RESEARCH Open Access

# Exploring the comorbidity mechanisms between asthma and idiopathic pulmonary fibrosis and the pharmacological mechanisms of Bu-Shen-Yi-Qi decoction therapy via network pharmacology

Yuanyuan Zhong<sup>1</sup>, Lingli Hu<sup>1</sup>, Wenjing Chen<sup>1</sup>, Bin Wang<sup>1</sup>, Jing Sun<sup>1,2\*</sup> and Jingcheng Dong<sup>1,2\*</sup>

## **Abstract**

**Backgrounds:** Asthma and idiopathic pulmonary fibrosis (IPF) are common chronic diseases of the respiratory system in clinical practice. However, the relationship and molecular links remain unclear, and the current treatment's efficacy is disappointing. Bu-Shen-Yi-Qi (BSYQ) decoction has proven effective in treating various chronic airway inflammatory diseases, including asthma and IPF. But the underlying pharmacological mechanisms are still to be elucidated.

**Methods:** This study searched the proteins related to asthma and IPF via TTD, CTD, and DisGeNET databases and then submitted to the STRING to establish the protein–protein interaction (PPI) network. The co-bioinformatics analysis was conducted by Metascape. The active ingredients of BSYQ decoction were screened from TCMSP, ETCM, BATMAN-TCM databases, and HPLC/MS experiment. The corresponding targets were predicted based on TCMSP, ETCM, and BATMAN-TCM databases. The shared targets for asthma and IPF treatment were recognized, and further GO and KEGG analyses were conducted with the DAVID platform. Finally, molecule docking via Autodock Vina was employed to predict the potential binding mode between core potential compounds and targets.

**Results:** Finally, 1333 asthma-related targets and 404 IPF-related proteins were retrieved, 120 were overlapped between them, and many of the asthma-related proteins fall into the same statistically significant GO terms with IPF. Moreover, 116 active ingredients of BSYQ decoction were acquired, and 1535 corresponding targets were retrieved. Eighty-three potential compounds and 56 potential targets were recognized for both asthma and IPF treatment. GO and KEGG analysis indicated that the inflammation response, cytokine production, leukocyte differentiation, oxygen level response, etc., were the common pathological processes in asthma and IPF, which were regulated by BSYQ decoction. Molecule docking further predicted the potential binding modes between the core potential compounds and targets.

Full list of author information is available at the end of the article



<sup>\*</sup>Correspondence: sjing0610@163.com; jcdong2004@126.com

<sup>&</sup>lt;sup>1</sup> Department of Integrative Medicine, Huashan Hospital, Fudan University, Shanghai, China

**Conclusion:** The current study successfully clarified the complex molecule links between asthma and IPF and found the potential common targets. Then we demonstrated the efficacy of BSYQ decoction for asthma and IPF treatment from the angle of network pharmacology, which may provide valuable references for further studies and clinical use.

Keywords: Asthma, IPF, BSYQ decoction, Network pharmacology, Molecule docking

#### Introduction

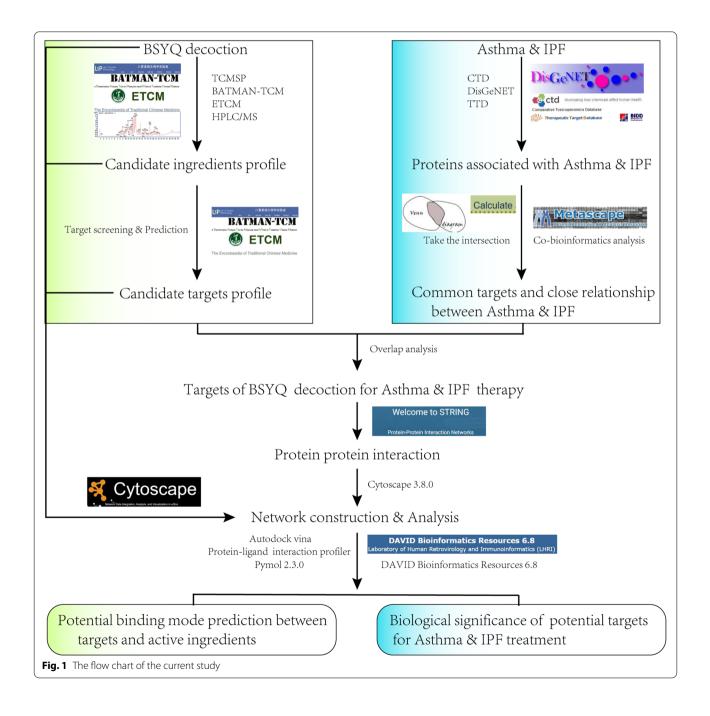
Asthma is one of the most common chronic non-communicable diseases, affecting about 334 million people worldwide and causing approximately 250,000 deaths [1]. It is characterized by airway hyperresponsiveness and reversible airflow restriction, with recurrent wheezing, shortness of breath, chest tightness, and cough. Idiopathic pulmonary fibrosis (IPF) is a chronic, progressive lung disease characterized by varying degrees of inflammation and fibrosis of the lung parenchyma with no definite cause [2]. The prevalence ranges from 10-60 instances per 100,000 persons, with an incidence of 2-30 cases per 100,000 person-years [3-10]. These numbers roughly equate to 130,000 persons in the United States, 300,000 in Europe, 640,000 in East Asia, and 3 million people worldwide. Both asthma and IPF are common chronic diseases of the respiratory system in clinical practice. However, the causal relationship and molecular links between them remain unclear.

Genetic susceptibility and environmental exposures are the common risk factors; persistent chronic inflammation and structural changes involving tissue remodeling and fibrosis are major features of asthma and IPF. Asthma and IPF are complex disorders resulting from risk factors and innumerate multidirectional interactions between the structural cells (bronchial epithelial cells, the alveolar epithelium fibroblasts and myofibroblasts, etc.), inflammatory cells (macrophages, neutrophils, eosinophils, and T/B lymphocytes, etc.) and extracellular matrix (ECM) whereby the relative contribution of each factor differs between individuals, different disease, and different disease subtypes. The current medications for asthma have limitations (such as glucocorticoid insensitivity, poor asthma control, side effects, etc.). Pirfenidone and nintedanib are recommended to manage IPF despite the limited efficacy in preventing disease progression and improving quality of life [11, 12]. Hence it is critical to evaluate both common pathological processes and those that are specific, recognize similarities and differences between asthma and IPF, and seek potential complementary and alternative medical treatments and strategies. Respiratory physicians search for potential novel drugs from the traditional Chinese medicine (TCM) library to treat asthma and IPF.

TCM has progressively gained wider attention worldwide due to its specific theory and long historical clinical practice [13]. Unlike modern medicine, in TCM theory, syndrome differentiation and treatment are the essential diagnosis and treatment principles for disease. TCM syndrome is a specific set of symptoms or a pattern of symptoms presenting the body's internal and external condition at a particular stage [14]. Lung-kidney deficiency is one of the prevalent clinical syndrome types in clinical practice for asthma and IPF patients, and tonify the kidney and replenishing qi is frequently-used treatment principle according to TCM theory [15, 16]. Bu-Shen-Yi-Qi formulae (BSYQ) consists of three herbs, including Epimrdii Herba (Yinyanghuo), Radix Astragali (Huangqi), and Radix Rehmanniae (Shengdihuang), has been demonstrated to be effective in the treatment of chronic airway inflammatory diseases based on our randomized, double-blind placebo-controlled parallel-group multicentre clinical trials [17, 18]. Experiment studies demonstrated that BSYQ decoction could relieve airway inflammation, airway hyperresponsiveness, and airway remodeling in the OVA-induced asthma mice model [19-21]. It can also reduce collagen deposition in lung tissue of bleomycin-induced pulmonary fibrosis mice model and improve pulmonary fibrosis (our unpublished data). However, it is still challenging to clarify the mechanisms of the BSYQ formula in the treatment of asthma and IPF via routine methods because TCM formulae is a complex system with multiple components, multiple targets, and synergistic interactions among its components [22].

Based on polypharmacology and systems biology, network pharmacology integrates various biological data information such as genomics, proteomics, metabolomics, and bioinformatics. It expounds on the occurrence and development of diseases from the perspective of biological network balance, understanding the interaction between the body and drugs, and guiding the rational design of drugs from the perspective of restoring or improving the balance of the biological network, which is considered to be the next-generation drug development paradigm [23–25]. At the same time, the guiding ideology of a holistic view and balance view of TCM and the overall synergistic mechanism of TCM prescription compatibility coincide with the drug research and development model advocated by network pharmacology. Therefore, integrating the emerging network pharmacology and TCM theory will provide new opportunities and methods to discover bioactive components and biomarkers, reveal their action mechanism, and explore the modern scientific connotation of TCM prescriptions based on complex biological systems [26]. Some studies have elucidated the scientific basis and systematic features of herbal medicine to treat diseases via network pharmacology, such as Xuefu Zhuyu decoction [27], Ma-huang decoction [28], Liu-Wei-Di-Huang pill [29], and Qingluoyin [30], etc.

In the present study, we first try to explore the potential molecule links between asthma and IPF and the possible therapeutic mechanisms of BSYQ formulae for asthma and IPF and then try to understand the modern scientific connotation of the TCM theory- the same treatment for different diseases from the angle of network pharmacology (Fig. 1 depicts a flowchart of the entire research procedure).



#### **Materials and methods**

#### Asthma & IPF-related protein screening

The known target proteins for asthma and IPF were screened from the Therapeutic Target Database (TTD, http://bidd.nus.edu.sg/group/cjttd/), which is publicly accessible and provides comprehensive information about the known therapeutic protein, nucleic acid targets described in the literature, and the corresponding drugs/ligands directed at each of these targets, etc. [31]. Then we further searched the Comparative Toxicogenomics Database CTD, http://ctdbase.org/ [32] and DisGeNET database [33] (https://www.disge net.org/) to collect the proteins related to asthma and IPF. The public CTD is an innovative digital ecosystem that connects toxicological information for chemicals, genes, phenotypes, diseases, and exposures. It now provides 45 million toxicogenomic relationships for over 16 300 chemicals, 51 300 genes, 5500 phenotypes, 7200 diseases, and 163 000 exposure events [32]. DisGeNET is a knowledge management platform integrating and standardizing data about disease-associated genes and variants from multiple sources. The latest release covers the full spectrum of human diseases (more than 24 000 diseases and traits, 17 000 genes, and 117 000 genomic variants) [33]. We searched the three databases with the keywords "asthma" or "idiopathic pulmonary fibrosis" and set the species to "Homo sapiens." Finally, we consolidated the information and removed duplicates. The common proteins of asthma and IPF were reserved for further analysis.

#### Bioactive ingredients collection and targets prediction

Potential active compounds of BSYQ decoction were screened from TCMSP (http://sm.nwsuaf.edu.cn/lsp/ tcmsp.php) [34], BATMAN-TCM (http://bionet.ncpsb. org/batman-tcm) [35], ETCM (http://www.nrc.ac.cn: 9090/ETCM/) database [36] and the data from our previous HPLC/MS study [37]. Then the candidate targets of the active compounds were predicted based on the three databases above. TCMSP consists of all the 499 Chinese herbs registered in the Chinese pharmacopeia with 29,384 ingredients, 3,311 targets, and 837 associated diseases, as well as the ADME-related properties such as oral bioavailability (OB), half-life (HL), drug-likeness (DL), and Lipinski's rule of five (MW, AlogP, TPSA, Hdon, Hacc), etc. [34] BATMAN-TCM is the first online bioinformatics analysis tool specially designed for the research of the molecular mechanism of TCM [35]. The ETCM database includes comprehensive and standardized information for the commonly used herbs and formulas of TCM and their ingredients. It can also provide predicted target genes of TCM ingredients, herbs, and formulas, according to the chemical fingerprint similarity between TCM ingredients and known drugs [36].

## Protein–protein interaction (PPI) network construction and analysis

We took the intersection of targets of BSYQ decoction and the common proteins between asthma and IPF, then uploaded them to STRING [38] (https://string-db.org/) to generate the PPI network, the minimum required interaction score was set to high confidence (0.7) and limited to Homo sapiens The STRING database aims to collect, score and integrate all publicly available sources of protein-protein interaction information, complement these with computational predictions and then achieve a comprehensive and objective global network, including direct (physical) as well as indirect (functional) interactions [38]. The final PPI network was established and visualized via Cytoscape 3.8.0 [39]. The network parameters were calculated by NetworkAnalyzer. The MCODE app (based on vertex weighting) in Cytoscape 3.8.0 was used to search the highly connected sub-networks in the PPI network [40].

#### Gene Ontology (GO) and pathway enrichment analysis

To further explore the mechanisms of BSYQ for asthma and IPF treatment, the intersection of targets of BSYQ and the common proteins between asthma and IPF were additionally performed GO enrichment and Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway analysis via the online platform DAVID 6.8 [41] (DAVID, https://david.ncifcrf.gov/) and Metascape [42] (https://metascape.org/).

### Molecule docking

AutoDock vina was used in this study to evaluate the potential molecular binding mode between ingredients and candidate targets. The PyMol 2.3.0 (http://www. pymol.org/) and the online platform PLIP 2021 [43] (https://plip-tool.biotec.tu-dresden.de) were employed to analyze the docked structures. The crystal structures of the target proteins were downloaded from the RCSB Protein Data Bank (www.rcsb.org). Water and hetero molecules were removed, and hydrogen atoms were added by AutoDock tools (1.5.6). The 3D chemical structures of active ingredients were retrieved from the PubChem compound database (NCBI, USA) and subjected to minimize the energy via molecular mechanics-2 (MM2) force field in Chem 3D Pro. The protein-ligand docking active site center was defined by the location of the original ligand, and the dimensions of the grids were set at  $30 \times 30 \times 30$  Å in the x, y, and Z directions, with a spacing of 0.375 Å between the grid points. The docked conformation corresponding to the lowest binding energy was selected as the most probable binding conformation.

#### **Results**

#### Asthma & IPF related proteins collecting and analyzing

One thousand three hundred thirty-three asthmarelated targets and 404 IPF-related targets were retrieved from the TTD, CTD, and DisGeNET database (Duplicates were removed and detailed in additional table S1). Asthma and IPF disease-specific PPI networks were established (Fig. 2A, B). The top 15 core proteins based on

two network topology parameters (degree and betweenness centrality) in asthma and IPF were displayed in additional table S2 and S3. Then we found that VEGFA, TP53, EGFR, AKT1, EGF, IL6, STAT3, and MYC occupied the core positions in asthma and IPF-specific PPI networks, indicating the essential roles of these proteins in the pathological process of asthma and IPF. To further explore the molecule links between asthma and IPF,

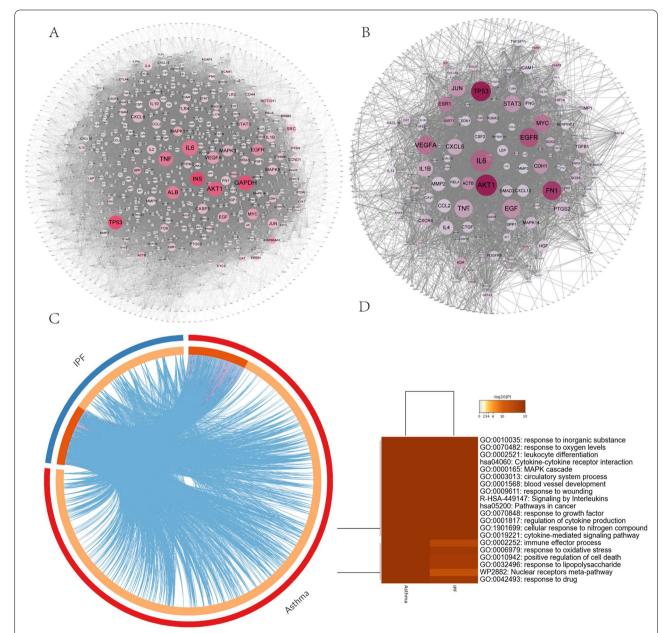


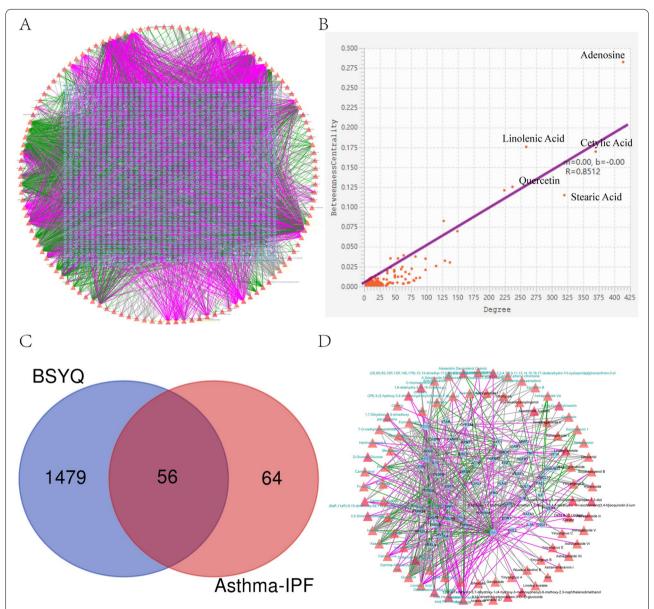
Fig. 2 Asthma and IPF specific PPI network and co-bioinformatics analysis for the two protein profiles via Metascape. The size and color of the node are proportional to the value of the degree and betweenness centrality (A and B). C Circos plot for the two groups of proteins. Purple lines connect proteins that appear in both protein profiles. Blue lines connect proteins that belong to the same ontology term. D Top 20 common GO terms or pathways enriched by the two protein profiles

a co-bioinformatics analysis was conducted by Metascape. One hundred twenty proteins were overlapped in the two groups of protein lists (Fig. 2C). Much of the asthma-related proteins fall into the same statistically significant GO terms (such as response to oxygen levels, leukocyte differentiation, MAPK cascades, signaling by interleukins, response to growth factors and regulation of cytokine production, etc.) with IPF-specific proteins (Fig. 2D), indicating the strong function association

between the two comparison cohorts. The 120 common proteins were used for further analysis.

## Active ingredients screening and corresponding targets prediction of BSYQ decoction

After removing duplicates, 175 active ingredients were acquired and further submitted to TCMSP, BATMAN-TCM, and ETCM databases to get the corresponding targets. Finally, except for 59 components predicted no



**Fig. 3** The analysis of the compounds and targets of BSYQ decoction and the potential targets screening for asthma and IPF treatment. **A** The compound-target (C-T) network, **B** the relationship between degree and betweenness centrality of the nodes in the C-T network, and the core compounds and targets of BSYQ decoction based on the two topology parameters were marked. **C** the Venn diagram between targets of BSYQ and common proteins of asthma and IPF. **D** The potential compound-potential target (PC-PT) network. The triangles and circles represent the compounds and targets, respectively

targets, 116 active compounds, and 1535 related targets were retrieved (additional table S4 and S5). The compound-target (C-T) network was constructed and analyzed via Cytoscape 3.8.0 (Fig. 3A, B). The C-T network consists of 1651 nodes (116 active compounds and 1535 potential targets) and 5255 edges. Two centrality indicators, degree and betweenness centrality, were calculated to identify the critical nodes within the network (Fig. 3B). Interestingly, both two types of centrality indicators uniformly confirmed the core 15 candidate compounds (including adenosine, cetylic acid, octadecanoic, linolenic acid and quercetin, etc.) and targets (including PTGS2,

NCOA2, AR, ESR1, and PTGS1, etc.) of BSYQ decoction (additional table S6 and S7).

# Potential ingredients and targets of BSYQ decoction for asthma and IPF therapy

To further explore the molecule mechanisms of BSYQ decoction for asthma and IPF therapy, we took the intersection of the targets profile of BSYQ decoction with the 120 common proteins between asthma and IPF. Finally, 56 potential targets were retrieved and were regarded as the potential targets for asthma and IPF treatment (Fig. 3C). Then a potential compound-potential target (PC-PT)

Table 1 Top 15 potential compounds in the PC-PT network according to degree and betweenness centrality

Ingredient Name	Degree	Betweenness Centrality	PubChem CID	Source	Ingredient Name	Degree	Betweenness Centrality	PubChem CID	Source
Quercetin	36	0.309833105	5,280,343	BATMAN- TCM, TCMSP	Quercetin	36	0.309833105	5,280,343	BATMAN- TCM, TCMSP
luteolin	20	0.074877417	5,280,445	TCMSP	Sucrose	6	0.104421054	5988	BATMAN- TCM, ETCM
Linolenic Acid	16	0.076791716	5,280,934	ETCM	Adenosine,Adenine Nucleoside	15	0.079102825	60,961	ETCM
Adenosine,Adenine Nucleo- side	15	0.079102825	60,961	ETCM	Linolenic Acid	16	0.076791716	5,280,934	ETCM
Kaempferol	14	0.048065771	5,280,863	BATMAN- TCM, TCMSP	luteolin	20	0.074877417	5,280,445	TCMSP
lsorhamnetin	11	0.010756097	5,281,654	BATMAN- TCM, TCMSP	Kaempferol	14	0.048065771	5,280,863	BATMAN- TCM, TCMSP
Rhamnocitrin	10	0.00855788	5,320,946	BATMAN- TCM	Canavanine	6	0.033126673	439,202	BATMAN- TCM
Pratensein	10	0.00855788	5,281,803	HPLC/MS	FA	6	0.029886082	6037	TCMSP
Formononetin	10	0.01317824	5,280,378	BATMAN- TCM, HPLC/ MS, TCMSP	Fructose	4	0.019612123	5984	ETCM
Beta-Sitosterol	9	0.019445351	222,284	BATMAN- TCM	Beta-Sitosterol	9	0.019445351	222,284	BATMAN- TCM
Cetylic Acid,Hexadecanoic Acid,Palmitic Acid	8	0.017682546	985	ETCM	Cetylic Acid,Hexadecanoic Acid,Palmitic Acid	8	0.017682546	985	ETCM
Kumatakenin	7	0.005434062	5,318,869	BATMAN- TCM, ETCM, TCMSP	Hentriacontanol-6	2	0.014492754	5,318,007	BATMAN- TCM
Canavanine	6	0.033126673	439,202	BATMAN- TCM	Medicarpin	6	0.013850015	336,327	ETCM
Sucrose	6	0.104421054	5988	BATMAN- TCM, ETCM	(6aR,11aR)-9,10-dimeth- oxy-6a,11a-dihydro- 6H-benzofurano[3,2-c] chromen-3-ol	6	0.013850015	14,077,830	TCMSP
Octadecanoic?Acid,Stearic Acid	6	0.010597854	5281	ETCM	Formononetin	10	0.01317824	5,280,378	BATMAN- TCM, HPLC/ MS, TCMSP

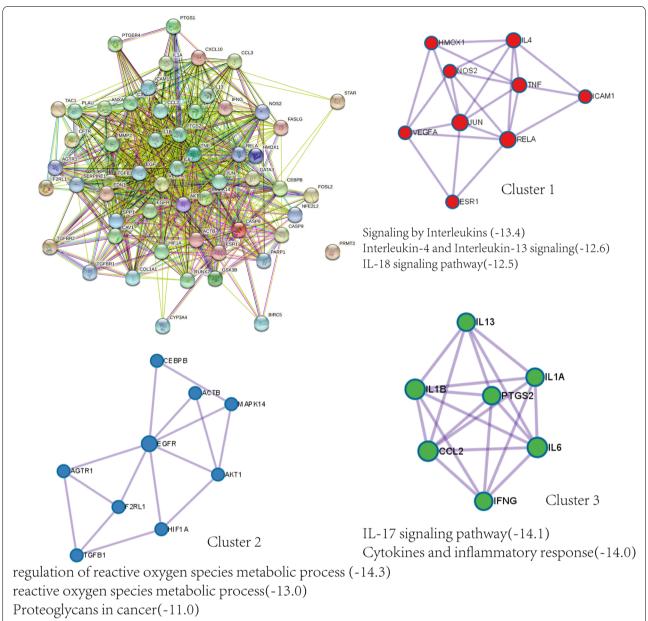
network was established and analyzed (Fig. 3D). The PC-PT network consists of 139 nodes (83 potential compounds and 56 potential targets) and 371 edges. The core potential ingredients and targets based on the two network parameters are shown in Tables 1 and 2. Quercetin, luteolin, linolenic acid, adenosine, kaempferol, etc., were considered the potential core compounds, and PTGS2, ESR1, PTGS1, NOS2, AKT1, etc. were the main potential targets of BSYQ for asthma and IPF therapy. We further constructed the PPI network with the 56 potential targets by STRING and searched the similar function clusters of the PPI network by MCODE analysis based on topology (Fig. 4). The top 15 core proteins based on the two topological parameters in the 56 potential targets PPI network are shown in Table 3. IL6, IL-1β, TNF, VEGFA, AKT1, etc., played an essential role in the PPI network, indicating the crucial roles in treating asthma and IPF. Similar function subnetworks were constructed, and function analysis showed that cluster 1 mainly participated in the interleukins signaling (Fig. 4B). Cluster 2 specifically regulates the reactive oxygen species (Fig. 4C). Cluster 3 mainly regulates the cytokines and inflammatory response (Fig. 4D). Then we performed the GO and KEGG analysis with the 56 potential targets (Fig. 5). KEGG pathway analysis showed that TNF signaling pathway, HIF-1 signaling pathway, cytokine-cytokine receptor interaction, toll-like receptor signaling pathway, MAPK signaling pathway, etc. were enriched and regulated by BSYQ decoction (Fig. 5A, 5B), indicating the underline comprehensive mechanisms of BSYQ decoction for asthma and IPF treatment. We found that the 56 potential targets mainly participate in the regulation of the inflammatory response, nitric oxide biosynthetic process, smooth muscle cell proliferation process, etc. (Fig. 5C).

# Molecule docking for the core potential ingredients and targets of BSYQ for asthma and IPF treatment

In the current study, the possible interaction modes between core ingredients and targets were predicted by Autodock vina. Molecule docking is a computational method that efficiently predicts the noncovalent binding of macromolecules or, more frequently, of a macromolecule (receptor) and a small molecule (ligand). It is generally believed that the lower the binding energy between ligand and receptor, the greater the possibility of interaction. Three core ingredients, including quercetin, luteolin, and kaempferol with four corresponding essential targets including AKT1, IL-6, PTGS2, and TNF, were docked and displayed to elucidate the exact binding mode (Fig. 6, A: kaempferol-AKT1; B: luteolin-AKT1; C: quercetin-AKT1; D: kaempferol-IL6; E: luteolin-IL6; F: quercetin-IL6; G: kaempferol-PTGS2; H: luteolin-PTGS2; I: quercetin-PTGS2; J: kaempferol-TNF; K: luteolin-TNF; L: quercetin-TNF). Specifically, taking the kaempferol with AKT1, for example, kaempferol was located within the interfaced pocket formed by active amino acid residues of AKT1, five conventional hydrogen bonds were formed between kaempferol and AKT1 by interacting with the vital amino acids, including SER205, THR211, and VAL271. Additionally, π-stacking between kaempferol and TRP80, hydrophobic interactions with TRP80, LEU210, and VAL270 were found in the active site, which helped stabilize the compound at the binding site (Fig. 6A). Six key hydrogen bonds with SER205A, THR211A, and VAL271A, hydrophobic interactions with TRP80A, LEU210A, LEU240A, and VAL270A, and π-Stacking interaction with TRP80A, were established between luteolin and AKT1 (Fig. 6B). Similarly, quercetin and AKT1 were shown to create

Table 2 Top 15 potential targets in the PC-PT network according to degree and betweenness centrality

Protein Name	Degree	Betweenness Centrality	Protein Name	Degree	Betweenness Centrality
PTGS2	54	0.277312173	PTGS2	54	0.277312173
ESR1	46	0.153120648	PTGS1	42	0.204507204
PTGS1	42	0.204507204	ESR1	46	0.153120648
NOS2	19	0.032615148	CXCR4	8	0.086469588
AKT1	15	0.01512124	TNF	12	0.060803175
GSK3B	13	0.009859331	CYP3A4	12	0.058290209
CYP3A4	12	0.058290209	HMOX1	6	0.035902065
TNF	12	0.060803175	HIF1A	4	0.035245261
ACTB	10	0.013813159	ICAM1	5	0.034843635
MAPK14	10	0.001209612	NOS2	19	0.032615148
CEBPB	9	0.007230542	IL1B	8	0.031406599
ANXA1	8	0.014493758	TGFBR2	4	0.029023858
CXCR4	8	0.086469588	IFNG	4	0.016903104
IL1B	8	0.031406599	AKT1	15	0.01512124
IL6	6	0.009704189	SERPINE1	5	0.014896122



**Fig. 4** The protein–protein Interaction (PPI) network of the 56 potential targets for asthma and IPF therapy based on STRING. Similar function subnetworks were analyzed by Metascape

five critical hydrogen bonds with SER205A, THR211A, and VAL271A, hydrophobic contacts with TRP80A, LEU210A, VAL270A, and ASP292A, and  $\pi$ -Stacking interaction with TRP80A (Fig. 6C). Between kaempferol and IL-6, seven important hydrogen bonds were discovered with ARG104A, GLU106A, SER108A, GLN156A, and ASP160A, as well as hydrophobic interactions with LYS46A and PHE105A, and  $\pi$ -Cation interactions with LYS46A were found (Fig. 6D). Five critical hydrogen bonds with THR43A, LYS46A, ARG 104A, GLU106A,

and THR163A, hydrophobic interactions with LYS46A, ARG104A, and PHE105A, and  $\pi$ -Cation interactions with LYS46A and ARG 104A were formed between luteolin and IL-6 (Fig. 6E). Quercetin and IL-6 formed seven critical hydrogen bonds with GLU42A, ARG104A, GLU106A, SER107A, SER108A, and GLN156A, as well as hydrophobic interactions with LYS46A and PHE105A, and  $\pi$ -Cation interactions with LYS46A (Fig. 6F). Between kaempferol and PTGS2, six critical hydrogen bonds with ARG44A, ILE124A, ASP125A, SER126A, and GLN372A, as well as

Table 3 Top 15 potential targets in the 56 potential targets PPI network

Protein Name	Degree	Betweenness Centrality	Protein Name	Degree	Betweenness Centrality	
IL6	51	0.059767121	IL6	51	0.059767121	
IL1B	46	0.034930701	EGFR	43	0.041706178	
TNF	46	0.030700549	VEGFA	46	0.038701872	
VEGFA	46	0.038701872	IL1B	46	0.034930701	
AKT1	45	0.034171523	AKT1	45	0.034171523	
PTGS2	44	0.027625229	TNF	46	0.030700549	
EGFR	43	0.041706178	ESR1	33	0.030204633	
EGF	42	0.022481519	PTGS2	44	0.027625229	
JUN	40	0.025683985	JUN	40	0.025683985	
CCL2	39	0.015488887	EGF	42	0.022481519	
MMP2	37	0.009780132	MAPK14	34	0.021812819	
IL4	36	0.013415867	CCL2	39	0.015488887	
TGFB1	35	0.009526729	IL4	36	0.013415867	
MAPK14	34	0.021812819	CEBPB	20	0.011576047	
ICAM1	33	0.005648341	CXCL10	27	0.011252843	

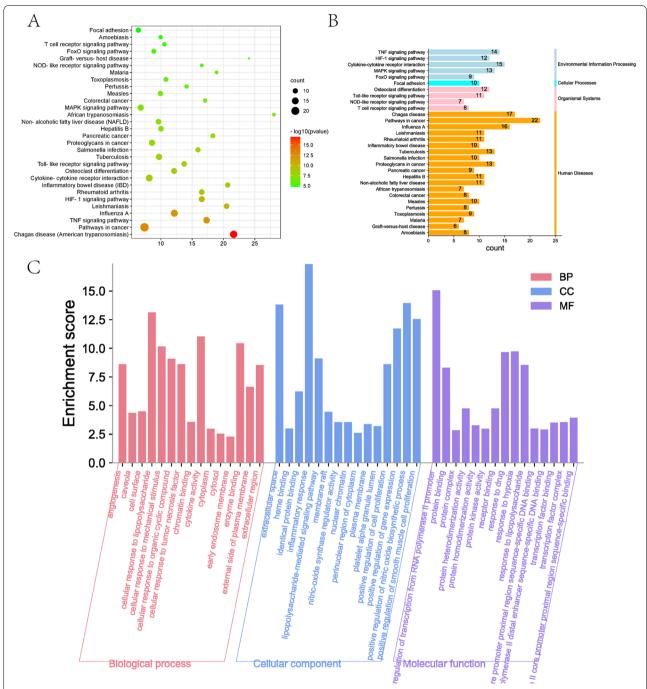
hydrophobic interactions with PRO542B and GLN543B, were discovered (Fig. 6G). Three key hydrogen bonds with SER126A and LYS546B, hydrophobic interactions with ARG44A, PRO542B, and GLN543B, were established between luteolin and PTGS2 (Fig. 6H). Quercetin and PTGS2 were shown to have three critical hydrogen bonds with ARG44A, SER126A, and LYS546B. Hydrophobic interactions with ARG44A, PRO542B, and GLN543B, and  $\pi$ -Cation interaction with ARG44A were predicted (Fig. 6I). Between kaempferol and TNF, four key hydrogen bonds with SER60B, GLN61A, TYR151A, and TYR151B, hydrophobic interactions with TYR59B and TYR119A, and π-Stacking interaction with TYR119A and TYR119B, were recognized (Fig. 6J). Five key hydrogen bonds with SER60B, LEU120B, GLY121A, TYR151A, and TYR151B, hydrophobic interactions with TYR59B and TYR119A, and  $\pi$ -Stacking interaction with TYR119A and TYR119B, were formed between luteolin and TNF (Fig. 6K). Quercetin and TNF established five critical hydrogen bonds with GLY121A, TYR151A, and TYR151B, hydrophobic interactions with TYR119A, and  $\pi$ -Stacking interactions with TYR119A (Fig. 6L). Taken together, hydrogen-bonding,  $\pi$ -stacking,  $\pi$ -cation, and hydrophobic interactions played key roles in the protein-ligand recognition and stability, which may be helpful for the activation or inhibition of the target proteins and is necessary for the pharmacology activities.

#### Discussion

Both asthma and IPF are inflammatory lung diseases characterized by airway injury, inflammation, bronchial and parenchymal remodeling [44]. The pathogenesis of asthma has not been fully defined, involving

immunology, neuroendocrinology, genetic factors, and environmental factors. Airway hyperresponsiveness and airflow restriction are the main pathological features, and chronic inflammation is the main trigger. IPF is now generally considered the result of the interactions of multiple genetic and environmental risk factors. The aging alveolar epithelial repetitive local micro-injuries trigger abnormal epithelial fibroblast communication, induce myofibroblasts and extracellular matrix accumulation, and pulmonary interstitial remodeling [45]. Chronic airway inflammation, epithelial-mesenchymal transformation (EMT), and oxidative stress also participate in the occurrence and development of IPF [46, 47]. Thus, the repeated airway epithelial injury, chronic airway inflammation, EMT, airway remolding, and their interactions play essential roles in the pathological process in asthma and IPF, indicating the similarity between the two diseases. Unfortunately, the anti-inflammatory therapy for asthma can only control symptoms, and the improvement of disease progression is limited. What's more, it did not improve the outcome in the treatment of IPF, and an immunosuppressive therapeutic strategy incorporating prednisolone and azathioprine was shown to increase mortality [48, 49]. The nintedanib and pirfenidone cannot completely prevent the progressive decline of pulmonary function [50]. So, seeking new alternative therapies for IPF and asthma treatment is highly urgent and of farreaching significance.

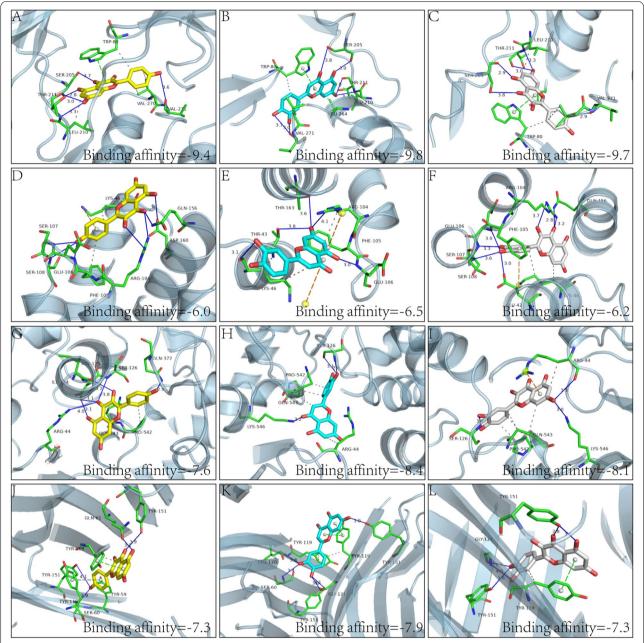
TCM is a comprehensive medicinal system that has been used in clinical practice for thousands of years and plays a vital role in the health maintenance of people all over the world [51, 52]. The validated curative effects of TCM make it a feasible alternative therapeutic agent for



**Fig. 5** GO and KEGG analysis for the potential targets of BSYQ decoction for asthma and IPF therapy by DAVID 6.8. **A** and **B** the top 30 enriched KEGG pathways of the 56 potential targets. **C** the top 15 enriched GO items, including biology process (BP), cellular compartment (CC), and molecular function (MF)

disease treatment [27]. Then, BSYQ decoction, proven effective, is regarded as the ideal joint therapy for asthma and IPF. The current study first explored the complex molecule links between asthma and IPF. We constructed asthma and IPF-specific PPI networks and compared

the two protein profiles. The co-bioinformatic analysis showed that inflammation response, cytokine production, leukocyte differentiation, oxygen level response, etc., commonly participate in the progress of asthma and IPF and found 120 proteins overlapped. Additionally,



**Fig. 6** Predicted binding mode within the active site of the drug-target complexes obtained from Autodock vina. **A** kaempferol-AKT1; **B** luteolin-AKT1; **C** quercetin-AKT1; **D** kaempferol-IL6; **E** luteolin-IL6; **F** quercetin-IL6; **G** kaempferol-PTGS2; **H** luteolin-PTGS2; **J** kaempferol-TNF; **K** luteolin-TNF; **L**-quercetin-TNF). The proteins were presented as cartoon modes, and molecules were presented as ball and stick models. Active site amino acid residues are represented as lines. Dotted blue lines in these pictures represent hydrogen bonds with distance units of ° A, dotted khaki lines represent π-Cation interactions, dotted red lines represent π-Stacking (parallel) interactions and dotted grey lines represent hydrophobic interaction. Other O and N atoms are colored red and blue, respectively

eight proteins, including VEGFA, TP53, EGFR, AKT1, EGF, IL6, STAT3, and MYC, played essential roles in asthma and IPF. Then we searched the active compounds and predicted the corresponding targets based on the TCMSP, BATMAN-TCM, and ETCM databases.

Finally, 175 active compounds (with 59 no predicted targets) and 1535 predicted targets were acquired. Then 83 potential targets anchored 56 common proteins between asthma and IPF, and the core potential compounds and targets were recognized. The additional GO and KEGG

analysis indicated that inflammatory response, nitric oxide biosynthetic process, smooth muscle cell proliferation, etc., were mainly regulated by BSYQ decoction both in asthma and IPF. We also constructed the PPI network based on the STRING database, searched the similar function clusters (the interleukins signaling, oxygen species metabolism, and the cytokines and inflammatory response), and further verified the potential binding mode between the potential compounds and targets via the molecule docking method. Unlike modern medicine anchored single targets, BSYQ decoction consists of 83 potential compounds and targets 56 common targets of asthma and IPF, regulated several pathways and biological processes, and showed a synthetic therapeutic effect.

IL-6, TNF, and AKT, which occupied an important position in asthma and IPF, are essential targets regulated by BSYQ decoction. IL-6 binds to sIL-6R and activates the membrane-bound glycoprotein 130 (gp130), then actives Jak/signal transducer and activator of transcription (STAT) signaling pathway [53], which is implicated in a variety of inflammatory processes, including IPF [54, 55] and asthma [56]. Increased levels of tumor necrosis factor (TNF) α have been linked to several pulmonary inflammatory diseases, including asthma, chronic obstructive pulmonary disease (COPD), acute lung injury (ALI), acute respiratory distress syndrome (ARDS), sarcoidosis, and IPF. TNF-α plays multiple roles in disease pathology by inducing an accumulation of inflammatory cells, stimulating the generation of inflammatory mediators, and causing oxidative and nitrosative stress, airway hyperresponsiveness, and tissue remodeling [44]. AKT regulates many processes, including metabolism, proliferation, cell survival, growth, and angiogenesis, and targeting the PI3K/AKT signal pathway effectively treats asthma and IPF [57, 58]. In the present study, we found that IL-6 was anchored by 6 potential ingredients of BSYQ decoction (astragaloside Viii, magnograndiolide, soyasaponin 1, luteolin, quercetin, and aucuboside), TNF was targeted by 12 potential ingredients (adenosine, linolenic acid, sucrose, alexandrin, astragaloside Viii, magnograndiolide, soyasaponin 1, kaempferol, luteolin, quercetin, aucuboside, and cetylic acid) and AKT was hit by 11 ingredients (adenosine, DFV, formononetin, isorhamnetin, kaempferol, kumatakenin, luteolin, patensein, quercetin, quercitrin, and rhamnocitrin). Multi compounds anchored IL-6, TNF, and AKT and then produced synergistic effects. Representative flavonoids, including quercetin, kaempferol, and luteolin, were regarded as the core compounds for asthma and IPF treatment of BSYQ prescription based on our study. The anti-inflammatory and immunomodulating properties of quercetin are effectively utilized in the treatment of late-phase, and late-late-phase bronchial asthma responses, which is more competent in inhibiting IL-8 than cromolyn [59]. It can regulate the Th1/Th2 stability and decrease the antigen-specific IgE antibody released by B cells [60]. At the same time, it can reverse bleomycin-induced pulmonary fibrosis and attenuate lethality, weight loss, and the expression of pulmonary senescence markers by promoting FasL receptor and caveolin-1 expression and inhibiting AKT activation [61]. Kaempferol is a flavonoid found in many edible plants. Its anti-oxidant/anti-inflammatory effects have been demonstrated in disease models such as diabetes and asthma [62]. It can alleviate airway inflammation by modulating the Tyk2-STAT1/3 signaling response in the endotoxin-exposed airway epithelium in asthmatic mice [63]. But the efficacy of IPF treatment has not been evaluated. We found that it may be a potential agent for IPF therapy for the first time. Luteolin can modulate OVA-induced airway bronchoconstriction and bronchial hyperreactivity [64], inhibit autophagy by activating PI3K/Akt/mTOR signaling and inhibit the Beclin-1-PI3KC3 complex [65]. It can reduce the weight index and hydroxyproline content, delay the process of pulmonary fibrosis and inhibit TGF-β1 mRNA expression in the bleomycin-induced pulmonary fibrosis model [66, 67]. Particularly, IL-13 (anchored by Linolenic Acid) is a key Th2 cytokine that induces airway inflammation and remodeling. It is recognized as a central mediator of allergic asthma [68, 69]. The anti-interleukin-4 receptor α monoclonal antibody (Dupilumab) that blocks both interleukin-4 and interleukin-13 signaling significantly decreased the rates of severe asthma exacerbation, as well as improved lung function and better asthma control [70]. It's also a stimulator of fibroblast proliferation and extracellular matrix synthesis in the process of IPF [71]. IL-13 and its receptors are elevated in IPF bronchoalveolar lavage fluid [72], while neutralization of IL-13 attenuated bleomycin-induced pulmonary fibrosis [73]. In summary, multiple active ingredients in BSYQ decoction can act on various targets to treat diseases and then play a synthetic therapeutic effect.

Despite the profound significance of this study, several limitations should be noted. Firstly, the network construction and analysis separated from biological entities cannot fully reflect the internal network regulation mechanisms and dynamic changes of disease. Secondly, there is a dose–effect relationship between drugs and diseases, and the current network pharmacology method is challenging to achieve the purpose of quantification.

#### Conclusion

The relationship between asthma and IPF is complicated, and clinical and experimental studies have proved the efficacy of BSYQ decoction for treating asthma and IPF. The current study successfully clarified the complex

molecule links between asthma and IPF and found the potential common targets. Then we demonstrated the efficacy of BSYQ decoction for asthma and IPF treatment from the angle of network pharmacology, which may provide valuable references for further studies and clinical use.

#### Abbreviations

IPF: Idiopathic pulmonary fibrosis; BSYQ decoction: Bu-Shen-Yi-Qi decoction; TTD: Therapeutic Target Database; CTD: Comparative Toxicogenomics Database; TCMSP: Traditional Chinese Medicine Systems Pharmacology Database and Analysis Platform; BATMAN-TCM: A Bioinformatics Analysis Tools for Molecular mechanism of Traditional Chinese Medicine; ETCM: The Encyclopedia of Traditional Chinese Medicine; TCM: Traditional Chinese Medicine; PPI: Protein–protein interaction; DAVID: The Database for Annotation, Visualization, and Integrated Discovery; AKT1: RAC-alpha serine/threonine-protein kinase; IL-6: Interleukin-6; PTGS2: Prostaglandin G/H synthase 2; TNF: Tumor necrosis factor; VEGFA: Vascular endothelial growth factor; TP53: Cellular tumor antigen p53; EGFR: Epidermal growth factor receptor; EGF: Pro-epidermal growth factor; STAT3: Signal transducer and activator of transcription 3; MYC: Myc proto-oncogene protein.

#### **Supplementary Information**

The online version contains supplementary material available at https://doi.org/10.1186/s12906-022-03637-7.

**Additional file 1: Supplementary Table S1.** Proteins related to asthma and IPF based on TTD, CTD and DisGeNET database.

**Additional file 2: Table S2.** Top 15 core proteins in the asthma-specific PPI network. **Table S3.** Top 15 core proteins in the IPF specific PPI network.

**Additional file 3: Supplementary Table S4.** Candidate compounds of BSYQ decoction.

**Additional file 4: Supplementary Table S5.** Candidate compunds and predicted targets based on TCMSP, ETCM and BATMAN-TCM.

**Additional file 5: Table S6.** Top 15 active compounds in the C-T network according to degree and betweenness centrality. **Table S7.** Top 15 candidate targets in the C-T network according to degree and betweenness centrality.

#### Acknowledgements

Not applicable.

#### Authors' contributions

Jingcheng Dong participated in the conception and design of the study. Yuanyuan Zhong, Lingli Hu, Wenjing Chen, and Bin Wang acquired and analyzed the data. Jing Sun drafted and revised the manuscript. The corresponding author and all the authors have read and approved the final submitted manuscript.

#### **Funding**

This work was supported by the National Natural Science Foundation of China (Grant Number: 81774074, 81973631), Shanghai Science and technology commission (Grant Number: 18401901800).

#### Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

#### **Declarations**

#### Ethics approval and consent to participate

Not applicable.

#### Consent for publication

Not applicable.

#### **Competing interests**

The authors declare that they have no conflicts of interest.

#### Author detail:

<sup>1</sup>Department of Integrative Medicine, Huashan Hospital, Fudan University, Shanghai, China. <sup>2</sup>Institutes of Integrative Medicine, Fudan University, Shanghai 200040, China.

Received: 12 September 2021 Accepted: 30 May 2022 Published online: 07 June 2022

#### References

- Vos T, Flaxman AD, Naghavi M, Lozano R, Michaud C, Ezzati M, Shibuya K, Salomon JA, Abdalla S, Aboyans V, et al. Years lived with disability (YLDs) for 1160 sequelae of 289 diseases and injuries 1990–2010: a systematic analysis for the Global Burden of Disease Study 2010. Lancet (London, England). 2012;380(9859):2163–96.
- Raghu G, Remy-Jardin M, Myers JL, Richeldi L, Ryerson CJ, Lederer DJ, Behr J, Cottin V, Danoff SK, Morell F, et al. Diagnosis of idiopathic pulmonary fibrosis. An official ATS/ERS/JRS/ALAT clinical practice guideline. Am J Respir Crit Care Med. 2018;198(5):e44–68.
- Baddini-Martinez J, Pereira CA. How many patients with idiopathic pulmonary fibrosis are there in Brazil? J Bras Pneumol. 2015;41(6):560–1.
- Esposito DB, Lanes S, Donneyong M, Holick CN, Lasky JA, Lederer D, Nathan SD, O'Quinn S, Parker J, Tran TN. Idiopathic pulmonary fibrosis in United States automated claims. Incidence, prevalence, and algorithm validation. Am J Respir Crit Care Med. 2015;192(10):1200–7.
- Harari S, Madotto F, Caminati A, Conti S, Cesana G. Epidemiology of idiopathic pulmonary fibrosis in northern italy. PLoS ONE. 2016;11(2):e0147072.
- Hopkins RB, Burke N, Fell C, Dion G, Kolb M. Epidemiology and survival of idiopathic pulmonary fibrosis from national data in Canada. Eur Respir J. 2016;48(1):187–95.
- Lee HE, Myong JP, Kim HR, Rhee CK, Yoon HK, Koo JW. Incidence and prevalence of idiopathic interstitial pneumonia and idiopathic pulmonary fibrosis in Korea. Int J Tuber Lung Dis. 2016;20(7):978–84.
- Natsuizaka M, Chiba H, Kuronuma K, Otsuka M, Kudo K, Mori M, Bando M, Sugiyama Y, Takahashi H. Epidemiologic survey of Japanese patients with idiopathic pulmonary fibrosis and investigation of ethnic differences. Am J Respir Crit Care Med. 2014;190(7):773–9.
- Raghu G, Chen SY, Hou Q, Yeh WS, Collard HR. Incidence and prevalence of idiopathic pulmonary fibrosis in US adults 18–64 years old. Eur Respir J. 2016;48(1):179–86.
- Raimundo K, Chang E, Broder MS, Alexander K, Zazzali J, Swigris JJ. Clinical and economic burden of idiopathic pulmonary fibrosis: a retrospective cohort study. BMC Pulm Med. 2016;16:2.
- Bando M, Yamauchi H, Ogura T, Taniguchi H, Watanabe K, Azuma A, Homma S, Sugiyama Y. Clinical Experience of the Long-term Use of Pirfenidone for Idiopathic Pulmonary Fibrosis. Inter Med (Tokyo, Japan). 2016;55(5):443–8.
- Galli JA, Pandya A, Vega-Olivo M, Dass C, Zhao H, Criner GJ. Pirfenidone and nintedanib for pulmonary fibrosis in clinical practice: Tolerability and adverse drug reactions. Respirology (Carlton, Vic). 2017;22(6):1171–8.
- Suo T, Wang H, Li Z. Application of proteomics in research on traditional Chinese medicine. Expert Rev Proteomics. 2016;13(9):873–81.
- 14. Zheng. A systems biology approach to diagnosis and treatments. Science. 2014;346(6216):S13–5.
- Dong J, Zhang H, Wei Y, Duan X, Liu B, Sun J, Lv Y, Chen M, Luo Q, Du Y, et al. Modern understanding and practice of lung and kidney qi deficiency syndrome based on airway chronic inflammatory diseases. Chinese J Integ Trad Western Med. 2015;35(05):521–8.
- Xu F, Cui W, Kong Q, Tang Z, Dong J. A Real-World Evidence Study for Distribution of Traditional Chinese Medicine Syndrome and Its Elements on Respiratory Disease. Evid-Based Complement altern Med: eCAM. 2018;2018:8305892.

- Kong L, Zhang H, Cao Y, Le J, Wu J, Liu B, Chen M, Du Y, Wang J, Wang G, et al. The Anti-Inflammatory Effects of Invigorating Kidney and Supplementing Qi Chinese Herbal Formulae in Asthma Patients. Evid-Based Complement altern Med: eCAM. 2017;2017:3754145.
- Wang G, Liu B, Cao Y, Du Y, Zhang H, Luo Q, Li B, Wu J, Lv Y, Sun J, et al. Effects
  of two Chinese herbal formulae for the treatment of moderate to severe
  stable chronic obstructive pulmonary disease: a multicenter, double-blind,
  randomized controlled trial. PLoS ONE. 2014;9(8):e103168.
- Cui J, Xu F, Tang Z, Wang W, Hu LL, Yan C, Luo Q, Gao H, Wei Y, Dong J. Bu-Shen-Yi-Qi formula ameliorates airway remodeling in murine chronic asthma by modulating airway inflammation and oxidative stress in the lung. Biomed Pharmacother. 2019;112:108694.
- Wang J, Wu J, Kong L, Nurahmat M, Chen M, Luo Q, Li B, Wu X, Dong J. BuShenYiQi Formula strengthens Th1 response and suppresses Th2-Th17 responses in RSV-induced asthma exacerbated mice. J Ethnopharmacol. 2014;154(1):131–47.
- Wei Y, Luo QL, Sun J, Chen MX, Liu F, Dong JC. Bu-Shen-Yi-Qi formulae suppress chronic airway inflammation and regulate Th17/Treg imbalance in the murine ovalbumin asthma model. J Ethnopharmacol. 2015;164:368–77.
- Zhang Q, Yu H, Qi J, Tang D, Chen X, Wan JB, Li P, Hu H, Wang YT, Hu Y. Natural formulas and the nature of formulas: Exploring potential therapeutic targets based on traditional Chinese herbal formulas. PLoS ONE. 2017;12(2):e0171628.
- 23. Hopkins AL. Network pharmacology: the next paradigm in drug discovery. Nat Chem Biol. 2008;4(11):682–90.
- 24. Zhou W. Research progress and development prospect of network pharmacology. Chinese J Pharmacol Toxicol. 2015;29(05):760–2.
- Zhou W, Chen X, Zhang Y. Network pharmacology- a new philosophy for understanding of drug action and discovery of new drugs. Chinese J Pharmacol Toxicol. 2012;26(01):4–9.
- Li S, Zhang B. Traditional Chinese medicine network pharmacology: theory, methodology and application. Chin J Nat Med. 2013;11(2):110–20.
- Zhong Y, Luo J, Tang T, Li P, Liu T, Cui H, Wang Y, Huang Z. Exploring Pharmacological Mechanisms of Xuefu Zhuyu Decoction in the Treatment of Traumatic Brain Injury via a Network Pharmacology Approach. Evid-Based complement Altern Med: eCAM. 2018;2018:8916938.
- Yao Y, Zhang X, Wang Z, Zheng C, Li P, Huang C, Tao W, Xiao W, Wang Y, Huang L, et al. Deciphering the combination principles of Traditional Chinese Medicine from a systems pharmacology perspective based on Ma-huang Decoction. J Ethnopharmacol. 2013;150(2):619–38.
- 29. Liang X, Li H, Li S. A novel network pharmacology approach to analyse traditional herbal formulae: the Liu-Wei-Di-Huang pill as a case study. Mol BioSyst. 2014;10(5):1014–22.
- Zhang B, Wang X, Li S. An Integrative Platform of TCM Network Pharmacology and Its Application on a Herbal Formula, Qing-Luo-Yin. Evid-Based complement Altern Med: eCAM. 2013;2013:456747.
- Wang Y, Zhang S, Li F, Zhou Y, Zhang Y, Wang Z, Zhang R, Zhu J, Ren Y, Tan Y, et al. Therapeutic target database 2020: enriched resource for facilitating research and early development of targeted therapeutics. Nucleic Acids Res. 2020;48(D1):D1031-d1041.
- Davis AP, Grondin CJ, Johnson RJ, Sciaky D, Wiegers J, Wiegers TC, Mattingly CJ. Comparative Toxicogenomics Database (CTD): update 2021. Nucleic Acids Res. 2021;49(D1):D1138-d1143.
- Piñero J, Ramírez-Anguita JM, Saüch-Pitarch J, Ronzano F, Centeno E, Sanz F, Furlong LI. The DisGeNET knowledge platform for disease genomics: 2019 update. Nucleic Acids Res. 2020;48(D1):D845-d855.
- Ru J, Li P, Wang J, Zhou W, Li B, Huang C, Li P, Guo Z, Tao W, Yang Y, et al. TCMSP: a database of systems pharmacology for drug discovery from herbal medicines. J Cheminformatics. 2014;6:13.
- Liu Z, Guo F, Wang Y, Li C, Zhang X, Li H, Diao L, Gu J, Wang W, Li D, et al. BATMAN-TCM: a Bioinformatics Analysis Tool for Molecular mechANism of Traditional Chinese Medicine. Sci Rep. 2016;6:21146.
- Xu HY, Zhang YQ, Liu ZM, Chen T, Lv CY, Tang SH, Zhang XB, Zhang W, Li ZY, Zhou RR, et al. ETCM: an encyclopaedia of traditional Chinese medicine. Nucleic Acids Res. 2019;47(D1):D976-d982.
- Luo QL, Nurahmat M, Li MH, Sun J, Chen MX, Liu F, Wei Y, Dong JC. Pharmacological investigation of a HPLC/MS standardized three herbal extracts containing formulae (Bu-Shen-Yi-Qi-Tang) on airway inflammation and hypothalamic-pituitary-adrenal axis activity in asthmatic mice. Phytomedicine. 2014;21(11):1439–50.

- Szklarczyk D, Gable AL, Lyon D, Junge A, Wyder S, Huerta-Cepas J, Simonovic M, Doncheva NT, Morris JH, Bork P, et al. STRING v11: protein-protein association networks with increased coverage, supporting functional discovery in genome-wide experimental datasets. Nucleic Acids Res. 2019;47(D1):D607-d613.
- Shannon P, Markiel A, Ozier O, Baliga NS, Wang JT, Ramage D, Amin N, Schwikowski B, Ideker T. Cytoscape: a software environment for integrated models of biomolecular interaction networks. Genome Res. 2003;13(11):2498–504.
- Bader GD, Hogue CW. An automated method for finding molecular complexes in large protein interaction networks. BMC Bioinformatics. 2003;4:2.
- da Huang W, Sherman BT, Lempicki RA. Systematic and integrative analysis
  of large gene lists using DAVID bioinformatics resources. Nat Protoc.
  2009;4(1):44–57.
- Zhou Y, Zhou B, Pache L, Chang M, Khodabakhshi AH, Tanaseichuk O, Benner C, Chanda SK. Metascape provides a biologist-oriented resource for the analysis of systems-level datasets. Nat Commun. 2019;10(1):1523.
- Adasme MF, Linnemann KL, Bolz SN, Kaiser F, Salentin S, Haupt VJ, Schroeder M. PLIP 2021: expanding the scope of the protein-ligand interaction profiler to DNA and RNA. Nucleic Acids Res. 2021;49(W1):W530-w534.
- Malaviya R, Laskin JD, Laskin DL. Anti-TNFα therapy in inflammatory lung diseases. Pharmacol Ther. 2017;180:90–8.
- Martinez FJ, Collard HR, Pardo A, Raghu G, Richeldi L, Selman M, Swigris JJ, Taniguchi H, Wells AU. Idiopathic pulmonary fibrosis. Nat Rev Dis Primers. 2017:3:17074.
- Kim DS, Collard HR, King TE Jr. Classification and natural history of the idiopathic interstitial pneumonias. Proc Am Thorac Soc. 2006;3(4):285–92.
- Lee JS, Song JW, Wolters PJ, Elicker BM, King TE Jr, Kim DS, Collard HR. Bronchoalveolar lavage pepsin in acute exacerbation of idiopathic pulmonary fibrosis. Eur Respir J. 2012;39(2):352–8.
- 48. Selman M, King TE, Pardo A. Idiopathic pulmonary fibrosis: prevailing and evolving hypotheses about its pathogenesis and implications for therapy. Ann Intern Med. 2001;134(2):136–51.
- Behr J. Prednisone, azathioprine, and N-acetylcysteine for pulmonary fibrosis. N Eng J Med. 2012;367(9):869 author reply 870-861.
- Saito S, Alkhatib A, Kolls JK, Kondoh Y, Lasky JA. Pharmacotherapy and adjunctive treatment for idiopathic pulmonary fibrosis (IPF). J Thorac Dis. 2019;11(Suppl 14):S1740-s1754.
- 51. Kong DX, Li XJ, Zhang HY. Where is the hope for drug discovery? Let history tell the future. Drug Discovery Today. 2009;14(3–4):115–9.
- 52. Cheung F. TCM: Made in China. Nature. 2011;480(7378):S82-83.
- Robinson MB, Deshpande DA, Chou J, Cui W, Smith S, Langefeld C, Hastie AT, Bleecker ER, Hawkins GA. IL-6 trans-signaling increases expression of airways disease genes in airway smooth muscle. Am J Physiol Lung Cell Mol Physiol. 2015;309(2):L129-138.
- O'Donoghue RJ, Knight DA, Richards CD, Prêle CM, Lau HL, Jarnicki AG, Jones J, Bozinovski S, Vlahos R, Thiem S, et al. Genetic partitioning of interleukin-6 signalling in mice dissociates Stat3 from Smad3-mediated lung fibrosis. EMBO Mol Med. 2012;4(9):939–51.
- Le TT, Karmouty-Quintana H, Melicoff E, Le TT, Weng T, Chen NY, Pedroza M, Zhou Y, Davies J, Philip K, et al. Blockade of IL-6 Trans signaling attenuates pulmonary fibrosis. J Immunol (Baltimore, Md: 1950). 2014;193(7):3755–68.
- Esnault S, Khosravi M, Kelly EA, Liu LY, Bochkov YA, Tattersall MC, Jarjour NN. Increased IL-6 and Potential IL-6 trans-signalling in the airways after an allergen challenge. Clin Exp Allergy. 2021;51(4):564–73.
- Wu D, Li S, Liu X, Xu J, Jiang A, Zhang Y, Liu Z, Wang J, Zhou E, Wei Z, et al. Alpinetin prevents inflammatory responses in OVA-induced allergic asthma through modulating PI3K/AKT/NF-kB and HO-1 signaling pathways in mice. Int Immunopharmacol. 2020;89(Pt A):107073.
- Qian W, Cai X, Qian Q, Zhang W, Wang D. Astragaloside IV modulates TGFβ1-dependent epithelial-mesenchymal transition in bleomycin-induced pulmonary fibrosis. J Cell Mol Med. 2018;22(9):4354–65.
- Mlcek J, Jurikova T, Skrovankova S, Sochor J: Quercetin and Its Anti-Allergic Immune Response. Mol (Basel, Switzerland). 2016;21(5):623.
- Jafarinia M, Sadat Hosseini M, Kasiri N, Fazel N, Fathi F, Ganjalikhani Hakemi M, Eskandari N. Quercetin with the potential effect on allergic diseases. J Can Soc Allergy Clin Immunol. 2020;16:36.
- Hohmann MS, Habiel DM, Coelho AL, Verri WA Jr, Hogaboam CM. Quercetin Enhances Ligand-induced Apoptosis in Senescent Idiopathic Pulmonary Fibrosis Fibroblasts and Reduces Lung Fibrosis In Vivo. Am J Respir Cell Mol Biol. 2019;60(1):28–40.

- Rajendran P, Rengarajan T, Nandakumar N, Palaniswami R, Nishigaki Y, Nishigaki I. Kaempferol, a potential cytostatic and cure for inflammatory disorders. Eur J Med Chem. 2014;86:103–12.
- Gong JH, Shin D, Han SY, Park SH, Kang MK, Kim JL, Kang YH. Blockade of Airway Inflammation by Kaempferol via Disturbing Tyk-STAT Signaling in Airway Epithelial Cells and in Asthmatic Mice. Evidence-Based Complement Altern Med: eCAM. 2013;2013;250725.
- Das M, Ram A, Ghosh B. Luteolin alleviates bronchoconstriction and airway hyperreactivity in ovalbumin sensitized mice. Inflamm Res. 2003;52(3):101–6.
- Wang S, Wuniqiemu T, Tang W, Teng F, Bian Q, Yi L, Qin J, Zhu X, Wei Y, Dong J. Luteolin inhibits autophagy in allergic asthma by activating PI3K/Akt/ mTOR signaling and inhibiting Beclin-1-PI3KC3 complex. Int Immunopharmacol. 2021;94: 107460.
- Gong G, Qian Z. Therapeutic effects of luteolin on rats pulmonary fibrosis. Chinesse J New Drugs. 2004;12:48–51.
- 67. Gong G, Qian Z, Zhou S. Effects of luteolin on expression of TGF-β1 mRNA in rats with pulmonary fibrosis. Chinese Pharmacol Bull. 2005;12:1466–9.
- Grünig G, Warnock M, Wakil AE, Venkayya R, Brombacher F, Rennick DM, Sheppard D, Mohrs M, Donaldson DD, Locksley RM, et al. Requirement for IL-13 independently of IL-4 in experimental asthma. Science (New York, NY). 1998;282(5397):2261–3.
- Wills-Karp M, Luyimbazi J, Xu X, Schofield B, Neben TY, Karp CL, Donaldson DD. Interleukin-13: central mediator of allergic asthma. Science (New York, NY). 1998;282(5397):2258–61.
- Castro M, Corren J, Pavord ID, Maspero J, Wenzel S, Rabe KF, Busse WW, Ford L, Sher L, FitzGerald JM, et al. Dupilumab Efficacy and Safety in Moderate-to-Severe Uncontrolled Asthma. N Engl J Med. 2018;378(26):2486–96.
- 71. Zhu Z, Homer RJ, Wang Z, Chen Q, Geba GP, Wang J, Zhang Y, Elias JA. Pulmonary expression of interleukin-13 causes inflammation, mucus hypersecretion, subepithelial fibrosis, physiologic abnormalities, and eotaxin production. J Clin Investig. 1999;103(6):779–88.
- Park SW, Ahn MH, Jang HK, Jang AS, Kim DJ, Koh ES, Park JS, Uh ST, Kim YH, Park JS, et al. Interleukin-13 and its receptors in idiopathic interstitial pneumonia: clinical implications for lung function. J Korean Med Sci. 2009;24(4):614–20.
- Belperio JA, Dy M, Burdick MD, Xue YY, Li K, Elias JA, Keane MP. Interaction of IL-13 and C10 in the pathogenesis of bleomycin-induced pulmonary fibrosis. Am J Respir Cell Mol Biol. 2002;27(4):419–27.

## **Publisher's Note**

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.