# Peer

## Segmental analysis by speckle-tracking echocardiography of the left ventricle response to isoproterenol in male and female mice

#### Elisabeth Walsh-Wilkinson, Marie Arsenault and Jacques Couet

Universite Laval, Groupe de recherche sur les valvulopathies, Centre de recherche de l'Institut universitaire de cardiologie et de pneumologie de Quebec, Quebec, Canada

## ABSTRACT

We studied by conventional and speckle-tracking echocardiography, the response of the left ventricle (LV) to a three-week continuous infusion of isoproterenol (Iso), a non-specific beta-adrenergic receptor agonist in male and female C57Bl6/J mice. Before and after Iso (30 mg/kg/day), we characterized LV morphology and function as well as global and segmental strain. We observed that Iso reduced LV ejection in both male (-8.7%) and female (-14.7%) mice. Several diastolic function parameters were negatively regulated in males and females such as E/A, E/E', isovolumetric relaxation time. Global longitudinal (GLS) and circumferential (GCS) strains were reduced by Iso in both sexes, GLS by 31% and GCS by about 20%. For the segmental LV analysis, we measured strain, strain rate, reverse strain rate, peak speckle displacement and peak speckle velocity in the parasternal long axis. We observed that radial strain of the LV posterior segments were more severely modulated by Iso than those of the anterior wall in males. In females, on the other hand, both posterior and anterior wall segments were negatively impacted by Iso. Longitudinal strain showed similar results to the radial strain for both sexes. Strain rate, on the other hand, was only moderately changed by Iso. Reverse strain rate measurements (an index of diastolic function) showed that posterior LV segments were negatively regulated by Iso. We then studied the animals 5 and 17 weeks after Iso treatment. Compared to control mice, LV dilation was still present in males. Ejection fraction was decreased in mice of both sex compared to control animals. Diastolic function parameters, on the other hand, were back to normal. Taken together, our study indicates that segmental strain analysis can identify LV regions that are more negatively affected by a cardiotoxic agent such as Iso. In addition, cessation of Iso was not accompanied with a complete restoration of cardiac function after four months.

**Subjects** Biochemistry, Cardiology, Radiology and Medical Imaging **Keywords** Speckle-tracking echocardiography, Isoproterenol, Takotsubo syndrome, Sex differences, Adrenergic suractivation, Strain, Mouse, Heart, Cardiac toxicity

## **INTRODUCTION**

Adrenergic overstimulation using isoproterenol (Iso) is often used to induce cardiac toxicity in small rodent models (*Gomes et al., 2013; Chang et al., 2018; Kudej et al., 1997*). Sometimes described as an HF model (systolic and/or diastolic), a cardiac hypertrophy model or a SIC model (Takotsubo-like syndrome) (*Puhl et al., 2016; Wallner et al., 2016*;

Submitted 13 November 2020 Accepted 18 February 2021 Published 12 March 2021

Corresponding author Jacques Couet, jacques.couet@med.ulaval.ca

Academic editor Gwyn Gould

Additional Information and Declarations can be found on page 17

DOI 10.7717/peerj.11085

© Copyright 2021 Walsh-Wilkinson et al.

Distributed under Creative Commons CC-BY 4.0

OPEN ACCESS

Sachdeva, Dai & Kloner, 2014; Shao et al., 2013a; Shao et al., 2013b; Angelini & Gamero, 2019), Iso treatment of rats or mice has been the object of an important amount of literature.

Many regimens of Iso administration to rodents have been used in past studies such as a single high dose bolus, several injections over days as well as continuous infusion using osmotic micro-pumps implanted subcutaneously or intra-peritoneally, or micro-pellets at different dosages and for various duration (*Chang et al., 2018; Shao et al., 2013b; An et al., 2016; Ali et al., 2019; Ma et al., 2011*). This diversity of experimental set-ups makes the comparison between studies sometimes difficult, as the goals pursued by the authors were often different. In addition, as for many preclinical studies, most experiments were conducted in male animals and less often in females. In spontaneously hypertensive rats, it was observed that Iso treatment was more prone in induce LV dilation in males than in females (*Michel et al., 2017*). In mice, more studies were performed using females but again, few studies studied in parallel both sexes (*Zhu et al., 2016*).

Sex-related differences in the Iso model have been studied using echocardiography in C57Bl6 mice infused for 7 or 14 days (10 mg/kg/day) but relatively few differences in either systolic or diastolic cardiac function were found. Most of the differences were identified at tissue level (*Zhu et al., 2016*). More recently, another study found no sex differences in mice treated with Iso for 14 days (*Grant et al., 2020*).

In the present study, we wished to document the left ventricular (LV) response to a beta-adrenergic receptor-mediated insult using the Iso mouse model and to identify possibly morphological and functional sex differences. We used a relatively high dosage of Iso (30 mg/kg/day) for a longer period (21 days). Using conventional and speckle tracking echocardiography (STE), we evaluated if LV dysfunction was present and then performed a regional study of this dysfunction. We also studied how male and female mice echo parameters evolved 5 and 17 weeks post-Iso.

Our results indicate that Iso induced both systolic and diastolic function impairments in mice and that only small sex differences were present in the extent of these. By STE, we showed that the response to Iso is not homogeneously distributed throughout the LV walls as the posterior LV wall seems more sensitive to Iso effects, at least in male mice. As for LV reverse remodeling taking place after Iso, dilation in males was still present four months later.

## **MATERIALS AND METHODS**

### Mouse model

The present study was conducted within the Mouse Animal model of Sex Differences and Aging in heart Failure (MASDAF) study, which follows longitudinally C57Bl6/J mice up to two years in order to investigate biological sex and aging effects on the LV response to an insult. In this sub-study, we compared the LV response of both males and females (age 8 weeks) from Jackson Laboratory (Bar Harbor, ME, USA). After a week of acclimatization, micro-osmotic pumps (cat. no.: 1004; Alzet, Cupertino, CA, USA), releasing isoproterenol (Iso: 30 mg/kg/day; Sigma-Aldrich, Mississauga, Ont, Canada; n = 10 mice/group) or

vehicle (saline; Ctrl. n = 8 mice/group) were implanted subcutaneously in the back of the neck and left for 21 days (*Roussel et al., 2018*). Experienced technicians for health and behavior monitored the animals daily during the protocol. The animals were weighed weekly. No mouse displayed markers associated with death or poor prognosis of quality of life, or specific signs of severe suffering or distress, which would have led to early and immediate euthanasia. Among those, significant loss or gain of weight, grooming and changes in behavior were monitored. The protocol was approved by the Université Laval's animal protection committee and followed the recommendations of the Canadian Council on Laboratory Animal Care (protocol #2019-360, VRR-19-075).

### Echocardiography

An echocardiography (Echo) exam was performed the day before Iso infusion started and three weeks later. Echo images were acquired on a Vevo 3100 imaging system (VisualSonics, FujiFilm, Toronto, Canada) by the same investigator and analyzed off-line using Vevo LAB software. The investigator was blinded for animal identification. Animals were anesthetized and placed in a supine position on a heated platform (39 °C). The concentration of isoflurane was maintained around 1.5–2.5%, so that the heart rate was kept over 400 beats/minute.

2D echo. M-mode images were recorded to measure diastolic LV walls thickness and chamber diameters from the parasternal long axis (PSLAX) view and the short axis (SAX) view at the papillary muscle level. LV mass was calculated by the VevoLab echo analysis software (VisualSonics) using the following equation:  $1.053 \times [(EDD + PW + IVSW)^3 - EDD^3] \times 0.8$ . EDD: end-diastolic LV diameter, PW: posterior diastolic wall thickness and IVSW: interventricular septal diastolic wall thickness. Fractional shortening from M-mode images was calculated using the following equation: (EDD - ESD)/EDD where: ESD end-systolic LV diameter. Mitral flow was measured by pulsed wave Doppler from an apical four-chamber view. Early diastolic peak filling velocity (E wave), peak filling velocity at atrial contraction (A wave), E wave deceleration time and the E/A ratio were obtained. The early-diastolic peak velocity (E'), the late-diastolic peak velocity (A') of the mitral valve annulus and E'/A' as well as E/ E' were obtained using tissue Doppler. LV volumes, ejection fraction (EF), stroke volume and cardiac output were calculated using the Simpson's rule method from LV chamber area tracings.

*Speckle-tracking echo.* 2D echo B-mode loops were acquired from the LV PSLAX and analyzed using Vevo Strain software (VisualSonics). Images were acquired at the highest frame rate possible (232 frames/s) and strain analysis was performed in the radial and longitudinal axes as previously described (*Walsh-Wilkinson et al., 2020*). Also calculated by the Vevo Strain software were the reverse strain rate (a diastolic function index), speckle displacement (mm) and velocity (mm/s). Figure 1A illustrates the various segments studied as well as the direction of strain or speckle displacement.



**Figure 1** Speckle tracking strain analysis. Radial and longitudinal strains can be obtained using the parasternal long axis (PSLAX) view. The six segments are also identified. Characters colors correspond to those used in the graphs for each of these LV segments. Ant, anterior, Post, posterior. Before after effects of Iso on the strain. Radial (A–B) and longitudinal (C–D) peak strains were obtained using the parasternal long axis view. The six segments are identified. Characters colors correspond to those used in the graphs for each of these LV segments. Ant, anterior, Post, posterior after effects of Iso on the strain. Radial (A–B) and longitudinal (C–D) peak strains were obtained using the parasternal long axis view. The six segments are identified. Characters colors correspond to those used in the graphs for each of these LV segments.(continued on next page...)

Full-size DOI: 10.7717/peerj.11085/fig-1

#### Figure 1 (... continued)

Radial (E–F and G–H) and longitudinal (I–J and K–L) peak strains were grouped either as from the base, the midsection or the apex or as from the anterior or the posterior segments in males (left panels) and females (right panels). Ant, anterior, Post, posterior. Males are represented on the left panels and females on the right. Results are represented as violin plots (n = 8-10). Inner black lines represent quartiles of the data. Significance between groups was calculated with paired Student's *T*-test. \*, p < 0.05, \*\*, p < 0.01, \*\*\*, p < 0.001 and \*\*\*\*, p < 0.001 between corresponding pre-Iso and Iso animals.

<b>There is a set a </b>
--

Parameters	M Ctrl, N = 8	M Iso, N = 10	F Ctrl, N = 8	F Iso, N = 10	Sex	Iso	Sex x Iso
BW, g	$25.0\pm0.31$	$27.9\pm0.53^{\rm b}$	$19.9\pm0.28$	$20.8\pm0.45$	< 0.0001	ns	ns
Tibia, mm	$21.0\pm0.08$	$21.7\pm0.10^{\rm d}$	$20.5\pm0.08$	$20.3\pm0.13$	< 0.0001	ns	ns
M-mode							
EDD, mm	$3.8\pm0.03$	$3.9\pm0.07$	$3.6\pm0.06$	$3.6\pm0.07$	< 0.0001	ns	ns
ESD, mm	$2.6\pm0.08$	$2.9 \pm 0.11^{a}$	$2.3\pm0.05$	$2.6\pm0.07^{b}$	< 0.01	< 0.001	ns
IVS, mm	$0.82\pm0.02$	$0.82\pm0.02$	$0.76\pm0.01$	$0.77\pm0.02$	< 0.001	ns	ns
PW, mm	$0.84\pm0.01$	$0.83\pm0.02$	$0.78\pm0.01$	$0.75\pm0.02$	< 0.0001	ns	ns
RWT	$0.44\pm0.01$	$0.42\pm0.01$	$0.43\pm0.01$	$0.42\pm0.01$	ns	ns	ns
FS, %	$32.2\pm1.69$	$24.8\pm1.52^{b}$	$35.4\pm0.98$	$27.5\pm0.86^{\rm c}$	< 0.05	< 0.0001	ns
LV, mg	$86 \pm 2.1$	$95 \pm 4.1$	$76 \pm 2.6$	$75 \pm 2.7$	< 0.0001	ns	ns
iLV, mg/g	$3.4\pm0.10$	$3.4 \pm 0.11$	$3.8\pm0.14$	$3.6 \pm 0.11$	< 0.01	ns	ns
Simpson's							
EDV, µl	$54.6 \pm 1.83$	$66.9\pm3.62^{\text{b}}$	$44.6\pm0.74$	$51.0\pm2.45$	< 0.0001	< 0.001	ns
ESV, μl	$23.4\pm0.72$	$34.8\pm2.78^{\text{b}}$	$17.1\pm0.34$	$27.4\pm2.13^{\rm b}$	< 0.01	< 0.0001	ns
SV, mm	$31.2\pm1.39$	$32.1\pm1.91$	$27.5\pm0.56$	$23.6\pm1.19$	< 0.001	ns	ns
EF, %	$57.1 \pm 1.01$	$48.4\pm2.19^{\rm a}$	$61.7\pm0.55$	$47.0 \pm 2.80^{\circ}$	ns	< 0.0001	ns
HR, bpm	$494 \pm 17.9$	$415 \pm 10.0^{b}$	$441 \pm 16.8$	$448\pm9.0$	ns	< 0.05	< 0.01
CO, ml/min	$15.3\pm0.69$	$13.3\pm0.73$	$12.2\pm0.56$	$10.6\pm0.60$	< 0.0001	< 0.01	ns

Notes.

Ctrl, control; Iso, isoproterenol treatment; BW, body weight; LV, left ventricle mass; EDD, end diastolic LV diameter; ESD, end-systolic diameter; IVS, interventricular septum; PW, posterior wall; RWT, relative wall thickness; FS, fractional shortening; EDV, end-diastolic volume; ESV, end-systolic volume; SV, stroke volume; EF, ejection fraction; HR, heart rate; bpm, beats per minute and CO: cardiac output.

Values are expressed as the mean  $\pm$  SEM. Two-way ANOVA statistical analysis results are displayed for each factor, sex and Iso, respectively. Intergroup *p* values were calculated using Holm-Sidak post-test.

 $^{a}p < 0.05.$ 

 ${}^{b}p < 0.01.$  ${}^{c}p < 0.001.$ 

 $^{d}p < 0.0001$  between Ctrl and Iso groups, respectively.

ns, not significant.

#### **Statistical analysis**

All data are expressed as mean  $\pm$  standard error of the mean (SEM). Statistical analyses were performed on the log<sub>10</sub> of the data. Normality was assessed using the Shapiro–Wilk test. Intergroup comparisons were conducted using Student's *T*-test using GraphPad Prism 8.4, (GraphPad Software Inc., La Jolla, CA, USA). Data from Tables 1–3 were analyzed using 2-way ANOVA and Holm-Sidak pos *t*-test. *P* < 0.05 was considered statistically significant.

## RESULTS

# Isoproterenol induces systolic and diastolic impairments in male and female mice

Before implantation of the micro-osmotic pump, a complete echocardiography exam was performed for each animal. Baseline values are listed in Table S1. With the exception of those related to the relative size of the animals depending on their biological sex, systolic and diastolic function baseline parameters were mostly similar between males and females.

Ten of these eighteen mice of each sex were then treated for three weeks with a continuous infusion of isoproterenol (Iso), a non-specific beta-adrenergic receptor agonist. On day 21, osmotic pumps were removed and the day after, a second echo exam was performed and data were compared to controls (Ctrl). As depicted in Table 1, Iso treatment had relatively similar effects in male and female mice on M-mode measurements. End systolic LV diameter (ESD) was increased in mice of both sexes resulting in a corresponding decrease in fractional shortening (-7.4% for males and -7.9% for females). SAX M-mode data were similar (not shown but included in the raw data). Both end-diastolic (EDV) and end-systolic volumes (ESV) were increased after Iso treatment, respectively by 23% and 49% in males and by 14% (not significant; ns) and 60% in females. This resulted in lowered ejection fraction for both male (-8.7%) and female (-14.7%) mice compared to control. Stroke volume was unchanged in males but tended to decrease in females (ns). Cardiac output was lower in Iso-treated mice.

Iso treatment also influenced diastolic echo parameters (Table 2). The E wave measured by pulse-wave Doppler was decreased in male mice but not in females. We then measured E' and A' waves from the mitral valve annulus movements by tissue Doppler. The E' wave was significantly reduced by 34% in males, whereas the E/E' ratio was significantly increased by Iso (30%). Only a tendency was registered for females (+20%). Isovolumetric relaxation time (IVRT) was significantly longer in both Iso groups compared to controls (+40% in males and +20% in females, respectively).

In Table 3 is illustrated the evolution of global longitudinal (GLS) and global circumferential (GCS) strains in male and female mice receiving Iso. Global strain measurements take into consideration the entire LV wall comparing the relative changes in LV inner contour length during the cardiac cycle. GLS is calculated from the PSLAX view and GCS from SAX. For both parameters, GLS and GCS, more negative values are associated with better fractional change. In control animals, GLS and GCS values were similar between the sexes. After 3 weeks of Iso infusion, GLS values became significantly less negative in both males (+31%) and females (+31%). As for GCS, it worsened similarly in males (+17%) and females (+23%) after Iso.

# Segmental analysis using speckle tracking echocardiography (STE) points toward a non-uniform LV response to Iso in males

The LV was divided into six segments for the PSLAX view, as described in Fig. 1A. This allowed us to investigate if the effects of Iso infusion were distributed globally along the LV endocardial wall or if one or many segments were more seriously affected than others were.

Parameters	M Ctrl, N = 8	M Iso, N = 10	F Ctrl, N = 8	F Iso, N = 10	Sex	Iso	Sex x Iso
E, mm/s	$662\pm23.6$	$551 \pm 18,4^{\circ}$	$578 \pm 15.6$	$579 \pm 14.2$	ns	< 0.01	< 0.01
A, mm/s	$430\pm21.7$	$380\pm9.0$	$363 \pm 10.1$	$375\pm16.9$	< 0.05	ns	ns
E/A	$1.55\pm0.03$	$1.45\pm0.03$	$1.60\pm0.05$	$1.57\pm0.07$	ns	ns	ns
E dec, ms	$20.0\pm1.20$	$25.2\pm1.65$	$18.8\pm0.70$	$22.4 \pm 1.88$	ns	<0.01	ns
E', mm/s	$27.6 \pm 1.49$	$18.3 \pm 1.04^{\rm d}$	$26.1 \pm 1.61$	$22.3\pm0.81$	ns	< 0.0001	ns
A', mm/s	$19.3\pm0.44$	$13.8\pm0.98^{\circ}$	$20.4\pm1.41$	$16.4\pm0.49^{a}$	ns	< 0.0001	ns
E/E'	$24.2\pm1.45$	$31.4\pm1.57^{\rm a}$	$22.3\pm1.42$	$30.5\pm2.54$	< 0.05	< 0.001	ns
E'/A'	$1.43\pm0.06$	$1.36\pm0.04$	$1.29\pm0.04$	$1.43\pm0.06$	ns	ns	ns
IVRT, ms	$15.1\pm0.32$	$21.1\pm0.57^{\rm d}$	$16.1\pm0.61$	$19.4\pm0.93^{\rm b}$	ns	< 0.0001	< 0.05

#### Table 2 Left ventricle diastolic parameters in male and female mice receiving isoproterenol or not for three weeks.

Notes.

Dec, deceleration time; IVRT, Isovolumetric relaxation time.

Values are expressed as the mean  $\pm$  SEM. Two-way ANOVA statistical analysis results are displayed for each factor, sex and Iso, respectively. Intergroup *p* values were calculated using Holm-Sidak post-test.

a p < 0.05.

 $^{b}p < 0.01.$ 

 $^{c}p < 0.001.$ 

 $^{d}p < 0.0001$  between Ctrl and Iso groups, respectively.

ns, not significant.

Table 3 Global LV strain parameters in male and female mice treated with isoproterenol or not for three weeks.

Parameters	M Ctrl, N = 8	M Iso, N = 10	F Ctrl, N = 8	F Iso, N = 10	Sex	Iso	Sex × Iso
GLS	$-19.1\pm0.6$	$-13.1\pm0.8^{d}$	$-20.3\pm0.8$	$-14.0\pm0.9^{\circ}$	ns	< 0.0001	ns
GCS	$-27.6\pm1.5$	$-18.3\pm1.0^{\rm d}$	$-26.1\pm1.6$	$-22.2\pm0.8$	ns	< 0.0001	< 0.05

Notes.

GLS, global longitudinal strain; GCS, global circumferential strain.

Values are expressed as the mean  $\pm$  SEM.

Two-way ANOVA statistical analysis results are displayed for each factor, sex and Iso, respectively.

Intergroup *p* values were calculated using Holm-Sidak post-test.

 $^{c}p < 0.001.$ 

 $d^{\hat{p}} < 0.0001$  between Ctrl and Iso groups, respectively.

ns, not significant.

Using segmental analysis, we compared the strain at baseline (Pre-Iso) and after Iso in male and female mice. We chose to use baseline values as control in order to limit the effects of intra-group variability between animals. As illustrated in Figs. 1B–1E, radial and longitudinal strains were reduced by Iso in males for all three posterior LV wall segments, whereas anterior segments strain was mostly unchanged. Radial strains of two posterior wall segments (base and mid) was also reduced in females. In addition, radial strain for all LV anterior wall segments was reduced. Average radial and longitudinal strains (all segments; in black) were negatively modulated by Iso for both male and female mice.

We grouped data from the segmental strain analysis as from either the base, the mid-ventricle or the apex regardless of the posterior or the anterior walls. We did the same for the anterior or the posterior wall data that were analyzed together regardless if they originated from the base, mid-ventricle or apex (*Kudej et al., 1997*). As illustrated in Figs. 1E–1M, both radial and longitudinal strains were reduced for the base LV segment

in male and female mice. Mid LV radial strain was reduced for both sexes. This was also the case for the longitudinal strain in males as well as the apex longitudinal strain in male and female animals. We then compared if the posterior wall strain was more affected by Iso than the anterior wall. In females, Iso decreased both radial and longitudinal strains to the same extent (-30%). In males, the radial strain was significantly decreased only for the posterior wall. The longitudinal strain was reduced for both walls but this decrease was more important for the posterior wall than the anterior one (-39% vs. -18%, respectively).

We measured the indexed time-to-peak (T2P) for strain data. This represents the time for strain to reach its maximal value from baseline (R-wave). We indexed the value for the duration of the cardiac cycle (1/HR) in order to take into account variations in heart rate between animals. Values are thus expressed as a percentage of a cardiac cycle for strain to reach its peak. As illustrated in Fig. S1, iT2P for each LV segment in radial and longitudinal remained mostly stable in males. In females however, a greater variation after Iso was observed suggesting that normal LV wall synchronicity was perturbed. We then calculated the standard deviation (SD) of iT2P for each 6 segments. In males (blue), these parameters remained stable after Iso but in females (orange), radial iT2P SD was increased.

As illustrated in Video S1 (males) and Video S2 (females), LV wall deformation after Iso was markedly reduced. When LV movement tendency was expressed using velocity vectors as illustrated in Fig. 2, it can be appreciated that in both systole and diastole, the reduction of the length and the changes in the orientation of velocity vectors induced by Iso.

Peak strain rate (SR) represents a systolic function index that indicates the maximal rate of deformation (strain) during systole. On the other hand, reverse peak SR happens during the passive LV filling phase of diastole (Fig. 3A) and has been proposed as a new index of diastolic function (*Schnelle et al., 2018*). Changes in radial and longitudinal strain rates (SR) caused by Iso were relatively minor and concentrated on the posterior segments in males and females (Figs. 3B–3E). Peak reverse SR showed a pattern reminiscent of those of strain and SR. Iso negatively modified LV posterior segments in both males and females (Figs. 3F–3H). Several anterior wall segments were also affected, mostly in females.

Peak speckle displacement and peak speckle velocity are analogous to strain and strain rate although they are not expressed relative to a second point in the myocardium as for the strain and SR. Original speckle position or speckle velocity is determined at the R-wave and is arbitrarily fixed to zero. Iso reduced radial speckle displacement and velocity mainly for the posterior wall segments in males and females compared to baseline values. Displacement was also decreased for the anterior wall in females. In the longitudinal direction, peak speckle displacement remained stable in males and was reduced for only one segment (anterior base) in females. A similar situation was observed for longitudinal velocity of the LV wall (Fig. S2).

After 21 days (week 3) of Iso treatment, the osmotic micro-pumps were removed and the LV reverse remodeling was studied by echo at week 8 and 20 or 5 and 17 weeks post-Iso, respectively. As illustrated in Figs. 4A–4J, LV end-systolic volume in males remained increased compared to control animals, whereas in females, no difference was present. Iso cessation helped reduce LV end-systolic volume in both males and females. However, only females returned to normal values at week 8 before increasing again 12 weeks later. This



**Figure 2 PSLAX LV wall trace tendency before and after Iso.** LV wall trace tendency is expressed as velocity vectors for >48 points around the endocardium in systole (left panels) and diastole (right panels) before and after Iso in males (top panels) and female mice (bottom panels). Images of velocity vectors corresponds to the maximal peak (systole) and minimal peak (diastole) of the average curve of all six segments curves for speckle velocity in PSLAX. As evidenced by these PSLAX B-mode views, velocity vector orientation and length varies during the cardiac cycle. Vertical (radial) and horizontal (longitudinal) components of each vector do not correspond necessarily to the respective peak value of each orthogonal component.

Full-size 🖾 DOI: 10.7717/peerj.11085/fig-2

Peer





Full-size DOI: 10.7717/peerj.11085/fig-3

#### Figure 3 (... continued)

As evidenced by this representation, the first SR peak (top yellow circle) corresponds to the maximal SR (1/s) whereas the second peak (bottom yellow circle) is inverted and happens during the early stage of LV relaxation as evidenced by the M-mode image underneath. A male mouse after three weeks of Iso is represented. B-H. Before after effects of Iso on strain rate (SR) and reverse strain rate (rSR). Radial (B–C) and longitudinal (D–E) peak SR and rSR (F–H) are illustrated. Characters colors correspond to those used in the graphs for each of these LV segments. Ant, anterior; Post, posterior; SW, septal wall and FW, free wall. Males are represented on the left panels and females on the right. Results are represented as violin plots (n = 8-10). Inner black lines represent quartiles of the data. Significance between groups was calculated with paired Student's *T*-test. \*, p < 0.05, \*\*, p < 0.01 and \*\*\*, p < 0.001 between corresponding pre-Iso and Iso groups.

resulted that both male and female mice had a decreased ejection fraction four months post-Iso. On the other hand, both diastolic parameter, E/E' and IVRT, were normalized 5 weeks after Iso treatment. At the end of the follow-up, GLS and GCS had returned to normal for males but only GCS for females (Figs. 4K–4L). Segmental strain analysis showed a similar picture for males where strain values were unchanged compared to age-matched controls. On the other hand, both radial and longitudinal strains showed abnormalities in females 17 weeks after Iso treatment (Fig. 4M). Posterior apical segment in radial and both apical segments showed diminished strain (Figs. 4O–4R). We also look at the reverse strain rate as a diastolic parameter. Compared to control animals, average reverse SR in Iso-treated animals at week 20 were normal (Fig. 4N).

## DISCUSSION

In the present study, we demonstrated that the LV response to beta-adrenergic suractivation using Iso, resulted in both systolic and diastolic function impairments in mice of both sexes. Using speckle-tracking echocardiography, we proceeded to a thorough investigation of the strain, strain rate and other related parameters. In males, we observed that the LV posterior wall was in general, more negatively affected by Iso than the anterior wall. This was true for the strain in both radial and longitudinal directions. In females, strain in the LV posterior wall was also negatively reduced but the anterior wall was also affected making the effects of Iso more global.

Most baseline values for the different STE parameters (strain, SR, and rSR) were similar between males and females. Radial peak speckle displacement and velocity for several segments were significantly smaller in females, which is likely related to a smaller heart size. To our knowledge, this study is the first to report segmental LV wall displacement and velocity data from normal young adult mice.

#### **Conventional echocardiography**

Systolic function as estimated by the ejection fraction (EF) was relatively more reduced in females than in males. Baseline EF values were higher in females, though. Various methods are available for the evaluation of EF by echo. The availability for small animals of four-dimensional (4D) echo LV chamber reconstruction over the cardiac cycle adds another way to estimate EF. In this study, we finally opted for the Simpson's method from LV chamber area tracings in PSLAX instead of 4D echo. Four-dimensional echo, in



**Figure 4 Reversibility of Iso treatment.** (A–J) Head-to-head longitudinal comparison of several echo parameters of control (open circles) and Iso-treated (closed circles) male (blue) and female (orange) mice. EDV, end-diastolic volume; ESV, end-systolic volume; EF, ejection fraction and IVRT, Isovolumetric relaxation time. \*, p < 0.05, \*\*, p < 0.01, \*\*\*, p < 0.001 and \*\*\*\*, p < 0.0001 between corresponding control and Iso animals using unpaired Student's *T*-test. #, p < 0.05; ##, p < 0.01; ###, p < 0.001 and ####, p < 0.0001 between measurements at different times using ANOVA and Tukey's pos *t*-test. K–L, Global longitudinal strain (GLS) and global circumferential strain (GCS) at week 20 in male and female mice treated or not with Iso for 3 weeks. M–N, Average radial and longitudinal peak strains at week 20. O–R, Radial and longitudinal peak strains at week 20 in male and female mice treated or not with Iso. The six segments are identified. Results are represented as violin plots (n = 8-10). Inner black lines represent quartiles of the data. Significance between groups was calculated with unpaired Student's *T*-test. \*, p < 0.05; \*\*, p < 0.01 and \*\*\*, p < 0.001 between corresponding control (Ctrl) and Iso animals. Full-size **E** DOI: 10.7717/peerj.11085/fig-4

our hands, probably underestimated LV volume, which lead to incorrect calculations of LV volumes, stroke volume, ejection fraction and so forth. Depending on the method, EF estimates went from being in the 65–70% range using M-mode, 57–62% range using Simpson's method to 55–60% (PSLAX) and 45–50% (SAX) using 4D echo in normal mice.

In addition, Iso effects seemed to be masked using 4D echo and variability was high using this method.

Several factors can limit acquisition of quality 4D echo scans in rodents as described before in several studies (*Grant et al., 2020*; *Grune et al., 2018*; *Damen et al., 2017*; *Rutledge et al., 2020*). One is interference from anatomical structures (sternum, ribs and lungs) that often obscure parts of the heart, making it difficult to visualize and to trace LV walls. Working with those low-quality 4D echo scans can significantly increase intra-observer variability and thus, reduce reproducibility.

Despite these discrepancies between the methods used to evaluate EF, Iso increased both systolic LV diameters and volumes in mice. A LV dilation was also apparent in 2D B-mode views, whereas the wall thickness seemed to remain remained stable (see Fig. 2, Videos S1 and S2). This LV remodeling helped preserve cardiac output although a trend for a decrease was present. It is difficult to conclude that mice were experiencing symptoms of HF, especially since cardiac output was mostly preserved. HF in small rodents is often recognized by increased lungs weight after euthanasia or decreased exercise performance (*Gomes et al., 2013*). We did not test resistance to forced exercise in our animals.

In the case of diastolic parameters measured by either pulse-wave or tissue Doppler, many of them were negatively modulated by Iso in male mice. In females, only those measured by tissue Doppler were affected by Iso and, to a lesser extent than for males. As in humans, defining diastolic dysfunction by echo is difficult although clearly, Iso treatment caused diastolic function impairments, especially in male mice.

Increased myocardial interstitial fibrosis is often related to diastolic dysfunction. Iso treatment has been shown to induce intertitial fibrosis in mice in other studies. Collagen production was shown to by more important in male mice receiving Iso than in females (*Zhu et al., 2016*). Among the other mechanisms that have been proposed to explain HF induced by Iso is an increased rate of cardiomyocyte apoptosis (*Zhuo et al., 2013*). Sur-activation of the beta-adrenergic system by Iso imposes an increased cardiac workload, which is associated with increased consumption of oxygen by the myocardium. This can lead to increased production of reactive oxygen species (*Ma et al., 2015*). An additional mechanism for negative effects of Iso is lipotoxicity. In an acute model of Iso-induced stress cardiomyopathy, rapid lipid accumulation was noticed as soon as two hours after injection. It is not clear if here, in our chronic setting, that this intracellular lipid accumulation is present or lasts for days but this does not exclude a possible toxicity of Iso for the cardiac myocytes leading to apoptosis (*Shao et al., 2013b*).

### Speckle-tracking echocardiography

In this study, conventional echo highlighted LV anomalies both morphological and functional. Our study design did not aim at testing if strain analysis was more sensitive for early detection of dysfunction as it was done previously (*An et al., 2016; Li et al., 2014; Peng et al., 2009; Szymczyk et al., 2013*). Our goal was to evaluate if additional and valuable information could be obtained using STE such as segmental strain and SR but also reverse SR, speckle displacement and speckle velocity.

Using the strain analysis, we observed that the LV posterior wall was in general, more negatively modulated by Iso than the anterior wall. The base and the mid-ventricle segments were the most affected and the apex, to a lesser extent. This trend was clearer in males although it was also present in females. Obviously, Iso treatment being longer and more severe in this study, triggered a compensatory response (*An et al., 2016*). LV dilatation had time to take place in males, which had to be accompanied by a concomitant extracellular matrix remodeling. Fibrosis has been described in other studies looking at Iso effects on the mouse myocardium (*Grant et al., 2020; Zhuo et al., 2013*). This long exposure to Iso also likely brought a general down regulation of beta-adrenergic receptors leaving non-receptor-mediated Iso effects play a significant part here.

The reason for the posterior wall being more sensitive to Iso than the anterior one is not clear. Differences in LV regional beta-receptor sub-types density have been reported before. Greater apical beta-adrenergic receptor density or responsiveness has been described in humans, dogs, rats, cats and rabbit hearts (*Mori et al., 1993; Kawano, Okada & Yano, 2003; Heather et al., 2009; Lathers, Levin & Spivey, 1986; Mantravadi et al., 2007*). Use of Iso infusion to create infarct-like damages and HF or for inducing Takotsubo-like syndrome selectively targets the LV apex (*Shao et al., 2013b; Lathers, Levin & Spivey, 1986; Mantravadi et al., 2007; Paur et al., 2012; Rona, 1959; Shao et al., 2013c*). This basal-apex gradient of beta-adrenergic receptor responsiveness has thus been well-described. Here, our observation made in the PSLAX now includes an additional axis for a possible anteroposterior gradient in beta-adrenergic receptor sensitivity to sur-activation. Unfortunately, we could only describe this observation without providing satisfactory explanations. The complex 3D architecture of the myocardium may provide clues, but more studies are needed to provide new insights about this intriguing observation.

Speckle peak displacement data at baseline show that in the radial direction, posterior segments are more mobile than anterior ones (Fig. S2). This is associated with larger radial wall velocities for these posterior LV segments. Here too, it was these posterior segments that were more negatively targeted by Iso for both peak displacement and peak velocity. In the longitudinal direction, baseline values were relatively similar to posterior and anterior segments.

As illustrated in Fig. 4, velocity vectors originating from the LV base (and the apex) show a more important longitudinal component than radial. For the mid-ventricle segments, it is the opposite and the radial component of the velocity vectors is more important. Strain and SR measurements are expressed relative to a baseline position at the initiation of systole (R-wave). They do not provide, however, a clear evaluation of the changes in the direction (more radial or more longitudinal) of these vectors after Iso treatment. These vectors also provide information about LV relaxation. This illustrates the high complexity of cardiac contraction and relaxation and the difficulty to assess regional LV dynamic response using only one dimension at a time, here radial vs. longitudinal.

Reverse strain rate has been proposed as an index of diastolic function in mice (*Schnelle et al., 2018*). Considering that most parameters measured by STE are systolic in nature, reverse strain rate can constitute an interesting window to the kinetics of LV relaxation, at least during its passive filling. It is interesting to notice that again, the posterior wall

segments in males are the ones with the more reduced reverse SR suggesting LV stiffening caused by Iso seemed to mainly target this region.

STE strain analysis has been performed in the past in Iso-treated mice. In male mice receiving Iso for either three or seven days, global radial and longitudinal strain and strain rate were reduced in PSLAX and only strain rates were reduced in SAX. When concentrating on LV wall segments in PSLAX, the authors found no regional differences suggesting that Iso effects on the LV after three or seven days were mostly global (*An et al., 2016*). Their dosage of Iso used was lower (5 vs. 30 mg/kg here) and duration of treatment shorter, making it difficult to make comparison with our work. It is probable that in the present study, more chronic mechanisms took place at the cellular and molecular levels, as mentioned above. Interestingly, after only three days, An and collaborators observed in their mice, increased myocardial fibrosis and hypertrophy. They did not mention if chamber dilatation was present but reported LV wall thickening. We did not observe this in our mice after three weeks of Iso (*An et al., 2016*).

We thus observe a clear reduction of global strain measurements in our animals and this was similar between males and females. These parameters are highly sensitive to detect cardiac dysfunction but do not provide regional information. Since most sex differences we observed were present at the regional level, global longitudinal and circumferential strains were not informative.

#### A Takotsubo cardiomyopathy model?

As mentioned above, isoproterenol has been used to develop animal models of Takotsubo syndrome. Takotsubo syndrome is a "transient LV dysfunction (hypokinesia, akinesia, or dyskinesia) presenting as apical, midventricular, basal, or focal ballooning" (*Napp* & *Bauersachs*, 2020). The adrenergic overstimulation is believed to be an important cause of this cardiomyopathy, which is more frequent in postmenopausal women suggesting possible protective roles for estrogens and/or for the male sex (*Sachdev, Merz & Mehta*, 2015).

The rat is usually the preferred animal model to study SIC. It has been studied acutely after a bolus administration of Iso or after a few days of treatment. These short regimens usually allow complete recuperation of systolic function in days or weeks following Iso (*Wallner et al., 2016; Shao et al., 2013a; Ma et al., 2011; Redfors et al., 2014; Willis et al., 2015*). Beta<sub>2</sub>- adrenergic receptor sarcolemmal localization was proposed for being responsible for the typical apical ballooning associated with Takotsubo cardiomyopathy (*Wright et al., 2018*). Since mid-ventricular and basal forms of this SIC exist, other mechanisms are involved.

One study had been conducted in mice using Iso (one single dose of 400 mg/kg) to induce a Takotsubo-like syndrome (*Shao et al., 2013b*). It did not result in global reduction of systolic function when assessed two hours after a bolus Iso injection. Segmental fractional wall thickening was measured in SAX view in these mice. Interestingly, two segments had their radial strain severely reduced namely the posterior wall and inferior free wall segments. These segments in SAX are part of the posterior wall in PSLAX. The four other LV segments had an increased strain to compensate in this study (*Shao et al., 2013b*). We did not observe this pattern of compensation in our mice. Most attempts to reproduce a Takotsubo-like syndrome in rodents have relied so far on acute administration of Iso or catecholamines. This relies on the hypothesis that an acute surge of circulating catecholamines levels or adrenergic overstimulation are important parts of SIC etiology in patients. Known triggering factors in humans such as the death of a loved one, divorce, financial loss, diagnosis of a serious disease all have an important chronic component. In addition, circulating levels of catecholamines are seldom elevated in SIC patients (*Shao et al., 2013b*). This can make our study relevant to mimic human SIC since mice received for a long period of time a beta-receptor agonist. One limitation is that Iso does not stimulate alpha receptors as catecholamines do, eliminating the vasospasm component of Takotsumo cardiomyopathy (*Shao et al., 2013b*).

Interestingly, we showed that cessation of Iso was accompanied with improvement of systolic and diastolic function parameters. Diastolic parameters quickly returned to normal 5 weeks post-Iso. It was not the case for the systolic function. In females, the apex region strain got worse with time after initial normalization 5 weeks post-Iso. This suggests that myocardial abnormalities remain long after Iso cessation and that may lead to an eventual degradation of heart function. The reversible nature of Takotsubo syndrome in patients has been questioned for some time in the literature (*Pelliccia et al., 2017*). If regional LV contraction and global ejection fraction recover completely, other parameters of cardiac function (*Dias et al., 2019*).

#### **Study limitations**

Among the limitations of this study, tissue data cannot be obtained since the animals were not euthanized at the end of Iso treatment as their longitudinal follow-up was continued for several months. Any tissue data in this context would have been from LVs having a long period of recuperation after Iso treatment. It will be interesting to evaluate how sex hormones and age can influence LV response to chronic Iso in the future. In addition, the study in short axis (SAX) at the level of the apex, the midsection and the base the strain and related parameters could be informative to correlate changes observed here in PSLAX.

## **CONCLUSION**

Segmental strain analysis in mice can provide information about the regional influence of a toxic cardiac insult such as Iso continuous infusion. We observed both similarities and sex-related differences in our male and female mice. Systolic and diastolic functions were negatively modulated by Iso. Differences between sexes were relatively subtle when studied by conventional echo. LV dilation was more important in males and remained present post-Iso. By STE, we observed Iso had a more global effect on the female LV whereas in males, the posterior wall was more specifically targeted. Normalization after Iso treatment was not complete for either male or female mice.

## ACKNOWLEDGEMENTS

The authors want to acknowledge the technical help of Ms. Marine Clisson and Mr. Thomas Couët.

## **ADDITIONAL INFORMATION AND DECLARATIONS**

### Funding

This work was supported by an operating grant to Jacques Couet and Marie Arsenault from the Canadian Institutes of Health Research (MOP-123186) and the IUCPQ Foundation. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

### **Grant Disclosures**

The following grant information was disclosed by the authors: Marie Arsenault from the Canadian Institutes of Health Research: MOP-123186. IUCPQ Foundation.

### **Competing Interests**

The authors declare there are no competing interests.

## **Author Contributions**

- Elisabeth Walsh-Wilkinson conceived and designed the experiments, performed the experiments, analyzed the data, prepared figures and/or tables, and approved the final draft.
- Marie Arsenault conceived and designed the experiments, authored or reviewed drafts of the paper, and approved the final draft.
- Jacques Couet conceived and designed the experiments, performed the experiments, analyzed the data, prepared figures and/or tables, authored or reviewed drafts of the paper, and approved the final draft.

### **Animal Ethics**

The following information was supplied relating to ethical approvals (i.e., approving body and any reference numbers):

The protocol was approved by the Université Laval's animal protection committee and followed the recommendations of the Canadian Council on Laboratory Animal Care (protocol #2019-360, VRR-19-075).

#### **Data Availability**

The following information was supplied regarding data availability:

All data for the different figures are available in the Supplemental Files.

#### **Supplemental Information**

Supplemental information for this article can be found online at http://dx.doi.org/10.7717/ peerj.11085#supplemental-information.

### REFERENCES

Ali A, Redfors B, Lundgren J, Alkhoury J, Oras J, Gan LM, Omerovic E. 2019. Effects of pretreatment with cardiostimulants and beta-blockers on isoprenaline-induced

takotsubo-like cardiac dysfunction in rats. *International Journal of Cardiology* **281**:99–104 DOI 10.1016/j.ijcard.2018.12.045.

- An X, Wang J, Li H, Lu Z, Bai Y, Xiao H, Zhang Y, Song Y. 2016. Speckle tracking based strain analysis is sensitive for early detection of pathological cardiac hypertrophy. *PLOS ONE* 11:e0149155 DOI 10.1371/journal.pone.0149155.
- Angelini P, Gamero MT. 2019. What can we learn from animal models of Takotsubo syndrome? *International Journal of Cardiology* 281:105–106 DOI 10.1016/j.ijcard.2019.01.064.
- Chang SC, Ren S, Rau CD, Wang JJ. 2018. Isoproterenol-induced heart failure mouse model using osmotic pump implantation. *Methods in Molecular Biology* 1816:207–220 DOI 10.1007/978-1-4939-8597-5-16.
- Damen FW, Berman AG, Soepriatna AH, Ellis JM, Buttars SD, Aasa KL, Goergen CJ. 2017. High-frequency 4-dimensional ultrasound (4DUS): a reliable method for assessing murine cardiac function. *Tomography* 3:180–187 DOI 10.18383/j.tom.2017.00016.
- Dias A, Núñez Gil IJ, Santoro F, Madias JE, Pelliccia F, Brunetti ND, Salmoirago-Blotcher E, Sharkey SW, Eitel I, Akashi YJ, El-Battrawy I, Franco E, Akin I, Jaguszewski M, Dawson D, Figueredo VM, Napp LC, Christensen TE, Hebert K, Ben-Dor I, Ozaki Y, García-Garcia HM, Kajita AH, Akasaka T, Kurisu S, Lerman A, Waksman R. 2019. Takotsubo syndrome: State-of-the-art review by an expert panel—Part 2. *Cardiovascular Revascularization Medicine* 20:153–166 DOI 10.1016/j.carrev.2018.11.016.
- Gomes AC, Falcao-Pires I, Pires AL, Bràs-Silva C, Leite-Moreira AF. 2013. Rodent models of heart failure: an updated review. *Heart Failure Reviews* 18:219–249 DOI 10.1007/s10741-012-9305-3.
- Grant MKO, Abdelgawad IY, Lewis CA, Seelig D, Zordoky BN. 2020. Lack of sexual dimorphism in a mouse model of isoproterenol-induced cardiac dysfunction. *PLOS ONE* 15:e0232507 DOI 10.1371/journal.pone.0232507.
- Grune J, Blumrich A, Brix S, Jeuthe S, Drescher C, Grune T, Foryst-Ludwig A, Messroghli D, Kuebler WM, Ott C, Kintscher U. 2018. Evaluation of a commercial multi-dimensional echocardiography technique for ventricular volumetry in small animals. *Cardiovascular Ultrasound* 16:10 DOI 10.1186/s12947-018-0128-9.
- Heather L, Catchpole A, Stuckey D, Cole M, Carr C, Clarke K. 2009. Isoproterenol induces in vivo functional and metabolic abnormalities; similar to those found in the infarcted rat heart. *Acta Physiologica Polonica* **12**:31–39.
- Kawano H, Okada R, Yano K. 2003. Histological study on the distribution of autonomic nerves in the human heart. *Heart and Vessels* 18:32–39 DOI 10.1007/s003800300005.
- Kudej RK, Iwase M, Uechi M, Vatner DE, Oka N, Ishikawa Y, Shannon RP, Bishop SP, Vatner SF. 1997. Effects of chronic beta-adrenergic receptor stimulation in mice. *Journal of Molecular and Cellular Cardiology* 29:2735–2746 DOI 10.1006/jmcc.1997.0508.

- **Lathers CM, Levin RM, Spivey WH. 1986.** Regional distribution of myocardial  $\beta$ -adrenoceptors in the cat. *European Journal of Pharmacology* **130**:111–117 DOI 10.1016/0014-2999(86)90189-5.
- Li RJ, Yang J, Yang Y, Ma N, Jiang B, Sun QW, Li YJ. 2014. Speckle tracking echocardiography in the diagnosis of early left ventricular systolic dysfunction in type II diabetic mice. *BMC Cardiovascular Disorders* 14:141 DOI 10.1186/1471-2261-14-141.
- Ma X, Fu Y, Xiao H, Song Y, Chen R, Shen J, An X, Shen Q, Li Z, Zhang Y. 2015. Cardiac fibrosis alleviated by exercise training is AMPK-dependent. *PLOS ONE* 10:e0129971 DOI 10.1371/journal.pone.0129971.
- Ma X, Song Y, Chen C, Fu Y, Shen Q, Li Z, Zhang Y. 2011. Distinct actions of intermittent and sustained  $\beta$ -adrenoceptor stimulation on cardiac remodeling. *Science China Life Sciences* 54:493–501 DOI 10.1007/s11427-011-4183-9.
- Mantravadi R, Gabris B, Liu T, Choi BR, De Groat WC, Ng GA, Salama G. 2007. Autonomic nerve stimulation reverses ventricular repolarization sequence in rabbit hearts. *Circulation Research* 100:e72–e80.
- Michel FS, Magubane M, Mokotedi L, Norton GR, Woodiwiss AJ. 2017. Sex-specific effects of adrenergic-induced left ventricular remodeling in spontaneously hypertensive rats. *Journal of Cardiac Failure* 23:161–168 DOI 10.1016/j.cardfail.2016.09.017.
- Mori H, Ishikawa S, Kojima S, Hayashi J, Watanabe Y, Hoffman JI, Okino H. 1993. Increased responsiveness of left ventricular apical myocardium to adrenergic stimuli. *Cardiovascular Research* 27:192–198 DOI 10.1093/cvr/27.2.192.
- Napp LC, Bauersachs J. 2020. Takotsubo syndrome: between evidence, myths, and misunderstandings. *Herz* 45:252–266 DOI 10.1007/s00059-020-04906-2.
- Paur H, Wright PT, Sikkel MB, Tranter MH, Mansfield C, O'Gara P, Stuckey DJ, Nikolaev VO, Diakonov I, Pannell L, Gong H, Sun H, Peters NS, Petrou M, Zheng Z, Gorelik J, Lyon AR, Harding SE. 2012. High levels of circulating epinephrine trigger apical cardiodepression in a  $\beta_2$ -adrenergic receptor/ G<sub>i</sub> dependent manner: a new model of Takotsubo cardiomyopathy. *Circulation* 126:697–706 DOI 10.1161/CIRCULATIONAHA.112.111591.
- Pelliccia F, Kaski JC, Crea F, Camici PG. 2017. Pathophysiology of Takotsubo syndrome. *Circulation* 135:2426–2441 DOI 10.1161/CIRCULATIONAHA.116.027121.
- Peng Y, Popovic ZB, Sopko N, Drinko J, Zhang Z, Thomas JD, Penn MS. 2009. Speckle tracking echocardiography in the assessment of mouse models of cardiac dysfunction. *The American Journal of Physiology-Heart and Circulatory Physiology* 297:H811–H820 DOI 10.1152/ajpheart.00385.2009.
- Puhl SL, Weeks KL, Ranieri A, Avkiran M. 2016. Assessing structural and functional responses of murine hearts to acute and sustained -adrenergic stimulation in vivo. *Journal of Pharmacological Toxicol Methods* 79:60–71 DOI 10.1016/j.vascn.2016.01.007.
- Redfors B, Shao Y, Wikström J, Lyon AR, Oldfors A, Gan LM, Omerovic E. 2014. Contrast echocardiography reveals apparently normal coronary perfusion in a rat model of stress-induced (Takotsubo) cardiomyopathy. *European Heart Journal: Cardiovascular Imaging* 15:152–157 DOI 10.1093/ehjci/jet079.

- **Rona G. 1959.** An infarct-like myocardial lesion and other toxic manifestations produced by isoproterenol in the rat. *Archives of Pathology* **67**:443–455.
- Roussel E, Drolet MC, Lavigne AM, Arsenault M, Couet J. 2018. Multiple short-chain dehydrogenases/reductases are regulated in pathological cardiac hypertrophy. *FEBS Open Bio* 8:1624–1635 DOI 10.1002/2211-5463.12506.
- Rutledge C, Cater G, McMahon B, Guo L, Nouraie SM, Wu Y, Villanueva F, Kaufman BA. 2020. Commercial 4-dimensional echocardiography for murine heart volumetric evaluation after myocardial infarction. *Cardiovascular Ultrasound* 18:1–10 DOI 10.1186/s12947-020-0185-8.
- Sachdev E, Merz CNB, Mehta PK. 2015. Takotsubo cardiomyopathy. *European Cardiology Review* 10:25–30 DOI 10.15420/ecr.2015.10.01.25.
- Sachdeva J, Dai W, Kloner RA. 2014. Functional and histological assessment of an experimental model of Takotsubo's cardiomyopathy. *Journal of the American Heart Association* 3:e000921 DOI 10.1161/jaha.114.000921.
- Schnelle M, Catibog N, Zhang M, Nabeebaccus AA, Anderson G, Richards DA, Sawyer G, Zhang X, Toischer K, Hasenfuss G, Monaghan MJ, Shah AM. 2018. Echocardiographic evaluation of diastolic function in mouse models of heart disease. *Journal of Molecular and Cellular Cardiology* 114:20–28 DOI 10.1016/j.yjmcc.2017.10.006.
- Shao Y, Redfors B, Scharin Täng M, Möllmann H, Troidl C, Szardien S, Hamm C, Nef H, Borén J, Omerovic E. 2013a. Novel rat model reveals important roles of -adrenoreceptors in stress-induced cardiomyopathy. *International Journal of Cardiology* 168:1943–1950 DOI 10.1016/j.ijcard.2012.12.092.
- Shao Y, Redfors B, Stahlman M, Täng MS, Miljanovic A, Möllmann H, Troidl C, Szardien S, Hamm C, Nef H, Borén J, Omerovic E. 2013b. A mouse model reveals an important role for catecholamine-induced lipotoxicity in the pathogenesis of stress-induced cardiomyopathy. *European Journal of Heart Failure* 15:9–22 DOI 10.1093/eurjhf/hfs161.
- Shao Y, Redfors B, Tang MS, Assarsson U, Omerovic E. 2013c. Novel simple approach for detection of regional perturbations of cardiac function in mouse models of cardiovascular disease. *Echocardiography* 30:843–849 DOI 10.1111/echo.12138.
- Szymczyk E, Lipiec P, Plewka M, Białas M, Olszewska M, Rozwadowska N, Kamiński K, Kurpisz M, Michalski B, Kasprzak JD. 2013. Feasibility of strain and strain rate evaluation by two-dimensional speckle tracking in murine model of myocardial infarction: comparison with tissue Doppler echocardiography. *Journal of Cardiovas-cular Medicine* 14:136–143 DOI 10.2459/JCM.0b013e328351dbe0.
- Wallner M, Duran JM, Mohsin S, Troupes CD, Vanhoutte D, Borghetti G, Vagnozzi RJ, Gross P, Yu D, Trappanese DM, Kubo H, Toib A, 3rd TESharp, Harper SC, Volkert MA, Starosta T, Feldsott EA, Berretta RM, Wang T, Barbe MF, Molkentin JD, Houser SR. 2016. Acute catecholamine exposure causes reversible myocyte injury without cardiac regeneration. *Circulation Research* 119:865–879 DOI 10.1161/circresaha.116.308687.

- Walsh-Wilkinson E, Drolet MC, Arsenault M, Couet J. 2020. Sex differences in the evolution of left ventricle remodeling in rats with severe volume overload. *BMC Cardiovascular Disorders* 20:51 DOI 10.1186/s12872-020-01360-0.
- Willis BC, Salazar-Cantú A, Silva-Platas C, Fernández-Sada E, Villegas CA, Rios-Argaiz E, González-Serrano P, Sánchez LA, Guerrero-Beltrán CE, García N, Torre-Amione G, García-Rivas GJ, Altamirano J. 2015. Impaired oxidative metabolism and calcium mishandling underlie cardiac dysfunction in a rat model of post-acute isoproterenol-induced cardiomyopathy. *The American Journal of Physiology-Heart and Circulatory Physiology* 308:H467–H477 DOI 10.1152/ajpheart.00734.2013.
- Wright PT, Bhogal NK, Diakonov I, Pannell LMK, Perera RK, Bork NI, Schobesberger S, Lucarelli C, Faggian G, Alvarez-Laviada A, Zaccolo M, Kamp TJ, Balijepalli RC, Lyon AR, Harding SE, Nikolaev VO, Gorelik J. 2018. Cardiomyocyte membrane structure and camp compartmentation produce anatomical variation in beta(2)AR-cAMP responsiveness in murine hearts. *Cell Reports* 23:459–469 DOI 10.1016/j.celrep.2018.03.053.
- **Zhu B, Liu K, Yang C, Qiao Y, Li Z. 2016.** Gender-related differences in β-adrenergic receptor-mediated cardiac remodeling. *Canadian Journal of Physiology and Pharma-cology* **94**:1349–1355 DOI 10.1139/cjpp-2016-0103.
- Zhuo XZ, Wu Y, Ni YJ, Liu JH, Gong M, Wang XH, Wei F, Wang TZ, Yuan Z, Ma AQ, Song P. 2013. Isoproterenol instigates cardiomyocyte apoptosis and heart failure via AMPK inactivation-mediated endoplasmic reticulum stress. *Apoptosis* 18:800–810 DOI 10.1007/s10495-013-0843-5.