

## Research Article

# Pharmacological Mechanism of Danggui-Sini Formula for Intervertebral Disc Degeneration: A Network Pharmacology Study

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Received 12 September 2021; Accepted 19 October 2021; Published 11 November 2021

Academic Editor: Pei Li

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**Background.** Intervertebral disc degeneration (IVDD) is the most significant cause of low back pain, the sixth-largest disease burden globally, and the leading cause of disability. This study is aimed at investigating the molecular biological mechanism of Danggui-Sini formula (DSF) mediated IVDD treatment. **Methods.** A potential gene set for DSF treatment of IVDD was identified through TCMSp, UniProt, and five disease gene databases. A protein interaction network of common targets between DSF and IVDD was established by using the STRING database. GO and KEGG enrichment analyses were performed using the R platform to discover the potential mechanism. Moreover, AutoDock Vina was used to verify molecular docking and calculate the binding energy. **Results.** A total of 119 active ingredients and 136 common genes were identified, including 10 core genes (AKT1, IL6, ALB, TNF, VEGFA, TP53, MAPK3, CASP3, JUN, and EGF). Enrichment analysis results showed that the therapeutic targets of DSF for diseases mainly focused on the AGE-RAGE signaling pathway involved in diabetic complications, IL-17 signaling pathway, TNF signaling pathway, Toll-like receptor signaling pathway, apoptosis, cellular senescence, PI3K-Akt signaling pathway, and FoxO signaling pathway. These biological processes are induced mainly in response to oxidative stress and reactive oxygen species and the regulation of apoptotic signaling pathways. Molecular docking showed that there was a stable affinity between the core genes and the key components. **Conclusions.** The combination of network pharmacology and molecular docking provides a practical way to analyze the molecular biological mechanism of DSF-mediated IVDD treatment, which confirms the “multicomponent, multitarget and multipathway” characteristics of DSF and provides an essential theoretical basis for clinical practice.

## 1. Introduction

Low back pain is the sixth largest disease burden globally and is the leading cause of disability, which has become a global health concern [1]. A previous study showed that 40% of low back pain is related to intervertebral disk degeneration (IVDD) [2]. The major pathological features of IVDD are the elevated expression of inflammatory mediators, increased senescence and apoptosis of nucleus pulposus cells (NPCs), and degradation of the extracellular matrix [3, 4]. Importantly, IVDD may bring about a range of clinical symptoms, such as pain, numbness, and even paralysis of the lower

extremities, causing enormous suffering and economic burden.

The initial clinical treatment for IVDD has focused on symptom relief, mainly with nonsteroidal anti-inflammatory drugs (NSAIDs) [5, 6]. However, studies on the use of NSAIDs in articular cartilage have shown detrimental effects [7–9]. Therefore, there is an urgent need for an effective and safe drug. Traditional Chinese medicine (TCM) has been used to treat various diseases for thousands of years [10]. However, due to the diversity of TCM components and the complexity of the body, systematic research on TCM has been limited.

Network pharmacology is a strategy for elucidating the synergistic effects and potential mechanism of multicomponent and multitarget drugs based on various complex “drug-ingredient-target gene” networks [11]. It may be a potential tool to systematically explore new applications for TCM. Recent studies demonstrated that network pharmacology could achieve high performance in the prediction of the mechanisms of TCM involved in treating diseases [12, 13]. We hypothesize that combining gene ontology (GO) and Kyoto Encyclopedia of Genes and Genomes (KEGG) analysis with network pharmacology may provide more valuable and complementary information, thereby further improving the prediction performance of potentially effective mechanisms.

The Danggui-Sini formula (DSF) is a classic prescription in the Treatise on Febrile Diseases and is mainly composed of seven herbs (Table 1): *Angelica sinensis* (Dang Gui, DG), *Cinnamomum cassia Presl* (Gui Zhi, GZ), *Cynanchum otophyllum Schneid* (Bai Shao, BS), *Asarum sieboldii Miq* (Xi Xin, XX), *Glycyrrhiza uralensis Fisch* (Gan Cao, GC), *Tetrapanax papyrifera* (Tong Cao, TC), and *Ziziphus jujuba Mill* (Da Zao, DZ). In recent years, pharmacological studies have shown that this decoction has positive anti-inflammatory and antioxidative effects, especially in the treatment of osteoarthritis [12, 13]. Although Chinese medicine has already used DSF to treat IVDD, its mechanism of action needs to be further studied [14].

This study integrated network pharmacology and molecular docking to explore the potentially effective mechanism and targets of DSF acting on IVDD. The research process is shown in Figure 1. The results highlight the potential of DSF in the treatment of IVDD.

## 2. Materials and Methods

**2.1. Active Ingredients and Target Genes of the Danggui-Sini Formula.** By searching the Traditional Chinese Medicine Systems Pharmacology (TCMSP) database (<https://tcmsp-e.com/>) [15], with the filter conditions of oral bioavailability (OB)  $\geq 30\%$  and drug likeness (DL)  $\geq 0.18$ , the active ingredients of each herb from DSF were acquired. At the same time, the corresponding target protein information of the active compounds was obtained through the DrugBank database (<https://go.drugbank.com/>) [16], and the conversion of gene symbols was completed through the UniProt database (<https://www.UniProt.org>) [17].

**2.2. Related Targets of IVDD and Common Gene Set.** To obtain the targets of IVDD, we searched a total of five databases, namely, Online Mendelian Inheritance in Man (OMIM) (<https://omim.org/>) [18], Genecards database (<https://www.genecards.org/>) [19], Comparative Toxicogenomics Database (CTD) (<http://ctdbase.org/>) [20], DrugBank database (<https://go.drugbank.com/>) [16], and DisGeNet database (<https://www.disgenet.org/>) [21]. In addition, all targets were standardized in the UniProt database [17]. A common gene set of “compound-targets-disease” comprises the potential target genes of DSF for IVDD and was built by creating a Venn diagram.

TABLE 1: Full scientific species names of herbs of Danggui-Sini formula.

Pin Yin	Latin name
Dang Gui (DG)	<i>Angelica sinensis</i>
Gui Zhi (GZ)	<i>Cinnamomum cassia Presl</i>
Bai Shao (BS)	<i>Cynanchum otophyllum Schneid</i>
Xi Xin (XX)	<i>Asarum sieboldii Miq</i>
Gan Cao (GC)	<i>Glycyrrhiza uralensis Fisch</i>
Tong Cao (TC)	<i>Tetrapanax papyrifera</i>
Dao Zao (DZ)	<i>Ziziphus jujuba Mill</i>

**2.3. Network Visualization and Enrichment Analysis.** “Active ingredients - potential targets” network and another related network for disease-core genes–active ingredients–herbs were established using the Cytoscape software [22]. To obtain more accurate gene annotation results, we utilized the clusterProfiler package of the R platform for gene ontology (GO) and Kyoto Encyclopedia of Genes and Genomes (KEGG) functional enrichment analysis [23]. GO enrichment analysis revealed the underlying mechanisms from biological processes (BPs), cell components (CCs), and molecular functions (MFs), while KEGG is a pathway-related database.

**2.4. Protein–Protein Interaction (PPI) Network Construction and Core Gene Selection.** We imported the common genes into the STRING database (<https://www.string-db.org/>) [24] to obtain the PPI network and TSV format file with the parameters of Organism = Homo sapiens and moderate confidence = 0.400. Then, TSV files were imported into the software to realize PPI network visualization and further analysis. The CytoHubba plug-in was used to identify the top 10 core genes by using 12 kinds of topological measures.

**2.5. Molecular Docking.** Molecular docking analysis focused on the proteins encoded by the 10 core genes and their corresponding active components. First, we downloaded the 3D structure of the protein from the RCSB PDB database (<https://www.rcsb.org/>) and used the PyMol 2.4.0 software to remove water and natural ligands from the protein. Second, the 2D structures of active components (small molecular ligands) were downloaded from the PubChem database (<https://pubchem.ncbi.nlm.nih.gov/>). Then, we used the ChemBio3D software to convert the molecular ligand into a 3D structure and performed energy minimization by the MM2 calculation method. Hydrogenation of proteins and small molecular ligands was prepared with the Autodock tool. Finally, Autodock Vina was used for molecular docking and calculating the binding energy [25].

## 3. Results

**3.1. Active Ingredients in DSF and Potential Genes.** Seven herbal medicines of the DSF were successively inputted into the TCMSP database, and a total of 155 active ingredients were obtained, among which 2 were from DG, 7 were from

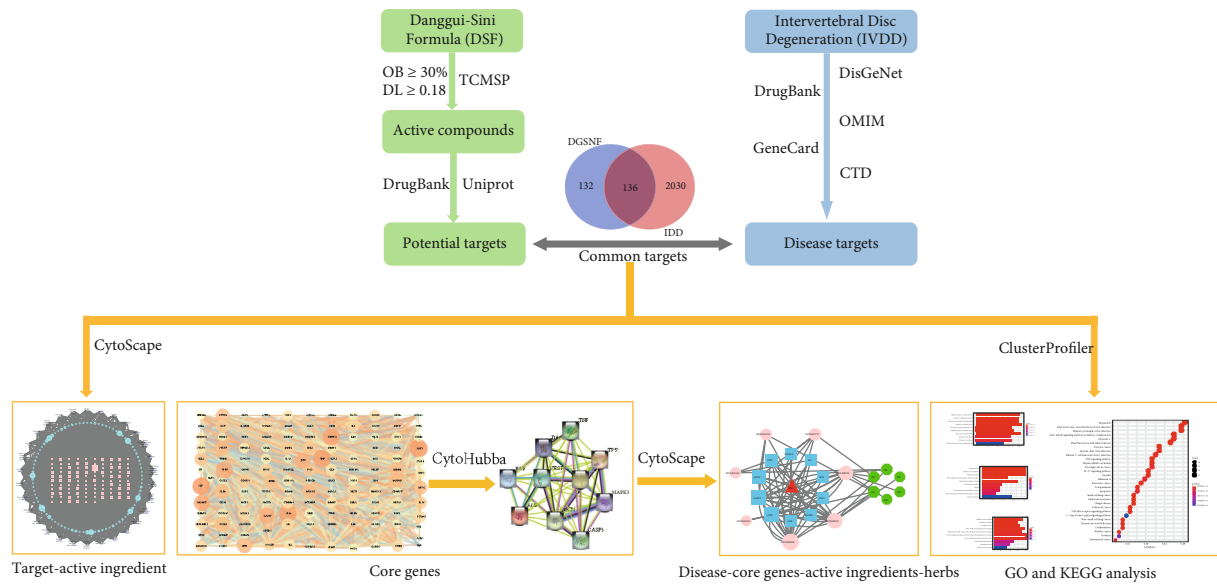


FIGURE 1: Flowchart to explore the possible mechanisms of the Danggui-Sini formula (DSF) for intervertebral disc degeneration (IVDD).

GZ, 13 were from BS, 8 were from XX, 92 were from GC, 4 were from TC, and 29 were from DZ (Supplementary Table 1). Then, we searched the DrugBank database for targets of each active ingredient and obtained a total of 268 genes after removing duplicates. All gene names were corrected through the UniProt database. Details of the above information are listed in Supplementary Table 2.

With the keyword “intervertebral disc degeneration,” we searched the five databases mentioned above and obtained a total of 2166 genes after deduplication. Using an online web service (<http://bioinformatics.psb.ugent.be/webtools/Venn/>), we obtained common genes that intersect between drug-target and disease-related genes. One hundred thirty-six genes were identified as potential genes for DSGF in the treatment of IVDD, and the Venn diagram is shown in Figure 2 and Supplementary Table 3.

**3.2. Active Ingredient-Potential Target Network.** After obtaining the common genes and their corresponding active compounds, a network of active ingredient-potential targets was drawn by using Cytoscape (Figure 3). The network has 243 nodes and 1214 edges. Furthermore, we found that one gene corresponds to multiple active ingredients and vice versa. According to the number of target nodes, we ranked the active ingredients, with quercetin (MOL000098) targeting the most genes.

**3.3. PPI Network and Core Genes.** To understand the relationship among common genes and obtain core genes, 136 common genes were first imported into the STRING database, and then, the PPI network and TSV format file were obtained (Figure 4(a)). Second, we visualized the PPI network by importing it into Cytoscape and identified the 10 core genes using Cytohubba plug-ins with 12 kinds of topological sorting. Finally, we found that the top 10 core genes

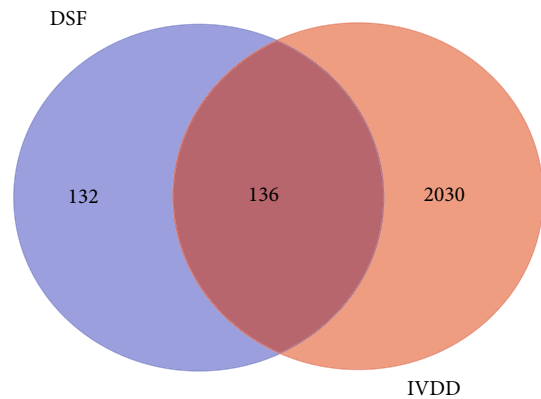


FIGURE 2: Venn diagram for DSGF-related genes and IVDD-related genes.

obtained by the two methods of maximum neighborhood component (MNC) and degree were the same, namely, AKT1, IL6, ALB, TNF, VEGFA, TP53, MAPK3, CASP3, JUN, and EGF (Figure 4(b)). We performed further cluster analysis of the PPI network with the MCODE plug-in and acquired 5 clusters (Figure 4(c), Table 2). The top 1 cluster contained 10 core genes, which further confirmed the criticality of core genes.

**3.4. GO and KEGG Analysis.** To further explore the interaction between common target genes and the mechanism by which DSGF may treat IVDD, GO functional analysis and KEGG pathway enrichment analysis were performed using the R platform.

A total of 2617 items were acquired: 2417 from BPs, 60 from CCs, and 140 from MFs (Supplementary Table 4). The top 10 of each category are shown in Figure 5. The results showed that the biological processes of the potential genes are

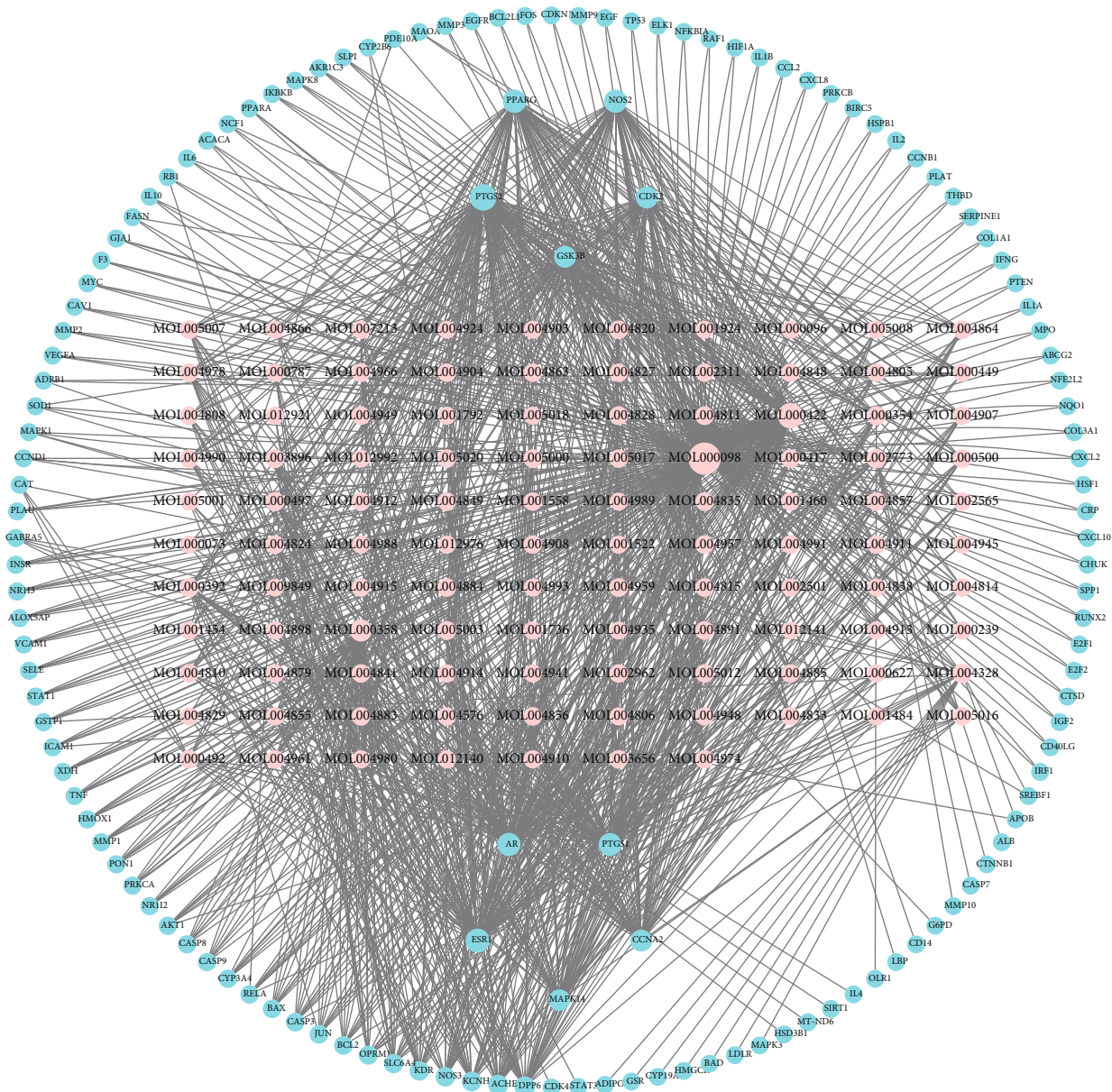


FIGURE 3: Construction of a common gene-active ingredient network. Blue nodes represent the common targets of IVDD and DSF; pink nodes represent the active ingredients related to the common targets. The line between two nodes represents the interaction; the size of each node represents the number of connections.

associated with oxidative stress, such as response to oxidative stress, response to reactive oxygen species, cellular response to oxidative stress, and cellular response to reactive oxygen species. The cellular component might be mainly activated in the membrane raft, membrane microdomain, membrane region, RNA polymerase II, transcription regulator complex, and transcription regulator complex. The molecular functions of common genes include cytokine receptor binding, phosphatase binding, cytokine activity, DNA binding, transcription factor binding, and RNA polymerase II-specific DNA-binding transcription factor binding.

Simultaneously, we obtained 166 KEGG pathways associated with potential genes (Supplement Table 4), of which

the top 30 are shown in Figure 5. The meaningful pathways contain the AGE-RAGE signaling pathway involved in diabetic complications, the IL-17 signaling pathway, the TNF signaling pathway, the Toll-like receptor signaling pathway, and apoptosis. Although cellular senescence, the PI3K-Akt signaling pathway, and the FoxO signaling pathway are not shown in the diagram, they were also selected for further analysis because they may be related to inflammation, apoptosis, senescence, and autophagy. The results confirmed that DSF alleviated IVDD disease by regulating antioxidant stress and inflammatory reactions. The top 30 pathway-common target-active ingredient networks are shown in Figure 6.

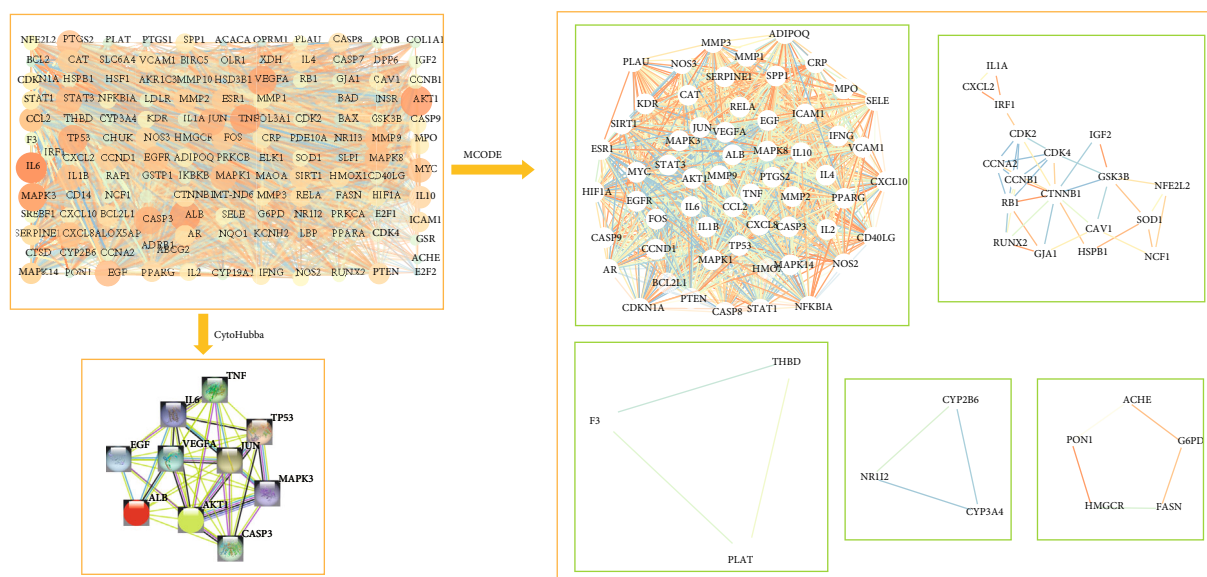


FIGURE 4: Protein–protein interaction (PPI) network and core genes: (a) the protein–protein interaction (PPI) network of potential targets of DSF in the treatment of IVDD; (b) PPI network of the core genes; (c) cluster analysis of the common targets.

TABLE 2: Cluster information of the protein-protein interaction (PPI) network for common genes.

Cluster	Score	Nodes	Edges	Gene symbol
1	48.483	59	1406	IL10, MAPK1, NFKBIA, MAPK14, CXCL10, MMP1, CASP9, CASP8, STAT1, CD40LG, PLAU, RELA, CDKN1A, HIF1A, CCND1, <i>MAPK3</i> *, BCL2L1, AR, STAT3, <i>CASP3</i> *, KDR, <i>VEGFA</i> *, <i>ALB</i> *, <i>TP53</i> *, SPP1, CXCL8, PPARG, <i>JUN</i> *, EGFR, PTEN, ESRI, MYC, <i>EGF</i> *, <i>AKT1</i> *, SIRT1, ADIPOQ, MPO, CAT, IL1B, <i>TNF</i> *, FOS, PTGS2, <i>IL6</i> *, HMOX1, MMP2, SERPINE1, CCL2, IL2, IFNG, IL4, CRP, ICAM1, VCAM1, NOS3, MMP3, NOS2, SELE, MMP9, MAPK8
2	4.706	18	40	IGF2, SOD1, CXCL2, NFE2L2, IL1A, IRF1, HSPB1, CCNB1, CDK4, CDK2, RB1, CCNA2, NCF1, GSK3B, CAV1, GJA1, CTNNB1, RUNX2
3	3	3	3	PLAT, THBD, F3
4	3	3	3	NRI12, CYP3A4, CYP2B6
5	2.5	5	5	G6PD, HMGCR, FASN, ACHE, PON1

\*Core genes are shown in italic.

3.5. *Disease-Core Genes–Active Ingredients–Herbs*. Based on the 10 core genes, we obtained seven active ingredients and six herbs. A network of disease core gene–active ingredient–herb interactions is shown in Figure 7 with 24 nodes and 70 edges. Quercetin (MOL000098) is the most crucial active ingredient with the largest degree, which is consistent with the results of the active ingredient-potential target network. JUN and CASP3 were the key genes with the highest degrees.

3.6. *Molecular Docking*. Ten key genes were selected for molecular docking with seven major active compounds to verify the binding ability between key active compounds and core gene interactions. Studies have shown that the lower the binding energy, the more stable the conformation of the compound binding to the protein, and the greater the possibility of interaction [26]. The docking results are shown in the heat map (Figures 8 and 9). It is generally believed that the binding energy between proteins and small molecule

compounds is relatively stable when it is less than -5.0 kcal/mol [26]. As predicted by network pharmacology, our binding energy results were all less than -5.0 kcal/mol, indicating that all active compounds could easily enter the active pocket of the protein and bind stably. In addition, all compounds could bind to multiple targets simultaneously, suggesting the mechanism of the multitarget role of decoction in the treatment of intervertebral disc degeneration. In addition, all compounds can bind to multiple targets simultaneously, suggesting the mechanism of the multitarget role of DSF in the treatment of IVDD.

#### 4. Discussion

IVDD is one of the common causes of low back pain. At present, the incidence of IVDD is increasing, which seriously aggravates patients’ mental and financial burden [27, 28]. Previous evidence has shown that the occurrence and development of IVDD are related to inflammation and oxidative

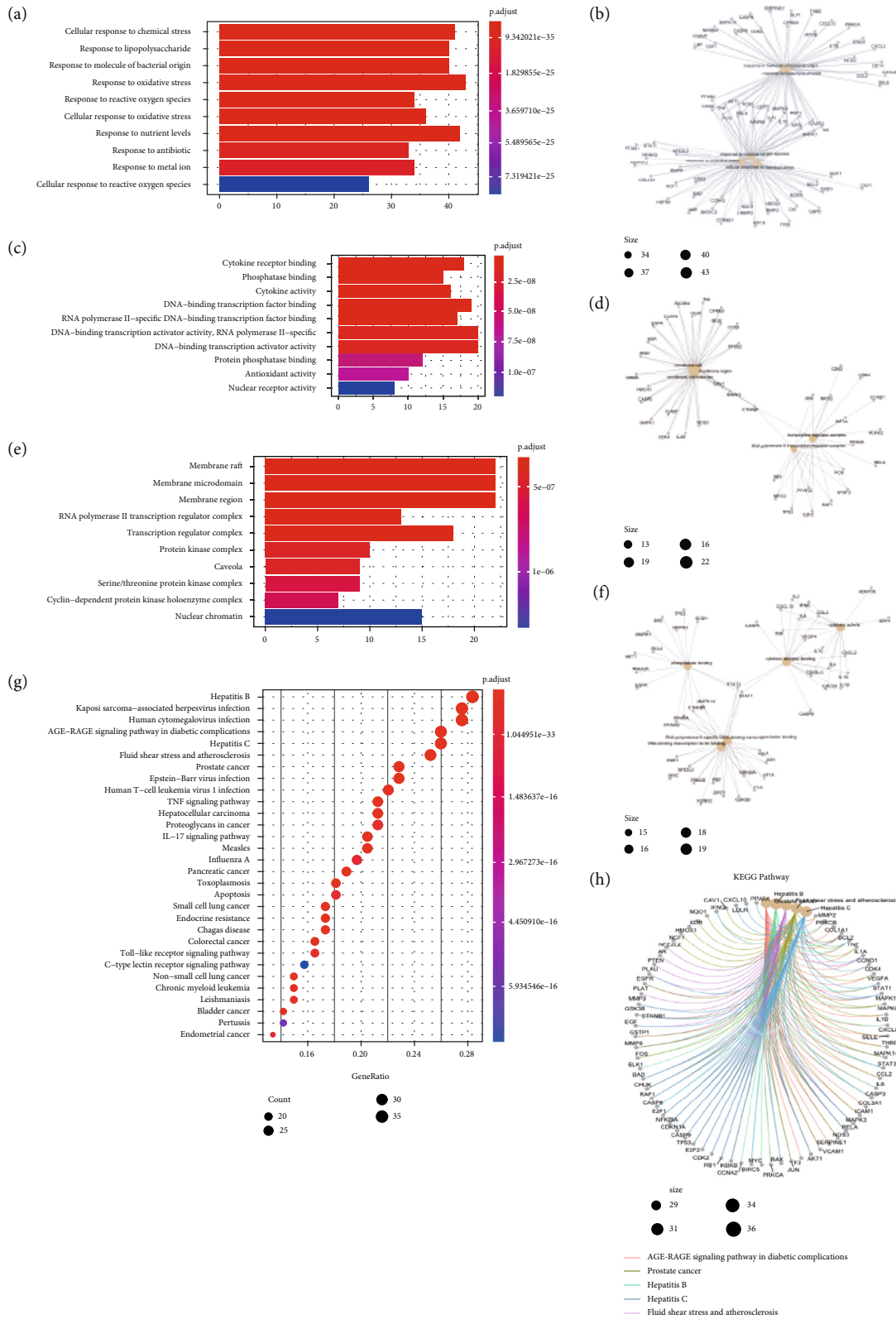


FIGURE 5: GO (including BP, MF, and CC) and KEGG analysis of common genes. (a) Top 10 significantly enriched terms in biological processes (BPs). (b) Subnetwork showing the top five BP terms and related genes. (c) Top 10 significantly enriched terms in cellular components (CCs). (d) Subnetwork showing the top five CC terms and related genes. (e) Top 10 significantly enriched terms in molecular functions (MFs). (f) Subnetwork showing the top five MF terms and related genes. (g) The 30 pathways with the lowest adjusted *p* values. The darker the color, the smaller the adjusted *p* value. The larger the circle, the greater the number of target genes in the term. (h) Subnetwork showing the top five KEGG pathways and related genes.

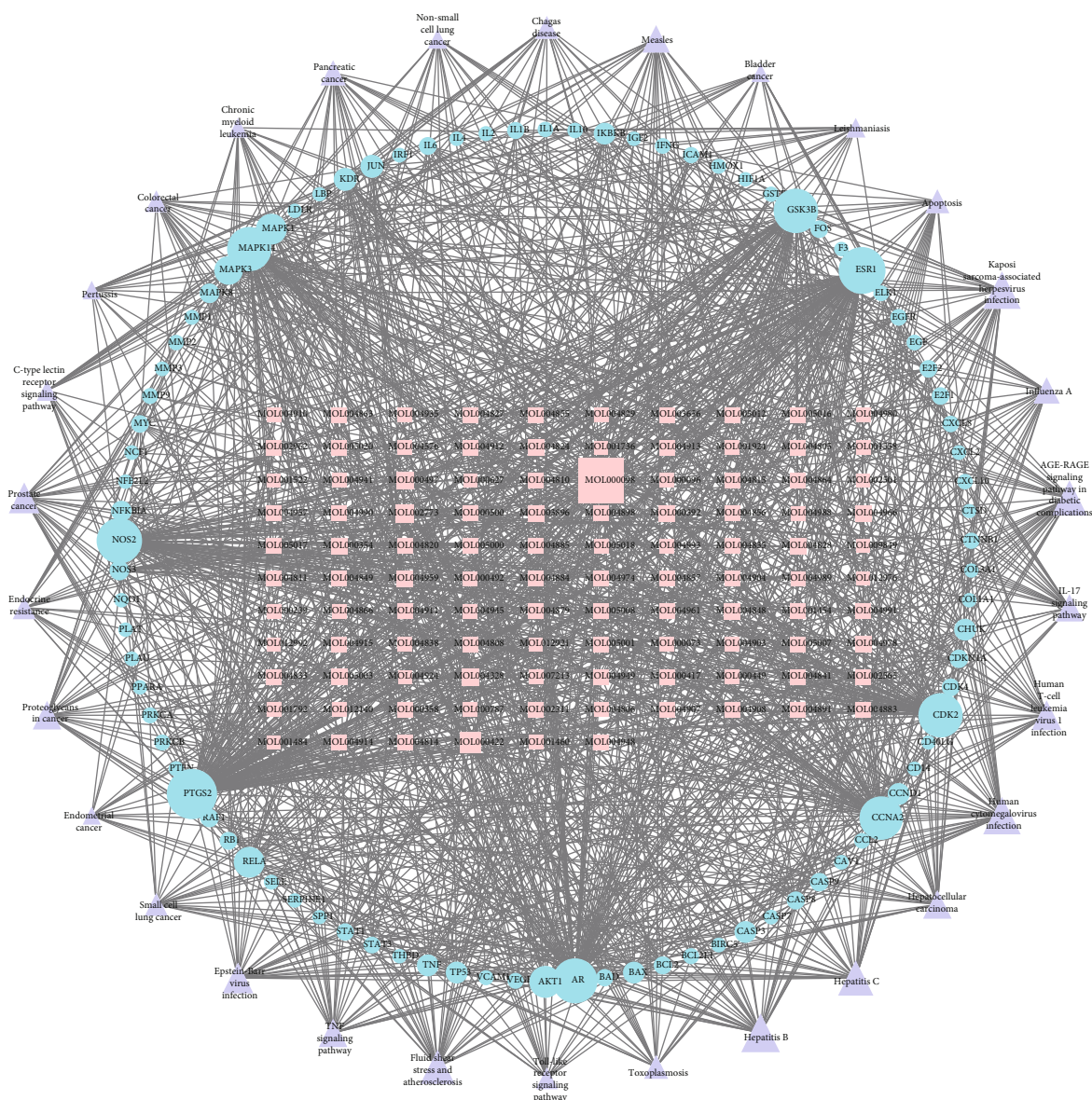


FIGURE 6: “Pathway-common target-active ingredient” network. Purple triangle nodes represent the top 30 IVDD-related pathways; blue circles represent common genes; pink rectangle represents the active ingredient of DSF.

stress, leading to apoptosis and senescence of nucleus pulposus cells [29–32]. Drugs currently used to treat IVDD are limited to nonsteroidal anti-inflammatory drugs or muscle relaxants to relieve symptoms [1, 33]. Notably, TCM has been used for more than 2,000 years to treat various diseases, including IVDD [34]. Compared with Western medicine, TCM has the advantages of mild and fewer side effects. To date, network pharmacology has been widely used to study the mechanisms of TCM. In our study, the results of the active ingredient-potential target network showed that the main active compounds, including quercetin, beta-sitosterol, kaempferol, naringenin, and formononetin, may have potential research value for the treatment of IVDD. In addition, enrichment analysis showed that DSF acted on various biological processes of IVDD and influenced the disease course through eight pathways, such as the AGE-RAGE sig-

nal pathway in diabetic complications, the IL-17 signaling pathway, the TNF signaling pathway, and the Toll-like receptor signaling pathway, which confirmed that DSF has multicomponent, multipathway, and multitarget characteristics.

Some of the main compounds in DSF identified in our study have anti-inflammatory and antioxidative stress effects, thereby inhibiting apoptosis and senescence. Quercetin and kaempferol are common ingredients in licorice. Previous studies suggested that quercetin has antisenescence and antiapoptotic effects, mainly by promoting SIRT1-dependent autophagy and inhibiting senescence-associated secreted phenotype factor expression and the senescence phenotype in nucleus pulposus cells to prevent and treat IVDD [35, 36]. In addition, Lu et al. found that quercetin could inhibit the expression and release of various

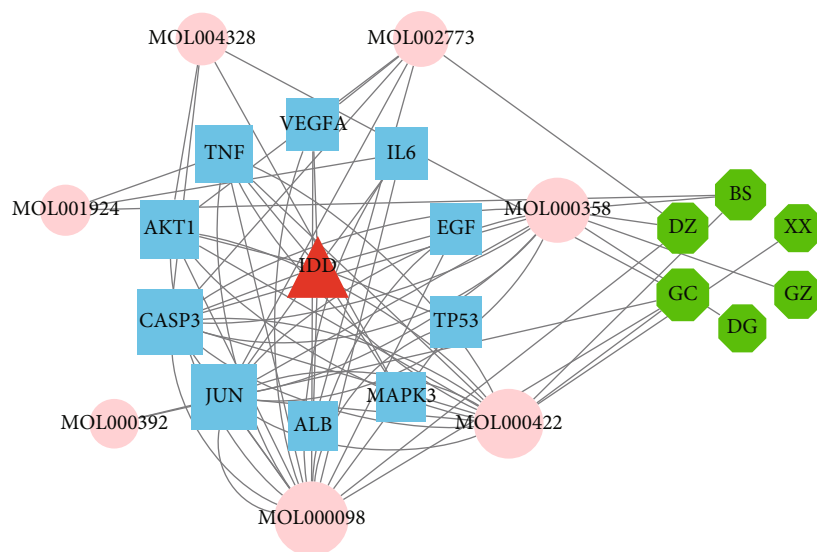


FIGURE 7: “IVDD-core gene-active ingredient-herb” network. Red triangle nodes represent disease, blue square nodes represent core genes, pink circle nodes represent the active ingredients related to the core genes, and the green diamond nodes represent herbs. The size of each node was set according to their degree value.

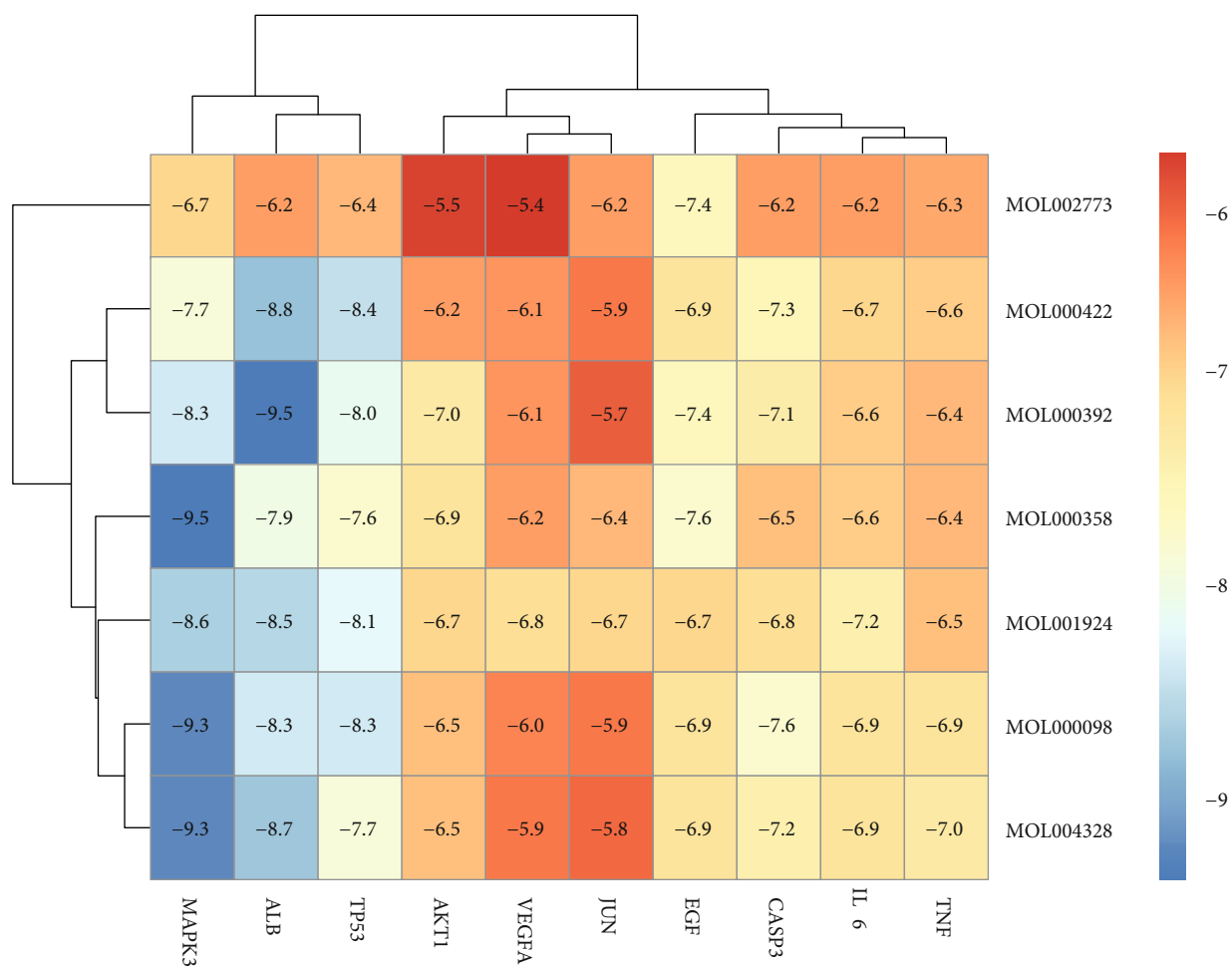


FIGURE 8: Heat map of molecular docking between 10 core genes and active ingredients of IVDD.



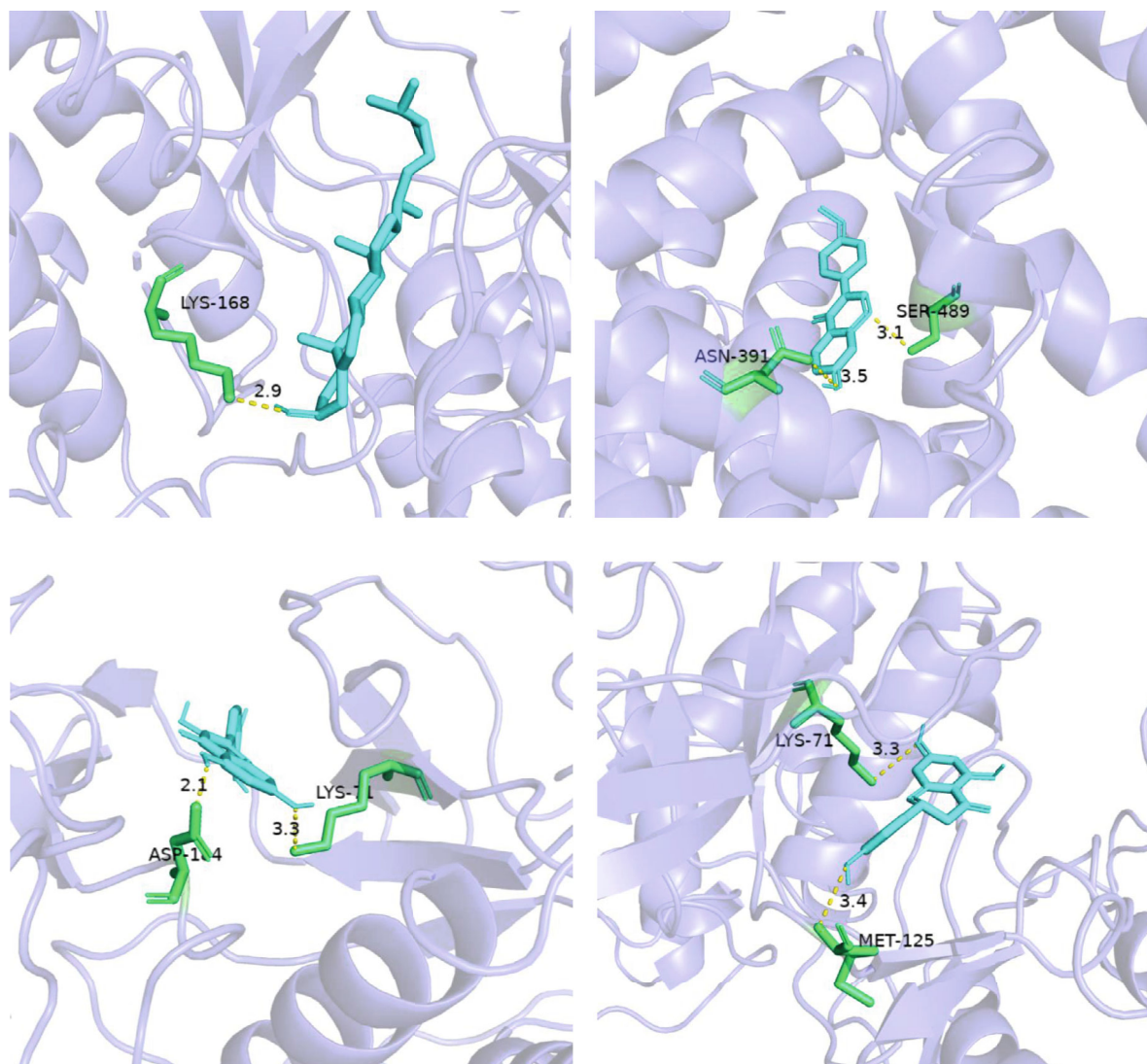


FIGURE 9: Molecular docking of the “bioactive compound-core gene”: (a) beta-sitosterol to MAPK3; (b) formononetin to ALB; (c) quercetin to MAPK3; (d) naringenin to MAPK3.

inflammatory factors, such as  $\text{TNF-}\alpha$ ,  $\text{IL-1}\beta$ , and  $\text{IL6}$ , by suppressing the activation of the  $\text{NF-}\kappa\text{B}$  pathway [37]. Kaempferol has been confirmed to modify the osteogenesis/adipogenesis balance and inhibit inflammation in BMSCs, making it a new target for the treatment of IVDD [38]. The common component in *Angelicae sinensis radix*, *cinnamomi ramulus*, *Paeoniae radix alba*, and *Jujubae fructus* is beta-sitosterol, which has anti-inflammatory and antioxidant effects [39]. Cho et al. found that formononetin may reduce the proteoglycan content used to treat musculoskeletal diseases [40].

By analyzing the PPI network of common genes, we found that  $\text{AKT1}$ ,  $\text{IL6}$ ,  $\text{ALB}$ ,  $\text{TNF}$ ,  $\text{VEGFA}$ ,  $\text{TP53}$ ,  $\text{MAPK3}$ ,  $\text{CASP3}$ ,  $\text{JUN}$ , and  $\text{EGF}$  might be the main potential targets for DSF therapy of IVDD. The apoptosis of nucleus pulposus cells (NPCs) plays a crucial role in the pathological process of IVDD, which is mainly mediated by inflammation and oxidative stress [3, 4].  $\text{IL6}$ ,  $\text{TNF}$ ,  $\text{TP53}$ , and  $\text{CASP3}$  are closely associated with the inflammatory response in the

process of IVDD, among which  $\text{TNF}$  and  $\text{IL6}$  are proinflammatory factors; moreover,  $\text{TP53}$  and  $\text{CASP3}$  induce apoptosis and senescence.  $\text{AKT1}$  encodes one of the three members of the human AKT serine-threonine protein kinase family, which are often referred to as protein kinase B alpha, beta, and gamma.  $\text{AKT/PI3K}$  is a key component of many signaling pathways, and AKT proteins regulate a wide variety of cellular functions, including cell proliferation, survival, metabolism, and angiogenesis, in both normal and malignant cells. Moreover, Zhan et al. found that the degree of intervertebral disc degeneration was related to the loss of vascular buds and the downregulation of  $\text{VEGFA}$  and its receptors [41].

GO and KEGG enrichment analysis results showed that the therapeutic targets of DSF for diseases mainly focused on the AGE-RAGE signaling pathway involved in diabetic complications,  $\text{IL-17}$  signaling pathway,  $\text{TNF}$  signaling pathway, Toll-like receptor signaling pathway, apoptosis, cellular senescence,  $\text{PI3K-Akt}$  signaling pathway, and  $\text{FoxO}$

signaling pathway. The biological processes are mainly induced in response to oxidative stress and reactive oxygen species and the regulation of apoptotic signaling pathways. Studies have shown that all of these factors play crucial roles in the progression of IVDD. The TNF and IL-17 pathways play a synergistic role in the progression of IVDD, mainly by promoting the release of inflammatory factors, the apoptosis of NPCs, and the degradation of extracellular matrix (ECM) [42–44]. mTOR is a serine/threonine protein kinase activated by PI3K/Akt, ERK, Wnt, TNF- $\alpha$ , IGF1 or low energy, low oxygen, and other factors. Then, mTOR activates downstream 4E-BP1 and p70S6K, which play a vital role in regulating the proliferation, apoptosis, and nutritional status of interdisc cells [45]. Furthermore, activation of the PI3K-Akt pathway may lead to a series of events, including reduction of ECM degradation, inhibition of apoptosis, and induction or suppression of autophagy, which can protect against IVDD [46, 47]. Recent studies have shown that FOXO is a critical regulator of cellular homeostasis during IVDD [48]. In addition, Gao et al. found that 17 $\beta$ -estradiol prevents ECM degradation by downregulating MMP3 expression via the PI3K/Akt/FOXO3 pathway [49]. At present, among the regulatory pathways of IVDD, few studies on the correlation between IVDD and other signaling pathways were screened out in our study by KEGG analysis. A study found that accumulation of AGE-MG-H1 was associated with endochondral ossifications, hypertrophy, and osteogenic differentiation in IVDD and that these are directly related to RAGE, suggesting that AGE/RAGE could be a potential therapeutic target [50].

However, our study also has some limitations. For example, the results of this study lacked in vitro validation, and further external validation with animals should be performed. Moreover, the screening of active ingredients and related genes through databases may not be sufficiently comprehensive.

## 5. Conclusions

The combination of network pharmacology and molecular docking provides a practical way to analyze the molecular biological mechanisms of DSF-mediated IVDD treatment. AKT1, IL6, ALB, TNF, VEGFA, TP53, MAPK3, CASP3, JUN, and EGF may be potential targets of DSF in treating IVDD, and enrichment analysis results showed that the therapeutic targets of DSF for diseases mainly focused on the AGE-RAGE signaling pathway in diabetic complications, IL-17 signaling pathway, TNF signaling pathway, Toll-like receptor signaling pathway, apoptosis, cellular senescence, PI3K-Akt signaling pathway, and FoxO signaling pathway, which confirms the multicomponent, multipathway, and multitarget characteristics of DSF and provides an essential theoretical basis for clinical practice.

## Abbreviations

IVDD: Intervertebral disk degeneration  
 NPCs: Nucleus pulposus cells  
 NSAIDs: Nonsteroidal anti-inflammatory drugs

TCM: Traditional Chinese medicine  
 DSF: Danggui-Sini formula  
 TCMSP: Traditional Chinese Medicine Systems Pharmacology  
 OB: Oral bioavailability  
 DL: Drug likeness  
 OMIM: Online Mendelian Inheritance in Man  
 CTD: Comparative Toxicogenomics Database  
 GO: Gene ontology  
 KEGG: Kyoto Encyclopedia of Genes and Genomes  
 BPs: Biological processes  
 CCs: Cell components  
 MFs: Molecular functions  
 PPI: Protein-protein interaction  
 NPCs: Nucleus pulposus cells  
 ECM: Extracellular matrix.

## Data Availability

The data used and analyzed during the current study was available from the corresponding author on reasonable request.

## Conflicts of Interest

The authors declare no conflict of interest.

## Authors' Contributions

Weishi Li contributed to the study conception and design. Acquisition of data, analysis, and interpretation of data were performed by Longjie Wang and Jialiang Lin. The figures were designed and drawn by Jialiang Lin. The article was written by Longjie Wang, and all authors commented on previous versions of the manuscript. All authors read and approved the final manuscript.

## Acknowledgments

This work was supported by the fellowship of China Postdoctoral Science Foundation (No. 2020M670002ZX).

## Supplementary Materials

Supplementary Table 1: detail information for active ingredients of Danggui-Sini formula. Supplementary Table 2: potential targets related to active ingredients. Supplementary Table 3: Common targets related to intervertebral disc degeneration. Supplementary Table 4: the results of GO and KEGG enrichment analysis. (*Supplementary Materials*)

## References

- [1] C. Maher, M. Underwood, and R. Buchbinder, "Non-specific low back pain," *The Lancet*, vol. 389, no. 10070, pp. 736–747, 2017.
- [2] M. L. Ji, H. Jiang, X. J. Zhang et al., "Preclinical development of a microRNA-based therapy for intervertebral disc degeneration," *Nature Communications*, vol. 9, no. 1, 2018.

- [3] J. Dowdell, M. Erwin, T. Choma, A. Vaccaro, J. Iatridis, and S. K. Cho, "Intervertebral disk degeneration and repair," *Neurosurgery*, vol. 80, no. 3s, pp. S46–S54, 2017.
- [4] M. A. Adams and P. J. Roughley, "What is intervertebral disc degeneration, and what causes it?," *Spine*, vol. 31, no. 18, pp. 2151–2161, 2006.
- [5] P. D. Roelofs, R. A. Deyo, B. W. Koes, R. J. Scholten, and M. W. van Tulder, "Nonsteroidal anti-inflammatory drugs for low back Pain," *Spine*, vol. 33, no. 16, pp. 1766–1774, 2008.
- [6] N. Henschke, C. G. Maher, K. M. Refshauge et al., "Prognosis in patients with recent onset low back pain in Australian primary care: inception cohort study," *BMJ*, vol. 337, 2008.
- [7] C. Ding, "Do NSAIDs affect the progression of osteoarthritis?," *Inflammation*, vol. 26, no. 3, pp. 139–142, 2002.
- [8] E. C. Huskisson, H. Berry, P. Gishen, R. W. Jubb, and J. Whitehead, "Effects of antiinflammatory drugs on the progression of osteoarthritis of the knee. LINK Study Group. Longitudinal Investigation of Nonsteroidal Antiinflammatory Drugs in Knee Osteoarthritis," *The Journal of Rheumatology*, vol. 22, no. 10, 1941.
- [9] K. L. Lapane, S. Yang, J. B. Driban et al., "Effects of prescription nonsteroidal antiinflammatory drugs on symptoms and disease progression among patients with knee osteoarthritis," *Arthritis & Rheumatology*, vol. 67, no. 3, pp. 724–732, 2015.
- [10] J. L. Tang, B. Y. Liu, and K. W. Ma, "Traditional Chinese medicine," *The Lancet*, vol. 372, no. 9654, pp. 1938–1940, 2008.
- [11] A. L. Hopkins, "Network pharmacology: the next paradigm in drug discovery," *Nature Chemical Biology*, vol. 4, no. 11, pp. 682–690, 2008.
- [12] R. Jiang, X. Zhang, Y. Li et al., "Identification of the molecular mechanisms of *Salvia miltiorrhiza* relevant to the treatment of osteoarthritis based on network pharmacology," *Discovery Medicine*, vol. 30, no. 160, 2020.
- [13] K. X. Wang, Y. Gao, C. Lu et al., "Uncovering the complexity mechanism of different formulas treatment for rheumatoid arthritis based on a novel network pharmacology model," *Frontiers in Pharmacology*, vol. 11, 2020.
- [14] C. Feng, M. Zhao, L. Jiang, Z. Hu, and X. Fan, "Mechanism of modified Danggui Sini Decoction for knee osteoarthritis based on network pharmacology and molecular docking," *Evidence-based Complementary and Alternative Medicine*, vol. 2021, Article ID 6680637, 11 pages, 2021.
- [15] J. Ru, P. Li, J. Wang et al., "TCMSP: a database of systems pharmacology for drug discovery from herbal medicines," *Journal of Cheminformatics*, vol. 6, no. 1, 2014.
- [16] D. S. Wishart, Y. D. Feunang, A. C. Guo et al., "DrugBank 5.0: a major update to the DrugBank database for 2018," *Nucleic Acids Research*, vol. 46, no. D1, pp. D1074–D1082, 2018.
- [17] UniProt Consortium, "UniProt: a hub for protein information," *Nucleic Acids Research*, vol. 43, 2015.
- [18] J. S. Amberger, C. A. Bocchini, F. Schiettecatte, A. F. Scott, and A. Hamosh, "OMIM.org: Online Mendelian Inheritance in Man (OMIM®), an online catalog of human genes and genetic disorders," *Nucleic Acids Research*, vol. 43, no. D1, pp. D789–D798, 2015.
- [19] M. Rebhan, V. Chalifa-Caspi, J. Prilusky, and D. Lancet, "GeneCards: integrating information about genes, proteins and diseases," *Trends in Genetics*, vol. 13, no. 4, 1997.
- [20] A. P. Davis, C. J. Grondin, R. J. Johnson et al., "The Comparative Toxicogenomics Database: update 2019," *Nucleic Acids Research*, vol. 47, no. D1, pp. D948–D954, 2019.
- [21] J. Piñero, À. Bravo, N. Queralt-Rosinach et al., "DisGeNET: a comprehensive platform integrating information on human disease-associated genes and variants," *Nucleic Acids Research*, vol. 45, no. D1, pp. D833–D839, 2017.
- [22] P. Shannon, A. Markiel, O. Ozier et al., "Cytoscape: a software environment for integrated models of biomolecular interaction networks," *Genome Research*, vol. 13, no. 11, pp. 2498–2504, 2003.
- [23] G. Yu, L. G. Wang, Y. Han, and Q. Y. He, "clusterProfiler: an R package for comparing biological themes among gene clusters," *Omic*s, vol. 16, no. 5, pp. 284–287, 2012.
- [24] D. Szklarczyk, A. L. Gable, D. Lyon et al., "STRING v11: protein-protein association networks with increased coverage, supporting functional discovery in genome-wide experimental datasets," *Nucleic Acids Research*, vol. 47, no. D1, pp. D607–D613, 2019.
- [25] O. Trott and A. J. Olson, "AutoDock Vina: improving the speed and accuracy of docking with a new scoring function, efficient optimization, and multithreading," *Journal of Computational Chemistry*, vol. 31, no. 2, 2009.
- [26] A. Khan, A. Diwan, H. K. Thabet, M. Imran, and M. A. Bakht, "Discovery of novel pyridazine-based cyclooxygenase-2 inhibitors with a promising gastric safety profile," *Molecules*, vol. 25, no. 9, 2020.
- [27] D. Hoy, C. Bain, G. Williams et al., "A systematic review of the global prevalence of low back pain," *Arthritis and Rheumatism*, vol. 64, no. 6, pp. 2028–2037, 2012.
- [28] C. E. Jensen, A. Riis, K. D. Petersen, M. B. Jensen, and K. M. Pedersen, "Economic evaluation of an implementation strategy for the management of low back pain in general practice," *Pain*, vol. 158, no. 5, pp. 891–899, 2017.
- [29] S. K. Tschoeke, M. Hellmuth, A. Hostmann et al., "Apoptosis of human intervertebral discs after trauma compares to degenerated discs involving both receptor-mediated and mitochondrial-dependent pathways," *Journal of Orthopaedic Research*, vol. 26, no. 7, pp. 999–1006, 2008.
- [30] H. E. Gruber, G. L. Hoelscher, S. Bethea, and E. N. Hanley Jr., "Mitochondrial membrane potential and nuclear and gene expression changes during human disc cell Apoptosis," *Spine*, vol. 40, no. 12, pp. 876–882, 2015.
- [31] F. Wang, F. Cai, R. Shi, X. H. Wang, and X. T. Wu, "Aging and age related stresses: a senescence mechanism of intervertebral disc degeneration," *Osteoarthritis and Cartilage*, vol. 24, no. 3, pp. 398–408, 2016.
- [32] H. Che, J. Li, Y. Li et al., "p16 deficiency attenuates intervertebral disc degeneration by adjusting oxidative stress and nucleus pulposus cell cycle," *eLife*, vol. 9, 2020.
- [33] H. C. Wenger and A. S. Cifu, "Treatment of low back pain," *JAMA*, vol. 318, no. 8, 2017.
- [34] L. Zhu, C. Yu, X. Zhang et al., "The treatment of intervertebral disc degeneration using Traditional Chinese Medicine," *Journal of Ethnopharmacology*, vol. 263, article 113117, 2020.
- [35] Z. Shao, B. Wang, Y. Shi et al., "Senolytic agent Quercetin ameliorates intervertebral disc degeneration via the Nrf2/NF- $\kappa$ B axis," *Osteoarthritis and Cartilage*, vol. 29, no. 3, pp. 413–422, 2021.
- [36] D. Wang, X. He, D. Wang et al., "Quercetin suppresses apoptosis and attenuates intervertebral disc degeneration via the SIRT1-autophagy pathway," *Frontiers in Cell and Development Biology*, vol. 8, article 613006, 2020.
- [37] J. Lu, D. M. Wu, Y. L. Zheng et al., "Quercetin activates AMP-activated protein kinase by reducing PP2C expression

- protecting old mouse brain against high cholesterol-induced neurotoxicity,” *The Journal of Pathology*, vol. 222, no. 2, pp. 199–212, 2010.
- [38] J. Zhu, H. Tang, Z. Zhang et al., “Kaempferol slows intervertebral disc degeneration by modifying LPS-induced osteogenesis/adipogenesis imbalance and inflammation response in BMSCs,” *International Immunopharmacology*, vol. 43, pp. 236–242, 2017.
- [39] F. Zhang, Z. Liu, X. He, Z. Li, B. Shi, and F. Cai, “ $\beta$ -Sitosterol-loaded solid lipid nanoparticles ameliorate complete Freund’s adjuvant-induced arthritis in rats: involvement of NF- $\kappa$ B and HO-1/Nrf-2 pathway,” *Drug Delivery*, vol. 27, no. 1, pp. 1329–1341, 2020.
- [40] I. A. Cho, T. H. Kim, H. Lim et al., “Formononetin antagonizes the interleukin-1 $\beta$ -induced catabolic effects through suppressing inflammation in primary rat chondrocytes,” *Inflammation*, vol. 42, no. 4, pp. 1426–1440, 2019.
- [41] J. W. Zhan, S. Q. Wang, M. S. Feng et al., “Effects of axial compression and distraction on vascular bud and VEGFA expression in the vertebral endplate of an ex vivo rabbit spinal motion segment culture model,” *Spine*, vol. 46, no. 7, pp. 421–432, 2021.
- [42] J. Zhang, X. Wang, H. Liu et al., “TNF- $\alpha$  enhances apoptosis by promoting chop expression in nucleus pulposus cells: role of the MAPK and NF- $\kappa$ B pathways,” *Journal of Orthopaedic Research*, vol. 37, no. 3, pp. 697–705, 2019.
- [43] L. Miguélez-Rivera, S. Pérez-Castrillo, M. L. González-Fernández et al., “Immunomodulation of mesenchymal stem cells in discogenic pain,” *The Spine Journal*, vol. 18, no. 2, pp. 330–342, 2018.
- [44] X. G. Liu, H. W. Hou, and Y. L. Liu, “Expression levels of IL-17 and TNF- $\alpha$  in degenerated lumbar intervertebral discs and their correlation,” *Experimental and Therapeutic Medicine*, vol. 11, no. 6, pp. 2333–2340, 2016.
- [45] N. Hay and N. Sonenberg, “Upstream and downstream of mTOR,” *Genes & Development*, vol. 18, no. 16, pp. 1926–1945, 2004.
- [46] Z. Liu, K. Zhou, W. Fu, and H. Zhang, “Insulin-like growth factor 1 activates PI3k/Akt signaling to antagonize lumbar disc degeneration,” *Cellular Physiology and Biochemistry*, vol. 37, no. 1, pp. 225–232, 2015.
- [47] Y. Xunlu, F. Minshan, Z. Liguó et al., “Integrative bioinformatics analysis reveals potential gene biomarkers and analysis of function in human degenerative disc annulus fibrosus cells,” *BioMed Research International*, vol. 2019, Article ID 9890279, 8 pages, 2019.
- [48] O. Alvarez-Garcia, T. Matsuzaki, M. Olmer et al., “FOXO are required for intervertebral disk homeostasis during aging and their deficiency promotes disk degeneration,” *Aging Cell*, vol. 17, no. 5, article e12800, 2018.
- [49] X. W. Gao, X. T. Su, Z. H. Lu, and J. Ou, “17 $\beta$ -estradiol prevents extracellular matrix degradation by downregulating MMP3 expression via PI3K/Akt/FOXO3 pathway,” *Spine*, vol. 45, no. 5, pp. 292–299, 2020.
- [50] S. Illien-Jünger, O. M. Torre, W. F. Kindschuh, X. Chen, D. M. Laudier, and J. C. Iatridis, “AGEs induce ectopic endochondral ossification in intervertebral discs,” *European Cells & Materials*, vol. 32, pp. 257–270, 2016.