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CAFFEOYLQUINIC ACID ENHANCES PROLIFERATION OF OLIGODENDROCYTE PRECURSOR CELLS

Abstract

This report evaluates the protective effect of caffeoylquinic acid (CA) injury to oligodendrocyte precursor cells (OPCs) by promoting the formation of oligodendrocytes. Neonatal rat brain was used to isolate primary OPCs and non-lethal CoCl2 was used to induce hypoxic stress to inhibit the differentiation of OPCs. Differentiation of OPCs was estimated by survival assay and the expressions of myelin-basic-protein (MBP). Moreover, the effect of CA on the Akt signanling pathway was also estimated in the presence and absence of LY294002 (PI3K/Akt inhibitor) and adrenomedullin (AM) receptor antagonist (AM22-52) by using western blot assay. It was observed that CA enhances the differentiation OPCs in CoCl2 induced hypoxic stress condition. However treatment with CA in presence of LY294002 and AM22-52 was not able to enhance the differentiation of OPCs. Moreover treatment with CA significantly enhances the phosphorylation of Akt and presence of LY294002 and AM22-52 inhibits it. This report concludes that CA effectively attenuates the injury of white matter (OPCs) by enhancing the differentiation of OPCs. It enhances the formation of oligodendrocytes by activating AM receptor and thereby accelerates the regeneration of neuron in pathological condition.

Keywords

Caffeoylguinic acid
Oligodendrocyte precursor cells
Myelin-basic-protein
Adrenomedullin

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Introduction

White matter of the central nervous system is made up of oligodendrocytes and these cells are responsible for the formation of myelin around axons [1]. It is very well documented that for neuronal homeostasis and effective conduction of nerve impulses, myelin sheath is required [2]. Oligodendrocytes mature during late embryonic development as oligodendrocyte precursor cells migrate and proliferate into oligodendrocytes [3, 4]. Moreover studies suggest that approximately 8% of adult brain cells as OPCs remain immature. These OPCs are responsible for the repair of neuronal injury in white matter by promoting myelinated neurons [5]. Literature reveals that degeneration of neurons and loss of myelin sheath from the neurons occurs in demyelinating diseases that causes permanent disability [6]. In the early stage of demyelinating disease proliferation of OPCs occur due to endogenous repair mechanism, but if disease progresses to an advanced stage, this neuron repair gets altered [7]. Therefore, for the treatment of demyelinating disease, promotion of proliferation of OPCs is a good target for the development of drug.

However, recently natural compounds have gained attention in the management of neuronal injury. Caffeoylquinic acid was isolated from the Chinese herb Aster tataricus, which belongs to the family Asteraceae [8]. It has been reported that caffeoylquinic acid is an ester of quinic and caffeic acid isolated from natural sources [9]. It has been reported have anti-oxidant, anti-inflammatory, to anti-cancer activities [10-12]. and 3,5-O-caffeoylquinic acid possesses strong anti-inflammatory activity, reducing the synthesis of cytokines, such as TNFα and IL-1β, and anti-oxidant properties [13]. Moreover, it reduces the generation of reactive oxygen species and, thus, reduces oxidative stress. This report estimates the effect of CA against demyelinating diseases.

Material and methods

Animals

Female Wistar rats were kept individually under observation for delivery in the cage. All the rats were housed under a controlled condition specified as per guidelines. All the experiments used in the given study are approved by animal ethical committee of the Tongji Medical College, Huazhong University of Science and Technology, Wuhan, China.

Cell Culture

A previously suggested method by Wang et al., 2011 was used for the proliferation of oligodendrocyte precursor cells. Cells were isolated from the cerebral cortex of pups by placing them in streptomycin (50 μ g/ml) and penicillin (50 μ g/ml) containing ice-cold DMEM/ F12 medium. Cell suspension was prepared and filtered with cell strainer (70 μ m). Later cell were centrifuged for the period of 10 min at 10000 rpm and collected cells were re suspended in DMEM/

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F12 medium and incubated it at 37 °C under controlled humid atmosphere for the duration of eight days. After every alternate day old medium was replaced with fresh medium. Later samples were kept on an orbital shaker to isolate the OPCs at 37 °C for the period of 60 min and the medium was replaced with fresh medium to remove the macrophages and microglial cells. The flask was again spun at 220 rpm for a period of 18 h by adding fresh DMEM/F12 medium (15 ml). Collected samples of cell suspension were kept in a Petri dish and incubated at 37 °C for 30 min. The cell suspension was then transferred to a 50 ml tube by passing it from the sieve of 15 μ m size and later centrifuged it for 10 min at 1000 rpm to separate the cells. Medium of DMEM/F12 that contains FGFb (20 ng/ml), PDGF-BB (20 ng/ ml) and 2 % B27 was used to re suspend the cells and thereafter placed it into 25 cm² flasks at a quantity of 10,000 cells/cm². Collected purified OPCs were differentiated.

CoCl, treatment

Cobalt chloride (CoCl2) was incubated with cells at a non-lethal concentration for the production of hypoxia. Differential media in the presence and absence of $CoCl_2$ (1 μ M) was used to culture the cells for 7 days. On 0, 3rd and 5th day OPCs culture media was replaced with fresh CoCl₂ containing media. Western blot assay, immunohistochemistry and survival assay was performed after the start of differentiation in OPCs i.e. on 7th day. Cells were separated into four groups such as control which do not receive the CoCl₂ CoCl₂ group which receives CoCl₂ (1 μ m) for the duration of 7 days, CoCl₂ +CA which receives both CoCl₂ (1 μ m) and CA (10 nm) for the duration of 7 days.

Survival assay

WST reduction assay was used for the estimation of proliferation of cell using Cell Counting Kit-8. All the cells were incubated at 37 °C for the duration of 60 min with WST solution (10 %). microplate reader was used for the estimation of absorbance of culture media at 630 nm for reference and 450 nm test wavelength.

Immunocytochemistry

Ice cold PBS solution was used for the washing of cells and thereafter with paraformaldehyde

(4%) for the duration of 15 min. Triton X-100 (1%) contained PBS was used to wash the cells thrice and then incubated for the duration of 60 min. primary antibodies against MBP was incubated with cells for the overnight at 4 °C. Afterwards cells with secondary antibodies were incubated at room temperature for the duration of 60 min after washing with PBS solution. All the cells were stained with DAPI.

Western blot assay

Cell lyses buffer was used to collect the cell after washing them with PBS buffer. PBS was added in the sample in an equal amount so that concentration of estimated protein remains the same. SDS (91%) and 2-mercaptoethanol (9%) containing buffer was added in an equal amount of sample and mix it. Later 4-20% Tris-glycine gels was used to load the samples after heating them for the duration of 5 min at 95 °C. Tris buffered saline solution used to block the nitrocellulose membranes to which samples were transferred. Later Akt antibody phosphorylated Akt (Ser473) antibody and myelin basic protein (MBP) antibody was incubated with membrane at 4 °C for the duration of overnight. Thereafter visualize it under the chemiluminescence.

Statistical analysis

Values of the given data represented as mean \pm SD (n=6) and the data of this study statistically analyzed by one-way ANOVA (Dunnett post hoc test). Level of significance was considered as p < 0.05.

Result

Effect of Caffeoylquinic acid on the proliferation and survival of OPCs

Effect of CA on the differentiation and proliferation of OPCs in CoCl₂ induced hypoxia caused injury in neonatal rats was shown in Fig.1. Expressions of MBP are related to differentiation of OPCs. It was observed that CoCl₂ group reduces the expressions of MBP and thereby decreases the differentiation of OPCs. However treatment with CA enhances the expressions of MBP as shown in Fig. 1 A. Fig 1B-C shows the effect of CA on the maturation of oligodendrocyte in hypoxia induced stress

condition by western blot study. Result suggested that maturation of oligodendrocyte was promoted in the CA treated group than $CoCl_2$ group. Moreover result of WST assay shows that both $CoCl_2$ and CA didn't reduces the count of OPCs in the culture medium.

Effect of Caffeoylquinic acid on signaling of Akt pathway

Effect of Caffeoylquinic acid on Akt signalling pathway in OPCs was shown in Fig. 2. It was observed that treatment with CA significantly enhances the level of phosphorylation of Akt in first 30 min and after 24 h it backs to normal in OPCs as shown in Fig. 2A-B. Moreover result suggested that treatment with AM ₂₂₋₅₂ and LY294002 inhibit the level of phosphorylation of Akt that was induced by CA as shown in Fig. 2C-D.

Effect of Caffeoylquinic acid on differentiation and survival of OPCs in presence AM ₂₂₋₅₂ and LY294002

Effect of Caffeoylquinic acid on differentiation and survival of OPCs in presence AM $_{22-52}$ and LY294002 was shown in Fig. 3. It was observed that treatment with CA not able to enhance the expressions of MBP in presence of AM $_{22-52}$ and LY294002 than CA alone. It means in presence of AM $_{22-52}$ and LY294002, CA is not able to differentiate the OPCs in hypoxic condition.

Discussion

This study evaluates the protective effect of CA against CoCl₂ induced hypoxia causing injury in OPCs. The effect of CA was evaluated by WST assay and MBP expressions in OPCs. Moreover a possible mechanism was postulated by evaluating its effect on Akt signaling.

Adrenomedullin receptors are widely distributed in the brain and AM is reported to act as a neurohormone and neurotransmitter [14]. Literature has also suggested that neuronal injury can be protected by activating the AM receptor, and thereby effectively aiding in the management of certain CNS disorders [15, 16]. In addition, several reports reveal that neurogenesis can be achieved by activating the AM receptor and it also helps in maturation of OPCs [17]. CA is a natural product, reported

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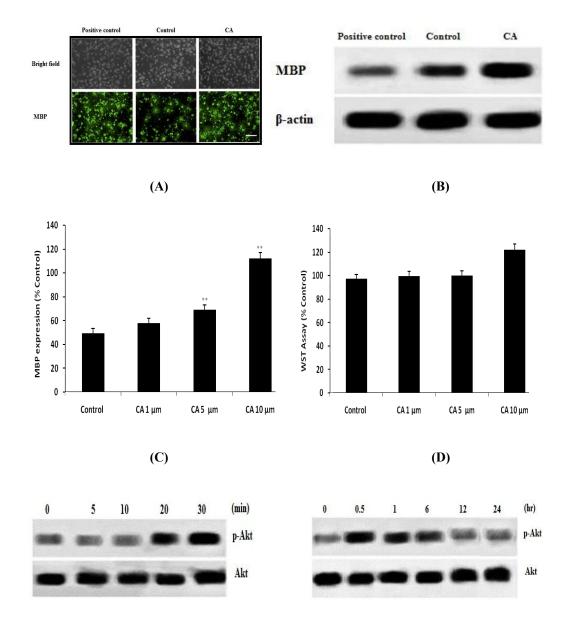


Fig. 1. Effect of Caffeoylquinic acid on the proliferation and survival of OPCs in CoCl₂ induced hypoxia. A. DAPI staining (Immunocytochemestry); B-C. Western blot for the expressions of MBP; D. WST assay for the assessment of cell death Values are expressed as Mean±SD (n=6) **p<0.01 Vs Control

to possess anti-oxidant, anti-inflammatory, and anti-cancer activities by reducing the generation of reactive oxygen species, TNF- α and IL [10-12]. Injury caused by hypoxia results in the activation of these factors.

The results of our study suggest that CA enhances the expression of MBP and survival of OPCs. Expression of MBP enhancement is related to the differentiation of OPCs in to oligodendrocytes [18]. Results suggest that the expressions of MBP in hypoxia or in an injury condition become reduced, and several drugs protect the white matter injury by promoting the expressions of MBP [19]. It was also observed that CA enhances the phosphorylation of Akt. However it will not become enhanced in the presence of AM receptor antagonist and PI3K inhibitor. Furthermore, similar results were observed in the expressions of MBP and the survival of OPCs in presence of them.

Conclusion

This report concludes that CA effectively attenuates the injury of white matter (OPCs) by enhancing the differentiation of OPCs. It enhances the formation of oligodendrocytes by activating AM receptor and thereby accelerates the regeneration of neurons in pathological conditions.

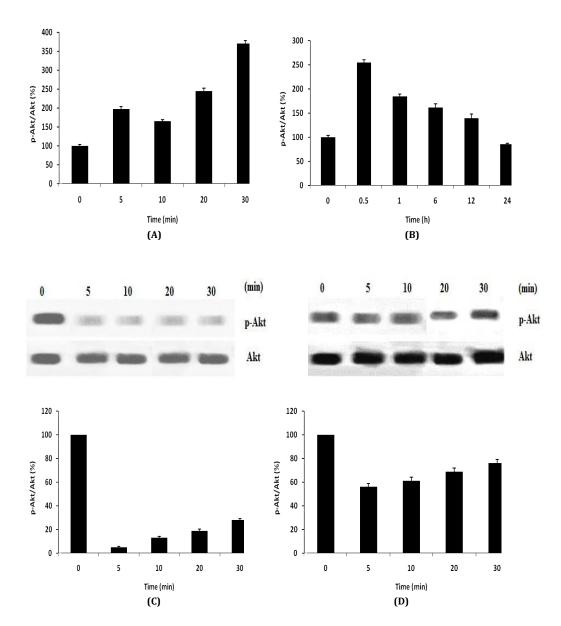


Fig. 2. Effect of Caffeoylquinic acid on Akt signalling pathway in OPCs. A-B, Effect of CA on the level of phosphorylation of Akt; C, Effect of CA on the level of phosphorylation of Akt in presence of LY294002 (PI3K inhibitor); D, Effect of CA on the level of phosphorylation of Akt in presence of AM receptor antagonist. Values are expressed as Mean±SD (n=6)

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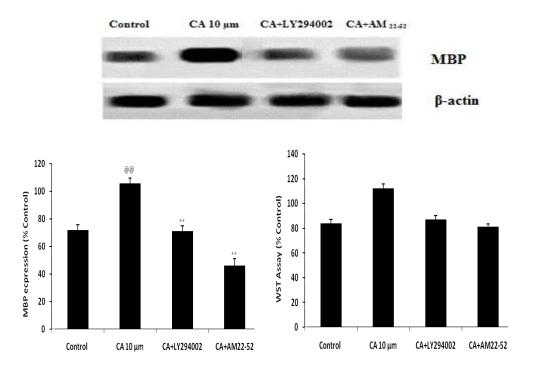


Fig. 3. Effect of Caffeoylquinic acid on differentiation and survival of OPCs in presence AM ₂₂₋₅₂ and LY294002 Values are expressed as Mean±SD (n=6) ^{@®}p<0.01 Vs Control; **p<0.01 Vs CA 10µm

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