REVIEW



The Status of Carbapenem Resistance in Cystic Fibrosis: A Systematic Review and Meta-Analysis

Mohamed J. Saadh^{*a,b*}, Armaghan Lohrasbi^{*c*}, Elaheh Ghasemian^{*d*}, Marzieh Hashemian^{*e*}, Anahita Etemad^{*e*}, Zahra Dargahi^{*f*}, and Vahab Hassan Kaviar^{*e*,*}

^aFaculty of Pharmacy, Middle East University, Amman, Jordan; ^bApplied Science Research Center, Applied Science Private University, Amman, Jordan; ^cDepartment of Biological and Biomedical Sciences, Glasgow Caledonian University, Glasgow, Scotland; ^dDepartment of Microbiology, School of Medicine, Kermanshah University of Medical Sciences, Tehran, Iran; ^cClinical Microbiology Research Center, Ilam University of Medical Sciences, Ilam, Iran; ^fDepartment of Microbiology, School of Medicine, Ahvaz Jundishapur University of Medical Sciences, Ahvaz, Iran

Background: Antibiotic resistance in cystic fibrosis (CF) is a well-known phenomenon. However, the comprehensive epidemiological impact of antibiotic resistance in CF is not clearly documented. So, this meta-analysis evaluated the proportion rates of carbapenem resistance (imipenem, meropenem, and doripenem) in CF based on publication date (1979-2000, 2001-2010, and 2011-2021), continents, pathogens, and antimicrobial susceptibility testing (AST). Methods: We searched studies in PubMed, Scopus, and Web of Science (until April 2021). Statistical analyses were conducted using STATA software (version 14.0). Results: The 110 studies included in the analysis were performed in 25 countries and investigated 13,324 pathogens associated with CF. The overall proportion of imipenem, meropenem, and doripenem resistance in CF were 43% (95% CI 36-49), 48% (95% CI 40-57), 28% (95% CI 23-33), and 45% (95% CI 32-59), respectively. Our meta-analysis showed that trends of imipenem, meropenem, and doripenem-resistance had gradual decreases over time (1979-2021). This could be due to the limited clinical effectiveness of these antibiotics to treat CF cases over time. Among the opportunistic pathogens associated with CF, the highest carbapenem resistance rates were shown in Stenotrophomonas maltophilia, Burkholderia spp., Pseudomonas aeruginosa, and Staphylococcus aureus. The highest and lowest carbapenem resistance rates among P. aeruginosa in CF patients were shown against meropenem (23%) and doripenem (39%). Conclusions: We showed that trends of carbapenem resistance had decreased over time (1979-2021). This could be due to the limited clinical effectiveness of these antibiotics to treat CF cases over time. Plans should be directed to fight biofilm-associated infections and prevent the emergence of mutational resistance. Systematic surveillance for carbapenemase-producing pathogens in CF by molecular surveillance is necessitated.

*To whom all correspondence should be addressed: Vahab Hassan Kaviar, PhD, Clinical Microbiology Research Center, Ilam University of Medical Sciences, Ilam, Iran; Email: vahab.kaviar@gmail.com.

Abbreviations: CF, cystic fibrosis; WPR, Weighted pooled resistance; AST, Antimicrobial susceptibility testing; AMR, Antimicrobial resistance; CI, Confidence interval.

Keywords: cystic fibrosis, antimicrobial resistance, carbapenem resistance, bacterial pathogens, systematic review and metaanalysis

INTRODUCTION

The autosomal recessive monogenetic disorder cystic fibrosis (CF) was once untreatable and deadly in childhood among those ethnically White/of European ancestry, occurring in ~1 in 3,400 live births; while in other ethnic groups, the rates are much lower [1-3]. Chronic suppurative airway infection is a hallmark feature of CF led by the opportunistic pathogens and their longterm persistence [4]. Human bacterial pathogens such as Pseudomonas aeruginosa, Staphylococcus aureus, Burkholderia, and Stenotrophomonas are the key contributors to morbidity and mortality in CF patients [4,5]. Carbapenems are a potent, broad-spectrum class of antibiotics that act as bactericidal inhibitors of the bacterial cell wall [6]. Carbapenems include imipenem and meropenem for the treatment of P. aeruginosa or S. aureus infections in people suffering CF [7]. However, antimicrobial resistance in CF is well-recognized phenomenon [8]. But, the global epidemiological impact of antimicrobial resistance in CF is not clearly understood. To answer this vexing question, the main objective of this review was to provide extensive data on the carbapenem resistance in CF.

METHODS

This review is reported in accordant with the Preferred Reporting Items for Systematic Reviews and Meta-Analyses guidelines (PRISMA) [9].

Search Strategy and Study Selection

We systematically searched for relevant articles in PubMed, Scopus, and Web of Science (until April 22, 2021) by using the related keywords: ("cystic fibrosis") AND ("antibiotic resistance" OR "drug resistance" OR "antimicrobial resistance" OR "carbapenems resistance") in the Title/Abstract/Keywords fields. No limitation was conducted while searching databases. The search strategy was designed and conducted by study investigators. Authors cross-checked included articles reference lists for any additional studies that may have been missed in the search. The records found through database searching were merged and the duplicates were removed using EndNote X8 (Thomson Reuters, New York, NY, USA). One of the team researchers randomly evaluated the search results and confirmed that no relevant study had been ignored. All these steps were done by three authors and any disagreements about article selection were resolved through discussion, and a fourth author acted as arbiter. Three reviewers screened all titles and abstracts separately and excluded irrelevant or duplicate articles first. Afterward, they evaluated the remaining articles for inclusion separately. Discrepancies were resolved by discussion.

Data Extraction

The following items were extracted from included studies: first author, year of study, year published, continent, country, number of pathogens (*P. aeruginosa, S. aureus, S. maltophilia, Burkholderia* species, nontuberculous *Mycobacteria* (NTM), and other bacterial species), number of carbapenem-resistant pathogens, and antimicrobial susceptibility testing (AST; automated system, disk diffusion, dilution methods, and MIX). The exclusion criteria were as follows: (1) studies that contained duplicate data or were overlapping articles; (2) animal research, reviews, meta-analysis and/or systematic review, and conference abstracts; and (3) carbapenem-resistance rates were not presented or reported.

Quality Assessment

The quality of the included studies was assessed using an adapted version of the tool proposed by the Newcastle-Ottawa assessment scale adapted for cross-sectional studies [10]. A score ranging from 0 to 7 points was attributed to each study (≥ 6 points: high quality, ≤ 5 points: low quality).

Statistical Analysis

The included studies presenting raw data on carbapenem resistance in CF was performed by computing the pooled using a random-effects model with Stata/SE software, v.14.1 (StataCorp. 2015. Stata Statistical Software: Release 14. College Station, TX: StataCorp LP.). Weighted pooled resistance rate (WPR: proportion of strains resistant to specific antimicrobial agents) was calculated based on the Freeman-Tukey double-arcsine transformation. The inconsistency across studies was examined by the forest plot as well as the I² statistic. Values of I² (25%, 50%, and 75%) were interpreted as the presence of low, medium, or high heterogeneity, respectively. So, the DerSimonian and Laird random effects models were used [11]. Publication bias was analyzed using Egger's linear regression test. All statistical interpretations were reported on a 95% confidence interval (CI) basis.

Study Outcomes

The main outcome of interest was the WPR rates of carbapenem resistance (imipenem, meropenem, and doripenem) in CF. A subgroup analysis was performed by; (1) publication date (1979-2000, 2001-2010, and 2011-2021), (2) geographic areas (continents), (3) pathogens, (4) AST.

RESULTS

Systematic Literature Search



Figure 1. Flow chart of study selection.

A total of 2,945 records were identified in the initial search. Among these, 2,724 articles were excluded after an initial screening of the title and abstract due to their irrelevance and duplication. The full texts of the remaining 200 articles were reviewed (Figure 1). Of 221 articles, 111 were excluded for the reasons as above mentioned. Finally, 110 studies [12-121] were included in this meta-analysis. The studies included in this meta-analysis evaluated antibiotic resistance to imipenem, meropenem, and doripenem. The final studies included in the analysis were performed in 25 countries and investigated 13,324 pathogens associated with CF. The WPR rates for each antimicrobial are shown in Table 1 and Figure 2.

META-ANALYSIS

Carbapenem Resistance

Imipenem: Susceptibility to imipenem was determined in 14,459 bacterial pathogens associated with CF; the WPR was 43% (95% CI 36-49) with substantial heterogeneity ($I^2=98.27\%$) (Table 1 and Figure 2). To analyze the trends for changes in the prevalence of imipenem-resistance in CF in more recent years, we performed

a subgroup analysis for three periods 1979-2000, 2001-2010, and 2011-2021) (Table 1, Figure 3). The subgroup analysis that compared the data from 1979-2000 (WPR 60%; 95% CI 38-59), 2001-2010 (WPR 46%; 95% CI 33-59), and 2011-2021 (WPR 38%; 95% CI 31-46) indicated a significant decrease in the resistance rate (P < 0.001) (Table 1, Figure 3). The highest imipenem resistance rate was shown in Europe, followed by North America (50%, 95% CI 42-57; 48%, 95% CI 25-72). Among 23 countries reporting resistance data for imipenem, 12 (52.1%) countries (China, Iraq, Scotland, Denmark, London, Turkey, Ireland, Italy, Germany, Argentina, US, and UK) reported that > 43% of isolates had imipenem resistance. The highest imipenem resistance rate was shown in S. maltophilia, non-tuberculous mycobacteria, and Burkholderia spp., followed by (99%, 95% CI 96-100; 75%, 95% CI 46-96; and 55%, 95% CI 42-68) (Table 1). Statistically, a significant difference was found in the subgroups AST, continents, and pathogens (P < 0.001).

Meropenem: Susceptibility to meropenem was determined in 17,032 bacterial pathogens associated with CF; the WPR was 28% (95% CI 23-33) with substantial heterogeneity (I²=97.55%) (Table 1). As shown in the Table 1 and Figure 3, the prevalence of meropenem resis-

| Variables | Proportion (%) (95% Cl) | n, N | Heterogeneity (I²) (%) | Egger test |
|-----------------------------|----------------------------|-------------|---------------------------|---------------|
| Imipenem | | | | |
| Overall | 43 (36, 49) | 6389, 14459 | 98.27 | 0.86 |
| 1979-2000 | 60 (38, 59) | 424, 1707 | 97.92 | |
| 2001-2010 | 46 (33, 59) | 3203, 6215 | 98.90 | |
| 2011-2021 | 38 (31, 46) | 2762, 6537 | 96.89 | |
| Nontuberculous mycobacteria | 75 (46, 96) | 177, 385 | 95.95 | |
| <i>Burkholderia</i> spp. | 55 (42, 68) | 898, 1219 | 94.52 | |
| P. aeruginosa | 34 (28, 40) | 3817, 9848 | 97.51 | |
| S. maltophilia | 99 (96, 100) | 1161, 1189 | 78.14 | |
| Others | 20 (10, 32) | 336, 1818 | 96.03 | |
| Asia | 22 (2, 49) | 217, 531 | 97.92 | |
| Europe | 50 (42, 57) | 4456, 10887 | 98.00 | |
| North America | 48 (25, 72) | 1372, 2165 | 99.15 | |
| South America | 10 (3, 21) | 103, 432 | 91.35 | |
| Oceania | 30 (4, 66) | 111, 273 | 98.11 | |
| Africa | 8 (1, 26) | 2, 25 | - | |
| Automated system | 12 (5, 20) | 23, 188 | - | |
| Disk diffusion | 43 (32, 54) | 1081, 2490 | 97.04 | |
| Dilution methods | 50 (40, 60) | 4972, 10909 | 98.91 | |
| MIX | 26 (15, 38) | 313, 872 | 92.04 | |
| Meropenem | | | | |
| Overall | 28 (23, 33) | 6410, 17032 | 97.55 | 0.17 |
| 1979-2000 | 7 (6, 9) | 78, 1051 | - | |
| 2001-2010 | 27 (20, 36) | 3282, 7936 | 97.77 | |
| 2011-2021 | 29 (23, 35) | 3050, 8045 | 96.69 | |
| Nontuberculous mycobacteria | - | - | - | |
| Burkholderia spp. | 38 (28, 47) | 1552, 3219 | 88.28 | |
| Streptococcus spp. | 7 (1, 15) | 7, 73 | - | |
| P. aeruginosa | 23 (18, 28) | 4101, 12478 | 97.59 | |
| S. aureus | 53 (28, 77) | 9, 17 | - | |
| S. maltophilia | 97 (92, 100) | 471, 487 | 66.76 | |
| Others | 18 (7, 33) | 270, 758 | 93.66 | |
| Asia | 14 (0, 51) | 154, 324 | 97.01 | |
| Europe | 35 (28, 41) | 4504, 12268 | 97.65 | |
| North America | 41 (29, 53) | 1520, 3453 | 96.13 | |
| South America | 14 (1, 34) | 65, 272 | 94.41 | |
| Oceania | 3 (0, 9) | 59, 274 | 92.19 | |
| Africa | 8 (1, 26) | 2, 25 | - | |
| Automated system | 6 (2, 14) | 5, 75 | - | |
| Disk diffusion | 25 (17, 34) | 1399, 3563 | 97.03 | |

Table 1. Prevalence of Carbapenem Resistance in Cystic Fibrosis Based on Years, Pathogens, AST, and Continents

| Dilution methods | 31 (24, 38) | 4721, 12585 | 98.12 | |
|------------------------------|-------------|-------------|-------|------|
| MIX | 27 (15, 40) | 285, 809 | 92.74 | |
| Doripenem | | | | - |
| Overall | 45 (32, 59) | 374, 795 | 90.76 | 0.91 |
| 1979-2000 | - | - | - | |
| 2001-2010 | 49 (45, 53) | 294, 600 | - | |
| 2011-2021 | 42 (13, 74) | 80, 195 | 93.65 | |
| Non-tuberculous mycobacteria | - | - | - | |
| Burkholderia spp. | 51 (31, 80) | 120, 231 | 74.07 | |
| Streptococcus spp. | - | - | - | |
| S. maltophilia | - | - | - | |
| P. aeruginosa | 39 (22, 58) | 254, 564 | 94.42 | |
| S. aureus | - | - | - | |
| Others | - | - | - | |
| Asia | 0 (0, 6) | 2, 52 | | |
| Europe | 63 (40, 84) | 78, 143 | 83.69 | |
| North America | 49 (45, 53) | 294, 600 | - | |
| South America | - | - | - | |
| Oceania | - | - | - | |
| Africa | - | - | - | |
| Automated system | - | - | - | |
| Disk diffusion | 75 (57, 90) | 23, 31 | - | |
| Dilution methods | 49 (43, 56) | 349, 712 | 66.52 | |
| MIX | 0 (0, 6) | 2, 52 | - | |

tance notably increased from 7% (95% CI 6-9) of 1,051 strains in 1979-2000 reaching 27% (95% CI 20-36) of 7,936 strains in 2001-2010. The frequency of meropenem resistance during the years 2011-2021 represents a gradual increase from the years 2001-2010. However, there was significant variation in the proportion of meropenem resistance isolates over time (P < 0.01). The highest meropenem resistance rate was shown in North America, followed by Europe (41%, 95% CI 29-33; 35%, 95% CI 28-41). Among 22 countries reporting resistance data for meropenem, 10 (~45.45%) countries (China, Scotland, Portugal, Spain, Turkey, Ireland, Italy, US, UK, and France) reported that > 28% of isolates had meropenem resistance. The highest meropenem resistance rate was shown in S. maltophilia followed by S. aureus (97%, 95% CI 93-100; 53%, 95% CI 28-77) (Table 1). Statistically, a significant difference was found in the subgroups AST, continents, and pathogens (P < 0.001).

Doripenem: Susceptibility to doripenem was determined in 795 bacterial pathogens associated with CF; the WPR was 45% (95% CI 32-59) with substantial heterogeneity (I²=90.76%) (Table 1). The subgroup analysis that compared the data from 2001-2010 (WPR 49%; 95% CI 45-53) and 2011-2021 (WPR 42%; 95% CI 13-74) indicated a minor decrease in the resistance rate (P < 0.001) (Table 1, Figure 3). The highest doripenem resistance rate was shown in Europe, followed by North America (63%, 95% CI 40-84; 49%, 95% CI 45-53). Among 27 countries reporting resistance data for doripenem, five countries (including the US, Argentina, and Denmark) reported that >45% of isolates had resistance. The highest doripenem resistance rate was shown in *Burkholderia* spp., followed by *P. aeruginosa* (51%, 95% CI 31-80; 39%, 95% CI 22-58) (Table 1). Statistically, a significant difference was found in the subgroups AST, continents, and pathogens (P < 0.001).

PUBLICATION BIAS

Egger's regression tests were performed to assess small study effect, and the results for each antibiotic were mentioned in Table 1. Egger's regression tests were conducted to evaluate the influence of small studies as publication bias. However, the *P*-value of Egger's test do not support the existence of publication bias for all antibiotics (P > 0.05).



Figure 2. The prevalence of carbapenem resistance in CF.

DISCUSSION

The crisis of AMR has potentially life-threatening concerns for management of infections, especially in chronic respiratory infection [5]. AMR can present major challenges in the treatment of CF lung infections [122]. This meta-analysis was conducted to consider the global status of carbapenem resistance (imipenem, meropenem, and doripenem) in CF. It is important to obtain further data about the carbapenem resistance profiles of circulating bacterial pathogens associated with CF. *P. aeruginosa, S. aureus, S. maltophilia,* and *Burkholderia* spp. remain the most frequent pathogens associated with CF patients [123].

Carbapenems such as imipenem, meropenem, and doripenem are potent broad-spectrum antimicrobial agents commonly used for treating of bacterial infections with these bacteria [6,124,125]. However, the increasing emergence and rapid development of resistance to these antibiotics, mainly among Gram-negative pathogens, dramatically limits treatment options in many countries and constitutes a major threat to global health [6,124,125]. Persistent colonization and infections with drug resistant pathogens have been associated with the progression of lung damage and raised morbidity and mortality among CF patients [126].

In 110 included studies, the prevalence of imipenem, meropenem, and doripenem resistance in CF was 43%, 28%, and 45%, respectively. Our meta-analysis showed that except meropenem, trends of imipenem and doripenem-resistance had decreased over time (1979-2021). This could be due to the limited clinical effectiveness of these antibiotics to treat CF cases over time. Regarding meropenem resistance, a huge increase was noted from the earlier time period of 1979-2000 to the period 2001-2010, but this was based on a single study [127]. Due to more frequent severe recessive disorder in people of European



Figure 3. The prevalence of carbapenem resistance in CF stratified by publication year.

descent, strong clinical awareness, and greater health facilities, it should be stated that most of reports were from Europe and North America countries [128]. Thus, our meta-analysis displays that the data are biased towards Europe and North America and the frequency of carbapenem-resistance of CF in Europe and North America were much higher than comparing with other continents. In Asian and African countries, the national CF registration system is mostly lacking, or it is individualized-based research due to civil war, poverty, malnutrition, and outbreaks of infectious diseases. The WHO has already listed CF as a significant disorder and requested all countries to update their data and reports on CF cases that underlines the urgent need for revitalization of national and global CF registration, worldwide [129]. Among the opportunistic pathogens associated with CF, the highest carbapenem-resistance rates were shown in S. maltophilia, Burkholderia spp., P. aeruginosa, and S. aureus. This could be due to their biofilm formation, their long-term persistence despite rigorous antibiotic therapy, high frequency of hypermutable or mutator microorganisms in CF chronic respiratory infection [5,8,122,130,131].

P. aeruginosa is the leading cause of morbidity and mortality in CF patients [132]. The highest and lowest carbapenem-resistance rates among *P. aeruginosa* in CF patients were shown against meropenem (23%) and doripenem (39%). Several mechanisms involved in carbapenems reduced susceptibility or resistance in *P. aeruginosa* isolates including target site mutations, carbapenem-hydrolyzing enzymes (mainly metallo-β-lactamases, AmpC chromosome-encoded cephalosporinase), altered permeability via deficiency in outer membrane porin oprD (through *parRS*, *mexS*, and *czcS* genes), overproduction of active efflux systems (overexpression of the MexAB-OprM, MexCD-OprJ, and MexXY-OprM), and environmental triggers [133-136].

CONCLUSION

We indicated that trends of imipenem, meropenem, and doripenem-resistance decreased over time (1979-2021). This could be due to the limited clinical effectiveness of these antibiotics to treat CF cases over time. The relatively high carbapenem-resistant rates were shown among the opportunistic pathogens associated with CF. With changing epidemiology and antimicrobial susceptibility trending among the opportunistic pathogens associated with CF, our meta-analysis underlines the importance of continuous monitoring, which could be applied by policymakers and health workers for good management of biofilm-associated infections. Plans should be directed to fight biofilm-associated infections and prevent the emergence of mutational resistance. Systematic surveillance for carbapenemase-producing pathogens in CF by molecular surveillance is necessitated.

Availability of data and materials: All the data in this review are included in the manuscript.

Competing interests: The authors declare that they have no competing interests.

Funding: Not applicable.

REFERENCES

- Goetz D, Ren CL. Review of Cystic Fibrosis. Pediatr Ann. 2019 Apr 1;48(4):e154-e161.
- Spoonhower KA, Davis PB. Epidemiology of Cystic Fibrosis. Clin Chest Med. 2016 Mar;37(1):1-8.
- Stephenson AL, Stanojevic S, Sykes J, Burgel PR. The changing epidemiology and demography of cystic fibrosis. Presse Med. 2017 Jun;46(6 Pt 2):e87-e95.
- Parkins MD, Floto RA. Emerging bacterial pathogens and changing concepts of bacterial pathogenesis in cystic fibrosis. J Cyst Fibros. 2015 May;14(3):293-304.
- Blanchard AC, Waters VJ, editors. Microbiology of cystic fibrosis airway disease. Seminars in respiratory and critical care medicine. Thieme Medical Publishers; 2019.
- Papp-Wallace KM, Endimiani A, Taracila MA, Bonomo RA. Carbapenems: past, present, and future. Antimicrob Agents Chemother. 2011 Nov;55(11):4943-60.
- Doi Y. Treatment options for carbapenem-resistant gram-negative bacterial infections. Clin Infect Dis. 2019;69(Supplement_7):S565-S75. https://doi.org/10.1093/ cid/ciz830.
- Kidd TJ, Canton R, Ekkelenkamp M, Johansen HK, Gilligan P, LiPuma JJ, Bell SC, Elborn JS, Flume PA, VanDevanter DR, Waters VJ; Antimicrobial Resistance in Cystic Fibrosis International Working Group. Defining antimicrobial resistance in cystic fibrosis. J Cyst Fibros. 2018 Nov;17(6):696-704.
- 9. Moher D, Liberati A, Tetzlaff J, Altman DG, med PGJP. Preferred reporting items for systematic reviews and meta-analyses: the PRISMA statement. PlosMed.

2009;6(7):e1000097.

- Modesti P, Reboldi G, Cappuccio FJ. [adapted for cross sectional studies]. Newcastle-Ottawa Quality Assessment Scale. 2016;11(1):e0147601.
- DerSimonian R, Laird N. Meta-analysis in clinical trials. Control Clin Trials. 1986 Sep;7(3):177–88.
- Alhanout K, Brunel JM, Raoult D, Rolain JM. In vitro antibacterial activity of aminosterols against multidrug-resistant bacteria from patients with cystic fibrosis. J Antimicrob Chemother. 2009 Oct;64(4):810–4.
- Amoureux L, Bador J, Siebor E, Taillefumier N, Fanton A, Neuwirth C. Epidemiology and resistance of Achromobacter xylosoxidans from cystic fibrosis patients in Dijon, Burgundy: first French data. J Cyst Fibros. 2013 Mar;12(2):170–6.
- 14. Ashish A, Shaw M, Winstanley C, Ledson MJ, Walshaw MJ. Increasing resistance of the Liverpool Epidemic Strain (LES) of *Pseudomonas aeruginosa* (Psa) to antibiotics in cystic fibrosis (CF)—a cause for concern? J Cyst Fibros. 2012 May;11(3):173–9.
- 15. Atkin SD, Abid S, Foster M, Bose M, Keller A, Hollaway R, et al. Multidrug-resistant *Pseudomonas aeruginosa* from sputum of patients with cystic fibrosis demonstrates a high rate of susceptibility to ceftazidime-avibactam. Infect Drug Resist. 2018 Sep;11:1499–510.
- 16. Balke B, Hogardt M, Schmoldt S, Hoy L, Weissbrodt H, Häussler S. Evaluation of the E test for the assessment of synergy of antibiotic combinations against multiresistant *Pseudomonas aeruginosa* isolates from cystic fibrosis patients. Eur J Clin Microbiol Infect Dis. 2006 Jan;25(1):25– 30.
- Balke B, Hoy L, Weissbrodt H, Häussler S. Comparison of the Micronaut Merlin automated broth microtiter system with the standard agar dilution method for antimicrobial susceptibility testing of mucoid and nonmucoid *Pseudomonas aeruginosa* isolates from cystic fibrosis patients. Eur J Clin Microbiol Infect Dis. 2004 Oct;23(10):765–71.
- Ballestero S, Fernández-Rodríguez A, Villaverde R, Escobar H, Pérez-Díaz JC, Baquero F. Carbapenem resistance in *Pseudomonas aeruginosa* from cystic fibrosis patients. J Antimicrob Chemother. 1996 Jul;38(1):39–45.
- Bevivino A, Dalmastri C, Tabacchioni S, Chiarini L, Belli ML, Piana S, et al. *Burkholderia* cepacia complex bacteria from clinical and environmental sources in Italy: genomovar status and distribution of traits related to virulence and transmissibility. J Clin Microbiol. 2002 Mar;40(3):846–51.
- Blondeau JM, Suter ME, Borsos S, Misfeldt C, Group TC. Canadian *Pseudomonas aeruginosa* susceptibility study from 48 medical centers: focus on ciprofloxacin. Int J Antimicrob Agents. 1998 Nov;10(4):297–302.
- Bradbury R, Champion A, Reid DW. Poor clinical outcomes associated with a multi-drug resistant clonal strain of *Pseudomonas aeruginosa* in the Tasmanian cystic fibrosis population. Respirology. 2008 Nov;13(6):886–92.
- 22. Bretonnière C, Maitte A, Caillon J, Potel G, Boutoille D, Jacqueline C, et al. MIC score, a new tool to compare bacterial susceptibility to antibiotics application to the comparison of susceptibility to different penems of clinical strains of *Pseudomonas aeruginosa*. J Antibiot (Tokyo). 2016 Nov;69(11):806–10.

- 23. Broda A, Jebbari H, Beaton K, Mitchell S, Drobniewski F. Comparative drug resistance of Mycobacterium abscessus and M. chelonae isolates from patients with and without cystic fibrosis in the United Kingdom. J Clin Microbiol. 2013 Jan;51(1):217–23.
- Burns JL. Burkholderia cepacia—a transmissible cystic fibrosis pathogen. J Pediatr. 2001 Nov;139(5):618–9.
- 25. Cantón R, Valdezate S, Vindel A, Sánchez Del Saz B, Maíz L, Baquero F. Antimicrobial susceptibility profile of molecular typed cystic fibrosis *Stenotrophomonas* maltophilia isolates and differences with noncystic fibrosis isolates. Pediatr Pulmonol. 2003 Feb;35(2):99–107.
- 26. Cardines R, Giufrè M, Pompilio A, Fiscarelli E, Ricciotti G, Di Bonaventura G, et al. Haemophilus influenzae in children with cystic fibrosis: antimicrobial susceptibility, molecular epidemiology, distribution of adhesins and bio-film formation. Int J Med Microbiol. 2012 Jan;302(1):45–52.
- Cardoso O, Alves AF, Leitão R. Metallo-β-lactamase VIM-2 in *Pseudomonas aeruginosa* isolates from a cystic fibrosis patient. Int J Antimicrob Agents. 2008 Apr;31(4):375–9.
- Chen Y, Garber E, Zhao Q, Ge Y, Wikler MA, Kaniga K, et al. In vitro activity of doripenem (S-4661) against multidrug-resistant gram-negative bacilli isolated from patients with cystic fibrosis. Antimicrob Agents Chemother. 2005 Jun;49(6):2510–1.
- Chiron R, Marchandin H, Counil F, Jumas-Bilak E, Freydière AM, Bellon G, et al. Clinical and microbiological features of Inquilinus sp. isolates from five patients with cystic fibrosis. J Clin Microbiol. 2005 Aug;43(8):3938–43.
- 30. Cipolla L, Rocca F, Martinez C, Aguerre L, Barrios R, Prieto M. Prevalence of *Burkholderia* cepacia complex species in cystic fibrosis patients in Argentina during the period 2011-2015. Enferm Infecc Microbiol Clin (Engl Ed). 2018;36(7):431–4.
- Courtois N, Caspar Y, Maurin M. Phenotypic and genetic resistance traits of *Pseudomonas aeruginosa* strains infecting cystic fibrosis patients: A French cohort study. Int J Antimicrob Agents. 2018 Sep;52(3):358–64.
- 32. Coward A, Kenna DT, Woodford N, Turton JF, Armstrong M, Auckland C, et al.; and members of the UK CF Surveillance Working Group., The UK CF Surveillance Working Group comprised. Structured surveillance of Achromobacter, Pandoraea and Ralstonia species from patients in England with cystic fibrosis. J Cyst Fibros. 2020 May;19(3):388–93.
- Crispino M, Boccia MC, Bagattini M, Villari P, Triassi M, Zarrilli R. Molecular epidemiology of *Stenotrophomonas* maltophilia in a university hospital. J Hosp Infect. 2002 Oct;52(2):88–92.
- 34. de Dios Caballero J, Pastor MD, Vindel A, Máiz L, Yagüe G, Salvador C, et al.; GEIFQ Study Group. Emergence of cfr-mediated linezolid resistance in a methicillin-resistant *Staphylococcus aureus* epidemic clone isolated from patients with cystic fibrosis. Antimicrob Agents Chemother. 2015 Dec;60(3):1878–82.
- 35. del Campo R, Morosini MI, de la Pedrosa EG, Fenoll A, Muñoz-Almagro C, Máiz L, et al.; Spanish Pneumococcal Infection Study Network. Population structure, antimicrobial resistance, and mutation frequencies of Streptococcus

pneumoniae isolates from cystic fibrosis patients. J Clin Microbiol. 2005 May;43(5):2207–14.

- 36. Bedir Demirdag T, Ozkaya Parlakay A, Aygar IS, Gulhan B, Kanik Yuksek S. Major Aspects of *Burkholderia* gladioli and *Burkholderia* cepacia Infections in Children. Pediatr Infect Dis J. 2020 May;39(5):374–8.
- Deredjian A, Colinon C, Brothier E, Favre-Bonté S, Cournoyer B, Nazaret S. Antibiotic and metal resistance among hospital and outdoor strains of *Pseudomonas aeruginosa*. Res Microbiol. 2011 Sep;162(7):689–700.
- Díez-Aguilar M, Ekkelenkamp M, Morosini MI, Merino I, de Dios Caballero J, Jones M, et al. Antimicrobial susceptibility of non-fermenting Gram-negative pathogens isolated from cystic fibrosis patients. Int J Antimicrob Agents. 2019 Jan;53(1):84–8.
- Digoy GP, Dunn JD, Stoner JA, Christie A, Jones DT. Bacteriology of the paranasal sinuses in pediatric cystic fibrosis patients. Int J Pediatr Otorhinolaryngol. 2012 Jul;76(7):934–8.
- 40. Ekkelenkamp MB, Cantón R, Díez-Aguilar M, Tunney MM, Gilpin DF, Bernardini F, et al. Susceptibility of *Pseudomonas aeruginosa* recovered from cystic fibrosis patients to murepavadin and 13 comparator antibiotics. Antimicrob Agents Chemother. 2020 Jan;64(2):e01541–19.
- Emerson J, McNamara S, Buccat AM, Worrell K, Burns JL. Changes in cystic fibrosis sputum microbiology in the United States between 1995 and 2008. Pediatr Pulmonol. 2010 Apr;45(4):363–70.
- 42. Feliziani S, Luján AM, Moyano AJ, Sola C, Bocco JL, Montanaro P, et al. Mucoidy, quorum sensing, mismatch repair and antibiotic resistance in *Pseudomonas aeruginosa* from cystic fibrosis chronic airways infections. PLoS One. 2010 Sep;5(9):e12669.
- 43. Filipic B, Malesevic M, Vasiljevic Z, Lukic J, Novovic K, Kojic M, et al. Uncovering differences in virulence markers associated with Achromobacter species of CF and non-CF origin. Front Cell Infect Microbiol. 2017 May;7:224.
- 44. Finklea JD, Hollaway R, Lowe K, Lee F, Le J, Jain R. Ceftolozane/tazobactam sensitivity patterns in *Pseudo-monas aeruginosa* isolates recovered from sputum of cystic fibrosis patients. Diagn Microbiol Infect Dis. 2018 Sep;92(1):75–7.
- 45. Forozsh FM, Irajian G, Moslehi TZ, Fazeli H, Salehi M, Rezania S. Drug resistance pattern of *Pseudomonas aeruginosa* strains isolated from cystic fibrosis patients at Isfahan AL Zahra hospital, Iran (2009-2010). Iran J Microbiol. 2012 Jun;4(2):94–7.
- 46. García-Castillo M, del Campo R, Baquero F, Morosini MI, Turrientes MC, Zamora J, et al. Stationary biofilm growth normalizes mutation frequencies and mutant prevention concentrations in *Pseudomonas aeruginosa* from cystic fibrosis patients. Clin Microbiol Infect. 2011 May;17(5):704–11.
- 47. Gherardi G, Linardos G, Pompilio A, Fiscarelli E, Di Bonaventura G. Evaluation of in vitro activity of ceftolozane-tazobactam compared to other antimicrobial agents against *Pseudomonas aeruginosa* isolates from cystic fibrosis patients. Diagn Microbiol Infect Dis. 2019 Jul;94(3):297–303.
- 48. Golini G, Cazzola G, Fontana R. Molecular epidemiology

and antibiotic susceptibility of *Burkholderia* cepacia-complex isolates from an Italian cystic fibrosis centre. Eur J Clin Microbiol Infect Dis. 2006 Mar;25(3):175–80.

- 49. Güzel ÇB, Gerçeker AA. In vitro activities of various antibiotics, alone and in combination with colistin methanesulfonate, against *Pseudomonas aeruginosa* strains isolated from cystic fibrosis patients. Chemotherapy. 2008;54(2):147–51.
- 50. Henwood CJ, Livermore DM, James D, Warner M; Pseudomonas Study Group. Antimicrobial susceptibility of *Pseudomonas aeruginosa*: results of a UK survey and evaluation of the British Society for Antimicrobial Chemotherapy disc susceptibility test. J Antimicrob Chemother. 2001 Jun;47(6):789–99.
- Høiby N, Bjarnsholt T, Givskov M, Molin S, Ciofu O. Antibiotic resistance of bacterial biofilms. Int J Antimicrob Agents. 2010 Apr;35(4):322–32.
- 52. Jansen G, Mahrt N, Tueffers L, Barbosa C, Harjes M, Adolph G, et al. Association between clinical antibiotic resistance and susceptibility of Pseudomonas in the cystic fibrosis lung. Evol Med Public Health. 2016 Jun;2016(1):182–94.
- 53. Lambiase A, Catania MR, Del Pezzo M, Rossano F, Terlizzi V, Sepe A, et al. Achromobacter xylosoxidans respiratory tract infection in cystic fibrosis patients. Eur J Clin Microbiol Infect Dis. 2011 Aug;30(8):973–80.
- Lambiase A, Del Pezzo M, Raia V, Sepe A, Ferri P, Rossano F. Chryseobacterium respiratory tract infections in patients with cystic fibrosis. J Infect. 2007 Dec;55(6):518–23.
- 55. Lambiase A, Raia V, Del Pezzo M, Sepe A, Carnovale V, Rossano F. Microbiology of airway disease in a cohort of patients with cystic fibrosis. BMC Infect Dis. 2006 Jan;6(1):4.
- 56. Lasko MJ, Huse HK, Nicolau DP, Kuti JL. Contemporary analysis of ETEST for antibiotic susceptibility and minimum inhibitory concentration agreement against *Pseudomonas aeruginosa* from patients with cystic fibrosis. Ann Clin Microbiol Antimicrob. 2021 Jan;20(1):9.
- 57. Leitão JH, Sousa SA, Cunha MV, Salgado MJ, Melo-Cristino J, Barreto MC, et al. Variation of the antimicrobial susceptibility profiles of *Burkholderia* cepacia complex clonal isolates obtained from chronically infected cystic fibrosis patients: a five-year survey in the major Portuguese treatment center. Eur J Clin Microbiol Infect Dis. 2008 Nov;27(11):1101–11.
- 58. Li Y, Zhang X, Wang C, Hu Y, Niu X, Pei D, et al. Characterization by phenotypic and genotypic methods of metallo-β-lactamase-producing *Pseudomonas aeruginosa* isolated from patients with cystic fibrosis. Mol Med Rep. 2015 Jan;11(1):494–8.
- Livermore DM, Mushtaq S, Ge Y, Warner M. Activity of cephalosporin CXA-101 (FR264205) against *Pseudomonas aeruginosa* and *Burkholderia* cepacia group strains and isolates. Int J Antimicrob Agents. 2009 Nov;34(5):402–6.
- 60. Llanes C, Pourcel C, Richardot C, Plésiat P, Fichant G, Cavallo JD, et al.; GERPA Study Group. Diversity of β-lactam resistance mechanisms in cystic fibrosis isolates of *Pseudomonas aeruginosa*: a French multicentre study. J Antimicrob Chemother. 2013 Aug;68(8):1763–71.
- 61. Logan C, Habington A, Lennon G, Grogan J, Byrne M,

O'Leary J, et al. Genetic relatedness of *Pseudomonas aeruginosa* isolates among a paediatric cystic fibrosis patient cohort in Ireland. J Med Microbiol. 2012 Jan;61(Pt 1):64–70.

- 62. López-Causapé C, de Dios-Caballero J, Cobo M, Escribano A, Asensio Ó, Oliver A, et al. Antibiotic resistance and population structure of cystic fibrosis *Pseudomonas aeruginosa* isolates from a Spanish multi-centre study. Int J Antimicrob Agents. 2017 Sep;50(3):334–41.
- 63. Lutz L, Pereira DC, Paiva RM, Zavascki AP, Barth AL. Macrolides decrease the minimal inhibitory concentration of anti-pseudomonal agents against *Pseudomonas aeruginosa* from cystic fibrosis patients in biofilm. BMC Microbiol. 2012 Sep;12(1):196.
- 64. Macdonald D, Cuthbertson L, Doherty C, Campana S, Ravenni N, Taccetti G, et al. Early *Pseudomonas aerugino-sa* infection in individuals with cystic fibrosis: is susceptibility testing justified? J Antimicrob Chemother. 2010 Nov;65(11):2373–5.
- 65. Maciá MD, Blanquer D, Togores B, Sauleda J, Pérez JL, Oliver A. Hypermutation is a key factor in development of multiple-antimicrobial resistance in *Pseudomonas aeruginosa* strains causing chronic lung infections. Antimicrob Agents Chemother. 2005 Aug;49(8):3382–6.
- 66. Macin S, Akarca M, Sener B, Akyon Y. Comparison of virulence factors and antibiotic resistance of *Pseudomonas aeruginosa* strains isolated from patients with and without cystic fibrosis. 2017. https://doi.org/10.1515/rrlm-2017-0027.
- 67. Manno G, Ugolotti E, Belli ML, Fenu ML, Romano L, Cruciani M. Use of the E test to assess synergy of antibiotic combinations against isolates of *Burkholderia* cepacia-complex from patients with cystic fibrosis. Eur J Clin Microbiol Infect Dis. 2003 Jan;22(1):28–34.
- 68. Martin I, Kenna DT, Morales S, Alton EW, Davies JC. Variability in bacteriophage and antibiotic sensitivity in serial *Pseudomonas aeruginosa* isolates from cystic fibrosis airway cultures over 12 months. Microorganisms. 2021 Mar;9(3):660.
- Martina PF, Martinez M, Rivas S, Leguizamón L, Von Specht M, Ferreras J. *Burkholderia* cepacia complex: 11 years of surveillance in patients with Cystic Fibrosis in Posadas, Argentina. Rev Argent Microbiol. 2020;52(3):176–82.
- 70. Masoud-Landgraf L, Badura A, Eber E, Feierl G, Posch J, Zarfel G, et al. Molecular epidemiology of *Pseudomonas aeruginosa* in cystic fibrosis patients from Southeast Austria. Wien Klin Wochenschr. 2012 Apr;124(7-8):262–5.
- 71. Mathy V, Grohs P, Compain F. In vitro activity of β-lactams in combination with avibactam against multidrug-resistant *Pseudomonas aeruginosa*, *Stenotrophomonas* maltophilia and Achromobacter xylosoxidans isolates from patients with cystic fibrosis. J Med Microbiol. 2018 Sep;67(9):1217–20.
- Medina-Pascual MJ, Valdezate S, Carrasco G, Villalón P, Garrido N, Saéz-Nieto JA. Increase in isolation of *Burk-holderia* contaminans from Spanish patients with cystic fibrosis. Clin Microbiol Infect. 2015 Feb;21(2):150–6.
- Medina-Pascual MJ, Valdezate S, Villalón P, Garrido N, Rubio V, Saéz-Nieto JA. Identification, molecular charac-

terisation and antimicrobial susceptibility of genomovars of the *Burkholderia* cepacia complex in Spain. Eur J Clin Microbiol Infect Dis. 2012 Dec;31(12):3385–96.

- 74. Milne KE, Gould IM. Combination antimicrobial susceptibility testing of multidrug-resistant *Stenotrophomonas* maltophilia from cystic fibrosis patients. Antimicrob Agents Chemother. 2012 Aug;56(8):4071–7.
- 75. Morosini MI, García-Castillo M, Loza E, Pérez-Vázquez M, Baquero F, Cantón R. Breakpoints for predicting *Pseudomonas aeruginosa* susceptibility to inhaled tobramycin in cystic fibrosis patients: use of high-range Etest strips. J Clin Microbiol. 2005 Sep;43(9):4480–5.
- Musafer HK, Kuchma SL, Naimie AA, Schwartzman JD, Al-Mathkhury HJ, O'Toole GA. Investigating the link between imipenem resistance and biofilm formation by *Pseudomonas aeruginosa*. Microb Ecol. 2014 Jul;68(1):111–20.
- Mutnick AH, Turner PJ, Jones RN. Emerging antimicrobial resistances among Proteus mirabilis in Europe: report from the MYSTIC program (1997-2001). J Chemother. 2002 Jun;14(3):253–8.
- Narayanaswamy VP, Giatpaiboon S, Baker SM, Wiesmann WP, LiPuma JJ, Townsend SM. Novel glycopolymer sensitizes *Burkholderia* cepacia complex isolates from cystic fibrosis patients to tobramycin and meropenem. PLoS One. 2017 Jun;12(6):e0179776.
- Nazik H, Ongen B, Erturan Z, Salcioğlu M. Genotype and antibiotic susceptibility patterns of *Pseudomonas aerugino*sa and *Stenotrophomonas* maltophilia isolated from cystic fibrosis patients. Jpn J Infect Dis. 2007 May;60(2-3):82–6.
- 80. Mazloomi Nobandegani N, Mahmoudi S, Pourakbari B, Hosseinpour Sadeghi R, Najafi Sani M, Farahmand F, et al. Antimicrobial susceptibility of microorganisms isolated from sputum culture of patients with cystic fibrosis: methicillin-resistant *Staphylococcus aureus* as a serious concern. Microb Pathog. 2016 Nov;100:201–4.
- Nolan PJ, Jain R, Cohen L, Finklea JD, Smith TT. In vitro activity of ceftolozane-tazobactam and ceftazidime-avibactam against *Pseudomonas aeruginosa* isolated from patients with cystic fibrosis. Diagn Microbiol Infect Dis. 2021 Feb;99(2):115204.
- 82. O'Carroll MR, Syrmis MW, Wainwright CE, Greer RM, Mitchell P, Coulter C, et al. Clonal strains of *Pseudomonas aeruginosa* in paediatric and adult cystic fibrosis units. Eur Respir J. 2004 Jul;24(1):101–6.
- Oermann CM, McCoy KS, Retsch-Bogart GZ, Gibson RL, McKevitt M, Montgomery AB. *Pseudomonas aeruginosa* antibiotic susceptibility during long-term use of aztreonam for inhalation solution (AZLI). J Antimicrob Chemother. 2011 Oct;66(10):2398–404.
- 84. Oliver A, Cantón R, Campo P, Baquero F, Blázquez J. High frequency of hypermutable *Pseudomonas aeruginosa* in cystic fibrosis lung infection. Science. 2000 May;288(5469):1251–4.
- 85. Paixão VA, Barros TF, Mota CM, Moreira TF, Santana MA, Reis JN. Prevalence and antimicrobial susceptibility of respiratory pathogens in patients with cystic fibrosis. Braz J Infect Dis. 2010;14(4):406–9.
- 86. Papp-Wallace KM, Becka SA, Zeiser ET, Ohuchi N, Mojica MF, Gatta JA, et al. Overcoming an extremely drug resistant (XDR) pathogen: avibactam restores suscepti-

bility to ceftazidime for *Burkholderia* cepacia complex isolates from cystic fibrosis patients. ACS Infect Dis. 2017 Jul;3(7):502–11.

- Pedersen SS, Pressler T, Høiby N, Bentzon MW, Koch C. Imipenem/cilastatin treatment of multiresistant *Pseudomo-nas aeruginosa* lung infection in cystic fibrosis. J Antimicrob Chemother. 1985 Nov;16(5):629–35.
- 88. Pereira RH, Leão RS, Carvalho-Assef AP, Albano RM, Rodrigues ER, Firmida MC, et al. Patterns of virulence factor expression and antimicrobial resistance in Achromobacter xylosoxidans and Achromobacter ruhlandii isolates from patients with cystic fibrosis. Epidemiol Infect. 2017 Feb;145(3):600–6.
- 89. Perez LR, Antunes AL, Freitas AL, Barth AL. When the resistance gets clingy: *pseudomonas aeruginosa* harboring metallo-β-lactamase gene shows high ability to produce biofilm. Eur J Clin Microbiol Infect Dis. 2012 May;31(5):711–4.
- 90. Perry JD, Laine L, Hughes S, Nicholson A, Galloway A, Gould FK. Recovery of antimicrobial-resistant *Pseudo-monas aeruginosa* from sputa of cystic fibrosis patients by culture on selective media. J Antimicrob Chemother. 2008 May;61(5):1057–61.
- 91. Pesavento G, Maggini V, Maida I, Nostro AL, Calonico C, Sassoli C, et al. Essential oil from Origanum vulgare completely inhibits the growth of multidrug-resistant cystic fibrosis pathogens. Natural Product Communications. 2016;11(6):1934578X1601100641. https://doi.org/10.1177/1934578X1601100641.
- 92. Phang SH, Greysson-Wong J, Somayaji R, Storey DG, Rabin HR, Surette MG, et al. Incidence, impact and natural history of Klebsiella species infections in cystic fibrosis: A longitudinal single center study. Canadian Journal of Respiratory, Critical Care, and Sleep Medicine. 2019;3(3):148–54.
- 93. Pivot D, Fanton A, Badell-Ocando E, Benouachkou M, Astruc K, Huet F, et al. Carriage of a single strain of nontoxigenic Corynebacterium diphtheriae bv. Belfanti (Corynebacterium belfantii) in four patients with cystic fibrosis. J Clin Microbiol. 2019 Apr;57(5):e00042–19.
- 94. Pollini S, Di Pilato V, Landini G, Di Maggio T, Cannatelli A, Sottotetti S, et al. In vitro activity of N-acetylcysteine against *Stenotrophomonas* maltophilia and *Burkholderia* cepacia complex grown in planktonic phase and biofilm. PLoS One. 2018 Oct;13(10):e0203941.
- 95. Pompilio A, Savini V, Fiscarelli E, Gherardi G, Di Bonaventura G. Clonal diversity, biofilm formation, and antimicrobial resistance among *Stenotrophomonas* maltophilia strains from cystic fibrosis and non-cystic fibrosis patients. Antibiotics (Basel). 2020 Jan;9(1):15.
- 96. Pournajaf A, Razavi S, Irajian G, Ardebili A, Erfani Y, Solgi S, et al. Integron types, antimicrobial resistance genes, virulence gene profile, alginate production and biofilm formation in Iranian cystic fibrosis *Pseudomonas aeruginosa* isolates. Infez Med. 2018 Sep;26(3):226–36.
- Rafiee R, Eftekhar F, Tabatabaii SA. Extended-Spectrum Beta-Lactamases in Cystic Fibrosis Isolates of Klebsiella pneumoniae. Jundishapur J Microbiol. 2018;11(1). https:// doi.org/10.5812/jjm.61086.
- 98. Raidt L, Idelevich EA, Dübbers A, Küster P, Drevinek

P, Peters G, et al. Increased prevalence and resistance of important pathogens recovered from respiratory specimens of cystic fibrosis patients during a decade. Pediatr Infect Dis J. 2015 Jul;34(7):700–5.

- Raja NS, Singh NN. Antimicrobial susceptibility pattern of clinical isolates of *Pseudomonas aeruginosa* in a tertiary care hospital. J Microbiol Immunol Infect. 2007 Feb;40(1):45–9.
- 100. Ramsay KA, Wardell SJ, Patrick WM, Brockway B, Reid DW, Winstanley C, et al. Genomic and phenotypic comparison of environmental and patient-derived isolates of *Pseudomonas aeruginosa* suggest that antimicrobial resistance is rare within the environment. bioRxiv. 2019:663674. https://doi.org/10.1101/663674.
- 101. Rees VE, Deveson Lucas DS, López-Causapé C, Huang Y, Kotsimbos T, Bulitta JB, et al. Characterization of hypermutator *Pseudomonas aeruginosa* isolates from patients with cystic fibrosis in Australia. Antimicrob Agents Chemother. 2019 Mar;63(4):e02538–18.
- 102. Rocha GA, Lima DF, Rodrigues ER, Leão RS, Folescu TW, Firmida MC, et al. Species distribution, sequence types and antimicrobial resistance of Acinetobacter spp. from cystic fibrosis patients. Epidemiol Infect. 2018 Mar;146(4):524–30.
- 103. Rodriguez-Nava V, Durupt S, Chyderiotis S, Freydière AM, Karsenty J, de Montclos M, et al. A French multicentric study and review of pulmonary Nocardia spp. in cystic fibrosis patients. Med Microbiol Immunol (Berl). 2015 Aug;204(4):493–504.
- 104. San Gabriel P, Zhou J, Tabibi S, Chen Y, Trauzzi M, Saiman L. Antimicrobial susceptibility and synergy studies of *Stenotrophomonas* maltophilia isolates from patients with cystic fibrosis. Antimicrob Agents Chemother. 2004 Jan;48(1):168–71.
- 105. Satana D, Erkose-Genc G, Tamay Z, Uzun M, Guler N, Erturan Z. Prevalence and drug resistance of *mycobacte-ria* in Turkish cystic fibrosis patients. Ann Clin Microbiol Antimicrob. 2014 Aug;13(1):28.
- 106. Shehata MM, Sayed AA. Genetic diversity and twitching motility of *Pseudomonas aeruginosa* strains isolated from different origins. Archives of Clinical Microbiology. 2011;2(5):0-.
- 107. Sherrard LJ, Graham KA, McGrath SJ, McIlreavey L, Hatch J, Muhlebach MS, et al. Antibiotic resistance in Prevotella species isolated from patients with cystic fibrosis. J Antimicrob Chemother. 2013 Oct;68(10):2369–74.
- 108. Smith WD, Bardin E, Cameron L, Edmondson CL, Farrant KV, Martin I, et al. Current and future therapies for *Pseudomonas aeruginosa* infection in patients with cystic fibrosis. FEMS Microbiol Lett. 2017 Aug;364(14). https:// doi.org/10.1093/femsle/fnx121.
- 109. Spencker FB, Staber L, Lietz T, Schille R, Rodloff AC. Development of resistance in *Pseudomonas aeruginosa* obtained from patients with cystic fibrosis at different times. Clin Microbiol Infect. 2003 May;9(5):370–9.
- 110. Spoletini G, Etherington C, Shaw N, Clifton IJ, Denton M, Whitaker P, et al. Use of ceftazidime/avibactam for the treatment of MDR *Pseudomonas aeruginosa* and *Burkholderia* cepacia complex infections in cystic fibrosis: a case series. J Antimicrob Chemother. 2019

May;74(5):1425-9.

- 111. Tabatabaei M, Dastbarsar M, Moslehi MA. Isolation and identification of Pandoraea spp. From bronchoalveolar lavage of cystic fibrosis patients in Iran. Ital J Pediatr. 2019 Sep;45(1):118.
- 112. Taccetti G, Campana S, Marianelli L. Multiresistant non-fermentative gram-negative bacteria in cystic fibrosis patients: the results of an Italian multicenter study. Italian Group for Cystic Fibrosis microbiology. Eur J Epidemiol. 1999 Jan;15(1):85–8.
- 113. Talebi G, Hakemi-Vala M. Survey on some carbapenems and colistin resistance genes among *Pseudomonas aeruginosa* isolates from burn and cystic fibrosis patients, Tehran, Iran. Arch Clin Infect Dis. 2019;14(5). https://doi. org/10.5812/archcid.93651.
- 114. Teri A, Sottotetti S, Arghittu M, Girelli D, Biffi A, D'Accico M, et al. Molecular characterization of Mycobacterium abscessus subspecies isolated from patients attending an Italian Cystic Fibrosis Centre. New Microbiol. 2020;43(3):127–132.
- 115. Tingpej P, Elkins M, Rose B, Hu H, Moriarty C, Manos J, et al. Clinical profile of adult cystic fibrosis patients with frequent epidemic clones of *Pseudomonas aeruginosa*. Respirology. 2010 Aug;15(6):923–9.
- 116. Traczewski MM, Brown SD. In vitro activity of doripenem against *Pseudomonas aeruginosa* and *Burkholderia* cepacia isolates from both cystic fibrosis and non-cystic fibrosis patients. Antimicrob Agents Chemother. 2006 Feb;50(2):819–21.
- 117. Valenza G, Tappe D, Turnwald D, Frosch M, König C, Hebestreit H, et al. Prevalence and antimicrobial susceptibility of microorganisms isolated from sputa of patients with cystic fibrosis. J Cyst Fibros. 2008 Mar;7(2):123–7.
- 118. Vali P, Shahcheraghi F, Seyfipour M, Zamani MA, Allahyar MR, Feizabadi MM. Phenotypic and genetic characterization of carbapenemase and ESBLs producing gram-negative bacteria (GNB) isolated from patients with cystic fibrosis (CF) in Tehran hospitals. J Clin Diagn Res. 2014 Jan;8(1):26–30.
- 119. Weiss K, Lapointe JR. Routine susceptibility testing of four antibiotic combinations for improvement of laboratory guide to therapy of cystic fibrosis infections caused by *Pseudomonas aeruginosa*. Antimicrob Agents Chemother. 1995 Nov;39(11):2411–4.
- 120. Worlitzsch D, Rintelen C, Böhm K, Wollschläger B, Merkel N, Borneff-Lipp M, et al. Antibiotic-resistant obligate anaerobes during exacerbations of cystic fibrosis patients. Clin Microbiol Infect. 2009 May;15(5):454–60.
- 121. Zhou J, Chen Y, Tabibi S, Alba L, Garber E, Saiman L. Antimicrobial susceptibility and synergy studies of *Burkholderia* cepacia complex isolated from patients with cystic fibrosis. Antimicrob Agents Chemother. 2007 Mar;51(3):1085–8.
- 122. Flume PA, Waters VJ, Bell SC, Van Devanter DR, Stuart Elborn J; Antimicrobial Resistance in Cystic Fibrosis International Working Group. Antimicrobial resistance in cystic fibrosis: Does it matter? J Cyst Fibros. 2018 Nov;17(6):687-689.
- 123. Salsgiver EL, Fink AK, Knapp EA, LiPuma JJ, Olivier KN, Marshall BC, Saiman L. Changing Epidemiology of

the Respiratory Bacteriology of Patients With Cystic Fibrosis. Chest. 2016 Feb;149(2):390-400.

- 124. Codjoe FS, Donkor ES. Carbapenem Resistance: A Review. Med Sci (Basel). 2017 Dec 21;6(1):1.
- 125. Meletis G. Carbapenem resistance: overview of the problem and future perspectives. Ther Adv Infect Dis. 2016 Feb;3(1):15-21.
- 126. Turcios NL. Cystic Fibrosis Lung Disease: An Overview. Respir Care. 2020 Feb;65(2):233-251.
- 127. Ballestero S, Fernández-Rodríguez A, Villaverde R, Escobar H, Pérez-Díaz JC, Baquero F. Carbapenem resistance in *Pseudomonas aeruginosa* from cystic fibrosis patients. J Antimicrob Chemother. 1996 Jul;38(1):39-45.
- 128. Bell SC, Mall MA, Gutierrez H, Macek M, Madge S, Davies JC, et al. The future of cystic fibrosis care: a global perspective. Lancet Respir Med. 2020 Jan;8(1):65-124. doi: 10.1016/S2213-2600(19)30337-6. Epub 2019 Sep 27. Erratum in: Lancet Respir Med. 2019 Dec;7(12):e40.
- 129. Mehta G, Macek M Jr, Mehta A; European Registry Working Group. Cystic fibrosis across Europe: EuroCare-CF analysis of demographic data from 35 countries. J Cyst Fibros. 2010 Dec;9 Suppl 2:S5-S21.
- López-Causapé C, Rojo-Molinero E, Macià MD, Oliver A. The problems of antibiotic resistance in cystic fibrosis and solutions. Expert Rev Respir Med. 2015 Feb;9(1):73-88.
- 131. Oliver A. Mutators in cystic fibrosis chronic lung infection: Prevalence, mechanisms, and consequences for antimicrobial therapy. Int J Med Microbiol. 2010 Dec;300(8):563-72.
- 132. Emerson J, Rosenfeld M, McNamara S, Ramsey B, Gibson RL. *Pseudomonas aeruginosa* and other predictors of mortality and morbidity in young children with cystic fibrosis. Pediatr Pulmonol. 2002 Aug;34(2):91-100.
- 133. Fusté E, López-Jiménez L, Segura C, Gainza E, Vinuesa T, Viñas M. Carbapenem-resistance mechanisms of multidrug-resistant *Pseudomonas aeruginosa*. J Med Microbiol. 2013 Sep;62(Pt 9):1317-1325.
- 134. Kamolvit W, Sidjabat HE, Paterson DL. Molecular Epidemiology and Mechanisms of Carbapenem Resistance of Acinetobacter spp. in Asia and Oceania. Microb Drug Resist. 2015 Aug;21(4):424-34.
- 135. Meletis G, Exindari M, Vavatsi N, Sofianou D, Diza E. Mechanisms responsible for the emergence of carbapenem resistance in *Pseudomonas aeruginosa*. Hippokratia. 2012 Oct;16(4):303-7.
- 136. Wang J, Zhou JY, Qu TT, Shen P, Wei ZQ, Yu YS, Li LJ. Molecular epidemiology and mechanisms of carbapenem resistance in *Pseudomonas aeruginosa* isolates from Chinese hospitals. Int J Antimicrob Agents. 2010 May;35(5):486-91.