



# Draft Genome Sequence of *Salmonella enterica* subsp. *enterica* Serovar Typhimurium Q1

Inga Eichhorn, Karsten Tedin, Marcus Fulde

Institute of Microbiology and Epizootics, Centre for Infection Medicine, Freie Universität Berlin, Berlin, Germany

**ABSTRACT** Here, we report the draft genome sequence of *Salmonella enterica* subsp. *enterica* serovar Typhimurium strain Q1. The draft genome contains 4,793,493 bp in 149 contigs.

The genus *Salmonella* comprises a heterogeneous group of enteric pathogenic bacteria that are considered to belong to either a specialist or a generalist class. The specialists are strongly host adapted and confer a pathology that goes beyond self-limiting gastrointestinal infections, leading to abortive and/or systemic clinical pathologies. Typical representatives are the human-specific *S. enterica* subsp. *enterica* serovars Typhi (the causative agent of typhoid fever) and Paratyphi, as well as bovine-adapted *S. enterica* subsp. *enterica* serovar Dublin and pig-restricted *S. enterica* subsp. *enterica* serovars Choleraesuis and Parasuis. In contrast, nontyphoidal *S. enterica* subsp. *enterica* (NTS) serovars, such as *S. Typhimurium* and *S. Enteritidis*, are routinely isolated from a variety of different warm- and cold-blooded host species (1, 2). Infections with NTS serovars generally lead to local and self-limiting gastrointestinal infections in immunocompetent patients, as well as to typhoid-fever-like symptoms in immunocompromised individuals, neonates, and infants (1, 3–5). Thus, due to its zoonotic potential and exceptional role as a foodborne pathogen, NTS serovars are a paradigm for the (<https://www.cdc.gov/onehealth/>).

Despite the large number of *Salmonella* serovars, clinical pictures, and disease severity, the pathogenic mechanisms are very similar and depend in large part on virulence factors encoded within two large genomic islands, termed *Salmonella* pathogenicity islands 1 and 2 (6–8). It is widely accepted that horizontal gene transfer (HGT) is the main driver of *Salmonella* pathogenicity evolution, bacterial fitness, and host adaptation and that lysogenic conversion of temperate phages has had a significant contribution. The most prominent examples of lysogenic phages in the genus *Salmonella* are the P2-like phages SopEΦ, Fels-1, and Fels-2 and the lambdoid prophages Gifsy-1, Gifsy-2, and Gifsy-3 (9–11).

Here, we provide the draft genome sequence of *S. Typhimurium* serovar Q1. Serologically, Q1 is a strain of *S. Typhimurium*, which was isolated from the feces of a human patient suffering from food poisoning and later cured of endogenous prophage (12). Q1 was believed to be phage and plasmid free (13), rendering it an excellent candidate strain for studies of HGT *in vitro* and *in vivo*. However, an uncharacterized, inducible bacteriophage infective for *S. enterica* subsp. *enterica* serovar Gallinarum was later observed (14). Using PHASTER, a Web-based online tool for identifying phage and phage-like sequences in bacterial genomes, we identified Gifsy-2 in the genome of *S. Typhimurium* Q1 (15, 16).

The 300 bp paired-end reads were generated using the Illumina MiSeq platform. The reads were *de novo* assembled into contigs with a minimum size of 200 bp using MIRA version 4.0 (17). A total of 149 contigs were generated ranging from 248 bp to 541,223 bp and resulted in a total genome size of 4,793,493 bp. The cumulative G+C

**Received** 13 September 2017 **Accepted** 18 September 2017 **Published** 19 October 2017

**Citation** Eichhorn I, Tedin K, Fulde M. 2017. Draft genome sequence of *Salmonella enterica* subsp. *enterica* serovar Typhimurium Q1. *Genome Announc* 5:e01151-17. <https://doi.org/10.1128/genomeA.01151-17>.

**Copyright** © 2017 Eichhorn et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Karsten Tedin, [karsten.tedin@fu-berlin.de](mailto:karsten.tedin@fu-berlin.de), or Marcus Fulde, [marcus.fulde@fu-berlin.de](mailto:marcus.fulde@fu-berlin.de).

content of the genome assembly was 52.2%. Gene annotation was performed using the RAST annotation server (18), which predicted 4,667 coding DNA sequences, 91 tRNAs, 42 rRNAs, and 1 transfer-messenger RNA in the draft genome. Strain Q1 was assigned to sequence type 19 using the multilocus sequence type service for total-genome-sequenced bacteria from the Center for Genomic Epidemiology (19). Using ResFinder (20) and PlasmidFinder (21), neither resistance genes nor plasmids were identified. Consistent with this observation, no phenotypic resistance against 11 different antibiotics was noted using the bioMérieux Vitek 2 system.

**Accession number(s).** This whole-genome shotgun project has been deposited at DDBJ/ENA/GenBank under the accession number [NNSK00000000](https://doi.org/10.1093/nar/gkw387). The version described in this paper is the first version, NNSK01000000.

## ACKNOWLEDGMENTS

We are grateful to Peter Schwerk (Institute of Microbiology and Epizootics, Freie Universität Berlin) for excellent technical support and the *Salmonella* Genetic Stock Center for providing the *S. Typhimurium* Q1 strain.

M.F. received support from the Freie Universität Berlin within the Excellence Initiative of the German Research Foundation.

## REFERENCES

- Bäumler A, Fang FC. 2013. Host specificity of bacterial pathogens. *Cold Spring Harb Perspect Med* 3:a010041. <https://doi.org/10.1101/cshperspect.a010041>.
- Whitley H, Gardner MG, Ross K. 2017. A review of *Salmonella* and squamates (lizards, snakes and amphisbians): implications for public health. *Pathogens* 6:3. <https://doi.org/10.3390/pathogens6030038>.
- Wotzka SY, Nguyen BD, Hardt WD. 2017. *Salmonella* Typhimurium diarrhea reveals basic principles of enteropathogen infection and disease-promoted DNA exchange. *Cell Host Microbe* 21:443–454. <https://doi.org/10.1016/j.chom.2017.03.009>.
- Keddy KH, Sooka A, Musekiwa A, Smith AM, Ismail H, Tau NP, Crowther-Gibson P, Angulo FJ, Klugman KP. 2015. Clinical and microbiological features of *Salmonella* meningitis in a South African population, 2003–2013. *Clin Infect Dis* 61(suppl 4):S272–S282. <https://doi.org/10.1093/cid/civ685>.
- Zhang K, Dupont A, Torow N, Gohde F, Leschner S, Lienenklaus S, Weiss S, Brinkmann MM, Kühnel M, Hensel M, Fulde M, Hornef MW. 2014. Age-dependent enterocyte invasion and microcolony formation by *Salmonella*. *PLoS Pathog* 10:e1004385. <https://doi.org/10.1371/journal.ppat.1004385>.
- Hansen-Wester I, Hensel M. 2001. *Salmonella* pathogenicity islands encoding type III secretion systems. *Microbes Infect* 3:549–559. [https://doi.org/10.1016/S1286-4579\(01\)01411-3](https://doi.org/10.1016/S1286-4579(01)01411-3).
- Jennings E, Thurston TLM, Holden DW. 2017. *Salmonella* SPI-2 type III secretion system effectors: molecular mechanisms and physiological consequences. *Cell Host Microbe* 22:217–231. <https://doi.org/10.1016/j.chom.2017.07.009>.
- Que F, Wu S, Huang R. 2013. *Salmonella* pathogenicity island 1 (SPI-1) at work. *Curr Microbiol* 66:582–587. <https://doi.org/10.1007/s00284-013-0307-8>.
- Diard M, Bakkeren E, Cornuault JK, Moor K, Hausmann A, Sellin ME, Loverdo C, Aertsen A, Ackermann M, De Paep M, Slack E, Hardt WD. 2017. Inflammation boosts bacteriophage transfer between *Salmonella*. *Science* 355:1211–1215. <https://doi.org/10.1126/science.aaf8451>.
- Brüssow H, Canchaya C, Hardt WD. 2004. Phages and the evolution of bacterial pathogens: from genomic rearrangements to lysogenic conversion. *Microbiol Mol Biol Rev* 68:560–602. <https://doi.org/10.1128/MMBR.68.3.560-602.2004>.
- Diard M, Hardt WD. 2017. Evolution of bacterial virulence. *FEMS Microbiol Rev* 22:fux023. <https://doi.org/10.1093/femsre/fux023>.
- Boyd JSK. 1956. Immunity of lysogenic bacteria. *Nature* 178:141. <https://doi.org/10.1038/178141a0>.
- Boyd JSK, Bidwell DE. 1957. The type A phages of *Salmonella typhimurium*: identification by a standardized cross-immunity test. *J Gen Microbiol* 16:217–228. <https://doi.org/10.1099/00221287-16-1-217>.
- Boyd JSK, Bidwell DE. 1959. The Q1 (A) strains of *Salmonella typhimurium*: induction phenomena. *J Gen Microbiol* 21:635–651. <https://doi.org/10.1099/00221287-21-3-635>.
- Arndt D, Grant J, Marcu A, Sajed T, Pon A, Liang Y, Wishart DS. 2016. PHASTER: a better, faster version of the PHAST phage search tool. *Nucleic Acids Res* 44:W16–W21. <https://doi.org/10.1093/nar/gkw387>.
- Zhou Y, Liang Y, Lynch KH, Dennis JJ, Wishart DS. 2011. PHAST: a fast phage search tool. *Nucleic Acids Res* 39:W347–W352. <https://doi.org/10.1093/nar/gkr485>.
- Chevreur B, Wetter T, Suhai S. 1999. Genome sequence assembly using trace signals and additional sequence information, p 45–56. *In* Computer science and biology: proceedings of the German Conference on Bioinformatics (GCB) '99. GCB, Hannover, Germany.
- Overbeek R, Olson R, Pusch GD, Olsen GJ, Davis JJ, Disz T, Edwards RA, Gerdes S, Parrello B, Shukla M, Vonstein V, Wattam AR, Xia F, Stevens R. 2014. The SEED and the Rapid Annotation of microbial genomes using Subsystems Technology (RAST). *Nucleic Acids Res* 42:D206–D214. <https://doi.org/10.1093/nar/gkt1226>.
- Larsen MV, Cosentino S, Rasmussen S, Friis C, Hasman H, Marvig RL, Jelsbak L, Sicheritz-Pontén T, Ussery DW, Aarestrup FM, Lund O. 2012. Multilocus sequence typing of total-genome-sequenced bacteria. *J Clin Microbiol* 50:1355–1361. <https://doi.org/10.1128/JCM.06094-11>.
- Zankari E, Hasman H, Cosentino S, Vestergaard M, Rasmussen S, Lund O, Aarestrup FM, Larsen MV. 2012. Identification of acquired antimicrobial resistance genes. *J Antimicrob Chemother* 67:2640–2644. <https://doi.org/10.1093/jac/dks261>.
- Carattoli A, Zankari E, García-Fernández A, Voldby Larsen M, Lund O, Villa L, Møller Aarestrup F, Hasman H. 2014. *In silico* detection and typing of plasmids using PlasmidFinder and plasmid multilocus sequence typing. *Antimicrob Agents Chemother* 58:3895–3903. <https://doi.org/10.1128/AAC.02412-14>.