# iScience



## Article

Metabolomic profiling of samples from pediatric patients with asthma unveils deficient nutrients in African Americans



lost significance after controlling for comorbid obesity.

Qu et al., iScience 25, 104650 July 15, 2022 © 2022 The Authors. https://doi.org/10.1016/ j.isci.2022.104650

## **iScience**

### Article

## Metabolomic profiling of samples from pediatric patients with asthma unveils deficient nutrients in African Americans

Hui-Qi Qu,<sup>1</sup> Joseph Glessner,<sup>1,2,3</sup> Jingchun Qu,<sup>1</sup> Steven Gilhool,<sup>1</sup> Frank Mentch,<sup>1</sup> Ian Campbell,<sup>1,3</sup> Patrick Sleiman,<sup>1,2,3</sup> John J. Connolly,<sup>1</sup> Hakon Hakonarson,<sup>1,2,3,4,5,6,\*</sup> and on behalf of the IHCC consortium

#### SUMMARY

Plasma metabolomics represents a potentially powerful approach to understand the biochemical mechanisms of nutrition and metabolism in asthma. This study aims to acquire knowledge on plasma metabolites in asthma, which may provide avenues for nutrition therapy, as well as explanations for the observed effects in existing therapies. This study investigated 249 metabolites from 18 metabolite groups in a large cohort of African American population, including 602 pediatric patients with asthma and 593 controls, using a nuclear magnetic resonance (NMR) metabolomics platform. Decreased levels of citrate, ketone bodies, and two amino acids histidine (His) and glutamine (Gln), were observed in asthma cases compared to controls. Metabolites for lipid metabolism lost significance after controlling for comorbid obesity. For the first time, this study depicts a broad panorama of lipid metabolism and nutrition in asthma. Supplementation or augmentation of nutrients that are deficient may be beneficial for asthma care.

#### INTRODUCTION

Asthma is a major health issue in African Americans. According to the Centers for Disease Control and Prevention (CDC), African Americans children have a 60% greater prevalence of asthma at 14.3%, and 7.1-fold greater risk of death due to asthma, than non-Hispanic white (Ebell et al., 2019).

With the rapid progress of omics technologies, plasma or serum metabolomics has become an accessible approach to probe the biomechanics of human diseases that may help uncovering noninvasive metabolites, in particular for nutritional/diet interventions (James and Parkinson, 2015). Important insights have been acquired about, for example, systemic inflammation and oxidative stress in asthma using metabolomics (Nambiar et al., 2020).

Diet and nutrition have been suggested as important risk factors for the development and management of obstructive lung diseases such as asthma (Berthon and Wood, 2015). In patients with asthma, diet manipulation is considered to have beneficial effects on systemic inflammation and oxidative stress, and may play protective roles in asthma development and improving asthma symptoms (Alwarith et al., 2020). However, our understanding of changes of plasma metabolites in asthma remains unclear.

The objective of this study is to acquire knowledge on plasma metabolites in asthma, which may provide novel avenues for nutrition therapy, as well as explanations for the observed effects in existing therapies. For this purpose, we performed a metabolomic study in a large cohort of pediatric patients with asthma of African American ancestry (n = 602) in comparison with 593 controls without asthma. This study used a nuclear magnetic resonance (NMR) metabolomic technology with high reproducibility, including 249 selected metabolites related to common human diseases from 18 metabolite groups, with 98 metabolites of lipoprotein subclasses, 70 metabolites of relative lipoprotein lipid concentrations, 18 metabolites of fatty acids, 10 metabolites of disease-related amino acids, as well as metabolites for glycolysis metabolites, ketone bodies, inflammation, etc. A number of nutrients correlated with lower levels of asthma, with beneficial effects being observed for the supplementation of these nutrients by previous studies (Hernández et al., 2019; Poynter et al., 2020; Smith et al., 2013).



<sup>1</sup>The Center for Applied Genomics, Children's Hospital of Philadelphia, 3615 Civic Center Blvd, Abramson Building, Philadelphia, PA 19104, USA

<sup>2</sup>Department of Pediatrics, The Perelman School of Medicine, University of Pennsylvania, Philadelphia, PA 19104, USA

<sup>3</sup>Division of Human Genetics, Children's Hospital of Philadelphia, Philadelphia, PA 19104, USA

<sup>4</sup>Division of Pulmonary Medicine, Children's Hospital of Philadelphia, Philadelphia, PA 19104, USA

<sup>5</sup>Faculty of Medicine, University of Iceland, 101 Reykjavik, Iceland

<sup>6</sup>Lead contact

\*Correspondence: hakonarson@chop.edu https://doi.org/10.1016/j.isci. 2022.104650

Table 1. Correlation between asthma and metabolite levels <sup>a</sup>								
	Controlled For Age & Sex		Controlled For Age, Sex, Obesity and VitD deficiency					
	r	P (2-tailed)	Df	r	P (2-tailed)	df	Group	Subgroup
Citrate	-0.136	2.48E-06	1191	-0.115	6.90E-05	1189	Glycolysis-related metabolites	
Acetoacetate	-0.092	0.002	1190	-0.083	0.004	1188	Ketone bodies	
β-Hydroxybutyrate	-0.087	0.003	1187	-0.082	0.005	1185	Ketone bodies	
Histidine	-0.093	0.001	1190	-0.072	0.013	1188	Amino acids	
Glutamine	-0.086	0.004	1158	-0.067	0.022	1156	Amino acids	
Ratio of saturated fatty acids to total fatty acids (SFA%)	-0.061	0.036	1190	-0.064	0.027	1188	Fatty acids	Fatty acid ratios
Free cholesterol to total lipids ratio in medium VLDL (FC% in mVLDL)	-0.06	0.039	1191	-0.062	0.033	1189	Relative lipoprotein lipid concentrations	Medium VLDL ratios
Acetate	-0.065	0.024	1189	-0.061	0.037	1187	Ketone bodies	

Beta: standardized beta coefficient; t: t-statistic; FC% in mVLDL: Free cholesterol to total lipids ratio in medium VLDL; VLDL: very low density lipoprotein; SFA%: Ratio of saturated fatty acids to total fatty acids.

<sup>a</sup>Point-Biserial partial correlation t-tests were used to measure the relationship between asthma and metabolite levels. r: Point-Biserial correlation coefficient; df: Degrees of Freedom; VitD: Vitamin D; VLDL: very low-density lipoprotein.

#### RESULTS

As shown by the Point-Biserial partial correlation between asthma and metabolite levels, and controlling for age and sex, 37 metabolites from 10 groups showed p < 0.05. Notably, a number of metabolites for lipid metabolisms are significant, e.g., 13 metabolites for relative lipoprotein lipid concentrations, 11 metabolites for lipoprotein subclasses, and several metabolites for cholesterol and fatty acids. In addition, many of these metabolites are for high-density lipoprotein (HDL) particles, and have lower levels in asthma cases.

Considering the clinical association of asthma and obesity, as well as the common comorbidities of obesity and VitD deficiency in asthma cases, we performed further partial correlation analysis between asthma and the metabolite levels, controlled for age, sex, obesity status, and VitD deficiency. As expected, significantly fewer (or only 8) metabolites with p < 0.05 in the initial test retained p < 0.05 after additionally controlled for obesity and VitD deficiency (Table 1).

#### Decreased citrate levels in asthma

Lower citrate is correlated with asthma with a p =  $6.90 \times 10^{-5}$ , and remains statistically significant after Bonferroni correction for multiple testing (i.e.,  $\alpha = 0.05/249 =$  significance threshold of 2.01 × 10<sup>-4</sup>). In addition, lower citrate is correlated with increased age, asthma, obesity, and VitD deficiency (Table 2).

#### Decreased levels of ketone bodies in asthma

The two main ketone bodies—acetoacetate and  $\beta$ -Hydroxybutyrate ( $\beta$ OHB)—as well as acetate, have lower levels in asthma cases with p < 0.05. Both age and asthma status are correlated with levels of the two main ketone bodies—i.e., lower levels in older individuals and asthma cases. In contrast, lower levels of acetate were seen in younger individuals and asthma cases (Table 2).

#### Decreased levels of two amino acids in asthma

Lower plasma levels of two amino acids histidine (His) and glutamine (Gln) were seen in asthma cases with p < 0.05. Both asthma and obesity are correlated with lower levels of His and Gln. In addition, older age is correlated with higher levels of His (Table 2).

#### Metabolites for lipid metabolism correlated with asthma

A diagnosis of asthma correlated with lower levels of FC% in mVLDL [Free cholesterol to total lipids ratio in medium very low-density lipoprotein (VLDL)], and lower ratio of saturated fatty acids (SFA) to total fatty

## iScience Article



Table 2. Linear regression between asthm	na and metabolite levels <sup>a</sup>					
Glycolysis related metabolites	Citrate	Standardi	Standardized Coefficients			
	Linear regression	Beta	t	Р		
	age	-0.171	-5.938	3.79E-09		
	sex	-0.036	-1.255	0.210		
	asthma	-0.115	-3.994	6.90E-05		
	Obesity	-0.083	-2.818	0.005		
	Vitamin D deficiency	-0.059	-2.012	0.044		
Ketone bodies	Acetate					
	age	0.105	3.555	3.94E-04		
	sex	-0.039	-1.325	0.185		
	asthma	-0.062	-2.090	0.037		
	Obesity	0.003	0.096	0.923		
	Vitamin D deficiency	-0.028	-0.925	0.355		
	Acetoacetate					
	age	-0.128	-4.353	1.46E-05		
	sex	-0.020	-0.705	0.481		
	asthma	-0.084	-2.858	0.004		
	Obesity	-0.013	-0.444	0.657		
	Vitamin D deficiency	-0.041	-1.379	0.168		
	β-Hydroxybutyrate					
	age	-0.090	-3.061	0.002		
	sex	-0.027	-0.938	0.348		
	asthma	-0.083	-2.816	0.005		
	Obesity	0.017	0.569	0.570		
	Vitamin D deficiency	-0.044	-1.456	0.146		
Amino acids	Histidine					
	age	0.172	5.962	3.28E-09		
	sex	-0.029	-1.002	0.317		
	asthma	-0.072	-2.494	0.013		
	Obesity	-0.115	-3.883	1.09E-04		
	Vitamin D deficiency	-0.033	-1.110	0.267		
	Glutamine					
	age	0.029	0.966	0.334		
	sex	-0.016	-0.553	0.581		
	asthma	-0.068	-2.283	0.023		
	Obesity	-0.099	-3.252	1.18E-03		
	Vitamin D deficiency	-0.034	-1.104	0.270		
Relative lipoprotein lipid concentrations	FC% in mVLDL					
	age	0.129	4.410	1.13E-05		
	sex	0.039	1.356	0.175		
	asthma	-0.063	-2.137	0.033		
	Obesity	0.045	1.505	0.133		
	Vitamin D deficiency	-0.020	-0.672	0.502		
Fatty acids	SFA%					
	age	-0.132	-4.513	7.02E-06		
	sex	-0.061	-2.105	0.035		

(Continued on next page)





Table 2. Continued				
Glycolysis related metabolites	Citrate	Standardized Coefficients		
	asthma	-0.065	-2.206	0.028
	Obesity	0.055	1.850	0.065
	Vitamin D deficiency	-0.022	-0.720	0.472

<sup>a</sup>Linear regression t-tests were used to estimate p values (two-tailed).

acids (SFA%). In addition, older age is correlated with both higher levels of FC% in mVLDL and lower levels of SFA% (Table 2).

#### DISCUSSION

#### Lower citrate in asthma

Levels of metabolites in the citrate cycle are altered in asthma related to hypoxia as reviewed by Kelly et al. (Kelly et al., 2017) Increased workload of inspiratory muscles and hypoxia, in addition to high turnover rate of the citrate cycle in asthma (Papamichael et al., 2021), may deplete energy stores and lower citrate level. We observed lower citrate levels in individuals with asthma with statistical significance after correction for multiple testing (Bonferroni corrected p = 0.017). No blood sample in this study was treated with citrate anticoagulant. Bone is the major source of citrate in plasma, which releases citrate into plasma during osteoclast bone resorption. Plasma citrate levels are mainly regulated by parathyroid hormone (PTH, upregulation of plasma citrate level) and calcitonin (CT, downregulation of plasma citrate level) (Costello and Franklin, 2016). The age effect observed in this study (i.e. higher citrate in younger age) may be explained by higher osteoclast activity and bone turnover in children than young adults (Jürimäe, 2010). The link between obesity and bone metabolism is complex (Cao, 2011; Hou et al., 2020; Viljakainen et al., 2014), which may explain the lower level of citrate in obesity. The correlation between VitD deficiency and lower citrate level of citrate in obesity. The correlation between VitD deficiency (Bikle, 2012).

The roles of citric acid cycle metabolites in the hyperresponsiveness and airway remodeling in asthma are related to the function of ten-eleven translocation (TET) enzymes in airway smooth muscle (SM) cells, as highlighted by a recent study (Huang, 2020; Yeung et al., 2020). TET1 has been demonstrated to prevent house dust mite (HDM)-induced allergic airway inflammation in a mouse model (Burleson et al., 2019). Hypocitricemia may lead to decreased TET activity, and thus contribute to the development of asthma. The finding of our study may suggest citrate supplementation as a potential new therapy for patients with asthma of African American ancestry.

#### Lower ketone bodies in asthma

Increased energy demands and hypoxia in asthma (Kelly et al., 2017) may increase the consumption of ketone bodies. The two main ketone bodies as a pair of redox couples, acetoacetate and βOHB, have lower levels in asthma cases as shown in this study. In addition to providing energy, signaling roles of ketone bodies in health and disease have been suggested by previous studies as reviewed by Puchalska and Crawford (Puchalska and Crawford, 2021). βOHB is a stress response molecule and plays key roles in maintaining redox homeostasis in response to environmental and metabolic challenges (Rojas-Morales et al., 2020). In particular, immunomodulatory and anti-inflammatory effects of βOHB by suppressing activation of the NLRP3 inflammasome have been demonstrated (Youm et al., 2015). NLRP3 (NOD-, LRR-, and pyrin domain-containing protein 3) is an intracellular sensor that detects variety of pathogen-associated molecular patterns (PAMPs) and danger-associated molecular patterns (DAMPs), and results in the formation and activation of the NLRP3 inflammasome (Swanson et al., 2019; Zhao and Zhao, 2020). Ketone body augmentation has been suggested to have therapeutic effects in HDM-induced asthma (Poynter et al., 2020).

In contrast to acetoacetate and βOHB being generated in liver, plasma acetate is from microbial-derived production in the colon, diet (e.g., vinegar), and endogenous acetogenesis from acetyl-CoA hydrolysis in cytoplasm (Moffett et al., 2020). Citrate, with lower levels among individuals with asthma as observed, is the

### iScience Article





#### Figure 1. Obesity and VitD deficiency in asthma cases and controls

Obesity and VitD deficiency are common in both cases and controls. For the cases, 22.9% have obesity, 10.5% have VitD deficiency, and 14.6% have both obesity and VitD deficiency. For the controls, 18.7% have obesity, 6.1% have VitD deficiency, and 7.9% have both obesity and VitD deficiency.

source of acetyl-CoA to generate acetate in cytoplasm in various tissues including liver, intestine, and brain (Hernández et al., 2019; Moffett et al., 2020). Acetate supplementation has been shown to increase the number and function of anti-inflammatory Treg cells and to decrease systemic pro-inflammatory cytokine levels (Hernández et al., 2019; Smith et al., 2013).

In this study, both ketone bodies are inversely correlated with age. An inverse relationship between age and blood ketone bodies in fasting status has been previously observed in children (Saudubray et al., 1981). Higher level of acetate is, however, correlated with increasing age as shown in these pediatric subjects. On the other hand, acetate levels were found to decline slightly in older adults (Skutches et al., 1979). These age effects highlight the importance of controlling for age in ketone body augmentation and acetate supplementation.

#### Lower His and Gln in asthma

Lower levels of plasma His and Gln were correlated with asthma. His is an essential amino acid (EAA). Lower His level in autoinflammatory states (e.g., rheumatoid arthritis) have been observed for a long time (Gerber et al., 1976). His supplementation for its anti-inflammatory and antioxidant effects have been investigated in a wide range of conditions with potential benefits, including autoinflammatory diseases like rheumatoid arthritis, inflammatory bowel disease, and atopic dermatitis (Holeček, 2020). The increased His level by age, as observed here, is supported by a previous study (Lepage et al., 1997). We observed a lower His level in obesity, while significantly lower serum His levels have been previously found in obese women (Niu et al., 2012). His supplementation reduces high-fat diet-induced body weight increase and ameliorates inflammation and oxidative stress of adipose tissue in female obese rat models (Sun et al., 2014).

It is important to emphasize that His is the precursor of histamine. Histamine plays a central role in allergic reactions mediated by mast cells (Amin, 2012). In asthma, histamine contributes to airway obstruction by inducing the bronchoconstriction of smooth muscle, secretion of mucous glycoprotein, and airway submucosal edema (Yamauchi and Ogasawara, 2019). His decarboxylase (HDC) catalyzes the generation of histamine from His, and the expression of HDC is elevated in asthma (Yamauchi, 1996). The potential effects of His supplementation on histamine reaction in asthma may warrant further investigation.

Gln is not an EAA, but rather n conditionally essential amino acid, showing deficiency in stress conditions (Morris et al., 2017). A trend of lower Gln level in asthma has been observed previously, albeit not meeting statistical significance (p = 0.07) (Fogarty et al., 2004). As reviewed by Oliveira et al., several studies have suggested that exogenous Gln administration might be beneficial in respiratory diseases, e.g., acute respiratory distress syndrome (ARDS) and asthma (Oliveira et al., 2016). We observed a lower Gln level in obesity. A study by Petrus et al. showed that reduced Gln levels are a metabolic signature of obese white adipose tissue in human (Petrus et al., 2020). Gln administration may attenuate adipose tissue inflammation in obese mouse model and human fat cell models (Petrus et al., 2020).





#### Lipid levels in asthma

Increased energy demands and hypoxia in asthma (Kelly et al., 2017) may increase lipid mobilization as an alternative energy source. Insulin resistance commonly seen in patients with asthma, especially in those with obesity (Arshi et al., 2010) is closely related to the pathogenesis of dyslipidemia. Dyslipidemia is a key driver of airway inflammation in asthma (Wood et al., 2003). Previous studies have shown that blood lipid levels are associated with childhood asthma, and higher levels of LDL cholesterol and triglyceride have been associated with concurrent asthma, airway obstruction, and aeroallergen sensitization, while higher levels of HDL cholesterol have been associated with improved airway resistance and decreased bronchial responsiveness and aeroallergen sensitization (Vinding et al., 2016). In this study, a number of metabolites for lipid metabolisms showed p < 0.05, including metabolites for relative lipoprotein lipid concentrations, metabolites for lipoprotein subclasses, and metabolites for cholesterol and fatty acids. However, obesity is a common comorbidity in asthma, and is the most common cause of lipid disorders. Here, we tested the correlation of metabolomic metabolites and asthma controlled for obesity. The associations of most of metabolites for lipoprotein subclasses were not significant after controlling for obesity, except two metabolites, free cholesterol to total lipids ratio in medium VLDL (FC% in mVLDL) and SFA%.

Levels of FC% in mVLDL were lower in asthma cases compared to controls. However, no significant change was observed for the level of free cholesterol in medium VLDL (M\_VLDL\_FC), nor was any change observed for total cholesterol (i.e., the sum of free cholesterol and cholesteryl esters) to total lipid ratios in medium VLDL (M\_VLDL\_C\_pct), or cholesteryl esters to total lipid ratios in medium VLDL (M\_VLDL\_C\_pct), or cholesteryl esters to total lipid ratios in medium VLDL (M\_VLDL\_C\_pct). Therefore, the change of FC% in mVLDL in asthma needs to be replicated in independent samples before we venture any biomedical explanation.

Increased SFA have been reported in patients with mild, moderate, and severe asthma by a previous study (Reinke et al., 2017). Instead, lower SFA% were observed in asthma cases in this study. In this regard, unsaturated fatty acids have been suggested to have health benefits, including anti-inflammatory effects (Delmastro-Greenwood et al., 2014). However, different unsaturated fatty acids have been shown to have different inflammatory properties. For example, the two essential nutrients of unsaturated fatty acids, the pro-inflammatory omega-6 (n-6), and the anti-inflammatory omega-3 (n-3) have opposite inflammatory properties in asthma (Wendell et al., 2014). The p = 0.027 of SFA% observed in this study, therefore, needs replication in an independent sample. If validated, further investigation would be warranted for the change of the components of unsaturated fatty acids and their pathogenic implications in asthma.

In conclusion, we observed decrease in citrate, ketone bodies, and the two amino acids, His and Gln, in the plasma of asthma cases undergoing metabolomic measurements. This suggests that augmentation or supplementation of these nutrients may be beneficial as asthma treatment. The change of citrate level is statistically significant after correction for multiple testing. Importantly, both the changes of the ketone bodies and the two amino acids lack statistical significance following correction for multiple testing. Thus, validation of these changes in independent samples is required. We do note, however, that a number of previous studies support these observations, while ketone body augmentation, or supplementation of acetate or administration of the two amino acids has been suggested to be potentially beneficial in treating inflammatory diseases such as asthma. Asthma is a complex disease, involving a combination of numerous genetic, environmental, and lifestyle factors (Arruda et al., 2005). Despite the small r values, the correlations identified in this study highlighted the decreased metabolites as risk factors and potential therapeutic targets of asthma, in addition to mechanistic knowledge gained on asthma. Finally, this study urges caution in ascribing changes in lipid metabolism in relation to asthma, as obesity is a common comorbidity, and the most common cause of lipid disorders. Nonetheless, obesity in asthma should not necessarily be regarded as a simple confounder. Obesity, the most common type of metabolic health problems in children and adults, has been suggested as both a major risk factor and a disease modifier for asthma in children and adults (Peters et al., 2018; Tesse et al., 2011).

#### Limitations of the study

Limited by the sample size of the current study, the reported metabolites warrant for replication in independent samples, including different populations other than African American. As a project of the International Hundred Thousand Plus Cohort Consortium, this study aims to assess the research application in asthma, as well as the clinical potential, of a cost-effective NMR metabolomic platform with high reproducibility. In contrast to an untargeted metabolomic platform, the Nightingale NMR platform used in this study





includes 249 selected metabolites. In this case, further study with an untargeted metabolomic technology or a targeted metabolomic platform with defined metabolite markers, e.g., assessing comprehensively the citrate cycle, will be helpful to validate and further clarify the change of related metabolic pathways.

#### **STAR\*METHODS**

Detailed methods are provided in the online version of this paper and include the following:

- KEY RESOURCES TABLE
- **RESOURCE AVAILABILITY** 
  - O Lead contact
  - Materials availability
  - Data and code availability
- EXPERIMENTAL MODEL AND SUBJECT DETAILS
  - O Patient recruitment and sample collection
- METHOD DETAILS
- QUANTIFICATION AND STATISTICAL ANALYSIS

#### SUPPLEMENTAL INFORMATION

Supplemental information can be found online at https://doi.org/10.1016/j.isci.2022.104650.

#### ACKNOWLEDGMENT

We thank the patients and their families for their participation in the study. We thank Nightingale for their services. Funding: The study was supported by: The International Hundred Thousand Plus Cohort Consortium (IHCC); CHOP funding: Institutional Development Funds from the Children's Hospital of Philadelphia to the Center for Applied Genomics, The Children's Hospital of Philadelphia Endowed Chair in Genomic Research (HH). Grant/award number: Not applicable.

#### **AUTHOR CONTRIBUTIONS**

Conceptualization, H.H. and H.Q.; literature search, H.Q.; Figures and Tables, H.Q.; data analysis, H.Q., J.G., J.Q., and F.M.; data interpretation, H.Q., J.G., J.Q., S.G., I.C., P.S., J.J.C., and H.H.; original draft writing, H.Q. and H.H.; review and revision, H.Q., J.J.C., and H.H.; supervision, H.H.

#### **DECLARATION OF INTERESTS**

The authors declare no competing interests.

Received: May 3, 2022 Revised: June 6, 2022 Accepted: June 16, 2022 Published: July 15, 2022

#### REFERENCES

Alwarith, J., Kahleova, H., Crosby, L., Brooks, A., Brandon, L., Levin, S.M., and Barnard, N.D. (2020). The role of nutrition in asthma prevention and treatment. Nutr. Rev. 78, 928–938. https://doi. org/10.1093/nutrit/nuaa005.

Amin, K. (2012). The role of mast cells in allergic inflammation. Respir. Med. 106, 9–14. https://doi. org/10.1016/j.rmed.2011.09.007.

Arruda, L.K., Solé, D., Baena-Cagnani, C.E., and Naspitz, C.K. (2005). Risk factors for asthma and atopy. Curr. Opin. Allergy Clin. Immunol. 5, 153–159. https://doi.org/10.1097/01.all. 0000162308.89857.6c.

Arshi, M., Cardinal, J., Hill, R.J., Davies, P.S., and Wainwright, C. (2010). Asthma and insulin resistance in children. Respirology 15, 779–784. https://doi.org/10.1111/j.1440-1843.2010. 01767.x.

Bener, A., Ehlayel, M.S., Tulic, M.K., and Hamid, Q. (2012). Vitamin D deficiency as a strong predictor of asthma in children. Int. Arch. Allergy Immunol. 157, 168–175. https://doi.org/10.1159/ 000323941.

Berthon, B.S., and Wood, L.G. (2015). Nutrition and respiratory health—feature review. Nutrients 7, 1618–1643. https://doi.org/10.3390/ nu7031618.

Beuther, D.A., Weiss, S.T., and Sutherland, E.R. (2006). Obesity and asthma. Am. J. Respir. Crit. Care Med. 174, 112–119. https://doi.org/10.1164/rccm.200602-231pp.

Bikle, D.D. (2012). Vitamin D and bone. Curr. Osteoporos. Rep. 10, 151–159. https://doi.org/ 10.1007/s11914-012-0098-z.

Burleson, J.D., Siniard, D., Yadagiri, V.K., Chen, X., Weirauch, M.T., Ruff, B.P., Brandt, E.B., Hershey, G.K.K., and Ji, H. (2019). TET1 contributes to allergic airway inflammation and regulates interferon and aryl hydrocarbon receptor signaling pathways in bronchial epithelial cells. Sci. Rep. 9, 7361. https://doi.org/ 10.1038/s41598-019-43767-6.

Cao, J.J. (2011). Effects of obesity on bone metabolism. J. Orthop. Surg. Res. 6, 30. https://doi.org/10.1186/1749-799x-6-30.

Costello, L.C., and Franklin, R.B. (2016). Plasma citrate homeostasis: how it is regulated; and its



physiological and clinical implications. An important, but neglected, relationship in medicine. HSOA J. Hum. Endocrinol. 1, 005. https://doi.org/10.24966/he-9640/100005.

Delmastro-Greenwood, M., Freeman, B.A., and Wendell, S.G. (2014). Redox-dependent antiinflammatory signaling actions of unsaturated fatty acids. Annu. Rev. Physiol. 76, 79–105. https:// doi.org/10.1146/annurev-physiol-021113-170341.

Ebell, M.H., Hall, S.P., Rustin, R.C., Powell-Threets, K., Munoz, L., Toodle, K., Meng, M.L., and O'Connor, J. (2019). A multicomponent, multi-trigger intervention to enhance asthma control in high-risk African American children. Prev. Chronic Dis. 16, E69. https://doi.org/10. 5888/pcd16.180387.

Fintini, D., Cianfarani, S., Cofini, M., Andreoletti, A., Ubertini, G.M., Cappa, M., and Manco, M. (2020). The bones of children with obesity. Front. Endocrinol. 11, 200. https://doi.org/10.3389/ fendo.2020.00200.

Fogarty, A., Broadfield, E., Lewis, S., Lawson, N., and Britton, J. (2004). Amino acids and asthma: a case-control study. Eur. Respir. J. 23, 565–568. https://doi.org/10.1183/09031936.04.00090404.

Gerber, D.A., Tanenbaum, L., and Ahrens, M. (1976). Free serum histidine levels in patients with rheumatoid arthritis and control subjects following an oral load of free L-histidine. Metabolism 25, 655–657. https://doi.org/10. 1016/0026-0495(76)90062-7.

Hardin, A.P., Hackell, J.M., Simon, G.R., Boudreau, A.D.A., Baker, C.N., Barden, G.A., Meade, K.E., Moore, S.B., and Richerson, J.; Committee on practice and ambulatory medicine (2017). Age limit of pediatrics. Pediatrics 140, e20172151. https://doi.org/10.1542/peds.2017-2151.

Hernández, M.A.G., Canfora, E.E., Jocken, J.W.E., and Blaak, E.E. (2019). The short-chain fatty acid acetate in body weight control and insulin sensitivity. Nutrients *11*, 1943. https://doi.org/10.3390/nu11081943.

Holeček, M. (2020). Histidine in health and disease: metabolism, physiological importance, and use as a supplement. Nutrients 12, 848. https://doi.org/10.3390/nu12030848.

Hou, J., He, C., He, W., Yang, M., Luo, X., and Li, C. (2020). Obesity and bone health: a complex link. Front. Cell Dev. Biol. *8*, 600181. https://doi. org/10.3389/fcell.2020.600181.

Huang, S.K. (2020). A fresh take on the "TCA" cycle: TETs, citrate, and asthma. Am. J. Respir. Cell Mol. Biol. 63, 1–3. https://doi.org/10.1165/ rcmb.2020-0101ED.

James, E.L., and Parkinson, E.K. (2015). Serum metabolomics in animal models and human disease. Curr. Opin. Clin. Nutr. Metab. Care 18, 478–483. https://doi.org/10.1097/mco. 0000000000000200.

Jürimäe, J. (2010). Interpretation and application of bone turnover markers in children and adolescents. Curr. Opin. Pediatr. 22, 494–500. https://doi.org/10.1097/MOP. 0b013e32833b0b9e. Kelly, R.S., Dahlin, A., McGeachie, M.J., Qiu, W., Sordillo, J., Wan, E.S., Wu, A.C., and Lasky-Su, J. (2017). Asthma metabolomics and the potential for integrative omics in research and the clinic. Chest 151, 262–277. https://doi.org/10.1016/j. chest.2016.10.008.

Lepage, N., McDonald, N., Dallaire, L., and Lambert, M. (1997). Age-specific distribution of plasma amino acid concentrations in a healthy pediatric population. Clin. Chem. 43, 2397–2402. https://doi.org/10.1093/clinchem/43.12.2397.

Moffett, J.R., Puthillathu, N., Vengilote, R., Jaworski, D.M., and Namboodiri, A.M. (2020). Acetate revisited: a key biomolecule at the nexus of metabolism, epigenetics and oncogenesis— Part 1: acetyl-CoA, acetogenesis and Acyl-CoA short-chain synthetases. Front. Physiol. 11, 580167. https://doi.org/10.3389/fphys.2020. 580167.

Morris, C.R., Hamilton-Reeves, J., Martindale, R.G., Sarav, M., and Ochoa Gautier, J.B. (2017). Acquired amino acid deficiencies: a focus on arginine and glutamine. Nutr. Clin. Pract. 32, 30S-47S. https://doi.org/10.1177/ 0844533617691250.

Nambiar, S., Bong How, S., Gummer, J., Trengove, R., and Moodley, Y. (2020). Metabolomics in chronic lung diseases. Respirology 25, 139–148. https://doi.org/10. 1111/resp.13530.

Niu, Y.-C., Feng, R.-N., Hou, Y., Li, K., Kang, Z., Wang, J., Sun, C.-H., and Li, Y. (2012). Histidine and arginine are associated with inflammation and oxidative stress in obese women. Br. J. Nutr. 108, 57–61. https://doi.org/10.1017/ S0007114511005289.

Oliveira, G.P., de Abreu, M.G., Pelosi, P., and Rocco, P.R.M. (2016). Exogenous glutamine in respiratory diseases: myth or reality? Nutrients *8*, 76. https://doi.org/10.3390/nu8020076.

Papamichael, M.M., Katsardis, C., Sarandi, E., Georgaki, S., Frima, E.-S., Varvarigou, A., and Tsoukalas, D. (2021). Application of metabolomics in pediatric asthma: prediction, diagnosis and personalized treatment. Metabolites 11, 251. https://doi.org/10.3390/ metabo11040251.

Peters, U., Dixon, A.E., and Forno, E. (2018). Obesity and asthma. J. Allergy Clin. Immunol. 141, 1169–1179. https://doi.org/10.1016/j.jaci. 2018.02.004.

Petrus, P., Lecoutre, S., Dollet, L., Wiel, C., Sulen, A., Gao, H., Tavira, B., Laurencikiene, J., Rooyackers, O., Checa, A., et al. (2020). Glutamine links obesity to inflammation in human white adipose tissue. Cell Metab. *31*, 375– 390.e11. https://doi.org/10.1016/j.cmet.2019.11. 019.

Poynter, M., Ather, J., Reed, L., Barup, M., and Mank, M. (2020). Ketone body augmentation decreases methacholine hyperresponsiveness in a mouse model of house dust mite-induced allergic asthma. In A31. Asthma Translational Studies (American Thoracic Society), p. A1297.

Puchalska, P., and Crawford, P.A. (2021). Metabolic and signaling roles of ketone bodies in health and disease. Annu. Rev. Nutr. 41, 49–77.

#### https://doi.org/10.1146/annurev-nutr-111120-111518.

**iScience** 

Article

Reinke, S.N., Gallart-Ayala, H., Gómez, C., Checa, A., Fauland, A., Naz, S., Kamleh, M.A., Djukanović, R., Hinks, T.S., and Wheelock, C.E. (2017). Metabolomics analysis identifies different metabotypes of asthma severity. Eur. Respir. J. 49, 1601740. https://doi.org/10.1183/13993003. 01740-2016.

Rojas-Morales, P., Pedraza-Chaverri, J., and Tapia, E. (2020). Ketone bodies, stress response, and redox homeostasis. Redox Biol. 29, 101395. https://doi.org/10.1016/j.redox.2019.101395.

Saudubray, J.M., Marsac, C., Limal, J.M., Dumurgier, E., Charpentier, C., Ogier, H., and Coudè, F.X. (1981). Variation in plasma ketone bodies during a 24-hour fast in normal and in hypoglycemic children: relationship to age. J. Pediatr. 98, 904–908. https://doi.org/10.1016/ S0022-3476(81)80583-5.

Skutches, C.L., Holroyde, C.P., Myers, R.N., Paul, P., and Reichard, G.A. (1979). Plasma acetate turnover and oxidation. J. Clin. Invest. 64, 708–713. https://doi.org/10.1172/jci109513.

Smith, P.M., Howitt, M.R., Panikov, N., Michaud, M., Gallini, C.A., Bohlooly-y, M., Glickman, J.N., and Garrett, W.S. (2013). The microbial metabolites, short-chain fatty acids, regulate colonic Treg cell homeostasis. Science 341, 569–573. https://doi.org/10.1126/science. 1241165.

Sun, X., Feng, R., Li, Y., Lin, S., Zhang, W., Li, Y., Sun, C., and Li, S. (2014). Histidine supplementation alleviates inflammation in the adipose tissue of high-fat diet-induced obese rats via the NF-κB-and PPARγ-involved pathways. Br. J. Nutr. *112*, 477–485. https://doi.org/10.1017/ s0007114514001056.

Swanson, K.V., Deng, M., and Ting, J.P.Y. (2019). The NLRP3 inflammasome: molecular activation and regulation to therapeutics. Nat. Rev. Immunol. 19, 477–489. https://doi.org/10.1038/ s41577-019-0165-0.

Tesse, R., Schieck, M., and Kabesch, M. (2011). Asthma and endocrine disorders: shared mechanisms and genetic pleiotropy. Mol. Cell. Endocrinol. 333, 103–111. https://doi.org/10. 1016/j.mce.2010.11.032.

Viljakainen, H., Ivaska, K.K., Paldánius, P., Lipsanen-Nyman, M., Saukkonen, T., Pietiläinen, K.H., Andersson, S., Laitinen, K., and Mäkitie, O. (2014). Suppressed bone turnover in obesity: a link to energy metabolism? A case-control study. J. Clin. Endocrinol. Metab. *9*, 2155–2163. https:// doi.org/10.1210/jc.2013-3097.

Vinding, R.K., Stokholm, J., Chawes, B.L.K., and Bisgaard, H. (2016). Blood lipid levels associate with childhood asthma, airway obstruction, bronchial hyperresponsiveness, and aeroallergen sensitization. J. Allergy Clin. Immunol. 137, 68– 74.e4. https://doi.org/10.1016/j.jaci.2015.05.033.

Wendell, S.G., Baffi, C., and Holguin, F. (2014). Fatty acids, inflammation, and asthma. J. Allergy Clin. Immunol. 133, 1255–1264. https://doi.org/ 10.1016/j.jaci.2013.12.1087.

Wood, L., Gibson, P., and Garg, M. (2003). Biomarkers of lipid peroxidation, airway

### iScience Article



inflammation and asthma. Eur. Respir. J. 21, 177–186. https://doi.org/10.1183/09031936.03. 00017003a.

Würtz, P., Kangas, A.J., Soininen, P., Lawlor, D.A., Davey Smith, G., and Ala-Korpela, M. (2017). Quantitative serum nuclear magnetic resonance metabolomics in large-scale epidemiology: a primer on-omic technologies. Am. J. Epidemiol. 186, 1084–1096. https://doi.org/10.1093/aje/ kwx016.

Yamauchi, K. (1996). Regulation of gene expression of L-histidine decarboxylase and histamine N-methyl-transferase, and its relevance to the pathogenesis of bronchial asthma. Nihon. Rinsho *54*, 377–388.

Yamauchi, K., and Ogasawara, M. (2019). The role of histamine in the pathophysiology of asthma and the clinical efficacy of antihistamines in asthma therapy. Int. J. Mol. Sci. 20, 1733. https://doi.org/10.3390/ ijms20071733.

Yeung, B.H.Y., Huang, J., An, S.S., Solway, J., Mitzner, W., and Tang, W.Y. (2020). Role of isocitrate dehydrogenase 2 on DNA hydroxymethylation in human airway smooth muscle cells. Am. J. Respir. Cell Mol. Biol. 63, **36–45**. https://doi.org/10.1165/rcmb.2019-0323OC.

Youm, Y.-H., Nguyen, K.Y., Grant, R.W., Goldberg, E.L., Bodogai, M., Kim, D., D'agostino, D., Planavsky, N., Lupfer, C., Kanneganti, T.D., et al. (2015). The ketone metabolite  $\beta$ -hydroxybutyrate blocks NLRP3 inflammasomemediated inflammatory disease. Nat. Med. 21, 263–269. https://doi.org/10.1038/nm.3804.

Zhao, C., and Zhao, W. (2020). NLRP3 inflammasome—a key player in antiviral responses. Front. Immunol. 11, 211. https://doi. org/10.3389/fimmu.2020.00211.





#### **STAR\*METHODS**

#### **KEY RESOURCES TABLE**

REAGENT or RESOURCE	SOURCE	IDENTIFIER
Biological samples		
Plasma	The Committees for the Protection of Human Subjects (IRB), The Children's Hospital of Philadelphia (CHOP)	IRB 16-013278; IRB 10-007590
Deposited data		
Metabolomics data	MetaboLights (Metabolomics experiments and derived information, https://www.ebi.ac.uk/ metabolights/)	MTBLS5024

#### **RESOURCE AVAILABILITY**

#### Lead contact

Further information and requests for resources and reagents should be directed to and will be fulfilled by the lead contact, Dr. Hakon Hakonarson (hakonarson@chop.edu).

#### **Materials availability**

This study did not generate new unique reagents.

#### Data and code availability

- The metabolomic data presented in this study is available through Metabolights (MTBLS5024, www.ebi. ac.uk/metabolights/MTBLS5024).
- There is no original code associated with this work.
- Any additional information required to reanalyze the data reported in this paper is available from the lead contact upon request.

#### **EXPERIMENTAL MODEL AND SUBJECT DETAILS**

#### Patient recruitment and sample collection

This study included 602 pediatric cases (with the upper age limit as 21 years (Hardin et al., 2017)) with asthma (299 males and 303 females, age range 6–21 years old, and 593 controls without asthma (265 males and 328 females, age range 6–21 years old. All cases were recruited at the Center for Applied Genomics (CAG) at the Children's Hospital of Philadelphia (CHOP), with the ancestry of African American confirmed by the genome-wide single nucleotide polymorphism (SNP) genotyping and principal component analysis of population structure. No individuals had a history of sickle cell disease, type 1 diabetes, 22q11.2 deletion syndrome, or human immunodeficiency virus (HIV) infection. Obesity and vitamin D (VitD) deficiency were common in both cases and controls (Figure 1, Table S1). As expected, both obesity and VitD deficiency were more common in asthma cases as both are related to the risk of asthma (Bener et al., 2012; Beuther et al., 2006). This study was approved by the Institutional Review Board (IRB) of the Children's Hospital of Philadelphia. Personal information is all encrypted to ensure no protected health information (PHI) is included and study participants are de-identified. All human subjects or their proxies provided written informed consent.

#### **METHOD DETAILS**

Plasma samples were collected and kept according to standard procedures for lipid testing. In a sample of 300 μL, metabolites are quantified in absolute concentrations (e.g. mmol/l) and percentages. 249 metabolites of 18 groups were measured using a NMR metabolomics platform (Table S2, Nightingale Health Ltd;





biomarker quantification version 2020). Absolute concentrations of the metabolites was based on the Bayesian modeling that converted the spectral information by integrated quality control in the NMR spectrometer (Würtz et al., 2017).

#### QUANTIFICATION AND STATISTICAL ANALYSIS

Metabolite levels were normalized with means and standard deviation (STD), i.e. normalized value =  $(X - \overline{X})/s$ , where: x = metabolite level,  $\overline{X}$  = mean, s = STD. Point-Biserial partial correlations between asthma and metabolite levels were controlled for age, sex, obesity, and VitD deficiency, as covariates (Table S3, Related to Table 1. Correlation between asthma and metabolite levels). Statistical analysis was done by the IBM SPSS Statistics Version 23 software.