RESEARCH ARTICLE

Leukocyte cell population data from the blood cell analyzer as a predictive marker for severity of acute pancreatitis

Yihui Wang MD¹ | Zhihong Xu MD, PhD¹ | Yuhua Zhou MD, PhD¹ | Mengqi Xie MD¹ | Xing Qi MD¹ | Zhiwei Xu MD² | Qi Cai MD³ | Huiqiu Sheng MD¹ | Erzhen Chen MD, PhD¹ | Bing Zhao MD, PhD¹ | Enqiang Mao MD, PhD¹

¹Department of Emergency, Ruijin Hospital, Shanghai Jiao Tong University School of Medicine, Shanghai, China

²Department of General Surgery, Pancreatic Disease Center, Ruijin Hospital, Shanghai Jiao Tong University School of Medicine, Shanghai, China

³Department of Laboratory Medicine, Ruijin Hospital, Shanghai Jiao Tong University School of Medicine, Shanghai, China

Correspondence

Enqiang Mao and Bing Zhao, Department of Emergency, Ruijin Hospital, Shanghai Jiao Tong University School of Medicine, 197 Ruijin 2nd Road, Huangpu District, Shanghai, China. Emails: maoeq@yeah.net; zhaobing124@163.com

Funding information

National Natural Science Foundation Youth Project of China, Grant/Award Number: 81601665; National Natural Science Foundation of China, Grant/ Award Number: 81600501

Abstract

Revised: 11 May 2021

Background: The prediction for severe acute pancreatitis (SAP) is the key to give timely targeted treatment. Leukocyte cell population data (CPD) have been widely applied in early prediction and diagnosis of many diseases, but their predictive ability for SAP remains unexplored. We aim to testify whether CPD could be an indicator of AP severity in the early stage of the disease.

Methods: The prospective observational study was conducted in the emergency department ward of a territory hospital in Shanghai. The enrolled AP patients should meet 2012 Atlanta guideline.

Results: Totally, 103 AP patients and 62 healthy controls were enrolled and patients were classified into mild AP (*n* = 30), moderate SAP (*n* = 42), and SAP (*n* = 31). Forty-two CPD parameters were examined in first 3 days of admission. Four CPD parameters were highest in SAP on admission and were constantly different among 3 groups during first 3 days of hospital stay. Eighteen CPD parameters were found correlated with the occurrence of SAP. Stepwise multivariate logistic regression analysis identified a scoring system of 4 parameters (SD_LALS_NE, MN_LALS_LY, SD_LMALS_MO, and SD_AL2_MO) with a sensitivity of 96.8%, specificity of 65.3%, and AUC of 0.87 for diagnostic accuracy on early identification of SAP. AUC of this scoring system was comparable with MCTSI, SOFA, APACHE II, MMS, BISAP, or biomarkers as CRP, PCT, and WBC in prediction of SAP and ICU transfer or death.

Conclusions: Several leukocyte CPD parameters have been identified different among MAP, MSAP, and SAP. They might be ultimately incorporated into a predictive system marker for severity of AP.

KEYWORDS

acute pancreatitis, cell population data, prediction, scoring system, severity

This is an open access article under the terms of the Creative Commons Attribution-NonCommercial-NoDerivs License, which permits use and distribution in any medium, provided the original work is properly cited, the use is non-commercial and no modifications or adaptations are made. © 2021 The Authors. *Journal of Clinical Laboratory Analysis* published by Wiley Periodicals LLC.

1 | INTRODUCTION

Acute pancreatitis (AP) is one of the most common gastrointestinal disorders requiring admission to hospital,¹⁻⁴ among which 15-20% of patients develop severe AP (SAP).^{3,4} The mortality rate in SAP has been reported as high as 20-40%.^{3,4} One of the key measures to improve the clinical outcome of AP is early evaluation and identification of SAP, which allows the physicians to perform promptly intensive care. According to the 2012 Atlanta guideline,⁵ SAP is characterized by persistent organ failure, which was assessed by modified Marshall score (MMS) >2 and the duration of organ failure last \geq 48 h. Therefore, determination of SAP usually requires 48 h after disease onset. This makes it difficult to predict SAP in an early stage. Previously, various biomarkers like C-reactive protein (CRP), D-dimer, procalcitonin, and interleukins have been tested for early prediction of AP outcomes.^{4,6-10} But none of them present a flawless performance in predicting SAP.

Leukocyte cell population data (CPD) consist of several leukocyte morphologic parameters, which are measured by automated hematology analyzers. The Coulter DxH 800 hematology analyzer (Beckman Coulter, Fullerton, CA) collects data directly from more than 8000 white blood cells. It can measure cell volume (V) for accurate cell size by direct current impedance, characterize conductivity (C) for the internal composition of each cell through radio frequency opacity, and measure light scatter (S) for cytoplasmic granularity and nuclear structure using a laser beam. Using these data, it can identify neutrophil, lymphocyte, monocyte, eosinophil, or basophil in each cell sample and generate a cell count for each cell type. Leukocyte CPD parameters have been widely applied in the early prediction and diagnosis of acute infection, malaria, and leukemia,¹¹⁻¹⁶ due to their easy and early accessibility. However, there is a lack of study on the application of VCS technology in acute pancreatitis.

Most SAP patients lead to the development of systemic inflammatory response syndrome (SIRS) in the early stage of the disease.¹⁷⁻¹⁹ SIRS is recognized as one of the most important indicators for the occurrence of persistent organ failure, which is responsible for morbidity and mortality in most of the SAP patients.²⁰⁻²³ In addition, some patients will progress to infection of the pancreas or abdomen, which is a fatal complication and they will need active intervention or to be transferred for specialist care.

In the current study, we aim at assessing whether leukocyte CPD parameters could represent an early indicator of SAP. We measured and compared leukocyte CPD parameters of MAP, MSAP, and SAP in the first 3 days of admission. By using stepwise logistic regression model, we screened for a set of CPD parameters to predict SAP and evaluated its diagnostic accuracy.

2 | MATERIALS AND METHODS

2.1 | Participants

This study was approved by the ethical committee of Ruijin Hospital and conducted according to Helsinki declaration. The prospective observational study was conducted from March 2019 to August 2020. Patients who were hospitalized in the emergency department including the intensive care unit and medical ward of Ruijin Hospital, Shanghai Jiao Tong University School of Medicine (Shanghai, China), and diagnosed as AP were recruited in the study. All patients were admitted within 72 h after the onset of the symptoms. Informed consent was obtained from all patients before enrollment into the study. Exclusion criteria were as follows: 1) aged <18 years; 2) patients with a history of chronic pancreatitis; 3) pregnancy or lactation; 4) known malignancy, hematological system diseases.

Sixty-two healthy people including 34 males and 28 females were chosen as healthy controls by routine health checkup in our hospital. Their median (Q1, Q3) of age was 46(33, 55). Exclusion criteria were as follows: 1) aged <18 years; 2) patients with a history of pancreatitis; 3) pregnancy or lactation; 4) known malignancy, hematological system diseases.

2.2 | Diagnosis

According to 2012 revised Atlanta guideline,⁵ acute pancreatitis is defined as the presence of at least two out of the following three criteria: 1) pain in the upper abdomen, 2) serum amylase or lipase concentration >3 times the upper limit of normal, or 3) imaging features of acute pancreatitis on computed tomography (CT) or magnetic resonance imaging (MRI).

The MMS system was used to evaluate the respiratory, renal, and circulation variables. The type and duration of organ failure were also recorded. Mild AP (MAP) was characterized as neither organ failure nor local or systemic complications. Moderate SAP (MSAP) was characterized as transient organ failure (<48 h) or local or systemic complications. SAP was characterized by persistent organ failure (>48 h). Etiologies including biliary,²⁴ lipidemic,²⁵ alcoholic, and idiopathic²⁶ were also determined.

2.3 | Data collection

General information including gender, age, body mass index (BMI), pre-existing comorbidities (cardiac diseases, pulmonary disease, liver disease, diabetes mellitus, hypertension, hyperlipidemia, renal disease, and other comorbidities), and substance abuse (alcohol and tobacco) were collected in the medical chart of AP patient. The daily intake of alcohol abuse did not meet the criteria of alcoholic pancreatitis.²⁶

On admission (day 1) and on the two following days (days 2 and 3), blood samples were obtained from AP patients for blood tests. Data included complete blood cell count (CBC), and the CPD parameters of neutrophils (NE), lymphocytes (LY), eosinophils (EO), and monocytes (MO), which were generated by each individual cell passing through the aperture and were optically and electronically measured by the Coulter DxH 800 hematology analyzer (Beckman Coulter, Fullerton, CA). The CPD parameters included volume (V), conductivity (C), median angle light scatter (MALS),

upper median angle light scatter (UMALS), lower median angle light scatter (LMALS), low angle light scatter (LALS), and axial light loss (AL2). Routine blood tests included serum concentrations of urea, creatinine, bilirubin, procalcitonin (PCT), C-reactive protein (CRP), serum activities of amylase, aspartate and alanine aminotransferases (AST, ALT), and plasma concentrations of D-dimer. The routine tests were conducted on the day of blood collection in the Central Laboratory of Ruijin Hospital using automatic analyzers and standard protocol.

Accordingly, the organ support (mechanical ventilation, renal replacement therapy, vasoactive agent) was recorded. Several severity scoring systems including acute physiology and chronic health evaluation II (APACHE II), sequential organ failure assessment (SOFA), bedside index for severity in acute pancreatitis (BISAP), modified Marshall score (MMS), and modified computed tomography severity index (MCTSI) were collected on 3 days. The outcome indicators including the length of stay (in hospital), surgery, and inhospital mortality were analyzed.

All AP patients received intensive management including controlled fluid resuscitation, support of organ function, and enteral nutrition by the same clinical team to reduce potential bias.²⁷

2.4 | Scoring system

We developed a scoring system based on 4 CPD parameters (SD_ LALS_NE, MN_LALS_LY, SD_LMALS_MO, SD_AL2_MO) to predict the occurrence of SAP. Each patient was assigned a score on a scale of 0 to 4 according to the presence or absence of SD_LALS_NE<36.17, MN_LALS_LY>34.50, SD_LMALS_MO<16.62, SD_AL2_MO<17.27.

2.5 | Statistical analysis

All statistical tests were conducted using SPSS 20.0 statistical software package (IBM Analytics, Armonk, NY) and R project v. 3.5.2 (The R Foundation for Statistical Computing, Vienna, Austria. http:// www.r-project.org). Nominal data were reported as number (percentage of the group). Quantitative data were reported as mean and standard deviation (SD) or median, lower and upper guartiles (Q1; Q3), depending on the normality of each variable's distribution (as assessed with Shapiro-Wilk test). Due to the non-normal distribution of most quantitative variables, Wilcoxon test or Kruskal-Wallis analysis of variance (with post hoc comparisons using Siegel and Castellan method) was applied when comparing two groups or three groups, respectively. The optimal value of cutoff of related indicators was decided using the analysis of time-dependent receiver operating characteristic (ROC) curve. The univariate logistic regression and the backward stepwise (entry and removal probability were 0.05 and 0.10, respectively) multivariate logistic regression model were fitted with SAP, and we calculated the odds ratio (OR) together with corresponding 95% confidence interval (CI). All analysis in this study

was performed two-sided at the 5% significance level. R package stats and FSA were applied to conduct Kruskal-Wallis analysis with post hoc comparison. MASS was used for logistic regression analysis. pROC and ggplot2 were applied for the analysis of ROC curve.

3 | RESULTS

3.1 | Clinical characteristics

The study included 103 patients with AP, 43 women, 60 men, with a median (Q1; Q3) age of 46.00 (34.50; 60.00) years. The median (Q1; Q3) disease onset time of AP patients was 24.00 (12.00; 36.00) hours. According to the 2012 Atlanta criteria, 30 patients (29.13%) were diagnosed with MAP, 42 (40.78%) with MSAP, and 31 (30.10%) with SAP. MAP, MSAP, and SAP patients did not differ significantly in terms of age, sex, BMI, onset time, percentage affected with comorbidities, substance abuse, and AP etiology (Table 1). We also collected the vital sign of AP patients. Compared with MAP patients, the temperature and heart rate were higher in MSAP and SAP patients. And the respiratory rate in MAP patients was lower than MSAP and SAP patients (Table 1).

In the operation, the percentage of percutaneous transhepatic gallbladder drainage was higher in SAP (16.13%) patients compared with that in MAP (0%) patients. Compared with MAP (0.0%) patients, the percentage of percutaneous peritoneal drainage was higher in MSAP (16.67%) and SAP (58.06%) patients. The percentage of surgery was higher in SAP (25.81%) patients compared with that in MAP (3.33%) and MSAP (0.00%) patients (Table 1).

In addition, we investigated clinical outcomes among three groups. The percentage of ICU admission and the length of hospital stay were higher among patients with MSAP and SAP. Five AP patients (16.13%) died, all in SAP group (Table 1).

On admission, the patients with SAP were characterized with significantly higher concentrations of amylase (810.00 U/L), aspartate aminotransferase (47.00 U/L), total bilirubin (29.60 μ mol/L), creatinine (71.00 μ mol/L), urea (7.20 mmol/L), and D-dimer (6.07 mg/L). The concentrations of inflammatory markers, serum procalcitonin, in SAP (2.66 ng/ml) patients were higher than in MAP (0.26 ng/ml) and MSAP (0.70 ng/ml) patients significantly, and the serum CRP showed no significant difference between MSAP (197.00 mg/L) and SAP (231.00 mg/L) patients, but both of them were higher than that of MAP (123.50 mg/L) patients (Table 2).

In terms of hematological variables, the count of lymphocyte, eosinophil, red blood cell, platelet, and the percentage of hematocrit were significantly lower among patients with SAP relatively. However, other hematological variables did not differ significantly between the MAP, MSAP and SAP patients (Table 2).

As shown in Table 3, several scoring systems including APACHE II, MMS, SOFA, BISAP, and MCTSI scores in SAP patients were significantly higher than in MAP and MSAP patients. This was also observed in the case of persistent (≥48 h) organ dysfunction.

4 of 17	
	VILE I
TABLE 1	Clinical characteristics of the study group according to the severity of acute pancreatitis (AP)

	, , , ,	<i>i i</i>		
Characteristic	MAP (<i>n</i> = 30)	MSAP (n = 42)	SAP (n = 31)	P-value
Demographics				
Male sex, n (%)	16 (53.33)	26 (61.90)	18 (58.06)	0.767
Age (Q1; Q3), years	53.00 (36.00; 62.75)	48.00 (34.25; 59.75)	44.00 (34.50; 50.00)	0.198
Body mass index (Q1; Q3), kg/m ²	24.87 (23.55; 27.67)	25.09 (23.77; 28.15)	28.40 (24.30; 31.82)	0.109
Onset time (Q1; Q3), h	24.00 (12.00; 30.00)	30.00 (24.00; 48.00)	24.00 (24.00; 42.00)	0.096
Pre-existing comorbidities				
Cardiac diseases, n (%)	7 (23.33)	4 (9.52)	3 (9.68)	0.181
Pulmonary disease, n (%)	2 (6.67)	3 (7.14)	3 (9.68)	0.905
Liver disease, n (%)	12 (40.00)	25 (59.52)	11 (35.48)	0.087
Diabetes mellitus, n (%)	9 (30.00)	13 (30.95)	4 (12.90)	0.166
Hypertension, n (%)	13 (43.33)	14 (33.33)	9 (29.03)	0.484
Hyperlipidemia, <i>n</i> (%)	11 (36.67)	24 (57.14)	16 (51.61)	0.222
Renal disease, n (%)	1 (3.33)	3 (7.14)	3 (9.68)	0.637
Other comorbidities, n (%)	7 (23.33)	8 (19.05)	5 (16.13)	0.774
Substance abuse				
Alcohol, n (%)	11 (36.67)	15 (35.71)	11 (35.48)	0.928
Tobacco, n (%)	11 (36.67)	15 (35.71)	10 (32.26)	0.928
Etiology				
Biliary, n (%)	15 (50.00)	14 (33.33)	17 (54.84)	0.600
Hypertriglyceridemia, n (%)	8 (26.67)	18 (42.86)	9 (29.03)	
Alcoholic, n (%)	5 (16.67)	8 (19.05)	4 (12.90)	
Other/idiopathic, n (%)	2 (6.67)	2 (4.76)	1 (3.23)	
Vital signs				
Temperature (Q1; Q3), °C	37.30 (37.00; 37.60)	38.10 (37.55; 38.40)	38.90 (38.40; 39.05)	<0.001 ^{a,b,c}
Heart rate (Q1; Q3), beats per minute	94.50 (82.75; 101.50)	106.50 (92.50; 119.00)	125.00 (114.50; 135.00)	<0.001 ^{a,b,c}
Respiratory rate (Q1;Q3), breaths per minute	20.00 (19.25; 21.75)	24.50 (20.25; 28.00)	30.00 (22.50; 35.50)	<0.001 ^{a,c}
Mean arterial pressure (Q1; Q3), mmHg	100.16 (90.33; 105.58)	102.98 (88.42; 110.53)	103.00 (94.84; 118.35)	0.362
Pulse oxygen saturation (Q1; Q3), %	100.00 (99.00; 100.00)	100.00 (99.25; 100.00)	98.00 (93.00; 100.00)	0.001 ^{a,c}
Operation				
Therapeutic ERCP, n (%)	0 (0.00)	2 (4.76)	2 (6.45)	0.558
Percutaneous transhepatic gallbladder drainage, n (%)	0 (0.00)	2 (4.76)	5 (16.13)	0.040 ^a
Percutaneous peritoneal drainage, n (%)	0 (0.00)	7 (16.67)	18 (58.06)	<0.001 ^{a,b,c}
Surgery, n (%)	1 (3.33)	0 (0.00)	8 (25.81)	<0.001 ^{a,b}
Outcome				
ICU admission, n (%)	2 (6.67)	30 (71.43)	31 (100.00)	<0.001 ^{a,b,c}
Hospital mortality, n (%)	0 (0.00)	0 (0.00)	5 (16.13)	0.004 ^{a,b}
Length of hospital stay, days	15.00 (11.25; 19.75)	23.00 (21.00; 29.50)	45.00 (39.50; 62.50)	<0.001 ^{a,b,c}

Abbreviations: ERCP, endoscopic retrograde cholangiopancreatography; ICU, intensive care unit; MAP, mild acute pancreatitis; MSAP, moderately severe acute pancreatitis; SAP, severe acute pancreatitis.

Note: Categorical variables presented as *n* (%), number and percentage; Continuous variables presented as median (Q1; Q3), Q1, lower quartile; Q3, upper quartile; *p*-value is reported for overall comparison between three groups (in Pearson chi-squared test or Kruskal-Wallis ANOVA), the letters in superscript indicate the results of post hoc tests: ^a significant difference between the MAP and SAP groups in post hoc comparison; ^b significant difference between the MAP and MSAP groups in post hoc comparison; ^c significant difference between the MAP and MSAP groups in post hoc comparison.

TABLE 2 The results of laboratory tests on admission according to the AP severity. Data are shown as median (Q1; Q3)

Variable	MAP (n=30)	MSAP (n=42)	SAP (n=31)	P-value
Amylase, U/L	312.00 (112.50; 669.00)	467.00 (193.50; 828.50)	810.00 (299.00; 1282.00)	0.027 ^a
Triglyceride, mmol/L	3.24 (1.19; 4.48)	2.78 (1.50; 6.16)	2.63 (1.40; 6.75)	0.856
ALT, U/L	26.50 (19.00; 69.50)	16.00 (12.00; 36.75)	27.00 (15.00; 57.50)	0.062
AST, U/L	22.00 (14.25; 54.50)	22.50 (18.00; 32.75)	47.00 (30.00; 58.00)	0.006 ^{a,b}
Total bilirubin, μmol/L	17.00 (12.62; 22.67)	17.80 (11.25; 24.80)	29.60 (19.65; 46.70)	<0.001 ^{a,b}
Direct bilirubin, µmol/L	2.70 (1.90; 4.38)	3.55 (2.02; 7.28)	10.40 (4.65; 16.35)	<0.001 ^{a,b}
Creatinine, µmol/L	64.50 (53.25; 74.50)	56.00 (48.25; 74.50)	71.00 (59.50; 138.00)	0.008 ^b
Urea, mmol/L	4.10 (3.15; 5.68)	4.55 (3.10; 6.00)	7.20 (4.50; 10.75)	<0.001 ^{a,b}
D-dimer, mg/L	0.82 (0.57; 2.05)	4.86 (3.05; 8.96)	6.07 (3.68; 10.52)	<0.001 ^{a,c}
Procalcitonin, ng/ml	0.26 (0.13; 0.62)	0.70 (0.36; 1.92)	2.66 (0.83; 6.44)	<0.001 ^{a,b,c}
CRP, mg/L	123.50 (49.02; 157.50)	197.00 (124.25; 272.25)	231.00 (147.50; 276.00)	<0.001 ^{a,c}
White blood cell count, *10 ⁹ /L	12.57 (10.79; 13.98)	11.46 (8.63; 14.37)	10.15 (7.55; 13.20)	0.415
Neutrophil count, *10 ⁹ /L	10.41 (8.55; 12.13)	9.73 (7.51; 12.74)	9.12 (6.85; 12.20)	0.899
Lymphocyte count, *10 ⁹ /L	1.41 (1.03; 1.66)	0.92 (0.71; 1.29)	0.85 (0.62; 1.06)	<0.001 ^{a,b}
Eosinophil count, *10 ⁹ /L	0.05 (0.01; 0.18)	0.03 (0.01; 0.09)	0.01 (0.00; 0.04)	0.019 ^a
Monocyte count, *10 ⁹ /L	0.58 (0.47; 0.72)	0.49 (0.35; 0.70)	0.42 (0.30; 0.70)	0.136
Basophil count, *10 ⁹ /L	0.02 (0.01; 0.03)	0.02 (0.01; 0.03)	0.02 (0.01; 0.02)	0.640
Red blood cell count, $^{*10^{12}}/L$	4.60 (4.02; 4.98)	3.99 (3.65; 4.37)	3.46 (2.86; 4.43)	<0.001 ^{a,c}
Hematocrit, %	41.50 (37.00; 45.50)	37.00 (33.00; 40.00)	31.00 (27.50; 39.50)	0.002 ^{a,c}
Platelet count, *10 ⁹ /L	197.50 (175.75; 229.50)	173.00 (133.25; 249.75)	131.00 (87.50; 206.50)	0.007 ^a

Abbreviations: ALT, alanine aminotransferase; AST, aspartate aminotransferase; CRP, C-reactive protein; MAP, mild acute pancreatitis; MSAP, moderately severe acute pancreatitis; SAP, severe acute pancreatitis.

Note: Categorical variables presented as *n* (%), number and percentage; Continuous variables presented as median (Q1; Q3), Q1, lower quartile; Q3, upper quartile; P-value is reported for overall comparison between three groups (in Pearson chi-squared test or Kruskal-Wallis ANOVA), the letters in superscript indicate the results of post hoc tests: ^a significant difference between the MAP and SAP groups in post hoc comparison; ^b significant difference between the MAP and MSAP groups in post hoc comparison; ^c significant difference between the MAP and MSAP groups in post hoc comparison; ^c significant difference between the MAP and MSAP groups in post hoc comparison.

TABLE 3 Comparison of severity and organ dysfunction among cohorts

Characteristic	MAP (n=30)	MSAP (n=42)	SAP (n=31)	p-Value
APACHE II score (Q1; Q3), points	4.00 (2.25; 6.00)	6.50 (3.00; 10.00)	11.00 (7.50; 14.50)	<0.001 ^{a,b}
Modified Marshall score (Q1; Q3), points	0.00 (0.00; 1.00)	2.00 (1.00; 2.00)	3.00 (2.00; 3.50)	<0.001 ^{a,b,c}
SOFA score (Q1; Q3), points	1.00 (0.00; 2.00)	2.00 (2.00; 3.75)	5.00 (3.50; 7.00)	<0.001 ^{a,b,c}
BISAP score (Q1; Q3), points	0.50 (0.00; 1.00)	2.00 (1.00; 2.00)	2.00 (2.00; 3.00)	<0.001 ^{a,b,c}
MCTSI score (Q1; Q3), points	2.00 (2.00; 2.00)	4.00 (4.00; 5.50)	6.00 (4.00; 8.00)	<0.001 ^{a,b,c}
Persistent (\geq 48 h) respiratory failure, n (%)	0 (0.0)	0 (0.0)	27 (87.10)	<0.001 ^{a,b}
Persistent (≥48 h) acute renal failure, n (%)	0 (0.0)	0 (0.0)	9 (29.03)	0.002 ^{a,b}
Persistent (≥48 h) circulatory failure, <i>n</i> (%)	0 (0.0)	0 (0.0)	6 (19.35)	0.008 ^{a,b}
Mechanical ventilation, n (%)	0 (0.0)	0 (0.0)	23 (74.19)	<0.001 ^{a,b}
Renal replacement therapy, n (%)	0 (0.0)	0 (0.0)	5 (16.13)	0.004 ^b
Use of vasoactive agent, n (%)	0 (0.0)	0 (0.0)	6 (19.35)	<0.001 ^{a,b}

Abbreviations: APACHE II, acute physiology and chronic health evaluation II; BISAP, bedside index for severity in acute pancreatitis; MAP, mild acute pancreatitis; MCTSI, modified computed tomography severity index; MSAP, moderately severe acute pancreatitis; SAP, severe acute pancreatitis; SOFA, sequential organ failure assessment.

Note: Categorical variables presented as *n* (%), number and percentage; Continuous variables presented as median (Q1; Q3), Q1, lower quartile; Q3, upper quartile; P-value is reported for overall comparison between three groups (in Pearson chi-squared test or Kruskal-Wallis ANOVA), the letters in superscript indicate the results of post hoc tests: ^a significant difference between the MAP and SAP groups in post hoc comparison; ^b significant difference between the MAP and MSAP groups in post hoc comparison; ^c significant difference between the MAP and MSAP groups in post hoc comparison.

'II FY

3.2 | Comparison of CPD parameters among MAP, MSAP, and SAP patients in first 3 days after admission

We examined 42 CPD parameters in total during first 3 days of admission: 14 CPD parameters for neutrophil, lymphocyte, and monocyte, respectively. On admission, as listed in Tables 4, S1 and S4, 18 CPD parameters were significantly different among MAP, MSAP, and SAP. Most of them showed higher levels in SAP than either MAP or MSAP. Seven (SD_V_NE, SD_LMALS_NE, SD_AL2_NE, SD_C_LY, SD_UMALS_LY, SD_LMALS_LY, and SD_AL2_LY) and 2 (MN_C_LY and MN_C_MO) were higher in SAP compared with either MAP or MSAP, respectively. Five of 18 (MN_V_NE, SD_LALS_NE, SD_ MALS_LY, SD_V_MO, and SD_AL2_MO) were highest in SAP than in both MAP and MSAP. The rest 4 (MN_MALS_NE, MN_UMALS_ NE, MN_LMALS_NE, and MN_LALS_LY) were lower in SAP than in MAP but not MSAP. On day 2 (Tables 5, S2 and S5), 11 of 42 CPD parameters were different among 3 groups. Five of 11 (MN_V_NE, SD_MALS_LY, SD_UMALS_LY, MN_V_MO, and SD_AL2_MO) were higher in SAP than in MAP and MSAP. Two (SD_V_NE and SD_V_ MO) were higher in SAP compared with that in MAP but not MSAP. SD_C_LY was higher in SAP than in MSAP but not MAP. On the contrary, another 3 (SD_C_NE, MN_MALS_NE, and MN_UMALS_NE) showed lower levels in SAP. On day 3 (Tables 6, S3 and S6), 16 CPD

parameters were significantly different. Nine (MN_V_NE, SD_V_LY, SD_C_LY, SD_MALS_LY, SD_UMALS_LY, SD_LMALS_LY, SD_AL2_LY, MN_V_MO, and SD_V_MO) were higher in SAP compared with that in MAP and MSAP. Three of 16 (SD_V_NE, MN_UMALS_LY, and SD_AL2_MO) showed higher levels in SAP compared with that in MAP, whereas 4 (MN_MALS_NE, MN_UMALS_NE, MN_LMALS_NE, and MN_LALS_LY) were lower in SAP. Taken together, as shown in Figure 1, 4 of 42 CPD parameters were highest in SAP on admission and were constantly different among MAP, MSAP, and SAP during all 3 days of hospital stay.

3.3 | Diagnosis value of CPD parameters for SAP and ICU transfer or death

Before screening for biomarkers for early identification of SAP, we first compared levels of all 42 CPD parameters between AP patients and healthy controls on admission. As shown in Table S7, majority of the CPD parameters had significantly different levels between 2 groups, whereas only 4 (MN_V_LY, MN_LMALS_LY, SD_C_MO, and MN_MALS_MO) showed no difference. We used univariate logistic regression analysis to select parameters correlated with the occurrence of SAP. As listed in Table 7, 17 CPD parameters were screened.

TABLE 4 Results of CPD parameters on day 1 according to the AP severity. Data are shown as median (Q1; Q3)

Variable	MAP (n = 30)	MSAP (n = 42)	SAP (n = 31)	χ ²	P-value
MN_V_NE	152.50 (147.00; 157.75)	158.00 (152.25; 164.75)	164.00 (159.00; 168.50)	21.11	0.001 ^{a,b,c}
SD_V_NE	19.60 (18.64; 22.36)	22.23 (20.73; 23.86)	22.01 (21.02; 25.54)	14.30	0.001 ^{a,c}
MN_MALS_NE	140.50 (135.00; 144.00)	137.00 (130.25; 141.75)	134.00 (129.50; 138.00)	15.31	0.001 ^{a,c}
MN_UMALS_NE	141.00 (138.25; 144.00)	141.00 (135.00; 144.75)	138.00 (133.50; 141.00)	6.78	0.034ª
MN_LMALS_NE	137.50 (130.25; 141.00)	130.50 (119.00; 136.75)	125.00 (121.00; 133.00)	16.41	0.001 ^{a,c}
SD_LMALS_NE	12.72 (12.10; 14.45)	15.50 (12.61; 18.50)	15.76 (13.53; 18.11)	8.33	0.016 ^{a,c}
SD_LALS_NE	31.50 (29.61; 35.04)	35.12 (31.80; 41.51)	41.97 (39.26; 45.42)	25.96	0.001 ^{a,b,c}
SD_AL2_NE	13.39 (12.52; 14.78)	14.67 (13.49; 16.67)	14.75 (13.51; 17.21)	8.97	0.011 ^{a,c}
MN_C_LY	116.00 (114.00; 119.50)	115.00 (113.00; 117.00)	118.00 (115.00; 120.00)	8.08	0.018 ^b
SD_C_LY	11.54 (10.00; 14.92)	12.28 (9.45; 14.68)	14.57 (11.11; 17.21)	6.59	0.037 ^a
SD_MALS_LY	18.05 (16.19; 19.81)	18.61 (17.39; 20.33)	21.03 (18.56; 22.20)	13.95	0.001 ^{a,b}
SD_UMALS_LY	19.83 (18.74; 21.99)	21.11 (19.55; 23.40)	22.98 (20.47; 24.12)	9.59	0.008ª
SD_LMALS_LY	21.05 (19.17; 22.49)	21.94 (20.27; 23.03)	23.32 (21.55; 24.63)	11.03	0.004 ^a
MN_LALS_LY	38.50 (35.00; 49.00)	36.50 (33.25; 45.75)	34.00 (32.00; 41.00)	7.52	0.023ª
SD_AL2_LY	10.47 (10.04; 11.36)	11.37 (10.29; 12.32)	12.39 (11.04; 12.96)	12.16	0.002ª
SD_V_MO	21.94 (19.98; 24.55)	24.53 (22.65; 26.66)	26.91 (23.86; 29.72)	22.70	0.001 ^{a,b,c}
MN_C_MO	124.00 (122.25; 126.75)	123.00 (121.00; 123.00)	125.00 (121.50; 126.00)	8.58	0.014 ^{b,c}
SD_AL2_MO	13.66 (12.73; 14.96)	15.38 (13.89; 17.54)	20.85 (18.22; 22.71)	44.93	0.001 ^{a,b,c}

Abbreviations: AL2, axial light loss; C, conductivity; CPD, cell population data; LALS, low angle light scatter; LMALS, lower median angle light scatter; LY, lymphocyte; MALS, median angle light scatter; MAP, mild acute pancreatitis; MN, mean; MO, monocyte; MSAP, moderately severe acute pancreatitis; NE, neutrophil; Q1, lower quartile; Q3, upper quartile; SAP, severe acute pancreatitis; SD, standard deviation; UMALS, upper median angle light scatter; V, volume.

Note: P-value is reported for overall comparison between three groups (in Kruskal-Wallis ANOVA), the letters in superscript indicate the results of post-hoc tests: ^a significant difference between the MAP and SAP groups in post hoc comparison; ^b significant difference between the MSAP and SAP groups in post hoc comparison; ^c significant difference between the MAP and MSAP groups in post hoc comparison.

 TABLE 5
 Results of CPD parameters on day 2 according to the AP severity. Data are shown as median (Q1; Q3)

Variable	MAP (n = 30)	MSAP (n = 42)	SAP (n = 31)	χ ²	P-value
MN_V_NE	150.00 (146.00; 154.75)	154.50 (149.00; 163.75)	161.00 (155.00; 166.50)	16.43	0.001 ^{a,b,c}
SD_V_NE	20.21 (18.29; 21.34)	21.44 (19.58; 22.68)	21.40 (20.18; 23.82)	9.71	0.007 ^{a,c}
SD_C_NE	6.47 (6.14; 6.94)	6.14 (5.63; 6.59)	5.74 (5.02; 6.16)	13.62	0.001 ^{a,b}
MN_MALS_NE	141.50 (136.00; 143.75)	137.50 (133.00; 141.75)	136.00 (132.50; 138.50)	9.20	0.010 ^{a,c}
MN_UMALS_NE	140.00 (138.00; 143.75)	139.50 (136.00; 141.00)	136.00 (134.00; 141.00)	8.10	0.017 ^a
SD_C_LY	12.17 (9.60; 14.35)	11.68 (9.69; 13.78)	13.99 (11.71; 16.06)	7.56	0.022 ^b
SD_MALS_LY	17.89 (16.84; 19.50)	18.33 (16.92; 20.34)	20.29 (19.12; 21.87)	10.33	0.005 ^{a,b}
SD_UMALS_LY	20.26 (19.39; 22.09)	20.35 (19.25; 22.33)	22.32 (21.64; 23.65)	8.58	0.013 ^{a,b}
MN_V_MO	168.00 (163.00; 172.75)	169.00 (164.00; 174.00)	176.00 (167.00; 182.00)	8.93	0.011 ^{a,b}
SD_V_MO	21.17 (19.69; 23.46)	24.30 (21.29; 26.59)	25.73 (23.72; 27.37)	19.80	0.001 ^{a,c}
SD_AL2_MO	13.63 (13.26; 15.01)	15.97 (14.36; 18.28)	17.68 (16.15; 19.40)	30.00	0.001 ^{a,b,c}

Abbreviations: AL2, axial light loss; C, conductivity; CPD, cell population data; LALS, low angle light scatter; LMALS, lower median angle light scatter; LY, lymphocyte; MALS, median angle light scatter; MAP, mild acute pancreatitis; MN, mean; MO, monocyte; MSAP, moderately severe acute pancreatitis; NE, neutrophil; Q1, lower quartile; Q3, upper quartile; SAP, severe acute pancreatitis; SD, standard deviation; UMALS, upper median angle light scatter; V, volume.

Note: P-value is reported for overall comparison between three groups (in Kruskal-Wallis ANOVA), the letters in superscript indicate the results of post hoc tests: ^a significant difference between the MAP and SAP groups in post hoc comparison; ^b significant difference between the MSAP and SAP groups in post hoc comparison; ^c significant difference between the MAP and MSAP groups in post hoc comparison.

Variable	MAP (n = 30)	MSAP (n = 42)	SAP (n = 31)	χ^2	P-Value
MN_V_NE	148.50 (144.25; 153.00)	153.00 (149.00; 159.75)	159.00 (153.00; 167.50)	16.71	0.002 ^{a,b,c}
SD_V_NE	19.05 (18.17; 22.08)	21.43 (19.30; 22.74)	21.81 (19.56; 23.34)	7.07	0.029 ^a
MN_MALS_NE	142.00 (138.25; 144.75)	137.00 (132.50; 141.75)	135.00 (133.00; 139.00)	13.27	0.001 ^{a,c}
MN_UMALS_NE	142.00 (139.25; 145.00)	139.00 (136.00; 141.00)	137.00 (134.00; 141.50)	9.88	0.007 ^{a,c}
MN_LMALS_NE	138.00 (135.25; 140.75)	132.00 (126.00; 138.75)	131.00 (126.50; 135.00)	9.41	0.009 ^{a,c}
SD_V_LY	15.72 (14.02; 17.71)	17.04 (15.95; 19.14)	19.63 (16.34; 21.72)	14.81	0.001 ^{a,b,c}
SD_C_LY	11.44 (9.02; 12.71)	12.48 (10.39; 14.00)	15.66 (12.55; 16.91)	19.32	0.001 ^{a,b}
SD_MALS_LY	17.76 (16.79; 19.32)	18.63 (17.74; 20.34)	20.34 (18.87; 21.81)	15.79	0.001 ^{a,b}
MN_UMALS_LY	70.50 (65.50; 75.75)	76.50 (67.00; 81.00)	77.00 (70.50; 83.00)	6.97	0.030 ^a
SD_UMALS_LY	20.13 (19.04; 21.88)	20.92 (19.21; 22.90)	22.15 (21.26; 24.16)	10.38	0.005 ^{a,b}
SD_LMALS_LY	21.07 (19.90; 22.74)	21.67 (20.70; 22.98)	23.44 (21.76; 24.24)	13.76	0.001 ^{a,b}
MN_LALS_LY	44.00 (37.00; 48.00)	36.00 (34.00; 46.75)	35.00 (32.50; 43.50)	9.86	0.007 ^{a,c}
SD_AL2_LY	10.65 (9.85; 11.41)	11.48 (10.41; 12.76)	12.70 (11.76; 13.97)	15.58	0.001 ^{a,b}
MN_V_MO	168.00 (162.00; 170.00)	169.00 (164.00; 172.00)	175.00 (168.50; 181.50)	16.38	0.001 ^{a,b}
SD_V_MO	20.60 (18.77; 21.88)	24.49 (21.76; 26.29)	25.86 (23.83; 29.45)	27.32	0.001 ^{a,b,c}
SD_AL2_MO	13.94 (12.52; 15.81)	14.65 (13.01; 17.57)	16.80 (14.67; 19.40)	10.50	0.005 ^a

TABLE 6 Results of CPD parameters on day 3 according to the AP severity. Data are shown as median (Q1; Q3)

Abbreviations: AL2, axial light loss; C, conductivity; CPD, cell population data; LALS, low angle light scatter; LMALS, lower median angle light scatter; LY, lymphocyte; MALS, median angle light scatter; MAP, mild acute pancreatitis; MN, mean, SD, standard deviation; MO, monocyte. Q1, lower quartile; MSAP, moderately severe acute pancreatitis; NE, neutrophil; Q3, upper quartile; SAP, severe acute pancreatitis; UMALS, upper median angle light scatter; V, volume.

Note: P-value is reported for overall comparison between three groups (in Kruskal-Wallis ANOVA), the letters in superscript indicate the results of post hoc tests: ^a significant difference between the MAP and SAP groups in post hoc comparison; ^b significant difference between the MSAP and SAP groups in post hoc comparison; ^c significant difference between the MAP and MSAP groups in post hoc comparison.

Then, we used stepwise multivariate logistic regression model to screen for optimal parameters and identified a set of 4 CPD parameters (SD_LALS_NE, MN_LALS_LY, SD_LMALS_MO, and SD_AL2_MO) that made the model best fitted. We then established a simple

scoring system using these 4 CPD parameters. As listed in Table 8, cutoff values of each CPD parameter were calculated by ROC curve analysis and the scoring system was optimized upon AUCs of different combination of 4 cutoff values. As a result, according to the



FIGURE 1 Four CPD parameters (MN_V_NE, SD_MALS_LY, SD_V_MO, and SD_AL2_MO) were highest in SAP on admission and were continuously different among MAP, MSAP, and SAP during all 3 days of hospital stay. Data are shown as median (Q1; Q3). CPD, cell population data; MAP, mild acute pancreatitis; MSAP, moderately severe acute pancreatitis; SAP, severe acute pancreatitis; MN, mean, SD, standard deviation; V, volume; MALS, median angle light scatter; AL2, axial light loss; NE, neutrophil; LY, lymphocyte; MO, monocyte

presence of SD_LALS_NE<36.17, MN_LALS_LY>34.50, SD_LMALS_ MO<16.62 and SD_AL2_MO<17.27, 1 point for each CPD parameter was assigned, which leads to a scale of 0 to 4 points for each patient. Median scores of SAP and non-SAP were 1 and 3, respectively.

By using this scoring system, this set of 4 CPD parameters had a sensitivity of 96.8%, specificity of 65.3% and AUC of 0.87 for diagnostic accuracy on early identification of SAP. We drew ROC curve of this set of 4 CPD parameters and compared its AUC with other criteria (MCTSI, SOFA, APACHE II, MMS, BISAP) or biomarkers (CRP, PCT, WBC) that had been widely applied in SAP identification. As shown in Figure 2, Tables 9 and 10, this set of 4 CPD parameters showed an AUC of 0.87 which was comparable with 0.72, 0.85, 0.85, 0.87, 0.80 of MCTSI, SOFA, APACHE II, MMS, and BISAP, respectively. It even showed a higher AUC compared with CRP (0.67), PCT (0.79), WBC (0.57), and combination of these 3 biomarkers (0.74).

We also evaluated this scoring system on the prediction of ICU transfer or death. It had a sensitivity of 74.6%, specificity of 79.5%, and AUC of 0.81 to predict ICU transfer or death (Tables 9 and 10). Its AUC was slightly lower than other criteria like MCTSI (0.86), SOFA (0.86), APACHE II (0.88), MMS (0.87), and BISAP (0.91) but higher than biomarkers as CRP (0.76), PCT (0.79), WBC (0.51), and combination of these 3 biomarkers (0.83) (Figure 2, Tables 9 and 10).

Finally, we performed onset time subgroup analysis on our scoring system. As shown in Table 11, AUCs of our scoring system remained stable among patients who admitted to hospital within 24, 24-48, and 48-72 h after disease onset for prediction of SAP (0.88, 0.88, and 0.75, respectively) and ICU transfer or death (0.82, 0.77, and 0.84, respectively).

DISCUSSION 4

It continues to be a challenge of early prognostic prediction in AP. If the precise and rapid determination of disease course in the early stage of AP can be achieved, appropriate therapeutic intervention will be introduced in time. That is the reason why the availability of accessible and practical parameters, for example, ones measured by modern hematological analyzers, could be a valuable perspective.28

Many biomarkers have been evaluated previously with respect to their value for predicting AP results.^{6-10,29} Unlike those biomarkers, the white blood cell (WBC) count is among the first laboratory tests available. Unfortunately, previous studies^{30,31} and our results (Figure 2) suggested that WBC count elevation alone was

TABLE 7Odds ratios (95% confidence
intervals) for CPD parameters in
prediction of unfavorable course of AP

	Univariate analysis		Multivariate analysis	
	OR (95% CI)	P-value	OR (95% CI)	P-value
MN_V_NE	1.09 (1.04; 1.16)	0.002		
SD_V_NE	1.40 (1.16; 1.75)	0.001		
MN_MALS_NE	0.88 (0.81; 0.94)	0.001	0.91 (0.78; 1.04)	0.180
MN_LMALS_NE	0.91 (0.85; 0.95)	0.001		
SD_LMALS_NE	1.24 (1.07; 1.50)	0.010		
MN_LALS_NE	0.99 (0.97; 1.00)	0.028	1.04 (1.00; 1.08)	0.055
SD_LALS_NE	1.15 (1.07; 1.25)	0.001	1.13 (1.03; 1.27)	0.021
SD_AL2_NE	1.34 (1.08; 1.71)	0.013		
SD_MALS_LY	1.24 (1.06; 1.50)	0.014		
SD_UMALS_LY	1.20 (1.04; 1.42)	0.020		
SD_LMALS_LY	1.26 (1.05; 1.55)	0.020		
MN_LALS_LY	0.94 (0.89; 1.00)	0.043	0.81 (0.68; 0.95)	0.013
SD_AL2_LY	1.55 (1.15; 2.21)	0.009		
SD_V_MO	1.35 (1.17; 1.61)	0.001		
SD_LMALS_MO	1.31 (1.06; 1.67)	0.019	0.63 (0.40; 0.93)	0.031
MN_LALS_MO	0.98 (0.96; 1.00)	0.045		
SD AL2 MO	1.57 (1.29; 2.03)	0.001	1.83 (1.43: 2.55)	0.001

Abbreviations: AL2, axial light loss; CI, confidence interval; CPD, cell population data; LALS, low angle light scatter; LMALS, lower median angle light scatter; LY, lymphocyte; MALS, median angle light scatter; MN, mean; MO, monocyte; NE, neutrophil; OR, odds ratio; SD, standard deviation; UMALS, upper median angle light scatter; V, volume.

 TABLE 8
 Scoring system of 4 CPD parameters for prediction of unfavorable course of AP

			Median (C	Q1; Q3)
	Cutoff	Score	SAP	Non-SAP
SD_LALS_NE	<36.17	1	1 (1; 2)	3 (2; 4)
MN_LALS_LY	>34.50	1		
SD_LMALS_MO	<16.62	1		
SD_AL2_MO	<17.27	1		

Abbreviations: AL2, axial light loss; CPD, cell population data; LALS, low angle light scatter; LMALS, lower median angle light scatter; LY, lymphocyte; MN, mean; MO, monocyte; NE, neutrophil; Non-SAP, mild acute pancreatitis, moderately severe acute pancreatitis; SAP, severe acute pancreatitis; SD, standard deviation.

nonspecific for SAP identification. However, several studies implied that the morphologic alteration of leukocytes also generates important diagnostic information characteristically.^{32,33}

Potential clinical utilities of leukocyte CPD have been widely studied in the last several years. Nevertheless, using CPD to measure morphologic alteration in acute pancreatitis has not been fully investigated. Evaluation of peripheral blood leukocyte CPD is analogous to the microscopic examination of the leukocyte morphology on a peripheral blood smear but uses the modern technology to automatically define the cellular morphology with increased accuracy and consistency.³⁴

In our study, SD_V_MO (standard deviation of monocyte volume), which is also named MDW (monocyte volume distribution width), increased significantly in SAP patients during the first 3 days of admission. Most recently, the MDW, cleared by FDA for emergency department clinicians to identify patients with sepsis or increased risk of developing sepsis, became the first leukocyte morphologic parameter for clinical diagnosis.^{33,35,36} SAP and sepsis have some similarities in the early stage, including organ function damage,⁵ SIRS (diagnosed by Sepsis-2 criteria³⁷) and a probability of infection.³⁸

It has been previously recognized that morphologic changes of circulating immune cells could be an early sign of infection. In response to microbial "danger signals," circulating immune cells, especially monocytes and neutrophils, are rapidly activated, which can be characterized by changes in their size and shape^{39,40} as well as the release of chemokines and cytokines.^{41,42} The circulating monocytes are first-line responders to infections, ^{43,44} and such response is proportional to the intensity of the exposure to either bacterial, fungal, or viral pathogens,⁴⁵ resulting in an acute increase in cell size.^{46,47} We observed the increase of SD_V_MO in SAP patients and posited that it may be related to the secondary infection and SIRS in the process of SAP and SD_V_MO reflects the inflammatory process in these patients. We also found that MN_V_NE and SD_V_NE are significantly higher in SAP patients than those in MAP and MSAP patients during the first 3 days of admission (Figure 1). These two parameters have been reported their wide use in the early prediction of bacterial



FIGURE 2 Receiver operating characteristic (ROC) curves for the the scoring system of 4 CPD parameters on day 1 in prediction of SAP (A and B), and ICU transfer or death (C and D). For comparison, ROC curves are shown for other scoring systems and biomarkers of AP severity measured on day 1. CPD, cell population data; MN, mean, SD, standard deviation; MALS, median angle light scatter; LALS, low angle light scatter; AL2, axial light loss; NE, neutrophil; LY, lymphocyte; MO, monocyte; SAP, severe acute pancreatitis; ICU, intensive care unit; MCTSI, modified computed tomography severity index; SOFA, sequential organ failure assessment; APACHE II, acute physiology and chronic health evaluation II; MMS, modified Marshall score; BISAP, bedside index for severity in acute pancreatitis; CRP, C-reactive protein; PCT, procalcitonin; WBC, white GA blood cell; CRP, C-reactive protein; PCT, procalcitonin; WBC, white blood cell

infection and sepsis,^{48,49} and their increase in SAP patients may also predict the infection.

The VCS technology measures the mean of cellular volume and conductivity as well as the light scatter.⁵⁰ The parameter combinations we selected to predict SAP were all light scatter correlation, including low angle light scatter (LALS), lower median angle light scatter (LMALS), and axial light loss measurement (AL2). Among them, LALS is an indicator of nuclear complexity, suggesting nuclear hypo-segmentation or chromatin condensation. LMALS implies cytoplasmic degranulation. AL2 suggests the change of cellular transparency or opacity.³⁴ These light scatter changes likely reflect the alterations corresponding to intrinsic biophysical properties of activated leukocytes either due to underlying inflammation or infection. Reviewing the previous studies, F Chaves et al reported the light scatter of neutrophil decreased significantly in acute bacterial infection patients.⁵¹ P Arora et al also found that mean neutrophil

scatter was significantly lower in cases as compared to that of the controls, and a significantly higher mean monocyte scatter was observed in sepsis patients than in controls.⁵² YJ Jung et al's research shows that all neutrophil light scatter parameters were lower in the children with viral infection relative to the normal controls, and SD_MALS_MO was higher in viral infection than tuberculosis.⁵³ DH Park reported that LALS of lymphocytes value has good sensitivity and specificity in the discrimination of fungemia from bacteremia.³² However, the exact pathological mechanisms of the light scatter changes seen among AP are not entirely clear. It can be confirmed that SIRS⁵ and a probability of infection³⁸ exist in the early stage of pancreatitis. Infection could cause a series of pathological change such as toxic vacuolization in cells to increase the complexity of cell internal structure, resulting in the increase of light scatter.⁵⁴ SIRS is closely related with immunological function change. Monocytes and lymphocytes are mainly types of cells in immune response,

	SAP				ICU transfer or death			
	AUC (95% CI)	Cutoff	Specificity	Sensitivity	AUC (95% CI)	Cutoff	Specificity	Sensitivity
SD_LALS_NE+MN_LALS_LY+SD_LMALS_ MO+SD_AL2_MO	0.87 (0.80; 0.93)	2.5	65.3%	96.8%	0.81 (0.72; 0.89)	2.5	79.5%	74.6%
MCTSI	0.72 (0.63; 0.82)	1.5	97.2%	48.4%	0.86 (0.80; 0.93)	3.0	61.5%	95.2%
SOFA	0.85 (0.78; 0.91)	3.5	54.2%	93.5%	0.86 (0.78; 0.93)	1.5	59.0%	96.8%
APACHE II	0.85 (0.78; 0.92)	5.0	83.3%	67.7%	0.88 (0.81; 0.95)	5.5	82.1%	85.7%
MMS	0.87 (0.80; 0.94)	4.5	90.3%	64.5%	0.87 (0.81; 0.94)	1.5	79.5%	76.2%
BISAP	0.80 (0.71; 0.89)	6.5	62.5%	87.1%	0.91 (0.85; 0.97)	1.5	89.7%	87.3%
Abbreviations: AL2, axial light loss; APACHE II, a	acute physiology and chrc	nic health eval	uation II; AUC, area	under the receiver	operating characteristic o	curve; BISAP, b	edside index for sev	erity in acute

pancreatitis; CI, confidence interval; CPD, cell population data; ICU, intensive care unit; LALS, low angle light scatter; LMALS, lower median angle light scatter; LY, lymphocyte; MCTSI, modified computed tomography severity index; MMS, modified Marshall score; MN, mean; MO, monocyte; NE, neutrophil; SAP, severe acute pancreatitis; SD, standard deviation; SOFA, sequential organ failure assessment.

ADEE TO COMPANISON OF MAGINOSILC ACCUR	acy of 4 CPD parallerers	scuring system		s ul Ar sevelity III	easured on uay I for pre		avul able coulse ul	ЧL
	SAP				ICU transfer or death			
	AUC (95% CI)	Cutoff	Specificity	Sensitivity	AUC (95% CI)	Cutoff	Specificity	Sensitivity
SD_LALS_NE+MN_LALS_LY+SD_LMALS_ MO+SD_AL2_MO	0.87 (0.80; 0.93)	2.5	65.3%	96.8%	0.81 (0.72; 0.89)	2.5	79.5%	74.6%
CRP	0.67 (0.56; 0.78)	216.5	72.2%	61.3%	0.76 (0.67; 0.86)	202.5	89.7%	58.7%
PCT	0.79 (0.69; 0.88)	1.1	76.1%	71.0%	0.79 (0.70; 0.88)	1.15	92.3%	57.1%
WBC	0.57 (0.44; 0.70)	9.2	72.2%	48.4%	0.51 (0.39; 0.62)	13.5	74.4%	36.5%
CRP+PCT+WBC	0.74 (0.64; 0.84)	0.3	73.2%	71.0%	0.83 (0.75; 0.91)	0.6	84.6%	69.8%

-÷ 2 ł L

Abbreviations: AL2, axial light loss; AUC, area under the receiver operating characteristic curve; Cl, confidence interval; CPD, cell population data; CRP, C-reactive protein (mg/L); ICU, intensive care unit; LALS, low angle light scatter; LMALS, lower median angle light scatter; LY, lymphocyte; MN, mean; MO, monocyte; NE, neutrophil; PCT, procalcitonin (ng/ml); SAP, severe acute pancreatitis; SD, standard deviation; WBC, white blood cell (*10⁹/L).

Wiley

TABLE 11 Onset time subgroup analysis of diagnostic accuracy of 4 CPD parameters' scoring system measured on day 1 for prediction of unfavorable course of AP

	SAP				ICU transfer or dea	ath		
Onset time	AUC (95% CI)	Cutoff	Specificity	Sensitivity	AUC (95% CI)	Cutoff	Specificity	Sensitivity
≤24 h	0.88 (0.80; 0.96)	2.5	72.5%	100.0%	0.82 (0.71; 0.93)	2.5	84.0%	75.8%
>24 h, ≤48 h	0.88 (0.76; 0.99)	1.5	85.7%	77.8%	0.77 (0.60; 0.95)	3.5	50.0%	90.0%
>48 h, ≤72 h	0.75 (0.47; 1.00)	2.5	72.7%	75.0%	0.84 (0.69; 0.99)	2.5	100.0%	60.0%

Abbreviations: AUC, area under the receiver operating characteristic curve; CI, confidence interval; CPD, cell population data; h, hour; ICU, intensive care unit; SAP, severe acute pancreatitis.

activation of monocytes and lymphocytes by SIRS, which can trigger cell death. During the process of cell death, signal transduction, nuclear, and protein synthesis will increase or decrease depending on different kind of genes, which accumulate to change on morphological level and lead to cell apoptosis.⁵⁵ And the light scatter can measure the morphology changes of the nuclear structure. Therefore, we speculate that these light scatter parameters may reflect the morphological changes of leukocyte in different types of infection and inflammation in the pathogenesis of SAP.

12 of 17

The combination of CPD parameters has been reported to indicate local bacterial infection in cancer patients.⁵⁴ In our study, the scoring system of 4 CPD parameters has the largest AUC compared with that of CRP, PCT, and WBC, also larger then MCTSI, SOFA, APACHE II, and BISAP scores. It could be clinically valuable not only because they show a good diagnostic accuracy on SAP identification but also are readily obtained by hematology analyzer during automated leukocyte sorting with no additional cost. Furthermore, this scoring system is easy to calculate manually or even faster by computer program automatically to trigger-specific test.

Additionally, it should be noted that SAP patients were not significantly higher in terms of BMI, age, pre-existing comorbidities, and triglyceride in our study. The association between these indicators and severity of AP was reported to be controversial. Some studies proved that age increases with AP severity,^{56,57} while other cohort studies reported no significant difference in age between non-SAP and SAP,⁵⁸⁻⁶⁰ which is also consistent with our current study and previous cohort study of 238 AP patients.⁶¹ Furthermore, central obesity, hypertriglyceridemia (HTG), hypertension (HT), and diabetes mellitus (DM), a set of comorbidities termed metabolic syndrome (MetS),⁶² were also reported ambiguous correlation with SAP.^{59,63} Explanation for these paradoxical findings can be varied. For example, previous opinion that BMI is a predictor of the severity of $AP^{64,65}$ was challenged, because BMI does not distinguish between truncal and visceral obesity,⁶⁴ leading to a weaker correlation with disease states not as strongly as intra-abdominal and visceral fat measurement like waist circumference.^{66,67} Alternatively, different etiologies of AP patients enrolled in study may also affect the conclusion. In the case of HTG, we did not find triglyceride increasing significantly in SAP, which was in line with that reported by Pothoulakis et al.⁶⁸ and Balachandra et al.⁶⁹ In contrast, others reported that the severity

of pancreatitis increases with elevated levels of triglycerides.^{70,71} This may because some of these studies only enrolled patients with hyperlipidemic AP or hyperlipidemia, but we enrolled AP patients with all common etiologies. Third, study design and the ability to control for confounding variables can be another reason. For the impact of DM on AP severity, a retrospective cohort study reported a higher risk of SAP in DM patients,⁷² whereas another retrospective cohort study suggested no significant difference on AP severity between those with and without DM by multivariate analysis.⁷³ Forth, few study has analyzed the effect of arterial HT on the severity of AP except Szentesi et al. reported it as an independent risk factor for severity.⁷⁴ Further investigation is needed. Finally, explanation for conflicting findings may be due to the variations among studies regarding race. Evidence showed that the effect of obesity on AP severity seemed to be worse in South Africa.⁷⁵ moderate in Mexico.⁷⁶ and least severe in Taiwan.⁷⁷

We acknowledge the limitations of this study. The main limitation of our study is the relative small sample size. We attempted to minimize these limitations by using strict inclusion and exclusion criteria, conducting the treatment for all patients by the same clinical team to avoid the bias. In addition, we used multivariate regression analysis to screen the diagnostic indicators of CPD statistically and excluded confounding factors. A prospective multicenter study is in warrant.

Another limitation is that we enrolled patients within 72 h of onset, for the course of the disease could change dynamically during the first three days, making an urgent need to diagnose disease severity in the early stage of AP. According to the 2012 Atlanta guideline, the early stage of AP is defined as the first week after abdominal pain. We enrolled AP patients with the median onset time of 24, 24, 30, and 24 h for overall, MAP, MSAP, and SAP, respectively. Furthermore, we performed subgroup analysis on our scoring system of CPD parameters and found comparable diagnostic efficiency among patients who admitted to hospital within 24, 48, and 72 h after onset for prediction of SAP (0.88, 0.88, and 0.75, respectively) and ICU transfer or death (0.82, 0.77, and 0.84, respectively).

In conclusion, the leukocyte CPD parameters that we studied, these objective, quantitative, and more sensitive parameters, can ultimately be incorporated into a predictive marker for the severity of acute pancreatitis.

ACKNOWLEDGMENTS

This study was supported by National Natural Science Foundation of China (No. 81600501) and National Natural Science Foundation Youth Project (No. 81601665).

CONFLICT OF INTEREST

The authors declare that they have no competing interests.

ETHICAL APPROVAL

The study was approved by the Institutional Ethics Committee of Ruijin Hospital. Written informed consent was obtained from each participant or their family members.

DATA AVAILABILITY STATEMENT

The data that support the findings of this study are available from the corresponding author upon reasonable request.

ORCID

Yihui Wang 🕩 https://orcid.org/0000-0001-5665-5557

REFERENCES

- Demcsák A, Soós A, Kincses L, et al. Acid suppression therapy, gastrointestinal bleeding and infection in acute pancreatitis – an international cohort study. *Pancreatology*. 2020;20(7):1323-1331.
- Paragomi P, Spagnolo DM, Breze CR, et al. Introduction and validation of a novel acute pancreatitis digital tool: interrogating large pooled data from 2 prospectively ascertained cohorts. *Pancreas*. 2020;49:1276–1282.
- Mederos MA, Reber HA, Girgis M. Acute pancreatitis: a review. JAMA. 2021;325(4):382-390.
- Boxhoorn L, Voermans RP, Bouwense SA, et al. Acute pancreatitis. Lancet. 2020;396(10252):726-734.
- Banks PA, Bollen TL, Dervenis C, et al. Classification of acute pancreatitis-2012: revision of the Atlanta classification and definitions by international consensus. *Gut.* 2012;62(1):102-111.
- Mikó A, Vigh É, Mátrai P, et al. Computed tomography severity index vs. other indices in the prediction of severity and mortality in acute pancreatitis: a predictive accuracy meta-analysis. Front Physiol. 2019;10:1002.
- Farkas N, Hanák L, Mikó A, et al. A multicenter, international cohort analysis of 1435 cases to support clinical trial design in acute pancreatitis. *Front Physiol*. 2019;10:1092.
- Tian F, Li H, Wang L, et al. The diagnostic value of serum Creactive protein, procalcitonin, interleukin-6 and lactate dehydrogenase in patients with severe acute pancreatitis. *Clin Chim Acta*. 2020;510:665-670.
- Gupta S, Shekhawat V, Kaushik G. D-dimer, a potential marker for the prediction of severity of acute pancreatitis. *Clin Lab.* 2015;61(9):1187-1195.
- Fisic E, Poropat G, Bilic-Zulle L, Licul V, Milic S, Stimac D. The Role of IL-6, 8, and 10, sTNFr, CRP, and pancreatic elastase in the prediction of systemic complications in patients with acute pancreatitis. *Gastroenterol Res Pract*. 2013;2013(2):1-6.
- Urrechaga E, Bóveda O, Aguirre U, García S, Pulido E. Neutrophil cell population data biomarkers for acute bacterial infection. *J Pathol Infect Dis.* 2018;1(1):1–7.
- Urrechaga E. Reviewing the value of leukocytes cell population data (CPD) in the management of sepsis. Ann Transl Med. 2020;8(15):953-963.

- Zhu Y, Cao X, Tao G, Xie W, Hu Z, Xu D. The lymph index: a potential hematological parameter for viral infection. *Int J Infect Dis.* 2013;17(7):e490-e493.
- 14. Buoro S, Manenti B, Seghezzi M, et al. Abnormal scattergrams and cell population data generated by fully automated hematological analyzers: new tools for screening malaria infection? *Int J Lab Hematol.* 2018;40(3):326-334.
- 15. Zeeshanhaider R, Shamsi TS, Ujjan ID. Cell population data-driven acute promyelocytic leukemia flagging through artificial neural network predictive modeling. *Transl Oncol.* 2020;13(1):11-16.
- Hong S, Jie Z, Yunhua L. The clinical application of neutrophil VCS parameters in the detection of bacterial infection and stringency diseases. *Lab Med.* 2012;27(12):1027-1030.
- Liu L, Wang C, Luo T, Li L. Effects of fluid resuscitation on organ failure and mortality in patients with acute pancreatitis and systemic inflammatory response syndrome- a retrospective cohort study. *Biomed J Sci Tech Res.* 2019;22(5):17021-17029.
- 18. Wilson J, Zarabi S. SIRS criteria as a way of predicting mortality in acute pancreatitis. *Emerg Med J.* 2017;34(9):621-622.
- Kumar A, Chari ST, Vege SS. Can the time course of systemic inflammatory response syndrome score predict future organ failure in acute pancreatitis? *Pancreas*. 2014;43(7):1101-1105.
- Johnson CD, Besselink MG, Carter R. Acute pancreatitis. BMJ. 2014;349(3):601-608.
- Chen Y, Ke L, Tong Z, Li W, Li J. Association between severity and the determinant-based classification, Atlanta 2012 and Atlanta 1992, in acute pancreatitis: a clinical retrospective study. *Medicine*. 2015;94(13):e638.
- Zhao J, Liao Q, Zhao Y, Hu Y. Mortality indicators and risk factors for intra-abdominal hypertension in severe acute pancreatitis. *Int Surg.* 2014;99(3):252-257.
- Gougol A, Paragomi P, Pothoulakis I, Talukdar R, Papachristou GI. Temporal relationship between SIRS, organ failure, and death in acute pancreatitis: data from a large, multicenter, international study (APPRENTICE Study Group). *Pancreas*. 2018;47(10):1389-1393.
- 24. van Geenen EJM, van der Peet DL, Bhagirath P, Mulder CJJ, Bruno MJ. Etiology and diagnosis of acute biliary pancreatitis. *Nat Rev Gastroenterol Hepatol.* 2010;7(9):495-502.
- Scherer J, Singh VP, Pitchumoni CS, Yadav D. Issues in hypertriglyceridemic pancreatitis: an update. J Clin Gastroenterol. 2014;48(3):195-203.
- Lee P, Papachristou G. New insights into acute pancreatitis. Nat Rev Gastroenterol Hepatol. 2019;16(8):479-496.
- Mao E. Intensive management of severe acute pancreatitis. Ann Transl Med. 2019;7(22):687.
- Micha L, GraYna R. Immature granulocytes predict severe acute pancreatitis independently of systemic inflammatory response syndrome. *Pancreatology*. 2017;12:140-144.
- Choudhuri A, Duggal S, Biswas P, Uppal R. A comparison of acute physiology and chronic health evaluation II score and serum procalcitonin change for predicting mortality in acute pancreatitis. *Indian J Crit Care Med.* 2020;24(3):190-194.
- Lipinski M, Rydzewska-Rosolowska A, Rydzewski A, Cicha M, Rydzewska G. Soluble urokinase-type plasminogen activator receptor (suPAR) in patients with acute pancreatitis (AP) - progress in prediction of AP severity. *Pancreatology*. 2016;17(1):24-29.
- Wang Y, Fuentes HE, Attar BM, Jaiswal P, Demetria M. Evaluation of the prognostic value of neutrophil to lymphocyte ratio in patients with hypertriglyceridemia-induced acute pancreatitis. *Pancreatology*. 2017;17(6):893-897.
- Park D-H, Park K, Park J, et al. Screening of sepsis using leukocyte cell population data from the Coulter automatic blood cell analyzer DxH800. *Int J Lab Hematol.* 2011;33(4):391-399.

^{14 of 17} WILEY

- Crouser ED, Parrillo JE, Seymour CW, et al. Monocyte distribution width: a novel indicator of sepsis-2 and sepsis-3 in high-risk emergency department patients. *Crit Care Med.* 2019;47(8):1018-1025.
- Sun T, Li J, Wu B, et al. Effects of blood storage on cell population data. Clin Lab. 2020;66:1501–1508.
- Crouser ED, Parrillo J, Martin GS, Huang DT, Tejidor L. Monocyte distribution width enhances early sepsis detection in the emergency department beyond SIRS. J Intens Care. 2020;8:33.
- Agnello L, Bivona G, Vidali M, et al. Monocyte distribution width (MDW) as a screening tool for sepsis in the Emergency Department. *Clin Chem Lab Med.* 2020;58(11):1951-1957.
- 37. American College of Chest Physicians/Society of Critical Care Medicine Consensus Conference. Definitions for sepsis and organ failure and guidelines for the use of innovative therapies in sepsis. *Crit Care Med.* 1992;20(6):864-874.
- Leppäniemi A, Tolonen M, Tarasconi A, et al. 2019 WSES guidelines for the management of severe acute pancreatitis. World J Emerg Surg. 2019;14(1):27.
- Kim MJ, Cheng G, Agrawal DK. Cl- channels are expressed in human normal monocytes: a functional role in migration, adhesion and volume change. *Clin Exp Immunol*. 2010;138(3):453-459.
- Leckie MJ. Automated quantitation of circulating neutrophil and eosinophil activation in asthmatic patients. *Thorax*. 2000;55(6):471-477.
- 41. Mifsud EJ, Tan ACL, Jackson DC. TLR agonists as modulators of the innate immune response and their potential as agents against infectious disease. *Front Immunol.* 2014;5:79.
- Mukherjee R, Kanti Barman P, Kumar Thatoi P, Tripathy R, Kumar Das B, Ravindran B. Non-classical monocytes display inflammatory features: validation in sepsis and systemic lupus erythematous. *Sci Rep.* 2015;5:13886.
- Henderson R, Hobbs J, Mathies M, Hogg N. Rapid recruitment of inflammatory monocytes is independent of neutrophil migration. *Blood*. 2003;102(1):328-335.
- Italiani P, Boraschi D. From monocytes to M1/M2 macrophages: phenotypical vs. Functional Differentiation. *Frontiers Immunol*. 2014;5(514):1-22.
- 45. Xu D. Clinical applications of leukocyte morphological parameters. Int J Pathol Clin Res. 2015;1:1-12.
- Mccullough KC, Basta S, Knötig S, et al. Intermediate stages in monocyte-macrophage differentiation modulate phenotype and susceptibility to virus infection. *Immunology*. 2010;98(2):203-212.
- Wang SY, Mak KL, Chen LY, Chou MP, Ho CK. Heterogeneity of human blood monocyte: two subpopulations with different sizes, phenotypes and functions. *Immunology*. 1992;77(2):298-303.
- Zhu Y, Cao X, Chen Y, et al. Neutrophil cell population data: useful indicators for postsurgical bacterial infection. *Int J Lab Hematol.* 2012;34(3):295-299.
- Bhargava M, Saluja S, Sindhuri U, Saraf A, Sharma P. Elevated mean neutrophil volume+CRP is a highly sensitive and specific predictor of neonatal sepsis. *Int J Lab Hematol.* 2014;36(1):e11-e14.
- Tang H, Jing J, Bo D, Xu D. Biological variations of leukocyte numerical and morphologic parameters determined by UniCel DxH 800 hematology analyzer. Arch Pathol Lab Med. 2012;136(11):1392-1396.
- 51. Fernando C, Bethany T, Dongsheng XU. Quantitative determination of neutrophil VCS parameters by the coulter automated hematology analyzer. *Am J Clin Pathol.* 2005;124(3):440-444.
- Arora P, Gupta PK, Lingaiah R, Mukhopadhyay AK. Volume, conductivity, and scatter parameters of leukocytes as early markers of sepsis and treatment response. J Lab Physicians. 2019;11(1):29-33.
- Jung Y-J, Kim J-H, Park Y-J, et al. Evaluation of cell population data on the UniCel DxH 800 Coulter Cellular Analysis system as a screening for viral infection in children. *Int J Lab Hematol.* 2012;34(3):283-289.

- 54. Li D, Zhou N, Liu L, Zeng Q, Song X. VCS parameters of neutrophils, monocytes and lymphocytes may indicate local bacterial infection in cancer patients who accepted cytotoxic chemotherapeutics. *Eur J Clin Microbiol Infect Dis.* 2016;35(1):41-48.
- 55. Shao R, Fang Y, Yu H, Zhao L, Jiang Z, Li CS. Monocyte programmed death ligand-1 expression after 3–4 days of sepsis is associated with risk stratification and mortality in septic patients: a prospective cohort study. *Crit Care.* 2016;20:3–4.
- 56. Márta K, Lazarescu A-M, Farkas N, et al. Aging and comorbidities in acute pancreatitis i: a meta-analysis and systematic review based on 194,702 patients. *Front Physiol*. 2019;10:328.
- Szakács Z, Gede N, Pécsi D, et al. Aging and comorbidities in acute pancreatitis II.: a cohort-analysis of 1203 prospectively collected cases. Front Physiol. 2019;9:1776.
- Zhang QI, Li LE, Chen H, et al. Soluble urokinase plasminogen activator receptor associates with higher risk, advanced disease severity as well as inflammation, and might serve as a prognostic biomarker of severe acute pancreatitis. J Clin Lab Anal. 2020;34(3). e23097
- Sawalhi S, Al-Maramhy H, Abdelrahman A, Allah S, Al-Jubori S. Does the presence of obesity and/or metabolic syndrome affect the course of acute pancreatitis?: A prospective study. *Pancreas*. 2014;43(4):565-570.
- 60. Wu Q, Zhong XI, Fu M, et al. High-density lipoprotein cholesterol to low-density lipoprotein cholesterol ratio in early assessment of disease severity and outcome in patients with acute pancreatitis admitted to the ICU. *BMC Gastroenterol.* 2020;20(1):164.
- 61. Zhao B, Sun S, Wang Y, et al. Cardiac indicator CK-MB might be a predictive marker for severity and organ failure development of acute pancreatitis. *Ann Transl Med.* 2021;9(5):368.
- 62. Alberti K, Eckel RH, Grundy SM, et al. Harmonizing the metabolic syndrome: a joint interim statement of the International Diabetes Federation Task Force on Epidemiology and Prevention; National Heart, Lung, and Blood Institute; American Heart Association; World Heart Federation; International Atherosclerosis Society; and International Association for the Study of Obesity. *Circulation*. 2009;120(16):1640-1645.
- Mikolasevic I, Milic S, Orlic L, et al. Metabolic syndrome and acute pancreatitis. *Eur J Intern Med*. 2016;32:79-83.
- Li LK, Dennison AR, Garcea G. Association of visceral adipose tissue on the incidence and severity of acute pancreatitis: a systematic review. *Pancreatology*. 2020;20:1056-1061.
- 65. Dobszai D, Mátrai P, Gyöngyi Z, et al. Body-mass index correlates with severity and mortality in acute pancreatitis: a meta-analysis. *World J Gastroenterol*. 2019;25(6):729-743.
- Després J. Health consequences of visceral obesity. Ann Med. 2001;33(8):534-541.
- Goodger R, Asrani V, Windsor J, Petrov M. Impact of metabolic comorbidities on outcomes of patients with acute pancreatitis: a scoping review. *Panminerva Med.* 2016;58(1):86-93.
- Pothoulakis I, Paragomi P, Archibugi L, et al. Clinical features of hypertriglyceridemia-induced acute pancreatitis in an international, multicenter, prospective cohort (APPRENTICE consortium). *Pancreatology*. 2020;20(3):325-330.
- Balachandra S, Virlos IT, King NKK, Siriwardana HPP, France MW, Siriwardena AK. Hyperlipidaemia and outcome in acute pancreatitis. Int J Clin Pract. 2010;60(2):156-159.
- Olesen S, Harakow A, Krogh K, Drewes A, Handberg A, Christensen P. Hypertriglyceridemia is often under recognized as an aetiologic risk factor for acute pancreatitis: a population-based cohort study. *Pancreatology*. 2021;21(2):334-341.
- Mosztbacher D, Hanák L, Farkas N, et al. Hypertriglyceridemiainduced acute pancreatitis: a prospective, multicenter, international cohort analysis of 716 acute pancreatitis cases. *Pancreatology*. 2020;20(4):608-616.

- 72. Shen H, Lu C, Li C. Effect of diabetes on severity and hospital mortality in patients with acute pancreatitis: a national populationbased study. *Diabetes Care*. 2012;35:1061-1066.
- Nawaz H, O'Connell M, Papachristou G, Yadav D. Severity and natural history of acute pancreatitis in diabetic patients. *Pancreatology*. 2015;15(3):247-252.
- 74. Szentesi A, Párniczky A, Vincze Á, et al. Multiple hits in acute pancreatitis: components of metabolic syndrome synergize each other's deteriorating effects. *Front Physiol*. 2019;10:1202.
- Funnell IC, Bornman PC, Weakley SP, Terblanche J, Marks IN. Obesity: an important prognostic factor in acute pancreatitis. Br J Surg. 1993;80(4):484-486.
- Suazo-Baráhona J, Carmona-Sánchez R, Robles-Díaz G, et al. Obesity: a risk factor for severe acute biliary and alcoholic pancreatitis. *Am J Gastroenterol.* 1998;93(8):1324-1328.
- 77. Tsai CJ. Is obesity a significant prognostic factor in acute pancreatitis? *Dig Dis Sci.* 1998;43(10):2251-2254.

SUPPORTING INFORMATION

Additional supporting information may be found online in the Supporting Information section.

How to cite this article: Wang Y, Xu Z, Zhou Y, et al. Leukocyte cell population data from the blood cell analyzer as a predictive marker for severity of acute pancreatitis. *J Clin Lab Anal*. 2021;35:e23863. <u>https://doi.org/10.1002/</u> jcla.23863