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SPECIALTY SECTION

This article was submitted to Pediatric Surgery, a section of the journal Frontiers in Surgery

RECEIVED 29 April 2022 ACCEPTED 08 August 2022 PUBLISHED 05 September 2022

CITATION

Lyu H, Ye Y, Lui VCH, Wu W, Chung PHY, Wong KKY, Li H-W, Wong MS, Tam PKH and Wang B (2022) Plasma amyloid-beta levels correlated with impaired hepatic functions: An adjuvant biomarker for the diagnosis of biliary atresia.

Front. Surg. 9:931637. doi: 10.3389/fsurg.2022.931637

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Plasma amyloid-beta levels correlated with impaired hepatic functions: An adjuvant biomarker for the diagnosis of biliary atresia

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Background: Biliary atresia (BA) is an infantile fibro-obstructive cholestatic disease with poor prognosis. An early diagnosis and timely Kasai portoenterostomy (KPE) improve clinical outcomes. Aggregation of amyloidbeta (A β) around hepatic bile ducts has been discovered as a factor for BA pathogenesis, yet whether plasma A β levels correlate with hepatic dysfunctions and could be a biomarker for BA remains unknown.

Method: Plasma samples of 11 BA and 24 controls were collected for liver function test, A β 40 and A β 42 measurement by enzyme-linked immunosorbent assay (ELISA). Pearson's chi-squared test or Mann–Whitney U test was performed to assess differences between groups. Correlation between A β 42/A β 40 and liver function parameters was performed using Pearson analysis. The area under the receiver-operative characteristic (ROC) curve (area under curve; AUC) was measured to evaluate the diagnostic power of A β 42/A β 40 for BA. Diagnostic enhancement was further evaluated by binary regression ROC analysis of A β 42/A β 40 combined with other hepatic function parameters.

Results: Plasma A β 42/A β 40 was elevated in BA patients. A β 42 displayed a weak positive correlation with γ -glutamyl transpeptidase (GGT) (Pearson's correlation = 0.349), while there was no correlation for A β 40 with hepatic functions. A β 42/A β 40 was moderately correlated with GGT, total bile acid (TBA), direct bilirubin (DBIL) (Pearson's correlation = 0.533, 0.475, 0.480), and weakly correlated with total bilirubin (TBIL) (Pearson's correlation = 0.337). A β 42/A β 40 showed an acceptable predictive power for cholestasis [AUC = 0.746 (95% CI: 0.552–0.941), p < 0.05]. Diagnostic powers of A β 42/A β 40 together with hepatic function parameters for cholestasis were markedly improved compared to any indicator alone. Neither A β 42/A β 40 nor hepatic function parameters of A β 42/A β 40 + GGT along with any other hepatic function parameters could differentiate BA from CC-cholestasis (AUC = 1.000, p < 0.05) with a cut-off value as 0.02371, -0.28387, -0.34583, 0.06224, 0.01040, 0.06808, and 0.05898, respectively.

Conclusion: $A\beta 42/A\beta 40$ is a good indicator for cholestasis, but alone is insufficient for a distinction of BA from non-BA. However, $A\beta 42/A\beta 40$ combined with GGT and one other hepatic function parameter displayed a high predictive power as a screening test for jaundiced neonates who are more likely to be BA, enabling them to early intraoperative cholangiography for BA confirmation and KPE to improve surgical outcomes. However, a multi-centers validation is needed before introduction into daily clinical practice.

KEYWORDS

biliary atresia, amyloid-beta, hepatic function, biomarker, diagnosis

Introduction

Biliary atresia (BA) is a progressive fibrosclerosing disease of the biliary tract, resulting in obstructive bile flow, cholestasis, and jaundice in young infants. BA affects all ethnicities with a noticeably higher incidence in the Asia–Pacific region (5– 20:100,000 live-births) (1, 2). If untreated, patients develop progressive hepatic fibrosis leading to cirrhosis, portal hypertension, liver failure, and death by the age of two (1, 3). Kasai portoenterostomy (KPE) is the most widely accepted primary treatment. A timely KPE can potentially restore bile flow in 30%–80% of BA patients, but complications occur in many patients (4). It is estimated that up to 56%–74% of post-KPE BA patients at 10 years of age require liver transplantation (LT) (5).

Despite significant advances in BA management, clinicians face major challenges in establishing an early diagnosis for BA and predicting post-KPE outcomes. Early diagnosis is important because early KPE correlates with the best chances of good operative outcomes, delaying or even avoiding the need for LT (3). Currently, preoperative percutaneous liver biopsy has a higher specificity and sensitivity in early diagnosis of BA (6); however, an accurate, confirming diagnosis can only be established by surgical exploration and intraoperative cholangiography (7-9). Biomarkers are urgently needed for BA diagnosis. Plasma matrix metalloproteinase-7 (MMP7) (10-12) and γ -glutamyl transpeptidase (GGT) (13-16) have been suggested as promising tools in the diagnosis of BA; however, these studies have limitations such as different quantitation methods and different cut-off values were used. Furthermore, diagnostic accuracies of GGT for BA vary considerably in different age groups (16).

The amyloid precursor protein (APP) is pivotal in the pathophysiology of Alzheimer's disease (AD) since its abnormal cleavage by β -secretase and γ -secretase generates the hydrophobic amyloid-beta (A β) peptides including the two major A β peptides amyloid-beta 40 (A β 40) and amyloid-beta 42 (A β 42), which aggregate into neurotoxic amyloid plaques in the brain tissues, one of the key pathological hallmarks of AD [for a review, see (17)]. Aggregation of the A β peptides into amyloids is conceived as the pathogenic trigger of a

cascade leading to tau accumulation into neurofibrillary tangles, neuronal loss, and clinical dementia in AD. Plasma A β 42 correlates with cerebrospinal fluid A β 42 in AD patients, and blood A β 42 has been proposed as an alternative biomarker for AD (18–20).

Using human and mouse liver organoid and transcriptomics, we found (i) human and mouse BA liver organoids exhibited aberrant morphology, disturbed apicalbasal organization, defective cholangiocyte development and altered Aβ-related gene expression; (ii) Aβ peptide deposition in bile ducts of BA livers; and (iii) AB induced the aberrant morphology in control organoids (21). The aberrant organoid morphology and periductal AB deposition are novel pathobiological and diagnostic features of BA. Our data identified Aß deposition, the main pathological feature of AD and cerebral amyloid angiopathy, around BA bile ducts, suggesting that BA could be grouped under amyloid diseases. Plasma AB levels associate with hepatic functions in adults with liver cirrhosis (22). However, whether or not plasma Aβ levels correlate with hepatic functions in neonates and could be a non-invasive biomarker for BA is not known.

In the current study, we performed statistical and correlation analysis of plasma levels of A β 40, A β 42, and A β 42/A β 40 in BA and non-BA neonates to determine if plasma levels of A β peptides correlate with hepatic functions and can be used as an adjuvant biomarker to enhance the diagnostic accuracy for BA.

Materials and methods

Patients

This study was conducted prospectively at Shenzhen Children's Hospital, China, based on a protocol developed by the Hong Kong-Macau research team. Infants diagnosed with BA (N = 11) were enrolled from November 2021 to February 2022. The non-BA control group subjects included infants who suffered from choledochal cysts (CC; N = 5) as well as those patients (N = 19) with conditions unrelated to the liver. Intraoperative cholangiography and histologic examination

confirmed the diagnosis of BA. The study was approved by the ethical committee of Shenzhen Children's Hospital and informed consent was obtained from all guardians or legal representatives of patients. Patients' information was tabulated and shown in **Supplementary Table S1**.

Collection and preparation of plasma

Peripheral blood was collected into a vacuum tube containing EDTA at the time of KPE (BA patients) or at admission (non-BA subjects) and centrifuged (1,600 rpm for 10 min at 4 °C) to collect plasma for clinical laboratory tests and storage at -80 °C until A β peptides level quantitation. The clinical laboratory tests included the alanine aminotransferase (ALT), aspartate aminotransferase (AST), GGT, total bile acids (TBAs), total bilirubin (TBIL), direct bilirubin (DBIL), and indirect bilirubin (IBIL) levels.

Plasma A β 40 and A β 42 measurement

Plasma Aβ40 and Aβ42 levels were measured using enzymelinked immunosorbent assay (ELISA) kits [no. 27,718 human amyloid β (1-40) (FL) Assay Kit and no. 27,719 human amyloid β (1-42) (FL) Assay Kit, IBL, Gunma, Japan] according to the manufacturer's instructions. In brief, 100 µl of undiluted plasma samples were added to each well of the assay plate and incubated overnight at 4 °C. After washing, 100 µl of labeled antibody solution was added to each well, and the plate was incubated for 1 h at 4 °C. After washing, color was developed by incubation with 100 µl of chromogen at room temperature in dark, and the reactions were stopped by the addition of stop solution. The absorbance was measured at 450 nm using a plate reader (Infinite F50, TECAN), and the concentrations of (pg/ml) were calculated with reference to standard curves. AB42/AB40 ratios were multiplied by 100 and log₂-transformed before being subjected for statistical analysis.

Statistical analysis

Variables were presented as mean \pm SEM if normally distributed, and otherwise as Median (Q1, Q3) values. Continuous data if normally distributed were compared using analysis of variance (ANOVA). The Pearson's chi-squared test was performed to assess differences between groups. The Mann–Whitney *U* test was performed for the continuous variables which were non-normally distribution. The correlation between Aβ42/Aβ40 and each of the liver function biochemical parameters was performed using Pearson (0.2–0.4: weak correlation; 0.4–0.7: moderate correlation; >0.7:

strong correlation). The area under the receiver-operative characteristic (ROC) curve (area under curve; AUC) was calculated for A β 42/A β 40. For A β 42/A β 40 combined with positively correlated parameters of hepatic functions to predict BA, a binary regression for each independent variable was performed first, then a Logit(*P*) equation was derived, followed by ROC analysis. The sensitivity, specificity, Youden index of diagnostic test were calculated by discriminant analysis. All statistical analyses were performed with SPSS 26. A *p*-value <0.05 was considered statistically significant.

Results

Demographic characteristics of subjects

The demographic data of BA and non-BA patients are shown in **Table 1**. The age of BA group was younger that the non-BA group, but there was no difference in gender between the two groups. Significantly higher levels of TBA, TBIL, DBIL, IBIL, ALT, AST, GGT were detected in BA plasma, which indicated impaired hepatic functions in BA patients. Aβ42 and Aβ40 were not significantly different, but Aβ42/ Aβ40 ratio was significantly elevated in BA as compared to the non-BA group (**Figure 1**).

Positive correlation of plasma A β 42/A β 40 with hepatic functions

Next, Pearson correlation analysis was performed to determine if plasma $A\beta42$, $A\beta40$, and $A\beta42/A\beta40$ ratios correlated with hepatic functions in neonates. As shown in **Table 2**, plasma $A\beta42$ displayed a weak positive correlation with GGT, but no positive correlation with hepatic functions was detected for $A\beta40$. In contrast, Pearson correlation analysis revealed a statistically significant moderate positive correlation between $A\beta42/A\beta40$ and GGT, TBA, DBIL, and weak positive correlation between $A\beta42/A\beta40$ and GGT, TBA, DBIL, and weak positive correlation between $A\beta42/A\beta40$ and TBIL (**Table 2**). Among children with non-liver related diseases, no significant correlation was identified between plasma $A\beta42$, plasma $A\beta40$, and plasma $A\beta42/A\beta40$ ratio and age, which suggested that age factor did not have much effect on their plasma levels (**Supplementary Table S2**).

Plasma A β 42/A β 40 for the diagnosis of cholestasis

Positive correlation of plasma $A\beta 42/A\beta 40$ with hepatic function parameters prompted us to examine if plasma $A\beta$ peptides could be a biomarker for cholestasis. We performed ROC analysis to evaluate the efficacies of $A\beta 42$, $A\beta 40$, $A\beta 42/$

	BA $(N = 11)$	Non-liver $(N = 19)$	CC $(N = 5)$	<i>p</i> -Value ^a	<i>p</i> -Value ^b	<i>p</i> -Value ^c
Age (month)	1.7 ± 0.2	3.9 ± 0.5	19.4 ± 6.0	0.001	0.118	0.170
Gender (male)	5	10	5	0.5	0.106	0.047
Aβ42 (pg/ml)	7.7 (5.5, 9.3)	4.5 (2.3, 9.3)	4.7 (3.1, 7.3)	0.171	0.086	0.956
Aβ40 (pg/ml)	286.9 (201.4, 375.6)	303.7 (240.5, 413.7)	355.6 (198.6, 405.2)	0.657	0.582	0.944
Αβ42/Αβ40 (×100)	3.1 (1.6, 4.0)	1.7 (1.3, 2.3)	1.4 (1.4, 2.1)	0.029	0.110	0.774
ALT IU/L (8–71)	124.0 (76.0, 249.0)	27.0 (20.0, 31.0)	223.0 (13.5, 294.0)	0.003	0.394	0.188
AST IU/L (21–80)	173.0 (100.0, 225.0)	42.0 (34.0, 51.0)	58.0 (28.5, 208.0)	0.001	0.169	0.318
GGT IU/L (29-80)	300.0 (168.0, 1011.0)	21.0 (16.0, 34.0)	375.0 (155.5, 621.0)	< 0.001	0.893	0.001
TBA μmol/L (0.5–10)	122.2 (99.2, 188.3)	12.5 (5.5, 26.1)	34.0 (6.0, 105.7)	< 0.001	0.060	0.274
TBIL μmol/L (0–17.1)	148 (114.9, 183.4)	7.9 (6.8, 14.5)	13.5 (9.0, 179.2)	< 0.001	0.178	0.105
DBIL µmol/L (0–6.8)	91.8 (80, 111.1)	2.9 (2.2, 5.6)	7.0 (3.1, 78.1)	< 0.001	0.065	0.142
IBIL µmol/L (2–17)	47.7 (24.5, 56.2)	5.3 (3.2, 9.4)	9.6 (4.4, 101.1)	< 0.001	0.222	0.161

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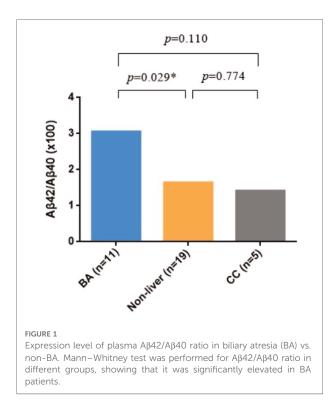
Values in parenthesis indicated normal range of hepatic function indicators. Levels of A β peptides and hepatic function indicators in patients were shown as Median (Q1, Q3). Age was shown as mean <u>+</u> SEM.

BA, biliary atresia; CC, choledochal cysts; ALT, alanine aminotransferase; AST, aspartate aminotransferase; GGT, γ-glutamyl transpeptidase; TBA, total bile acid; TBIL, total bilirubin; DBIL, direct bilirubin; IBIL, indirect bilirubin.

^aBA vs. non-liver.

^bBA vs. CC.

^cNon-liver vs. CC.



Aβ40, and hepatic function parameters in the diagnosis of cholestasis. Plasma Aβ42/Aβ40 had an acceptable predictive power for cholestasis [AUC = 0.746 (95% CI: 0.552–0.941), p < 0.05]; however, Aβ40 and Aβ42 did not show a significant predictive power for cholestasis (**Table 3**). Plasma ALT, AST, GGT, TBA, TBIL, DBIL, and IBIL had variable predictive

power for cholestasis [AUC ranging from 0.746 to 0.962; (95% CI: 0.604–1.000), p < 0.05]. ROC analysis for Aβ42/Aβ40 alone and in combinations with different hepatic function parameters showed that the diagnostic powers of Aβ42/Aβ40 and each of the hepatic function parameters for cholestasis were markedly improved if combined (Table 3 and Figure 2).

Plasma A β 42/A β 40 together with hepatic function parameters for the diagnosis of biliary atresia

Neonatal pathological cholestasis can be caused by a number of disorders such as BA or non-BA cholestasis like CC, viral infections like cytomegalovirus (CMV), and metabolic liver diseases or genetic disorders like Alagille syndrome. Since BA was the most severe cholestatic disease, we next sought to test if AB42/AB40 and hepatic function parameters either alone or in combinations could well discriminate BA cholestasis from non-BA cholestasis by performing ROC analysis on the 11 BA patients and the two CC patients with cholestasis (patient nos. 12 and 16; Supplementary Table S1). Though AB42/AB40 was found to be a good indicator for cholestasis, it did not display sufficient power in differentiating BA cholestasis from CC-cholestasis, neither did any other hepatic function parameters (Tables 4, 5). However, when A\u00e342/A\u00e340 combined with GGT, a liver enzyme elevated in patients with biliary tract obstruction, it improved the efficiency to prone to BA (AUC = 0.955, p =0.048). While combination of Aβ42/Aβ40 and GGT, and then

		Αβ42	Αβ40	Αβ42/Αβ40	ALT	AST	GGT	TBA	TBIL	DBIL	IBIL
Αβ42	Pearson correlation	1	0.544**	0.233	-0.001	0.111	0.349*	0.157	0.276	0.241	0.211
	Sig. (two-tailed)		0.001	0.178	0.998	0.524	0.040	0.384	0.109	0.163	0.224
	Ν	35	35	35	35	35	35	33	35	35	35
Αβ40	Pearson correlation	0.544**	1	-0.312	-0.172	-0.128	-0.236	-0.230	-0.117	-0.190	-0.004
	Sig. (two-tailed)	0.001		0.068	0.324	0.462	0.172	0.199	0.504	0.274	0.983
	Ν	35	35	35	35	35	35	33	35	35	35
Αβ42/Αβ40	Pearson correlation	0.233	-0.312	1	0.283	0.272	0.533**	0.475**	0.337*	0.480**	0.078
	Sig. (two-tailed)	0.178	0.068		0.099	0.114	0.001	0.005	0.048	0.004	0.657
	Ν	35	35	35	35	35	35	33	35	35	35
ALT	Pearson correlation	-0.001	-0.172	0.283	1	0.869**	0.439**	0.604**	0.309	0.566**	-0.052
	Sig. (two-tailed)	0.998	0.324	0.099		0.000	0.008	0.000	0.071	0.000	0.767
	Ν	35	35	35	35	35	35	33	35	35	35
AST	Pearson correlation	0.111	-0.128	0.272	0.869**	1	0.437**	0.846**	0.509**	0.789**	0.055
	Sig. (two-tailed)	0.524	0.462	0.114	0.000		0.009	0.000	0.002	0.000	0.755
	N	35	35	35	35	35	35	33	35	35	35
GGT	Pearson correlation	0.349*	-0.236	0.533**	0.439**	0.437**	1	0.477**	0.690**	0.641**	0.491**
	Sig. (two-tailed)	0.040	0.172	0.001	0.008	0.009		0.005	0.000	0.000	0.003
	Ν	35	35	35	35	35	35	33	35	35	35
TBA	Pearson correlation	0.157	-0.230	0.475**	0.604**	0.846**	0.477**	1	0.648**	0.926**	0.146
	Sig. (two-tailed)	0.384	0.199	0.005	0.000	0.000	0.005		0.000	0.000	0.417
	N	33	33	33	33	33	33	33	33	33	33
TBIL	Pearson correlation	0.276	-0.117	0.337*	0.309	0.509**	0.690**	0.648**	1	0.812**	0.824**
	Sig. (two-tailed)	0.109	0.504	0.048	0.071	0.002	0.000	0.000		0.000	0.000
	N	35	35	35	35	35	35	33	35	35	35
DBIL	Pearson correlation	0.241	-0.190	0.480**	0.566**	0.789**	0.641**	0.926**	0.812**	1	0.339**
	Sig. (two-tailed)	0.163	0.274	0.004	0.000	0.000	0.000	0.000	0.000		0.046
	N	35	35	35	35	35	35	33	35	35	35
IBIL	Pearson correlation	0.211	-0.004	0.078	-0.052	0.055	0.491**	0.146	0.824**	0.339**	1
	Sig. (two-tailed)	0.224	0.983	0.657	0.767	0.755	0.003	0.417	0.000	0.046	
	N	35	35	35	35	35	35	33	35	35	35

TABLE 2 Pearson correlation analysis of variables.

ALT, alanine aminotransferase; AST, aspartate aminotransferase; GGT, γ-glutamyl transpeptidase; TBA, total bile acid; TBIL, total bilirubin; DBIL, direct bilirubin; IBIL, indirect bilirubin.

*Correlation is significant at the 0.05 level (two-tailed)

**Correlation is significant at the 0.01 level (two-tailed).

with any one liver function parameters could further enhance the power to inform cholestasis patients to be BA or non-BA (AUC = 1.000, p < 0.05) (**Tables 4, 6**, and **Supplementary Table S3**), which had higher power than the same combinations but without A β 42/A β 40 (**Table 6**), indicating the role of A β 42/A β 40 together with other parameters could be used as an adjuvant indicator to enhance the accuracy in distinguishing BA from non-BA.

Discussion

Neonatal jaundice is common, and up to two-thirds of all newborns develop this problem within the first 2 weeks of life (23). While most cases are physiological jaundice that are mild, transient, and self-limiting, more severe cases of pathological jaundice are not uncommon. Diseases including BA and CC are common pathological causes of neonatal cholestasis requiring timely intervention (24). However, BA is challenging to diagnose because many of the clinical and imaging features of this condition overlap with those of other causes of neonatal cholestasis. In this study, we found that plasma A β 42/A β 40 correlated with hepatic function parameters, and combinations of plasma A β 42/A β 40 with GGT, TBA, TBIL, or DBIL displayed sensitivity and specificity for the diagnosis of BA.

Plasma levels of liver enzymes (ALT, AST, and GGT) and various forms of bilirubin (TBIL, DBIL, and IBIL) are hepatic function indicators, which are used to assist the diagnosis of cholestatic liver disease liver. In line with the use of these hepatic function indicators in the diagnosis of cholestasis, we have observed that plasma levels of ALT, AST, GGT, TBIL, DBIL, and IBIL have predictive power for cholestasis. Pearson correlation analysis revealed positive correlations of plasma Aβ42/Aβ40 with GGT, TBA, DBIL, and TBIL. In line with the positive correlation between Aβ42/Aβ40 and hepatic function indicators, plasma Aβ42/Aβ40 also has a predictive power for cholestasis [AUC = 0.746 (95% CI: 0.552–0.941), p < 0.05]. More importantly, combinations of Aβ42/Aβ40 and hepatic function indicators markedly improved the diagnostic

Test result variable(s)	Area	Std. error ^a	Asymptotic sig. ^b	Asymptotic 95% confidence interval		
				Lower bound	Upper bound	
ALT	0.788	0.094	0.007	0.604	0.972	
AST	0.833	0.084	0.002	0.669	0.997	
GGT	0.898	0.053	0.000	0.795	1.000	
TBA	0.928	0.051	0.000	0.828	1.000	
TBIL	0.902	0.054	0.000	0.795	1.000	
DBIL	0.962	0.037	0.000	0.889	1.000	
IBIL	0.856	0.068	0.001	0.723	0.989	
Αβ42	0.682	0.090	0.088	0.506	0.858	
Αβ40	0.439	0.110	0.570	0.224	0.655	
Αβ42/Αβ40	0.746	0.099	0.021	0.552	0.941	
$A\beta 42/A\beta 40 + GGT$	0.875	0.063	0.000	0.751	0.999	
$A\beta 42/A\beta 40 + TBA$	0.983	0.019	0.000	0.944	1.000	
$A\beta 42/A\beta 40 + TBIL$	0.939	0.039	0.000	0.863	1.000	
$A\beta 42/A\beta 40 + DBIL$	0.977	0.024	0.000	0.930	1.000	
$A\beta 42/A\beta 40 + GGT + TBA$	0.983	0.019	0.000	0.944	1.000	
$A\beta 42/A\beta 40 + GGT + TBIL$	0.939	0.039	0.000	0.863	1.000	
$A\beta 42/A\beta 40 + GGT + DBIL$	0.977	0.024	0.000	0.930	1.000	
$A\beta 42/A\beta 40 + TBA + TBIL$	0.978	0.023	0.000	0.933	1.000	
$A\beta 42/A\beta 40 + TBA + DBIL$	0.978	0.023	0.000	0.933	1.000	
$A\beta 42/A\beta 40 + TBIL + DBIL$	0.973	0.027	0.000	0.920	1.000	
$A\beta 42/A\beta 40+GGT+TBA+TBIL$	0.978	0.023	0.000	0.933	1.000	
$A\beta 42/A\beta 40 + GGT + TBA + DBIL$	0.978	0.023	0.000	0.933	1.000	
$A\beta 42/A\beta 40 + GGT + TBIL + DBIL$	0.973	0.027	0.000	0.920	1.000	
$A\beta 42/A\beta 40 + TBA + TBIL + DBIL$	0.978	0.023	0.000	0.933	1.000	
$A\beta42/A\beta40 + GGT + TBA + TBIL + DBIL$	0.983	0.019	0.000	0.944	1.000	

TABLE 3 Area under curve of Aβ42, Aβ40, Aβ42/Aβ40, and hepatic function parameters for the diagnosis of cholestasis.

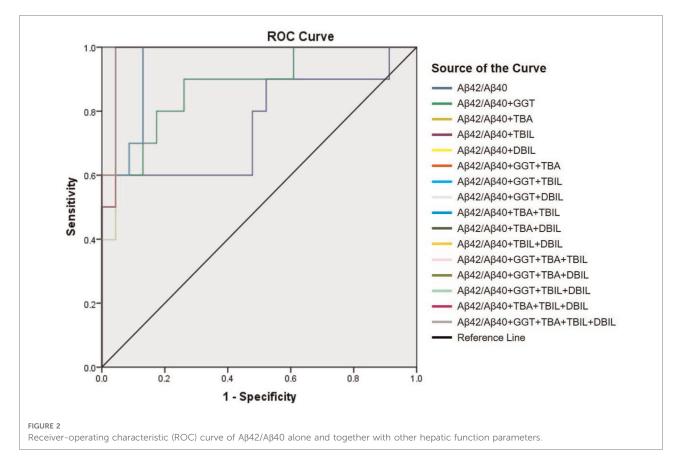
ALT, alanine aminotransferase; AST, aspartate aminotransferase; GGT, γ -glutamyl transpeptidase; TBA, total bile acid; TBIL, total bilirubin; DBIL, direct bilirubin; IBIL, indirect bilirubin.

^aUnder the nonparametric assumption

^bNull hypothesis: true area = 0.5.

accuracies of A β 42/A β 40 and each of the hepatic function parameters for cholestasis, suggesting that plasma A β 42/A β 40 can be an additional biomarker enhancing the accuracies of those common hepatic function indicators for the diagnosis of cholestatic liver disease. There is no current evidence to show that A β 42/A β 40 varies with ages. In our study, we also found no correlation between age and plasma A β 42, A β 40, and A β 42/A β 40, indicating that age may not be a determining factor in their plasma levels.

Since neonatal cholestasis has many different causes including BA, the most severe and poorest prognosis, and other non-BA cholestasis such as viral infection including CMV and herpes viruses; metabolic liver diseases or genetic disorders such as alpha-1-antitrypsin deficiency and Alagille syndrome (25). Cholestasis caused by BA or non-BA diseases is difficult to differentiate clinically. Currently, liver function test parameters may have an indication for the diagnosis of BA. For example, serum $DBIL \ge 1.0 \text{ mg/dl}$ had a better detection for BA with a sensitivity of 100% and specificity of 77.3% in infants aged from 3 to 60 days, but with a low positive predictive value (2.7%-5.4%) to distinguish BA from non-BA (26, 27). Hence it is important that new non-invasive biomarkers are discovered to allow early screening of BA from non-BA in jaundiced infants. Though AB42/AB40 was an indicator for liver dysfunction from our data, it was not sufficient on its own to distinguish BA from non-BA cholestasis. Similarly, the hepatic function parameters singly or collectively were insufficient for BA and non-BA differentiation. However, the combination of A\beta42/A\beta40 and GGT improved the efficiency of BA prediction compared to the use of either index alone. GGT, a liver enzyme elevated in patients with biliary tract obstruction, has been shown to have a contributory though not definitive role in the diagnosis of BA (16, 28). Addition of one of the other liver function



Test result variable (s)	Area	Std. error ^a	Asymptotic sig ^b	Asymptotic 95% confidence Interval		
				Lower bound	Upper bound	
GGT	0.364	0.151	0.554	0.068	0.659	
Αβ42/Αβ40	0.545	0.150	0.844	0.251	0.840	
$A\beta 42/A\beta 40 + GGT$	0.955	0.062	0.048	0.833	1.000	
$A\beta42/A\beta40+GGT+TBA$	1.000	0.000	0.032	1.000	1.000	
$A\beta42/A\beta40+GGT+TBIL$	1.000	0.000	0.030	1.000	1.000	
$A\beta42/A\beta40 + GGT + DBIL$	1.000	0.000	0.030	1.000	1.000	
$A\beta42/A\beta40+GGT+TBA+TBIL$	1.000	0.000	0.032	1.000	1.000	
$A\beta42/A\beta40+GGT+TBA+DBIL$	1.000	0.000	0.032	1.000	1.000	
$A\beta42/A\beta40+GGT+TBIL+DBIL$	1.000	0.000	0.030	1.000	1.000	
$A\beta42/A\beta40+GGT+TBA+TBIL+DBIL$	1.000	0.000	0.032	1.000	1.000	
ALT	0.636	0.221	0.554	0.204	1.000	
AST	0.682	0.242	0.430	0.208	1.000	
TBA	0.575	0.253	0.747	0.080	1.000	
TBIL	0.227	0.126	0.236	0.000	0.474	
DBIL	0.545	0.326	0.844	0.000	1.000	
IBIL	0.182	0.123	0.167	0.000	0.424	
Αβ42	0.682	0.242	0.430	0.208	1.000	
Αβ40	0.545	0.273	0.844	0.011	1.000	

(continued)

Test result variable (s)	Area	Std. error ^a	Asymptotic sig ^b	Asymptotic 95% confidence Interval		
				Lower bound	Upper bound	
Αβ42/Αβ40 + ΤΒΑ	0.750	0.159	0.283	0.439	1.000	
$A\beta 42/A\beta 40 + TBIL$	0.727	0.134	0.324	0.464	0.990	
$A\beta 42/A\beta 40 + DBIL$	0.727	0.214	0.324	0.309	1.000	
$A\beta42/A\beta40+TBA+TBIL$	0.800	0.134	0.197	0.537	1.000	
$A\beta42/A\beta40 + TBA + DBIL$	0.750	0.202	0.283	0.355	1.000	
$A\beta 42/A\beta 40 + TBIL + DBIL$	0.773	0.185	0.236	0.410	1.000	
$A\beta42/A\beta40+TBA+TBIL+DBIL$	0.800	0.170	0.197	0.466	1.000	

TABLE 4 Continued

ALT, alanine aminotransferase; AST, aspartate aminotransferase; GGT, γ-glutamyl transpeptidase; TBA, total bile acid; TBIL, total bilirubin; DBIL, direct bilirubin; IBIL, indirect bilirubin.

^aUnder the nonparametric assumption.

^bNull hypothesis: true area = 0.5.

TABLE 5 Sensitivity and specificity of A β 42/A β 40 and γ -glutamyl transpeptidase in combinations with any one hepatic function parameters in the diagnosis of biliary atresia vs. non-biliary atresia cholestasis.

Parameters	Cut-off value	Sensitivity (%)	Specificity (%)	Equation
$A\beta42/A\beta40+GGT+TBA$	-0.34583	100.00	100.00	22.549 + 35.326 * Ab42/Ab40 – 0.298 * GGT – 0.332 * TBA
$A\beta42/A\beta40+GGT+TBIL$	0.02371	100.00	100.00	$-526.079 + 287.021 * A\beta 42 / A\beta 40 - 2.250 * GGT + 1.555 * TBIL$
$A\beta42/A\beta40+GGT+DBIL$	-0.28387	100.00	100.00	83.795 + 98.348 *vA β42/Aβ40 – 0.805 * GGT – 1.383 * DBIL
$A\beta42/A\beta40+GGT+TBA+TBIL$	0.01040	100.00	100.00	$-48.789 + 25.050 * A\beta 42 / A\beta 40 - 0.396 * GGT - 0.453 * TBA + 0.974 * TBIL$
$A\beta42/A\beta40+GGT+TBA+DBIL$	0.06808	100.00	100.00	14.881 + 31.888 *vA β42/Aβ40 $-$ 0.267 * GGT $-$ 0.343 * TBA + 0.106 * DBIL
$A\beta42/A\beta40+GGT+TBIL+DBIL$	0.06224	100.00	100.00	$-86.646 + 38.587 * A\beta 42 / A\beta 40 - 0.585 * GGT + 1.631 * TBIL - 1.143 * DBIL$
Aβ42/Aβ40 + GGT + TBA + TBIL + DBIL	0.05898	100.00	100.00	$-43.153 + 24.039 * A\beta 42 / A\beta 40 - 0.378 * GGT - 0.403 * TBA + 0.934 * TBIL - 0.092 * DBIL$

GGT, y-glutamyl transpeptidase; TBA, total bile acid; TBIL, total bilirubin; DBIL, direct bilirubin; IBIL, indirect bilirubin.

TABLE 6 Area under the curve of γ -glutamyl transpeptidase and other hepatic function parameters with/without A β 42/A β 40 for the diagnosis of biliary atresia.

Test result variable(s)		Area	Std. error ^a	Asymptotic sig. ^b	Asymptotic 95% confidence interval		
					Lower bound	Upper bound	
GGT + TBA	Without Aβ42/Aβ40	0.600	0.239	0.667	0.132	1.000	
	With Aβ42/Aβ40	1.000	0.000	0.032	1.000	1.000	
GGT + TBIL	Without Aβ42/Aβ40	0.636	0.151	0.554	0.341	0.932	
	With Aβ42/Aβ40	1.000	0.000	0.030	1.000	1.000	
GGT + DBIL	Without Aβ42/Aβ40	0.545	0.326	0.844	0.000	1.000	
	With Aβ42/Aβ40	1.000	0.000	0.030	1.000	1.000	
GGT + TBA + TBIL	Without Aβ42/Aβ40	0.750	0.136	0.283	0.483	1.000	
	With Aβ42/Aβ40	1.000	0.000	0.032	1.000	1.000	
GGT + TBA + DBIL	Without Aβ42/Aβ40	0.550	0.323	0.830	0.000	1.000	
	With Aβ42/Aβ40	1.000	0.000	0.032	1.000	1.000	
GGT + TBIL + DBIL	Without Aβ42/Aβ40	0.636	0.221	0.554	0.204	1.000	
	With Aβ42/Aβ40	1.000	0.000	0.030	1.000	1.000	
GGT + TBA + TBIL + DBIL	Without Aβ42/Aβ40	0.700	0.232	0.390	0.245	1.000	
	With Aβ42/Aβ40	1.000	0.000	0.032	1.000	1.000	

GGT, y-glutamyl transpeptidase; TBA, total bile acid; TBIL, total bilirubin; DBIL, direct bilirubin; IBIL, indirect bilirubin.

^aUnder the nonparametric assumption.

^bNull hypothesis: true area = 0.5.

parameters to the combination of Aβ42/Aβ40 and GGT could further enhance the power to inform the cause of cholestasis as BA or non-BA (AUC = 1.000, p < 0.05). Taken all the above, our preliminary findings suggested that plasma Aβ42/ Aβ40 could be a valuable adjuvant biomarker for BA diagnosis.

We acknowledge there are limitations in our study. As BA is a relatively rare disease, the sample size of our study is modest. Our plasma $A\beta 42/A\beta 40$ data are encouraging and corroborate with the tissue and organoids findings of our previous study (20). Nevertheless, the diagnostic accuracy and usefulness of $A\beta 42/A\beta 40$ as an adjuvant non-invasive biomarker in combination with other liver function parameters for BA needs to be further evaluated and validated in a separate larger patient cohort of BA, non-BA cholestatic liver diseases, and conditions unrelated to the liver in multiple centers before introduction into daily clinical practice.

In conclusion, by combining $A\beta 42/A\beta 40$ as an adjuvant biomarker with other liver function parameters, we improve the sensitivity and the specificity of BA diagnosis. As an early screening tool, this may allow the identification of jaundiced neonates who are more likely to be suffering from BA to undergo early surgical exploration and intraoperative cholangiography for BA confirmation and KPE, thus improving the surgical outcome.

Data availability statement

The original contributions presented in the study are included in the article/Supplementary Material, further inquiries can be directed to the corresponding authors.

Ethics statement

The studies involving human participants were reviewed and approved by Ethical committee of Shenzhen Children's Hospital. Written informed consent to participate in this study was provided by the participants' legal guardian/next of kin.

Author contributions

HL, YY, and VL: study conception and design. HL and YY: data collection. HL, YY, and VL: data analysis and

References

interpretation. HL, YY, VL, and PC: drafting of the manuscript. All authors contributed to the article and approved the submitted version.

Funding

Authors declare all the funding supports are for research only and have no role in the data procurement and interpretation, and in the article preparation and submission. This work was supported by the Shenzhen Science and Technology Innovation Commission (grant number JCYJ20210324134202007), Sanming Project of Medicine in Shenzhen (grant number SZSM201812055), and Theme-based Research Scheme (T12-712/21-R) RGC Hong Kong SAR Government, Hong Kong SAR, China.

Acknowledgements

Authors thank all the patients who have participated in this study.

Conflict of interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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Supplementary material

The Supplementary Material for this article can be found online at: https://www.frontiersin.org/articles/10.3389/fsurg. 2022.931637/full#supplementary-material.

^{1.} Chung PHY, Zheng S, Tam PKH. Biliary atresia: east versus west. Semin Pediatr Surg. (2020) 29(4):150950. doi: 10.1016/j.sempedsurg.2020. 150950

^{2.} Sanchez-Valle A, Kassira N, Varela VC, Radu SC, Paidas C, Kirby RS. Biliary atresia: epidemiology, genetics, clinical update, and public health perspective. *Adv Pediatr.* (2017) 64(1):285–305. doi: 10.1016/j.yapd.2017.03.012

3. Tam PKH, Chung PHY, St Peter SD, Gayer CP, Ford HR, Tam GCH, et al. Advances in paediatric gastroenterology. *Lancet.* (2017) 390(10099):1072–82. doi: 10.1016/S0140-6736(17)32284-5

4. Mack CL, Sokol RJ. Unraveling the pathogenesis and etiology of biliary atresia. *Pediatr Res.* (2005) 57(5 Pt 2):87r-94r. doi: 10.1203/01.PDR.0000159569. 57354.47

5. Kelay A, Davenport M. Long-term outlook in biliary atresia. Semin Pediatr Surg. (2017) 26(5):295–300. doi: 10.1053/j.sempedsurg.2017.09.003

6. Lee JY, Sullivan K, El Demellawy D, Nasr A. The value of preoperative liver biopsy in the diagnosis of extrahepatic biliary atresia: a systematic review and meta-analysis. *J Pediatr Surg.* (2016) 51(5):753–61. doi: 10.1016/j.jpedsurg.2016. 02.016

7. Alisi A, de Vito R, Monti L, Nobili V. Liver fibrosis in paediatric liver diseases. Best Pract Res Clin Gastroenterol. (2011) 25(2):259-68. doi: 10.1016/j.bpg.2011.02. 008

8. Golden J, Zagory JA, Fenlon M, Goodhue CJ, Xiao Y, Fu X, et al. Liquid chromatography-mass spectroscopy in the diagnosis of biliary atresia in children with hyperbilirubinemia. *J Surg Res.* (2018) 228:228–37. doi: 10.1016/j. jss.2018.03.021

9. Shen WJ, Chen G, Wang M, Zheng S. Liver fibrosis in biliary atresia. *World J Pediatr.* (2019) 15(2):117–23. doi: 10.1007/s12519-018-0203-1

10. Yang L, Zhou Y, Xu PP, Mourya R, Lei HY, Cao GQ, et al. Diagnostic accuracy of serum matrix metalloproteinase-7 for biliary atresia. *Hepatology*. (2018) 68(6):2069–77. doi: 10.1002/hep.30234

11. Wu JF, Jeng YM, Chen HL, Ni YH, Hsu HY, Chang MH. Quantification of serum matrix metallopeptide 7 levels may assist in the diagnosis and predict the outcome for patients with biliary atresia. *J Pediatr.* (2019) 208:30–7.e1. doi: 10. 1016/j.jpeds.2018.12.006

12. Jiang J, Wang J, Shen Z, Lu X, Chen G, Huang Y, et al. Serum MMP-7 in the diagnosis of biliary atresia. *Pediatrics*. (2019) 144(5):e20190902. doi: 10.1542/peds. 2019-0902

13. Rendon-Macias ME, Villasis-Keever MA, Castaneda-Mucino G, Sandoval-Mex AM. Improvement in accuracy of gamma-glutamyl transferase for differential diagnosis of biliary atresia by correlation with age. *Turk J Pediatr.* (2008) 50(3):253–9.

14. Lertudomphonwanit C, Mourya R, Fei L, Zhang Y, Gutta S, Yang L, et al. Large-scale proteomics identifies MMP-7 as a sentinel of epithelial injury and of biliary atresia. *Sci Transl Med.* (2017) 9(417):eaan8462. doi: 10.1126/scitranslmed.aan8462

15. Dong R, Jiang J, Zhang S, Shen Z, Chen G, Huang Y, et al. Development and validation of novel diagnostic models for biliary atresia in a large cohort of Chinese patients. *EBioMedicine*. (2018) 34:223–30. doi: 10.1016/j.ebiom.2018.07.025

16. Chen X, Dong R, Shen Z, Yan W, Zheng S. Value of gamma-glutamyl transpeptidase for diagnosis of biliary atresia by correlation with age. *J Pediatr Gastroenterol Nutr.* (2016) 63(3):370–3. doi: 10.1097/MPG.000000000001168

17. Chen GF, Xu TH, Yan Y, Zhou YR, Jiang Y, Melcher K, et al. Amyloid beta: structure, biology and structure-based therapeutic development. *Acta Pharmacol Sin.* (2017) 38(9):1205–35. doi: 10.1038/aps.2017.28

18. Wang MJ, Yi S, Han JY, Park SY, Jang JW, Chun IK, et al. Oligomeric forms of amyloid-beta protein in plasma as a potential blood-based biomarker for Alzheimer's disease. *Alzheimers Res Ther.* (2017) 9(1):98. doi: 10.1186/s13195-017-0324-0

19. Nabers A, Perna L, Lange J, Mons U, Schartner J, Guldenhaupt J, et al. Amyloid blood biomarker detects Alzheimer's disease. *EMBO Mol Med.* (2018) 10(5):e8763. doi: 10.15252/emmm.201708763

20. Counts SE, Ikonomovic MD, Mercado N, Vega IE, Mufson EJ. Biomarkers for the early detection and progression of Alzheimer's disease. *Neurotherapeutics*. (2017) 14(1):35–53. doi: 10.1007/s13311-016-0481-z

21. Babu RO, Lui VCH, Chen Y, Yiu RSW, Ye Y, Niu B, et al. Beta-amyloid deposition around hepatic bile ducts is a novel pathobiological and diagnostic feature of biliary atresia. *J Hepatol.* (2020) 73(6):1391–403. doi: 10.1016/j.jhep. 2020.06.012

22. Wang YR, Wang QH, Zhang T, Liu YH, Yao XQ, Zeng F, et al. Associations between hepatic functions and plasma amyloid-beta levels-implications for the capacity of liver in peripheral amyloid-Beta clearance. *Mol Neurobiol.* (2017) 54 (3):2338-44. doi: 10.1007/s12035-016-9826-1

23. Maisels MJ. What's in a name? Physiologic and pathologic jaundice: the conundrum of defining normal bilirubin levels in the newborn. *Pediatrics*. (2006) 118(2):805-7. doi: 10.1542/peds.2006-0675

24. Pan DH, Rivas Y. Jaundice: newborn to age 2 months. *Pediatr Rev.* (2017) 38 (11):499–510. doi: 10.1542/pir.2015-0132

25. Feldman AG, Sokol RJ. Neonatal cholestasis: emerging molecular diagnostics and potential novel therapeutics. *Nat Rev Gastroenterol Hepatol.* (2019) 16 (6):346–60. doi: 10.1038/s41575-019-0132-z

26. Harpavat S, Garcia-Prats JA, Anaya C, Brandt ML, Lupo PJ, Finegold MJ, et al. Diagnostic yield of newborn screening for biliary atresia using direct or conjugated bilirubin measurements. *JAMA*. (2020) 323(12):1141–50. doi: 10. 1001/jama.2020.0837

27. Liao FM, Chang KC, Wu JF, Chen HL, Ni YH, Chang MH. Direct bilirubin and risk of biliary atresia. *Pediatrics*. (2022) 149(6):e2021053073. doi: 10.1542/peds.2021-053073

28. Liu J, Dai S, Chen G, Sun S, Jiang J, Zheng S, et al. Diagnostic value and effectiveness of an artificial neural network in biliary atresia. *Front Pediatr.* (2020) 8:409. doi: 10.3389/fped.2020.00409