

RESEARCH

Open Access

Cytoplasmic-targeted parvalbumin blocks the proliferation of multipotent mesenchymal stromal cells in prophase

Carolina Soares Barros Melo, Jerusa Araújo Quintão Arantes Faria, Natássia Caroline Resende Corrêa, Carolina de Andrade, Juliana Lott Carvalho, Alfredo M Goes, Michele A Rodrigues and Dawidson Assis Gomes*

Abstract

Introduction: Multipotent mesenchymal stromal cells (MSCs) have gained considerable interest because of their potential use in the treatment of a variety of diseases and injuries. Although remarkable advancements have been made in clinical studies, substantial concerns still regard the safety of MSCs. Some evidence suggests that MSCs can spontaneously generate a population of cells with tumorigenic potential. Thus, studying the molecular mechanisms that control the proliferation of MSCs may be a necessary step toward the development of strategies for safe clinical practice. Ca^{2+} is a second messenger that mediates a wide range of cellular responses, including the regulation of cell proliferation, but little is known about its function in MSCs. The aim of this study was to investigate the effects of targeted Ca^{2+} buffering on MSCs proliferation *in vitro*.

Methods: Here, we used an adenoviral (Ad) vector encoding the Ca^{2+} chelator protein parvalbumin (PV) fused to a nuclear exclusion signal (NES) and the *Discosoma* red fluorescent protein (DsRed) to investigate the function of cytoplasmic Ca^{2+} signals on MSC proliferation. Confocal microscopy was used to demonstrate that PV-NES-DsRed was expressed in the cytoplasm. Ca^{2+} signaling was monitored by using Fluo-4-AM. Fluorescence-activated cell sorting (FACS) analysis of cells that were stained with propidium iodide was used as a quantitative measure of cell death. The mitotic index was assessed by immunofluorescence, and the expression of cyclins was examined with Western blot.

Results: Our results show that the Ad-PV-NES-DsRed fusion protein decreased serum-induced Ca^{2+} signaling and blocked the proliferation of rat adipose-derived MSCs (AT-MSCs) in prophase. FACS analysis revealed that Ad-PV-NES-DsRed did not induce cell death in AT-MSCs. Furthermore, Western blot analysis demonstrated that Ad-PV-NES-DsRed reduced extracellular signal-regulated kinase (Erk1/2) phosphorylation and cyclin B1 expression. Buffering cytosolic Ca^{2+} did not alter the expression of cyclins A/D1/D2/D3/E and E2.

Conclusions: Our results show that cytoplasmic Ca^{2+} signals are important for AT-MSCs progression beyond prophase because of their effects on Erk phosphorylation and cyclin B1 expression.

Keywords: Mesenchymal stem cells, Targeted parvalbumin, Calcium signaling, Proliferation, Cyclins

Introduction

Multipotent mesenchymal stromal cells (MSCs) are nonhematopoietic stromal cells that have generated a great amount of interest in the field of regenerative medicine because of their unique biological properties. These cells give rise to diverse tissues, including bone,

cartilage, tendon, muscle, and adipose tissue [1], and they have been isolated from different sources, such as bone marrow, adipose tissue, peripheral blood, muscle, umbilical blood, placenta, and other sites [2,3]. These cells are relatively easy to obtain and have a remarkable capacity for extensive *in vitro* expansion, which allows them rapidly to reach the cell number required for *in vivo* therapy. In addition to their secretion of multiple bioactive molecules

* Correspondence: dawidson.gomes@gmail.com

Department of Biochemistry and Immunology, Universidade Federal de Minas Gerais, Av. Antonio Carlos 6627, Belo Horizonte-MG ZIP Code: 31270-901, Brazil

with trophic effects, MSCs are able to migrate and to exert immunomodulatory activities [4].

Over the past decade, many publications on MSCs reported experimental and clinical applications for these cells and demonstrated encouraging results. Although tremendous advancements have been made in clinical studies, substantial challenges remain and must be overcome before MSCs therapy can fulfill its promise in wider clinical practice [5].

The first major obstacle is definitively to determine the safety of MSCs. A few studies support the idea that MSCs suppress tumor growth, whereas others state that MSCs may contribute indirectly to cancer by antiapoptotic effects that protect tumors and by the promotion of tumor progression, metastasis, and drug resistance. Alternatively, MSCs may be directly involved in cancer development through malignant transformation [6].

Some evidence suggests that these cells can spontaneously transform to generate a population of cells with tumorigenic potential through the acquisition of point mutations [7]. Some of these mutations are involved in the expression of molecules that regulate the cell cycle and cell proliferation [8]. Thus, studying the molecular mechanisms involved in the cell cycle and proliferation of MSCs may be the first step toward the development of control strategies that prevent the proliferation of MSCs that have undergone tumorigenic transformation after transplantation for safe clinical practice.

Ca^{2+} is a second messenger that contributes to the cell cycle and cell proliferation. In mammalian somatic cells, the importance of intracellular Ca^{2+} signaling during cell-cycle progression is well established [9], but little is known about the effects of Ca^{2+} signals in the cell cycle and proliferation of MSCs. The first studies in this area used the microinjection of dextran-linked Ca^{2+} buffers, but a more-efficient approach was subsequently developed that used the targeted expression of Ca^{2+} -buffering proteins, such as parvalbumin (PV) or calretinin [10,11]. Here, we used a construct encoding PV that was targeted to the cytoplasm by a nuclear export signal (PV-NES), and we delivered these constructs to MSCs by using an adenovirus (Ad) expression system to achieve high-efficiency gene delivery. We used this strategy to examine the effects of targeted Ca^{2+} buffering on cell populations. The PV protein was fused to the *Discosoma* red fluorescent protein (DsRed) to monitor its expression and subcellular localization. The aim of this study was to investigate the effects of Ad-PV-NES-DsRed on MSCs proliferation *in vitro*.

We found that Ad-PV-NES-DsRed could block MSC proliferation during prophase because of its effects on Erk phosphorylation and cyclin B1 expression. Thus, this tool might provide a new perspective for understanding and controlling MSC proliferation.

Methods

Cell isolation and culture

Male Wistar rats (6 to 8 weeks old), which were obtained from the Centro de Bioterismo da Universidade Federal de Minas Gerais (CEBIO), were used for all of the studies. Rat inguinal adipose tissue was dissected into small pieces, digested with 0.15% collagenase B (Roche Applied Science, Indianapolis, IN, USA) in 0.1 M phosphate-buffered saline (PBS) for 40 minutes at 37°C. Mature adipocytes and connective tissues were separated from the cell pellet by centrifugation at 274.4 g for 10 minutes at room temperature. The cell pellet was resuspended in high-glucose Dulbecco modified Eagle medium (DMEM) (Sigma-Aldrich, St. Louis, MO, USA) with 10% fetal bovine serum (FBS) and 100 U/ml penicillin/streptomycin (Life Technologies, Carlsbad, CA, USA). The current protocol was adapted from a previously described protocol [12].

The initial passage of the primary cell culture was referred to as passage 0. We used cells in passage 3 only, except in the indicated experiments. The cells were maintained in complete media (DMEM + 10% FBS + penicillin/streptomycin) until they achieved 75% to 90% confluence. The cells were then replated in T75 tissue-culture flasks. The identity of the MSCs was defined by the criteria proposed by the International Society for Cellular Therapy [13]. The experiments were approved by the local ethical committee on animal experimentation (CETEA/UFMG; protocol number 239/10).

Phenotypic analysis with flow cytometry

MSCs were recognized by using specific antibodies against CD54, CD73, and CD90. In addition, we assessed the lack of expression of the CD45 hematopoietic marker. All antibodies were purchased from Becton Dickinson (BD) Biosciences (San Jose, CA, USA). The secondary antibodies used were conjugated to Alexa Fluor 488 (Life Technologies).

For immunophenotypic analysis, the MSCs were detached by incubation with 0.05% trypsin-EDTA (Life Technologies) for 5 minutes, immediately resuspended in DMEM plus 10% FBS, and plated in 96-well culture plates at a density of 2×10^5 cells/well. Next, the cells were washed with PBS, and the cell suspension (100 μl) was incubated at 4°C for 30 minutes with the primary antibody (1:50) or with an isotype antibody (1:50) as a negative control. Then, the plate was centrifuged at 274.4 g for 10 minutes at 10°C. After two washes with PBS, the cell suspension was incubated at 4°C for 30 minutes with the secondary antibody (1:200). Subsequently, the cells were washed with PBS and analyzed with FACSscan flow cytometry by using CELLQuest software (BD Biosciences). All fluorescence-activated cell-sorting (FACS) data were analyzed by using FlowJo software (Tree Star, San Carlos, CA, USA).

Adenoviral constructs

Adenoviral constructs encoding DsRed (Ad-DsRed) and the PV-DsRed fusion protein that was targeted to the cytosol (Ad-PV-NES-DsRed) were obtained as previously described [10]. The adenoviruses were amplified in HEK-293 cells, a cell line that constitutively expresses the E1 protein. Both viruses (Ad-DsRed and Ad-PV-NES-DsRed) were generated by using standard procedures and stored at -80°C in PBS plus 7% glycerol. Stocks of the adenoviral particles were quantified as plaque-forming units (pfus) by using plaque assays in HEK-293 cells. On day 1 of culture, each adenovirus construct (at a multiplicity of infection (MOI) of 100) was diluted in regular culture medium (DMEM) containing 10% FBS. The experiments were performed at 36 to 48 hours, when approximately 70% of the cells were transduced.

Ca²⁺ measurements

MSCs were cultured in 96-well culture plates at a density of 1×10^4 cells/well and then transduced with the adenoviral constructs. After 24 hours, the cells were synchronized to G₀ by serum starvation overnight. The next morning, the cells were washed in PBS and resuspended in HEPES buffer containing 3 μM Fluo-4-AM (Life Technologies) for 30 minutes at 37°C . Next, the cells were washed, resuspended in HEPES, and then stimulated with HEPES containing 10% FBS. The resultant fluorescent signal was monitored for the whole well over time by using a Synergy 2 multimode plate reader (BioTek, Winooski, VT, USA). The wavelength for excitation was 485/20 nm; emission was measured at 530/25 nm, and the dichroic that was used was the top 50%. The experiments were performed at 37°C . The mean peaks of the maximum and minimum fluorescence were analyzed by using Excel and plotted by using GraphPad software (La Jolla, CA, USA).

Analysis of cell viability

To assess the cell viability of the MSCs, we used the 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assay (Life Technologies), as previously described [14]. In brief, the MSCs were plated in 24-well culture plates at a density of 1×10^5 cells/cm² per well. MTT (5 mg/ml) was added to each well of the monolayer cultures, and the cultures were incubated in a humidified 5% CO₂ incubator at 37°C . Two hours later, the cell morphologies and formazan salts were visualized by using an inverted optical microscope. The formazan salts were dissolved with 10% SDS-HCl overnight, and the optical density was measured at 595 nm by using a microplate reader (Elx800; BioTek). Staurosporine was used as a positive control for cell death at a concentration of 100 nM in DMEM containing 10% FBS. The values from the samples are expressed as percentages of the control. All data were analyzed by using GraphPad software.

Analysis of cell death by flow cytometry

A flow-cytometric DNA fragmentation assay was used as a quantitative measure of cell death [15]. Forty-eight hours after transducing the MSCs with the adenoviral constructs, the cells were collected by centrifugation and lysed with 300 μl of a hypotonic solution containing 0.1% sodium citrate, 0.1% Triton X-100, and 20 $\mu\text{g}/\text{ml}$ propidium iodide (PI; Life Technologies). Then the cells were incubated at room temperature for 1 hour and analyzed with a FACScan flow cytometer (BD Biosciences) for shifts in PI fluorescence, which are indicative of nuclei with hypodiploid DNA content. All data were analyzed by using FlowJo software (Tree Star).

Mitotic index measurements by immunofluorescence

Confocal immunofluorescence was performed as previously described [10,16]. In brief, the cells were labeled with an anti-phospho-histone-3 monoclonal antibody (1:500; Millipore, Billerica, MA, USA) and then incubated with an Alexa Fluor 488-conjugated secondary antibody (1:1,000; Life Technologies) and the Hoechst nuclear stain (200 ng/ml; Life Technologies). Images were captured by using a Zeiss LSM 510 confocal microscope with a 63 \times /1.4 NA objective lens. The samples were excited at 488 nm and observed at 505 to 550 nm to detect Alexa Fluor 488, excited at 405 nm, and observed at 420 to 460 nm to detect Hoechst staining. Mitotic indexes were scored for the cells in each phase of mitosis according to the phospho-histone-3 distribution and DNA condensation pattern [10].

Immunoblotting

Immunoblotting was performed by using standard methods [10,16]. In brief, the cells, which were grown in T75 tissue-culture flasks, were washed 3 times with PBS and lysed in buffer containing 150 mM NaCl, 1 mM EDTA, 20 mM Tris-HCl, 0.5% Nonidet P-40, and a protease-inhibitor mixture (Sigma-Aldrich). The protein concentration was determined spectrophotometrically by using the Bradford method (Sigma-Aldrich), and 40 μg of protein was separated by electrophoresis in a 4% to 15% polyacrylamide gel and then transferred to a polyvinylidene difluoride membrane (Millipore). The membrane was blocked with 5% skim milk in Tris-buffered saline (TBS) plus 0.5% Tween 20 (TBST) for 60 minutes and then incubated with primary antibodies against total Erk1/2 or phospho-Erk1/2 (1:1,000; Cell Signaling Technology, Beverly, MA, USA), cyclins A, B1, D1, D2, D3, E, or E2 (1:500 to 1,000, Cell Signaling Technology) or GAPDH (1:500; Santa Cruz Biotechnology, Santa Cruz, CA, USA). The incubations with the primary antibodies were carried out overnight. After three 5-minute washes with TBST, the membranes were incubated with the appropriate horseradish peroxidase-conjugated secondary antibody (1:5,000) (Sigma-Aldrich)

for 1 hour at room temperature. After three additional 5-minute washes with TBST, the membranes were developed by using enhanced chemiluminescence (ECL Plus; GE Healthcare Life Sciences, Piscataway, NJ, USA). Subsequently, the films were scanned and analyzed by using Image J software.

Statistical analysis

All of the data shown represent at least three independent experiments and are expressed as the mean \pm standard error of the mean (SEM). The statistical analyses were performed with GraphPad software. The experiments with more than two data groups were compared by using one-way ANOVA and the Bonferroni posttest. *P* values of <0.05 were considered to be statistically significant.

Results

Characterization of rat MSCs

Rat MSCs were characterized according to the three criteria proposed by the Mesenchymal and Tissue Stem Cell Committee of the International Society for Cellular Therapy [13]. First, the cell-surface antigen profile was ascertained by staining the cells with rat-specific monoclonal antibodies, followed by flow-cytometric analyses, as shown in Figure 1. We investigated the expression of the cell markers CD54 ($99.1\% \pm 0.4\%$), CD73 ($99.5\% \pm 0.2\%$), CD90 ($90.3\% \pm 0.6\%$), and CD45 ($10.3\% \pm 0.3\%$). Second, we demonstrated that the MSCs were plastic-adherent when maintained in standard culture conditions, and fibroblast-like, as they appeared polygon-like or spindle-like with processes (data not shown). Third, we showed that the MSCs were capable of differentiating into osteoblasts and adipocytes (data not shown).

Cytoplasmic-targeted parvalbumin decreases agonist-induced Ca^{2+} signals

The relative role of cytoplasmic Ca^{2+} signals in specific cell functions can be determined by selectively attenuating Ca^{2+} increases in the cytoplasm with Ad-PV-NES-DsRed, as shown previously [10,17]. In this study, confocal microscopy demonstrated that PV-NES-DsRed was expressed in the cytoplasm and that DsRed alone was expressed in both the cytoplasm and nucleus (Figure 2A). We used 10% serum stimulation to test the efficiency of Ad-PV-NES-DsRed in attenuating agonist-induced Ca^{2+} signaling in MSCs. The expression of Ad-PV-NES-DsRed was able to attenuate locally $74\% \pm 2\%$ of the cytoplasmic Ca^{2+} signals (Figure 2B).

Cytosolic Ca^{2+} blocks MSC proliferation at prophase

To determine the involvement of cytoplasmic Ca^{2+} in MSC proliferation, the effect of the expression of our Ca^{2+} chelator fusion protein, PV-NES-DsRed, in MSCs was examined. First, the cells were synchronized to G_0 by

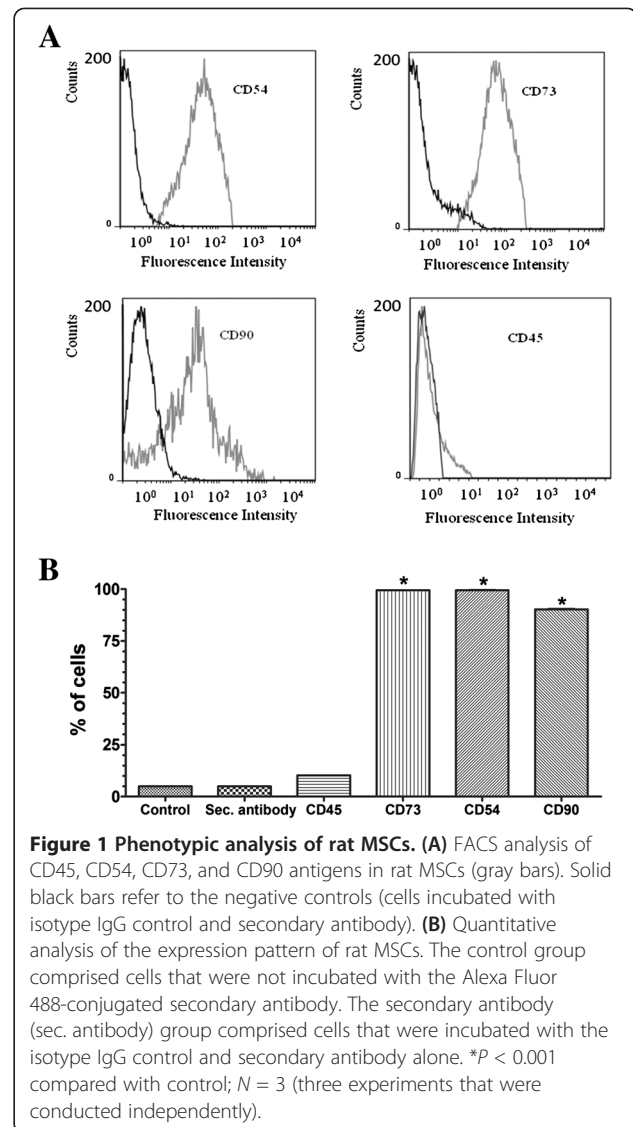
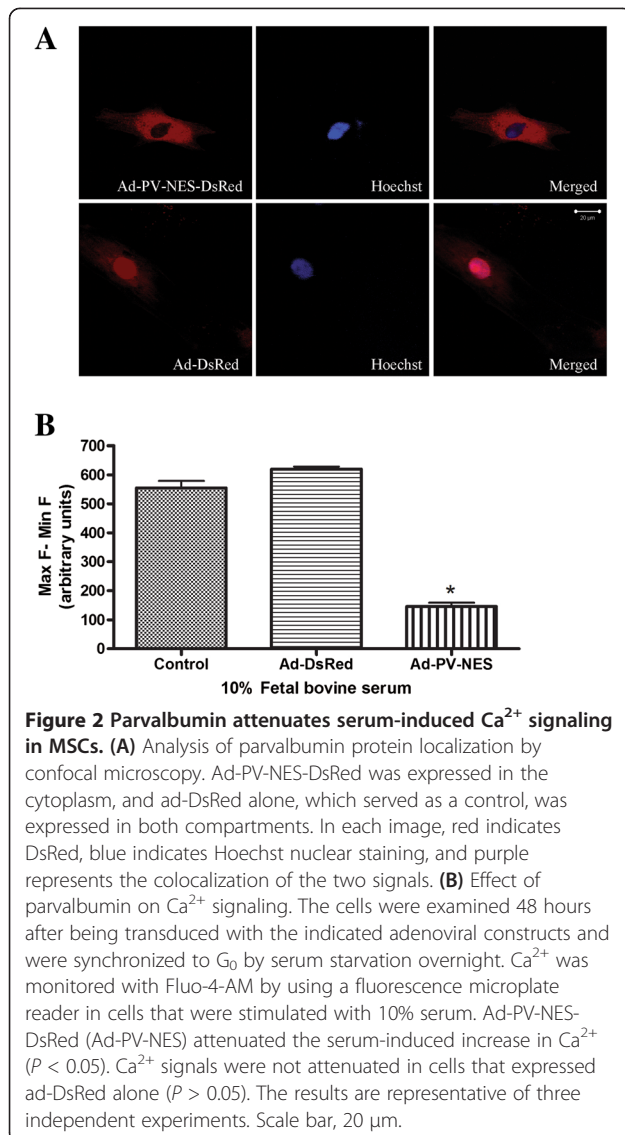


Figure 1 Phenotypic analysis of rat MSCs. (A) FACS analysis of CD45, CD54, CD73, and CD90 antigens in rat MSCs (gray bars). Solid black bars refer to the negative controls (cells incubated with isotype IgG control and secondary antibody). **(B)** Quantitative analysis of the expression pattern of rat MSCs. The control group comprised cells that were not incubated with the Alexa Fluor 488-conjugated secondary antibody. The secondary antibody (sec. antibody) group comprised cells that were incubated with the isotype IgG control and secondary antibody alone. **P* < 0.001 compared with control; *N* = 3 (three experiments that were conducted independently).

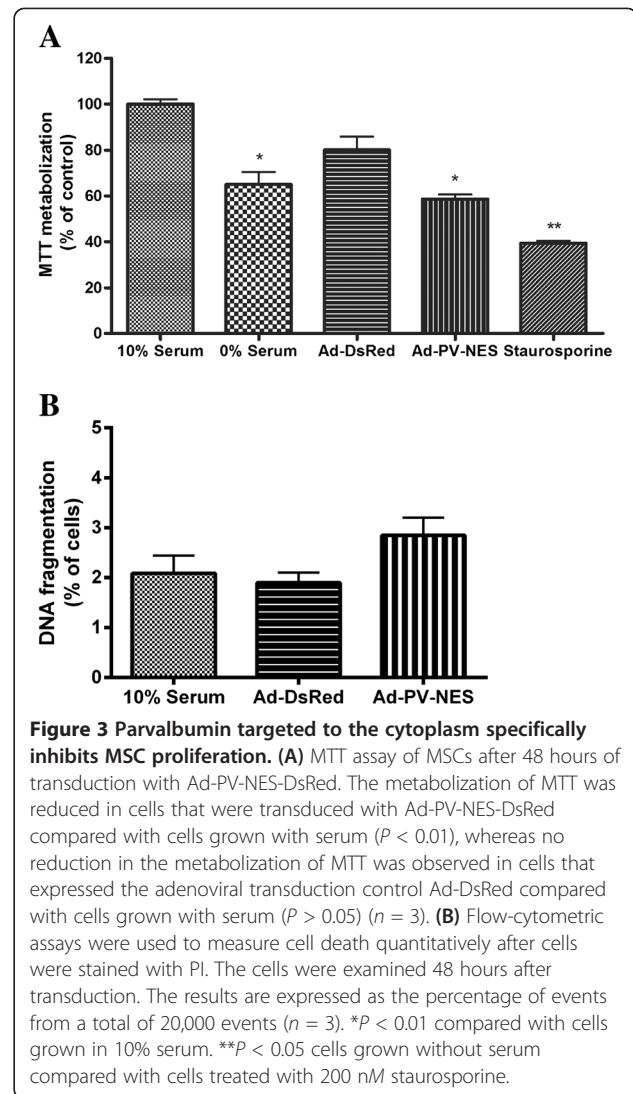
serum starvation and then transduced with the adenoviral constructs. Next, the cells were stimulated with complete medium to reenter the cell cycle. After 48 hours, the metabolization of MTT was reduced in cells that were transduced with Ad-PV-NES-DsRed compared with cells grown without serum, whereas no reduction in the metabolization of MTT was observed in cells that expressed the adenoviral transduction control Ad-DsRed compared with cells grown with serum (Figure 3A). Furthermore, flow-cytometric analysis of cells stained with propidium iodide showed that Ad-PV-NES had no effect on cell death (Figure 3B). These results suggest that the effects of Ad-PV-NES were specific to MSCs proliferation.

Second, to investigate the effects of cytoplasmic Ca^{2+} on mitosis, the MSCs were labeled with the mitotic marker phospho-histone-3. After 48 hours of adenoviral



transduction, the cells were examined with confocal microscopy to determine the fraction of cells in mitosis (Figure 4B). The mitotic index was increased to $61\% \pm 2\%$ in cells with buffered cytoplasmic Ca^{2+} ($P < 0.001$). In addition, 49.9% of the cells in mitosis were in prophase (Figure 5B). No significant effect was observed upon comparison of uninfected cells with Ad-DsRed-transduced cells (Figure 4B). These results demonstrate that cytoplasmic Ca^{2+} is important for the cell-cycle transition of MSCs beyond prophase.

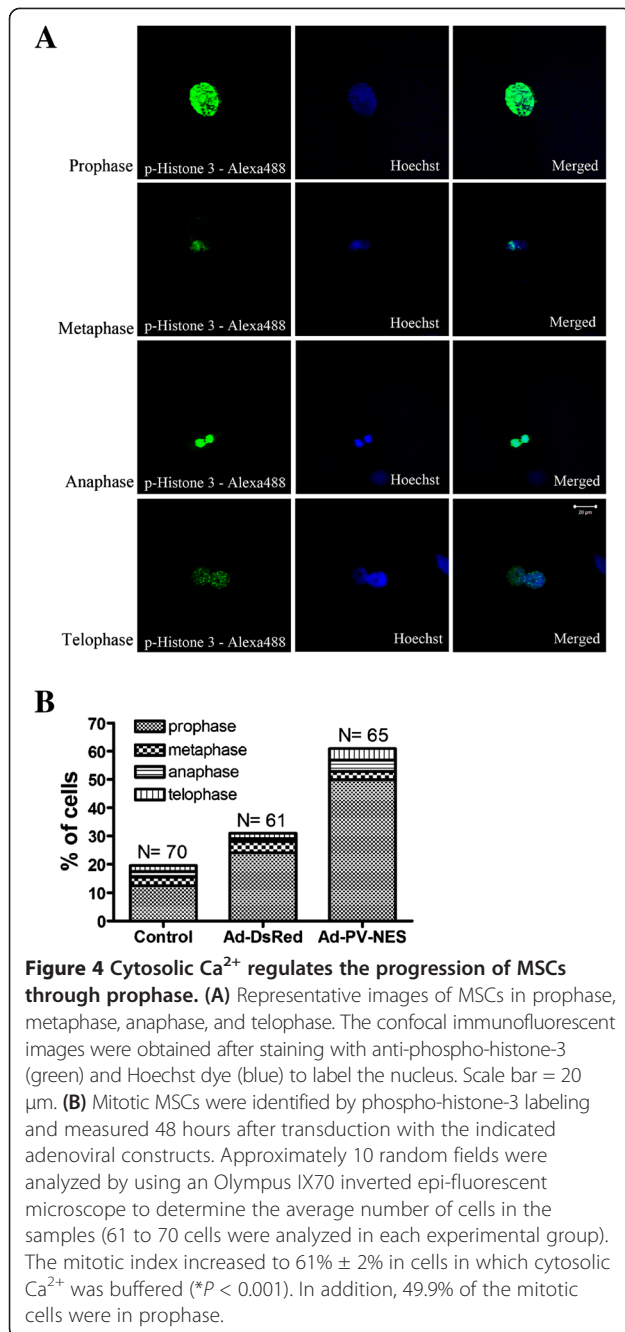
To determine whether the expression of Ad-PV-NES-DsRed specifically affected the G_2/M transition, we examined the expression of cyclin B1, a G_2/M -checkpoint protein. The expression of total cyclin B1 was decreased in cells expressing Ad-PV-NES-DsRed (Figure 5A and B). To confirm that Ad-PV-NES-DsRed expression does not



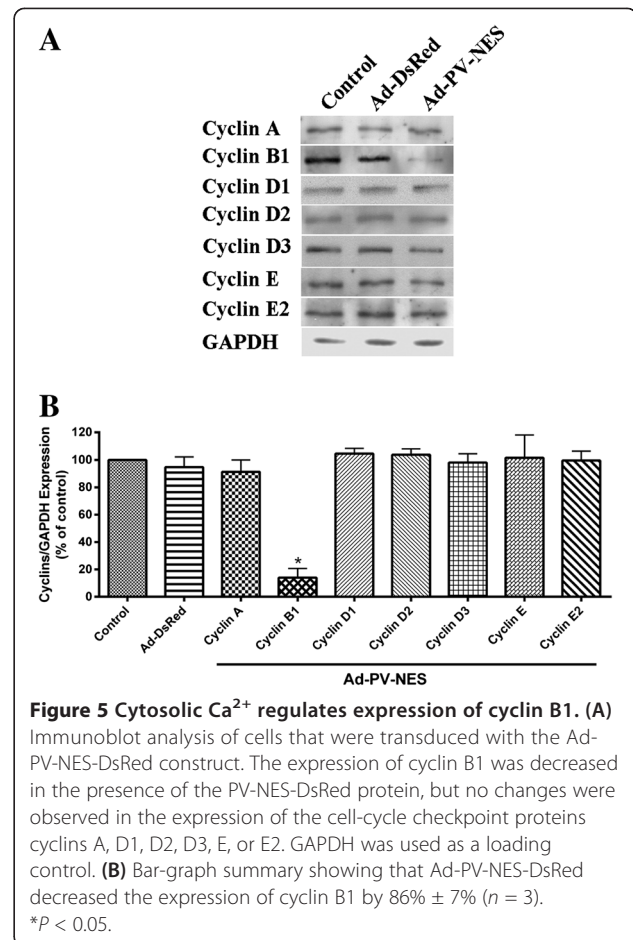
inhibit progression through the earlier phases of the cell cycle, we investigated the expression of G_1/S and S/G_2 checkpoint proteins, cyclins A/D1/D2/D3, and cyclins E/E2, respectively. Buffering cytosolic Ca^{2+} did not alter the expression of any of these checkpoint proteins (Figure 6A and B). To examine further the effects of cytosolic Ca^{2+} on cell proliferation, the effect of PV-NES on the expression level of the growth-related kinase Erk1/2 was examined, and buffering cytoplasmic Ca^{2+} was found to decrease the level of phospho-Erk1/2 (Figure 6A and B). Together, these observations suggest that Ad-PV-NES-DsRed acts during, but not before, mitosis to regulate MSCs proliferation.

Discussion

Recent clinical trials that tested MSCs for the treatment of debilitating disorders, such as osteogenesis imperfecta

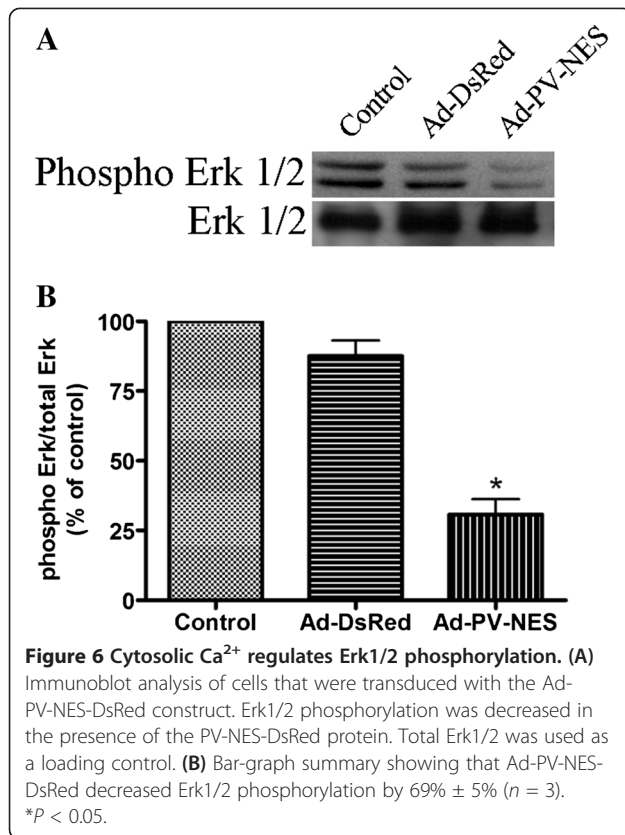


and myocardial infarction, have shown promising results [18]. Numerous preclinical studies have established the therapeutic potential of MSCs in tissue engineering and as cellular protein factories for the delivery of cytokines and anticancer agents [19]. However, although MSC therapy has gained popularity among practitioners and researchers, reports indicated the adverse effects of MSCs, especially in the context of tumor modulation and malignant transformation [6]. Here, we demonstrated that parvalbumin that was targeted to the cytoplasm induced cell-cycle



arrest at prophase in rat MSCs. Genetically modified MSCs have been successfully evaluated in animal models for diabetes, skeletal defects, and myocardial infarction [20]. We speculate that genetically modified MSCs, such as the MSCs expressing Ad-PV-NES-DsRed in our study, could be a valuable tool for understanding and controlling MSC proliferation.

The concept that Ca²⁺ signals regulate rat MSC proliferation at the level of the prophase-to-metaphase transition is interesting and novel. We used a cytoplasmic parvalbumin fusion protein as an alternative for intracellular Ca²⁺ chelators, such as BAPTA-AM. PV constructs have the ability to attenuate Ca²⁺ signals specifically to avoid cell death, which is normally observed after long periods of treatment with BAPTA-AM [21]. Increases in the free Ca²⁺ concentration in the cytosol are associated with progression through the cell cycle [22-24]. Increasing evidence suggests that the spatial and temporal patterns of Ca²⁺ signals may determine their specificity for the transitions between each phase of the cell cycle. The cell-cycle arrest observed at prophase in cells expressing Ad-PV-NES-DsRed could be attributed to the ability of this protein to attenuate Ca²⁺ signals specifically.



Previous studies in cell lines and in the liver showed that targeted PV constructs can efficiently alter Ca^{2+} signal profiles and arrest cell-cycle progression at mitosis, but the effects that we observed in the expression of cyclin B1 were not observed in MSCs or other cell models to date [10,17,25]. The cyclin B1/Cdk1 complex regulates many of the dramatic cellular rearrangements observed during mitosis [26]. Cdk1 activation is a multi-step process that begins when it binds to its regulatory subunit cyclin B, which increases in expression during G_2 and peaks in mitosis [27]. Before mitosis, cyclin B/Cdk1 complexes are held in an inactive state by Cdk1 phosphorylation, which is catalyzed by the protein kinases Wee1 and Myt1. Dephosphorylation of Cdk1 is carried out by the protein phosphatase cell-division cycle 25 homolog C (Cdc25C).

Activation of Cdc25C requires the phosphorylation of several sites in the Cdc25C amino-terminal domain [27]. Although the phosphorylation of cyclin B does not appear to regulate the enzymatic activity of the complex, the subcellular localization of the cyclin B1/Cdk1 complex is controlled through the phosphorylation of four serines (Ser 94, 96, 101, and 113 in *Xenopus*) within a region of cyclin B1 that is known as the cytoplasmic retention sequence (CRS) [26]. The phosphorylation of the first two CRS serines (Ser 94 and 96) is catalyzed by

Erk in the *Xenopus* system [26]. During mitosis, these cyclin B1 phosphorylations trigger a decrease in export and an increase in import, resulting in nuclear translocation. In late prophase, most cyclin B1/Cdk1 complexes are rapidly translocated from the cytoplasm to the nucleus, after which the nuclear envelope breaks down [27].

In this study, we demonstrated that the attenuation of Ca^{2+} signals by Ad-PV-NES-DsRed decreased cyclin B1 expression and Erk phosphorylation, and we hypothesize that these effects caused cell-cycle arrest at prophase. The immediate-early gene transcription and early phosphorylation of Erk and cAMP response element-binding (CREB) have been reported as crucial events for cell-cycle progression and are dependent on cytosolic Ca^{2+} signaling [10,17,28].

Therefore, this tool may represent a new method to control MSC proliferation *in vitro*. Further research is required to identify the targets of cytosolic Ca^{2+} signals for MSC proliferation.

Conclusions

Cytoplasmic Ca^{2+} signals are important for the cell-cycle progression of MSCs beyond prophase because of their effects on Erk phosphorylation and cyclin B1 expression.

Abbreviations

Ad: Adenovirus/adenoviral; DMEM: Dulbecco modified Eagle medium; DsRed: *Discosoma red* fluorescent protein; Erk1/2: Extracellular signal-regulated kinase; FACS: Fluorescence-activated cell sorting; FBS: Fetal bovine serum; MOI: Multiplicity of infection; MSC: Adipose-derived MSC; MSC: Multipotent mesenchymal stromal cell; MTT: 3-(4,5-Dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide; NES: Nuclear exclusion signal; PBS: Phosphate-buffered saline; PI: Propidium iodide; PV: Parvalbumin; PV-NES-DsRed: Parvalbumin fusion protein targeted to the cytosol; SEM: Standard error of the mean; TBS: Tris-buffered saline.

Competing interests

The authors declare that they have no competing interests.

Authors' contributions

CSBM performed collection and assembly of data, data analysis, writing of the manuscript, and final approval of the manuscript; JAQAF performed adenovirus amplification and purification, and final approval of the manuscript; NCRC performed qPCR and final approval of the manuscript; CA performed cell isolation and final approval of the manuscript; JLC performed MSC isolation and characterization and final approval of the manuscript; AMG performed study conception and design, provision of study materials, and final approval of the manuscript; MAR performed study conception and design, data analysis and interpretation, provision of study materials, and final approval of the manuscript; DAG performed study conception and design, data analysis and interpretation, provision of study materials, and manuscript writing. All authors read and approved the final manuscript.

Acknowledgements

This work was supported by NIH grant 1R03TW008709 and by grants from CAPES, FAPEMIG, and CNPq. We thank Dr. Michael H. Nathanson for the parvalbumin constructs (Yale University). Confocal laser-scanning microscopy was performed at the Centro de Microscopia Eletrônica do Instituto de Ciências Biológicas (CEMEL). The authors also thank Pró-Reitoria de Pesquisa da Universidade Federal de Minas Gerais for their financial sponsorship.

Received: 10 December 2012 Revised: 28 May 2012
Accepted: 2 August 2013 Published: 8 August 2013

References

1. Pittenger MF, Mackay AM, Beck SC, Jaiswal RK, Douglas R, Mosca JD, Moorman MA, Simonetti DW, Craig S, Marshak DR: **Multilineage potential of adult human mesenchymal stem cells.** *Science* 1999, **284**:143–147.
2. Hass R, Kasper C, Bohm S, Jacobs R: **Different populations and sources of human mesenchymal stem cells (MSC): a comparison of adult and neonatal tissue-derived MSC.** *Cell Commun Signal* 2011, **9**:12.
3. Pountos I, Giannoudis PV: **Biology of mesenchymal stem cells.** *Injury* 2005, **36**:S8–S12.
4. In't Anker PS, Scherjon SA, Kleijburg-van der Keur C, de Groot-Swings GM, Claas FH, Fibbe WE, Kanhai HH: **Isolation of mesenchymal stem cells of fetal or maternal origin from human placenta.** *Stem Cells* 2004, **22**:1338–1345.
5. Wang S, Qu X, Zhao RC: **Clinical applications of mesenchymal stem cells.** *J Hematol Oncol* 2012, **5**:19.
6. Wong RS: **Mesenchymal stem cells: angels or demons?** *J Biomed Biotechnol* 2011, **201**:459510.
7. Rubio R, Gutierrez-Aranda I, Sáez-Castillo AI, Labarga A, Rosu-Myles M, Gonzalez-Garcia S, Toribio ML, Menendez P, Rodriguez R: **The differentiation stage of p53-Rb-deficient bone marrow mesenchymal stem cells imposes the phenotype of in vivo sarcoma development.** *Oncogene* 2012. doi: 10.1038/onc.2012.507.
8. Rubio D, Garcia S, Paz MF, De la Cueva T, Lopez-Fernandez LA, Lloyd AC, Garcia-Castro J, Bernad A: **Molecular characterization of spontaneous mesenchymal stem cell transformation.** *PLoS One* 2008, **3**:e1398.
9. Machaca K: **Ca(2+) signaling, genes and the cell cycle.** *Cell Calcium* 2010, **48**:243–250.
10. Rodrigues MA, Gomes DA, Leite MF, Grant W, Zhang L, Lam W, Cheng YC, Bennett AM, Nathanson MH: **Nucleoplasmic calcium is required for cell proliferation.** *J Biol Chem* 2007, **282**:17061–17068.
11. Grant W, Roth RJ, Gomes DA, Ehrlich BE, Nathanson MH, Bennett AM: **Suppression of nuclear and cytosolic calcium signaling by targeted expression of calretinin.** *Calcium Binding Proteins* 2006, **1**:30–35.
12. Mosna F, Sensebe L, Krampere M: **Human bone marrow and adipose tissue mesenchymal stem cells: a user's guide.** *Stem Cells Dev* 2010, **19**:1449–1470.
13. Dominici M, Le Blanc K, Mueller I, Slaper-Cortenbach I, Marini F, Krause D, Deans R, Keating A, Prockop DJ, Horwitz E: **Minimal criteria for defining multipotent mesenchymal stromal cells. The International Society for Cellular Therapy position statement.** *Cytotherapy* 2006, **8**:315–317.
14. Mosmann T: **Rapid colorimetric assay for cellular growth and survival: application to proliferation and cytotoxicity assays.** *J Immunol Methods* 1983, **65**:55–63.
15. Riccardi C, Nicoletti I: **Analysis of apoptosis by propidium iodide staining and flow cytometry.** *Nat Protoc* 2006, **1**:1458–1461.
16. Gomes DA, Rodrigues MA, Leite MF, Gomez MV, Varnai P, Balla T, Bennett AM, Nathanson MH: **c-Met must translocate to the nucleus to initiate calcium signals.** *J Biol Chem* 2008, **283**:4344–4351.
17. Lagoudakis L, Garcin I, Julien B, Nahum K, Gomes DA, Combettes L, Nathanson MH, Tordjmann T: **Cytosolic calcium regulates liver regeneration in the rat.** *Hepatology* 2010, **52**:602–611.
18. Salem HK, Thiemermann C: **Mesenchymal stromal cells: current understanding and clinical status.** *Stem Cells* 2010, **28**:585–596.
19. Uccelli A, Moretta L, Pistoia V: **Mesenchymal stem cells in health and disease.** *Nat Rev Immunol* 2008, **8**:726–736.
20. Wagner J, Kean T, Young R, Dennis JE, Caplan AI: **Optimizing mesenchymal stem cell-based therapeutics.** *Curr Opin Biotechnol* 2009, **20**:531–536.
21. Grant S, Freemeran AJ, Gregory PC, Martin HA, Turner AJ, Mikkelsen R, Chelliah J, Yanovich S, Jarvis WD: **Induction of apoptotic DNA fragmentation and c-jun downregulation in human myeloid leukemia cells by the permeant Ca²⁺ chelator BAPTA/AM.** *Oncol Res* 1995, **7**:381–392.
22. Santella L, Ercolano E, Nusco GA: **The cell cycle: a new entry in the field of Ca²⁺ signaling.** *Cell Mol Life Sci* 2005, **62**:2405–2413.
23. Munaron L, Antonioti S, Lovisolio D: **Intracellular calcium signals and control of cell proliferation: how many mechanisms?** *J Cell Mol Med* 2004, **8**:161–168.
24. Berridge MJ: **Calcium signalling and cell proliferation.** *Bioessays* 1995, **17**:491–500.
25. Soliman EM, Rodrigues MA, Gomes DA, Sheung N, Yu J, Amaya MJ, Nathanson MH, Dranoff JA: **Intracellular calcium signals regulate growth of hepatic stellate cells via specific effects on cell cycle progression.** *Cell Calcium* 2009, **45**:284–292.
26. Walsh S, Margolis SS, Kornbluth S: **Phosphorylation of the cyclin B1 cytoplasmic retention sequence by mitogen-activated protein kinase and Plx.** *Mol Cancer Res* 2003, **1**:280–289.
27. Takizawa CG, Morgan DO: **Control of mitosis by changes in the subcellular location of cyclin-B1-Cdk1 and Cdc25C.** *Curr Opin Cell Biol* 2000, **12**:658–665.
28. Hardingham GE, Chawla S, Johnson CM, Bading H: **Distinct functions of nuclear and cytoplasmic calcium in the control of gene expression.** *Nature* 1997, **385**:260–265.

doi:10.1186/scrt291

Cite this article as: Melo et al.: Cytoplasmic-targeted parvalbumin blocks the proliferation of multipotent mesenchymal stromal cells in prophase. *Stem Cell Research & Therapy* 2013 **4**:92.

Submit your next manuscript to BioMed Central and take full advantage of:

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at
www.biomedcentral.com/submit

