse EAK, Legac J, Kezetas JB, Lenta BN, Boyom FF, Dimo T, Mbacham WF, Gut J, Rosenthal PJ: **Compounds from *Sorindeia juglandifolia* (Anacardiaceae) exhibit potent antiplasmodial activities in vitro and in vivo**. *Malar J* 2012, **11**:382–389.
72. Asres K, Bucar F, Knauder E, Yardley V, Kendrick H, Croft SL: **In vitro antiprotozoal activity of extract and compounds from the stem bark of *Cobretum molle***. *Phytother Res* 2001, **15**:613–617.
73. Banzouzi JT, Prado R, Menan H, Valentin A, Roumestan C, Mallie M, Pelissier Y, Blache Y: **In vitro antiplasmodial activity of extracts of *Alchornea cordifolia* and identification of an active constituent: ellagic acid**. *J Ethnopharmacol* 2002, **81**:399–401.
74. Cheplogoi PK, Mulholland DA, Coombes PH, Randrianarivojosia M: **An azole, an amide and a limonoid from *Vepris uguenensis* (Rutaceae)**. *Phytochemistry* 2008, **69**:1384–1388.
75. Lannang AM, Louh GN, Lontsi D, Specht S, Sarite SR, Flörke U, Hussain H, Hoerauf A, Krohn K: **Antimalarial compounds from the root bark of *Garcinia polyantha* Oliv.** *J Antibiot* 2008, **61**:518–523.
76. Deck LM, Royer RE, Chamblee BB, Hernandez VM, Malone RR, Torres JE, Hunsaker LA, Piper RC, Makler MT, Vander Jagt DL: **Selective inhibitors of human lactate dehydrogenases and lactate dehydrogenase from the malarial parasite *Plasmodium falciparum***. *J Med Chem* 1998, **41**:3879–3887.
77. Senn M, Gunzenhauser S, Brun R, Sequin U: **Antiprotozoal polyacetylenes from the Tanzanian medicinal plant *Cussonia zimmermannii***. *J Nat Prod* 2007, **70**:1565–1569.
78. Achenbach H, Waibel R, Nkunya MHH, Weenen H: **Antimalarial compounds from *Hoslundia opposita***. *Phytochemistry* 1992, **31**:3781–3784.
79. Ajaiyeoba EO, Ashidi JS, Okpako LC, Houghton PJ, Wright CW: **Antiplasmodial compounds from *Cassia siamea* stem bark extract**. *Phytother Res* 2008, **22**:254–255.
80. Lenta BN, Devkota PK, Ngouela S, Boyom FF, Naz Q, Choudhary MI, Tsamo E, Rosenthal PJ, Sewald N: **Anti-plasmodial and cholinesterase inhibiting activities of some constituents of *Psorospermum glaberrimum***. *Chem Pharm Bull* 2008, **56**:222–226.
81. Lenta BN, Ngouela S, Boyom FF, Tantangmo F, Tchouya GRF, Tsamo E, Gut J, Rosenthal PJ, Connolly JD: **Anti-plasmodial activity of some constituents of the root bark of *Harungana madagascariensis* LAM. (Hypericaceae)**. *Chem Pharm Bull* 2007, **55**:464–467.
82. Makinde JM, Amusan OOG, Adesogan EK: **The antimalarial activity of *Spathodea campanulata* stem bark extract on *Plasmodium berghei* in mice**. *Planta Med* 1988, **54**:122–125.
83. Dagne E, Steglich W: **Knipholone: a unique anthraquinone derivative from *Kniphofia foliosa***. *Phytochemistry* 1984, **23**:1729–1731.
84. Bringmann G, Menche D, Bezabih M, Abegaz BM, Kaminsky R: **Antiplasmodial activity of knipholone and related natural phenylanthraquinones**. *Planta Med* 1999, **65**:757–758.
85. Weiss CR, Moideen SVK, Croft SL, Houghton PJ: **Activity of extracts and isolated naphthoquinones from *Kigelia pinnata* against *Plasmodium falciparum***. *J Nat Prod* 2000, **63**:1306–1309.
86. Cubukcu B, Bray DH, Warhurst DC, Mericli AH, Ozhatay N, Sariyar G: **In vitro antimalarial activity of crude extracts and compounds from *Artemisia abrotanum* L.** *Phytother Res* 1990, **4**:203–204.
87. Noster S, Kraus L: **In vitro antimalarial activity of *Coutarea latiflora* and *Exostema caribaeum* extracts on *Plasmodium falciparum***. *Planta Med* 1990, **56**:63–65.
88. Oketch-Rabah HA, Mwangi JW, Lisgarten J, Mberu EK: **A new antiplasmodial coumarin from *Toddalia asiatica* roots**. *Fitoterapia* 2000, **71**:636–640.
89. Mthembu XS: **A phytochemical study of *Schefflera umbellifera* and *Elephantorrhiza elephantina***. MSc thesis. Pietermaritzburg, South Africa: School of Chemistry, University of KwaZulu-Natal; 2007.
90. Zofou D, Kowa TK, Wabo HK, Ngemenya MN, Tane P, Titanji VPK: ***Hypericum lanceolatum* (Hypericaceae) as a potential source of new anti-malarial agents: a bioassay-guided fractionation of the stem bark**. *Malar J* 2011, **10**:167.
91. Azebaze AGB, Meyer M, Valentin A, Nguemfo EL, Fomum ZT, Nkengfack AE: **Prenylated xanthone derivatives with antiplasmodial activity from *Allanblackia monticola* STANER L.C.** *Chem Pharm Bull* 2006, **54**:111–113.
92. Ngouela S, Lenta BN, Nongoué DT, Ngoupayo J, Boyom FF, Tsamo E, Gut J, Rosenthal PJ, Connolly JD: **Anti-plasmodial and antioxidant activities of constituents of the seed shells of *Symphonia globulifera* Linn f.** *Phytochemistry* 2006, **67**:302–306.
93. Lenta BN, Kamdem LM, Ngouela S, Tantangmo F, Devkota KP, Boyom FF, Rosenthal PJ, Tsamo E: **Antiplasmodial constituents from the fruit pericarp of *Pentadesma butyrea***. *Planta Med* 2011, **77**:377–379.
94. Kuria KAM, Chepkwony H, Govaerts C, Roets E, Busson R, de Witte P, Zupko I, Hoornaert G, Quiryren L, Maes L, Janssens L, Hoogmartens J, Laekeman G: **The antiplasmodial activity of isolates from *Ajuga remota***. *J Nat Prod* 2002, **65**:789–793.
95. Ohigashi H, Hoffman MA, Izutsu D, Koshimizu K, Kawanaka M, Sugiyama H, Kirby GC, Warhurst DC, Allen D, Wright CW, Phillipson JD, Timon-David P, Delmas F, Elias R, Balansard G: **Toward the chemical ecology of medicinal plant use in chimpanzees: the case of *Vernonia amygdalina*, a plant used by wild chimpanzees possibly for parasite-related diseases**. *J Chem Ecol* 1994, **20**:541–553.
96. Ramalhete C, Abrantes M, Mil-Homens T, Duarte N, Lopes D, Cravo P, Madureira MC, Ascenso J, Ferreira MJU: **Search for antimalarial compounds from *Pycnanthus angolensis***. *Planta Med* 2007, **73**:P160.
97. Oketch-Rabah HA, Dossaji SF, Christensen SB, Frydenvang K, Lemmich E: **Antiprotozoal compounds from *Asparagus africanus***. *J Nat Prod* 1997, **60**:1017–1022.
98. Ludere MT, Van Teunis Ree T, Robert Vleggaar R: **Isolation and relative stereochemistry of lippialactone, a new antimalarial compound from *Lippia javanica***. *Fitoterapia* 2013, **86**:188–192.
99. Jakupovic J, Zdero C, Grenz M, Tschirritiz F, Lehmann L, Hashemi-Nejad SM, Bohlmann F: **Twenty-one acylphloroglucinol and further constituents from South African *Helichrysum* species**. *Phytochemistry* 1989, **28**:1119–1131.
100. Van Vuuren SF, Viljoen AM, Van Zyl RL, Van Heerden FR, Baser KHC: **The antimicrobial and toxicity profiles of helihumulone, leaf essential oil and extracts of *Helichrysum cymosum* (L.) D. Don subsp. *cymosum***. *S Afr J Bot* 2006, **72**:287–290.
101. Pillay P, Vleggaar R, Maharaj VJ, Smith PJ, Lategan CA, Chouteau F, Chibale K: **Antiplasmodial hirsutinolides from *Vernonia staehelinoides* and their utilization towards a simplified pharmacophore**. *Phytochemistry* 2007, **68**(8):1200–1205.
102. Mbah JA, Tane P, Ngadjui BT, Connolly JD, Okunji CC, Iwu MM, Schuster BM: **Antiplasmodial agents from the leaves of *Glossocalyx brevipes***. *Planta Med* 2004, **70**:437–440.
103. Okunji CO, Iwu MM, Jackson JE, Tally JD: **Biological activity of saponins from two *Dracaena* species**. *Adv Exp Med Biol* 1996, **404**:415–428.
104. Gatsing D, Mbah JA, Garba IH, Tane P, Djemgou P, Njih-Nkah B: **An antisalmoneal agent from the leaves of *Glossocalyx brevipes* Benth (Monimiaceae)**. *Pak J Biol Sci* 2006, **9**:84–87.
105. Willcox M: **Improved traditional phytomedicines in current use for the clinical treatment of malaria**. *Planta Med* 2011, **77**:662–671.
106. Klebe G: **Virtual ligand screening: strategies, perspectives and limitations**. *Drug Discov Today* 2006, **11**:580–594.
107. Quinn RJ, Carroll AR, Pham MB, Baron P, Palframan ME, Suraweera L, Pierens GK, Muresan S: **Developing a drug-like natural product library**. *J Nat Prod* 2008, **71**:464–468.



108. Ntie-Kang F, Mbah JA, Mbaze LM, Lifongo LL, Scharfe M, Ngo Hanna J, Cho-Ngwya F, Onguéné PA, Owono LCO, Megnassan E, Sippl W, Efange SMN: **CamMedNP: building the Cameroonian 3D structural natural products database for virtual screening.** *BMC Complement Altern Med* 2013, **13**:88.
109. Ntie-Kang F, Onguéné PA, Scharfe M, Owono LCO, Megnassan E, Mbaze LM, Sippl W, Efange SMN: **ConMedNP: a natural product library from Central African medicinal plants for drug discovery.** *RSC Adv* 2014, **4**:409–419.
110. Ntie-Kang F, Zofou D, Babiaka SB, Meudom R, Scharfe M, Lifongo LL, Mbah JA, Mbaze LM, Sippl W, Efange SMN: **AfroDb: a select highly potent and diverse natural product library from African medicinal plants.** *PLoS ONE* 2013, **8**:e78085.
111. Ntie-Kang F, Mbah JA, Lifongo LL, Owono LCO, Megnassan E, Mbaze LM, Judson PN, Sippl W, Efange SMN: **Assessing the pharmacokinetic profile of the CamMedNP natural products database: an *in silico* approach.** *Org Med Chem Lett* 2013, **3**:10.
112. Ntie-Kang F, Lifongo LL, Mbah JA, Owono LCO, Megnassan E, Mbaze LM, Judson PN, Sippl W, Efange SMN: ***In silico* drug metabolism and pharmacokinetic profiles of natural products from medicinal plants in the Congo basin.** *In Silico Pharmacol* 2013, **1**:12.
113. Koehn FE, Carter GT: **The evolving role of natural products in drug discovery.** *Nat Rev Drug Discov* 2005, **4**:206–220.
114. Harvey AL: **Natural products in drug discovery.** *Drug Discov Today* 2008, **13**:894–901.
115. Newman DJ: **Natural products as leads to potential drugs: an old process or the new hope for drug discovery?** *J Med Chem* 2008, **51**:2589–2599.
116. Li JWH, Vederas JC: **Drug discovery and natural products: end of an era or an endless frontier?** *Science* 2009, **325**:161–165.
117. Lipinski CA, Lombardo F, Dominy BW, Feeney PJ: **Experimental and computational approaches to estimate solubility and permeability in drug discovery and development settings.** *Adv Drug Delivery Rev* 1997, **23**:3–25.
118. DiMasi JA, Hansen RW, Grabowski HG: **The price of innovation: new estimates of drug development costs.** *J Health Econ* 2003, **22**:151–185.

doi:10.1186/1475-2875-13-81

**Cite this article as:** Ntie-Kang et al.: The potential of anti-malarial compounds derived from African medicinal plants, part II: a pharmacological evaluation of non-alkaloids and non-terpenoids. *Malaria Journal* 2014 **13**:81.

**Submit your next manuscript to BioMed Central and take full advantage of:**

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at  
www.biomedcentral.com/submit

