

Piperazinobenzodiazepinones: New Encephalitic Alphavirus Inhibitors via Ring Expansion of 2-Dichloromethylquinazolinones

Michael C. Ryan, Eunjung Kim, Xufeng Cao, Walter Reichard, Tyler J. Ogorek, Pronay Das, Colleen B. Jonsson, Jerome Baudry, Donghoon Chung, and Jennifer E. Golden*



ABSTRACT: Venezuelan and eastern equine encephalitis viruses are disease-causing, neuropathic pathogens with no approved treatment options in humans. While expanding the pharmacophoric model of antialphaviral amidines prepared via a quinazolinone rearrangement, we discovered that diamine-treated, 2-dihalomethylquinolinones unexpectedly afforded ring-expanded piperazine-fused benzodiazepinones. Notably, this new chemotype (19 examples) showed potent, submicromolar inhibition of virus-induced cell death, >7-log reduction of viral yield, and tractable structure—activity relationships across both viruses. Antiviral activity was confirmed in primary human neuronal cells. A mechanistic rationale for product formation is proposed, and key structural elements were comparatively modeled between a similarly substituted antiviral amidine and piperazinobenzodiazepinone prototypes to guide future antiviral development.

KEYWORDS: Quinazolinone, rearrangement, alphavirus, benzodiazepinones, encephalitis

Vere energy energy and eastern equine encephalitis viruses, VEEV and EEEV, respectively, are neuroinvasive RNA alphaviruses that are transmitted through the bite of infected mosquitoes. These pathogens cause significant human disease that ranges in severity from a febrile, self-resolving flu-like illness to more debilitating cases characterized by disorientation, seizure, coma, and death. While the human mortality rate associated with VEE is generally less than 1%,¹ EEE affects children and the elderly population more severely² and has an overall case fatality rate of approximately 30–50%.³ In 2019, 38 cases of EEE were confirmed in the United States by the Center for Disease Control and Prevention, resulting in 19 fatalities.^{4,5}

Live, attenuated vaccines are available for veterinary use and at-risk lab or military personnel; however, they are considered unacceptable for broader human use due to high reactogenicity, limited seroconversion, and narrow viral strain specificity.^{6,7} While other therapeutic modalities^{8–10} also have been investigated, treatment options to date for VEEV and EEEV infection in humans are limited to supportive care. This scenario, coupled with the bioweapon potential of these viruses, along with reports that survivors of encephalitis often have neurological sequelae that results in permanent physical,

behavioral, and cognitive impairment,^{11,12} underscores the need for safe and effective countermeasures.^{13,14}

Following the discovery of a novel 2-chloromethylquinazolinone rearrangement¹⁵ that afforded potent, antialphaviral benzamidines **1** (Scheme 1, panel a), we have continued to explore this chemistry to elucidate the benzamidine pharmacophore and build structure–activity and structure– property relationships (SAR and SPR, respectively).^{16–18} Consequently, we discovered several quinazolinone-based transformations leading to differentiated, bioactive heterocyclic frameworks.^{19,20}

For instance, 2-chloroquinazolinones, when treated with 1,2dimethylethanediamine, undergo a similar rearrangement to generate benzylguanidines 2 with improved plasma stability compared to the benzamidine series 1.¹⁹ Herein, we describe

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Scheme 1. Structurally Divergent, Bioactive Heterocycles from the Rearrangement of 2-Substituted Quinazolinones a





"Reagents and conditions: (a) NEt₃, CH₃CN, μ W, 150 °C, 2 h; (b) K₂CO₃, CH₃CN, 50 °C, 2 h; (c) KOAc, DMSO, μ W, 60 °C, 2 h.

the discovery of piperazine-fused benzodiazepinones 4 that result from the ring expansion of 2-dichloromethylquinazolinones 3 when treated with N,N'-dialkylethane-1,2-diamines (Scheme 1, panel b). Further, we show that these products potently inhibit cellular VEEV and EEEV replication and provide an opportunity for differentiated scaffold development from the benzamidine template.

As part of an ongoing medicinal chemistry program aimed at developing antivirals against VEEV and EEEV, we attempted to prepare fluorobenzamidine **6** from 2-fluorochloromethylquinazolinone **5** (Scheme 2, path a). We anticipated that *N*,*N*-dimethylethane-1,2-diamine might displace the chlorine atom of a 2-dihaloalkylquinazolinone, leading to intramolecular cyclization with subsequent ring-opening to provide fluorobenzamidine **6**. While none of the benzamidines were observed, we isolated and characterized a small amount (~5%) of piperazine-fused benzodiazepinone **7a**. The

Scheme 2. Discovery of Piperazinobenzodiazepinones and Similarity to Bis-Methylaminobenzodiazepine-*N*-oxide Formation^{*a*}



^{*a*}Reagents and conditions: (a) CH₃NH(CH₂)₂NHCH₃, K₂CO₃, KI, DMF, μ W, 150 °C, 2 h; (b) NH₂CH₃, CH₃OH, r.t., 16 h, no yields reported; PMP = *p*-methoxyphenyl.

conversion of 2-dihaloalkylquinazolinones to benzodiazepinones bears some resemblance to the pioneering work of Dr. Leo Sternbach²¹ who is credited with the discovery of benzodiazepines.²² Specifically relevant is a Roche patent²³ which showed that dichloromethylquinazoline-*N*-oxides such as **8**, when treated with methylamine (15% in MeOH), afforded bis-methylaminobenzodiazepine-*N*-oxide **9** (Scheme 2, path b). Notably, diazepinone 7a contained benzamidine structural features known to be important for anti-VEEV activity. When 7a was evaluated in a cell-based cytopathic effect (CPE) assay employing VEEV TC83, we obtained a remarkable EC₅₀ of 0.041 μ M without observable cytotoxicity (CC₅₀ > 30 μ M). As such, we set out to improve the transformation yield and assess the antiviral SAR of the new chemotype for VEEV and EEEV.

We studied the rearrangement using a 2-dichloromethylquinazolinone because the 2,2-dichloroacetyl chloride required for the quinazolinone assembly was more economical than 2fluoro-2-chloroacetyl chloride, the same products were expected, and chloride as a leaving group was likely more favorable than the loss of fluoride, possibly improving the yield. Furthermore, we also knew from the benzamidines' optimizations that incorporating an amido-N-3-fluoro-4-methoxyphenyl group (i.e., amide-PMP group in 6) typically provided analogs with a better overall profile, and reaction optimization could then be more easily monitored by ¹⁹F NMR. Taking these considerations into account, reaction optimization with 2-(dichloromethyl)-3-(3-fluoro-4-methoxyphenyl)-6-nitroquinolin-4(3H)-one was undertaken (Table S1 in the Supporting Information). Yield and conversion of the transformation were evaluated as a function of solvent, reaction time, and the stoichiometry and identity of the base used. When the reaction was performed in DMSO with 1.5 equiv of diamine at 60 °C for 2 h, yields of the desired product were inferior (3-15%) by ¹⁹F NMR), and the most significant identifiable byproduct, 2amino-N-(3-fluoro-4-methoxyphenyl)-5-nitro-benzamide, resulted from starting material fragmentation. However, after screening a variety of exogenous bases, we found that the desired product was maximally generated in 36% under these conditions when KOAc was added (¹⁹F NMR analysis). As a result, these conditions were used to generate a piperazinobenzodiazepinone collection that was synthesized by the protocols shown in Scheme 3.

Briefly, aminobenzamides 11a-j were used without purification after being prepared by one of three methods. Peptide coupling conditions were used with commercially available anthranilic acids 10a-c to afford 11a-h. Treatment of 5-fluoroisatoic anhydride 12 with iodine and aniline afforded derivative 11i, while 11j was prepared in a threestep protocol from methyl 2-amino-5-cyanobenzoate 13. Employing a modified version of our previously published procedure²⁴ for quinazolinone core assembly, the 2-dichloromethylquinazolinones 14a-j were constructed using 2,2dichloroacetyl chloride to form bis-amides *in situ* (not shown) that underwent TMSCl/NEt₃-mediated ring-closure in yields ranging from 18 to 81%.

The 2-dichloromethylquinazolinone rearrangement was carried out under the optimized conditions involving treatment of 14a-j (1.0 equiv) with diamine (1.5 equiv) and KOAc (2.0 equiv) in DMSO at 60 °C for 2 h. Structural changes in each of the three essential components, i.e., anthranilic acid, aniline, and diamine, were surveyed with selection predominately driven by a known benzamidine SAR.¹⁵ For example, our

Scheme 3. Synthesis of Piperazinobenzodiazepinones^a



^aReagents and conditions: (a) ArNH₂, *i*-Pr₂NEt, EDC HCl, HOBt, DMF, rt, 56–100%; (b) I₂, PhNH₂, EtOH, reflux, 17 h, 47%; (c) Br₂, AcOH, rt, 4 h, 89%; (d) LiOH, THF, H₂O, 0 °C–rt, 17 h, 82%; (e) Cl₂CHCOCl, NEt₃, DCM or DMF, 0 °C–rt, 17 h, then TMSCl, NEt₃, rt, 17 h, 18–81%; (f) R₃NH(CH₂)₂NHR₄ or other diamine, KOAc, DMSO, μ W, 60 °C, 2 h, 8–66%; (g) For 7n, morpholine, 10 mol % Pd XPhos G3, Cs₂CO₃, toluene, reflux, 24 h, 32% (66% brsm); (h) For 7o, 10 mol % Pd(PPh₃)₄, Cs₂CO₃, boronic acid, dioxane, H₂O, 85 °C, 17 h, 69%.

benzamidine-related studies showed that a nitro group at the C5 position (see 6, Scheme 2) was critical for retaining substantial antialphavirus activity. As such, many diazepinones contain this structural feature at the analogous C8 position of the new core, but other substitutions were also examined. Improved activity and solubility resulted with benzamidines bearing an amido-*N*-4-methoxyphenyl ring, so analogs with this component were also made (Scheme 4).

In general, piperazinobenzodiazepinones containing a nitro group and constructed with N,N'-dimethylethane-1,2-diamine were isolated from the rearrangement reaction in modest yield (8-38%), most likely due to the increased reactivity of the nitro-containing substrate (see yields for 7a-e, Scheme 4). Accordingly, moving the nitro group from the C8-position to the C7-position of the piperazinobenzodiazepinone core resulted in an 18% increase in yield, comparing 7i (38%) to 7e (20%). Notably, the rearrangement step performed appreciably better for presumably less activated substrates, including C8-cyano-substituted analogs such as 7k-7m with isolated yields in the 49-66% range. Performing the rearrangement on a 2.2 mmol (1 g) scale generated 7m in an improved 63% yield. The use of bulkier, unsymmetrical diamines on the reaction efficiency and regioselectivity was also examined. Using N,N'-dibenzylethane-1,2-diamine provided benzodiazepinone 7h in 50% yield. Interestingly, using

Scheme 4. Yield of 2-Dichloroquinazolinone Ring Expansion for Various Piperazinobenzodiazepinones^{*a,b*}



^{*a*}Reagents and conditions: (a) diamine, KOAc, DMSO, 60 °C, 2 h; (b) For **7n**, morpholine, 10 mol % Pd XPhos G3, Cs₂CO₃, toluene, reflux, 24 h; (c) For **7o**, 10 mol % Pd(PPh₃)₄, Cs₂CO₃, (6methylpyridin-3-yl)boronic acid, dioxane, H₂O, 85 °C, 17 h. ^{*b*}Isolated yields; ring expansion step performed on a 0.5 mmol scale, unless noted. ^{*c*}Used bis-HCl diamine salt, 2 equiv of K₂CO₃ added. ^{*d*}Scale: 0.37 mmol due to limited starting material. ^{*e*}Scale: 2.2 mmol (1 g) of 2-dichloromethylquinazolinone. ^{*f*}Parenthetical yield for aryl bromide derivatization. ^{*g*}Major isomer shown.

unsymmetrical N,N'-benzylmethylethane-1,2-diamine gave benzodiazepinone 7g in 29% yield as the sole observed regioisomer. Similarly, N,N'-cyclohexylmethylethane-1,2-diamine afforded an analogous benzodiazepinone 7f as the only regioisomer, though only in 8% yield, likely due to the increased bulkiness of the cyclohexyl group. The assigned regiochemistry of both 7f and 7g was determined by 2D NOESY NMR. Additionally, each enantiomer of N,N'dimethylcylohexyl-1,2-diamine was separately employed in the reaction, affording benzodiazepinones 7p and 7q in 30 and 34% yields, respectively, and each in 14:1 d.r., the latter of which was confirmed by NOESY NMR. Lastly, bromide 7m was derivatized by a Buchwald-Hartwig coupling to afford morpholine analog 7n (32% yield, 66% brsm), while Suzuki coupling of 7m gave pyridyl analog 7o (69% yield).

Piperazinobenzodiazepinone formation could result from several plausible mechanisms. Considering an $S_N 2$ substitution reaction between a 2-dichloromethylquinazolinone and a nucleophilic diamine, α -chloroalkylamine **A** may result (Scheme 5, path a), facilitated by KOAc.^{25–27} In this

Scheme 5. Mechanistic Rationale for Diazepinone Formation

(a) S_N2/double ring expansion mechanism



mechanism, iminium ion **B** formation from α -chloroalkylamine **A** may be trapped in an intramolecular fashion by the appended secondary amine to form aminal **C**. Ring expansion of the aminal through neighboring nitrogen atom assistance would then form zwitterionic spirocyclic iminium ion **D**, a species that is similar to those implicated for similar rearrangements. Benzodiazepinone 7 could then be formed through either a synchronous or asynchronous event involving the amide portion of the structure. Specifically, the negatively charged quinazolinone nitrogen could promote a second ring expansion via a 1,2-*N*-acyl migration to the iminium ion carbon or by a sequential 6-membered ring-opening to a 7-membered ring-forming process. An intramolecular 7-membered ring formation involving an *NH*-amide is precedented in peptidomimetic syntheses.²⁸

Attempts to intercept iminium ions via reductive amination were unsuccessful, but the $S_N 2$ mechanisms account for the observed regiochemical outcomes. Specifically, unsymmetrical

diamines yielded products in which the bulkier group resided on the iminium ion nitrogen that was further away from the quinazolinone core in intermediate **D**. Alternatively, we considered a mechanism similar to the Sternbach rearrangement of 2-chloromethylquinazoline-3-oxides that, under treatment with an alkyl amine, affords aminodiazepoxides.²⁹ In our case, nucleophilic addition of the amine to the *N*-acyl iminelike carbon of the quinazolinone core would form **E** (Scheme 5, path b). Subsequent ring-opening/ring-forming or an *N*-acyl shift would generate a ring-expanded chlorobenzodiazepinone **F** that would undergo intramolecular chloride displacement by the tethered amine. At this point, these mechanisms are not differentiated, and multiple scenarios may be relevant depending on the quinazolinone substitution and the diamine partner.

Two orthogonal antiviral, cell-based assays using VEEV and EEEV were implemented to survey the antiviral effects of these compounds (Table 1).¹⁷ Viability of infected Vero 76 cells was measured as a function of compound concentration in a CPE assay, and the reduction of viral yield was measured at a concentration of 5 μ M for compounds with submicromolar CPE EC₅₀ values. Several notable outcomes emerged from these assays. First, data between the VEEV and EEEV CPE assays were strongly congruent (<2.1-fold difference), revealing similar SAR across these two alphaviruses in this assay. Second, data showed that compounds whose viral yield reduction potential was assessed were consistently better by about 2-4-fold against EEEV compared to that of VEEV. Third, we observed that several advantageous structural modifications implemented with the amidine series were also beneficial on the piperazinobenzodiazepinones in terms of antiviral activity. For instance, an N-4-methoxyphenyl amide substituent, in combination with a nitro group present at R¹, resulted in potent inhibition of CPE caused by either VEEV or EEEV (Table 1, entries 1 and 2, $EC_{50} = 27-48$ nM). Potency was eroded when the 4-methoxyphenyl substituent was exchanged for a fluorine or hydrogen atom at that paraposition or if the methoxy group was migrated to the 3position (entries 3-5), though the compounds still retained submicromolar activity. Substitution of one or more of the ring-fused piperazine nitrogen atoms with nonmethyl groups such as a benzyl or cyclohexyl moiety produced inferior results, with the N,N'-dibenzyl derivative 7h faring the worst (analogs 7f-h). Changing the piperazine ring to bicyclic, ring-fused variants (7p-q) while incorporating the *N*-methylpiperazine substitutions provided hit-like potencies in the range of 5 μ M, suggesting that structural modifications may be permissible in this region.

Relocating the R^1 nitro group to the R^2 position (entry 9, 7i) or replacing it with a fluorine atom (entry 10, 7j) was not tolerated (EC₅₀ > 30 μ M). In fact, a limitation of the amidine series was the need for a C5-nitro group (see Scheme 2, compound 6) to achieve exceptional potency in antiviral assays, and despite a survey of many isosteres in that series, only the C5 substitution with a nitrile moiety afforded analogs with low micromolar potency. Analogously, substituting the diazepinone C8 nitro group with a nitrile functionality resulted in a 100- and 75-fold loss in antiviral activity for VEEV and EEEV, respectively (cf., 7a versus 7l); however, the retention of low micromolar activity in these assays provided hope that structural changes made elsewhere on the scaffold may remedy these losses. From a design standpoint, substitution at R³ of the diazepinone core (see general structure of 7, Table 1) seemed reasonable as a hydrogen bond acceptor in that region may

Table 1. Survey of Piperazinobenzodiazepinones on VEEV and EEEV Cytopathic Effect and Viral Yield Reduction



								VEEV INH9813		EEEV VIUS		
compd	\mathbb{R}^1	R ²	R ³	R ⁴	R ⁵	R ⁶	\mathbb{R}^7	$\begin{array}{c} \text{CPE}^a \text{ EC}_{50} \\ (\mu \text{M}) \end{array}$	viral yield redn ^b	$\begin{array}{c} \text{CPE}^a \text{ EC}_{50} \\ (\mu \text{M}) \end{array}$	viral yield redn ^b	CC_{50} $(\mu M)^c$
7a	NO_2	Н	Н	CH_3	CH_3	Н	OCH_3	0.041 ± 0.9	5.5 ± 1.2	0.033 ± 0.002	7.9 ± 0.5	>30
7b	NO_2	Н	Н	CH_3	CH_3	F	OCH_3	0.048 ± 0.004	3.7 ± 1.1	0.027 ± 0.003	7.7 ± 0.8	>30
7c	NO_2	Н	Н	CH_3	CH_3	Н	F	0.15 ± 0.01	2.9 ± 0.9	0.11 ± 0.004	6.1 ± 1.8	>30
7d	NO_2	Н	Н	CH_3	CH_3	OCH_3	Н	0.53 ± 0.03	2.4 ± 0.4	0.50 ± 0.03	5.7 ± 1.8	>30
7e	NO_2	Н	Н	CH_3	CH_3	Н	Н	0.12 ± 0.02	2.6 ± 0.7	0.059 ± 0.01	5.7 ± 1.4	>30
7 f	NO_2	Н	Н	CH_3	Су	Н	OCH_3	7.0 ± 0.8		6.2 ± 0.2		>30
7g	NO_2	Н	Н	CH_3	Bn	Н	OCH_3	3.0 ± 0.1		3.2 ± 0.2		>30
7h	NO_2	Н	Н	Bn	Bn	Н	OCH_3	19.8 ± 3.5		13.8 ± 1.5		>30
7i	Н	NO_2	Н	CH_3	CH_3	Н	Н	>30		>30		>30
7j	F	Η	Н	CH_3	CH_3	Н	Н	>30		>30		>30
7k	CN	Н	Н	CH_3	CH_3	Н	Н	7.0 ± 1.1		4.4 ± 0.2		>30
71	CN	Η	Н	CH_3	CH_3	Н	OCH_3	4.2 ± 0.4		2.5 ± 0.1		>30
7 m	CN	Η	Br	CH_3	CH_3	F	OCH_3	4.1 ± 0.1		3.2 ± 0.1		>30
7 n	CN	Н	morpholine	CH_3	CH_3	F	OCH_3	2.5 ± 0.3	2.5 ± 0.9	1.9 ± 0.1	4.0 ± 1.1	>30
7 o	CN	Н	6-CH ₃ -3- pyridyl	CH_3	CH_3	F	OCH ₃	0.24 ± 0.03	5.2 ± 1.8	0.16 ± 0.003	6.9 ± 1.3	23.3
ent-70 ^d	CN	Н	6-CH ₃ -3- pyridyl	CH_3	CH_3	F	OCH ₃	0.50 ± 0.27	6.1 ± 0.7	0.30 ± 0.06	5.2 ± 2.2	>30
ent-' 7 0 ^d	CN	Н	6-CH ₃ -3- pyridyl	CH_3	CH_3	F	OCH ₃	0.33 ± 0.03	2.7 ± 0.6	0.26 ± 0.03	5.9 ± 0.8	>30
7p		see sti	see structure at top of table (major isomer shown)					4.7 ± 0.3		4.5 ± 0.3		>30
7q		see str	ucture at top of	table (ma	ijor isome	er shown)		5.6 ± 0.8		3.8 ± 0.4		>30
	compd 7a 7b 7c 7d 7e 7f 7g 7h 7i 7j 7k 7l 7m 7n 70 ent-70 ^{e1} 7p 7q	compd R ¹ 7a NO2 7b NO2 7c NO2 7c NO2 7d NO2 7d NO2 7d NO2 7f H 7j F 7k CN 7n CN 7n CN 7n CN 7n CN 7p CN 7p Y 7p Y	compd \mathbb{R}^1 \mathbb{R}^2 7a NO2 H 7b NO2 H 7b NO2 H 7c NO2 H 7c NO2 H 7d NO2 H 7d NO2 H 7f NO3 H 7j F H 7k CN H 7n CN H 7n CN H 7o CN H 7n CN H 7od CN H 7p See str See str 7p See str See str	compd R^1 R^2 R^3 7a NO2 H H 7b NO2 H H 7b NO2 H H 7c NO2 H H 7d NO2 H H 7d NO2 H H 7d NO2 H H 7f NO2 H H 7g F H H 7i CN H H 7m CN H Br 7m CN H Br 7m CN H 6-CH_3-3- pyridyl ent-70 ^{ef} CN H 6-CH_3-3- pyridyl fp CN H 6-CH_3-3- pyridyl <td< td=""><td>compd \mathbb{R}^1 \mathbb{R}^2 \mathbb{R}^3 \mathbb{R}^4 7a NO2 H H CH3 7b NO2 H H CH3 7c NO2 H H CH3 7c NO2 H H CH3 7d NO2 H H CH3 7d NO2 H H CH3 7f NO3 H H CH3 7f CN H Br CH3 7m CN H 6-CH3-3- CH3<td>compd R¹ R² R³ R⁴ R⁵ 7a NO2 H H CH3 CH3 7b NO2 H H CH3 CH3 7b NO2 H H CH3 CH3 7c NO2 H H CH3 CH3 7d NO2 H H CH3 CH3 7d NO2 H H CH3 CH3 7e NO2 H H CH3 CH3 7f NO3 H H CH3 CH3 7t NO4 H H CH3 CH3 7t CN H</td><td>compdR¹R²R³R⁴R⁵R⁶7aNO2HHCH3CH3H7bNO2HHCH3CH3F7cNO2HHCH3CH3H7dNO2HHCH3CH3H7dNO2HHCH3CH3H7dNO2HHCH3CH3H7fNO2HHCH3CH3H7gNO2HHCH3SHH7gNO2HHCH3HH7gNO2HHCH3H7gFHHCH3H7hCNHHCH3H7hCNHBrCH3H7nCNHBrCH3H7nCNHBrCH3F7nCNHServerlageF7nCNHServerlageCH3F7nCNHServerlageCH3F7nCNHServerlageCH3F7nCNHServerlageCH3F7nCNHServerlageCH3F7nCNHServerlageCH3F7nCNHServerlageCH3F7nCNHServerlageServerlageSer</td><td>compdR¹R²R³R⁴R⁵R⁶R⁷7aNO2HHCH3CH3HOCH37bNO2HHCH3CH3FOCH37cNO2HHCH3CH3HF7dNO2HHCH3CH3HF7dNO2HHCH3CH3HH7eNO2HHCH3CH3HOCH37gNO2HHCH3CH3HOCH37gNO2HHCH3SnHOCH37gNO2HHCH3CH3HH7fFHHCH3CH3HH7iFHHCH3CH3HH7iCNHHCH3CH3HH7iCNHHCH3CH3HH7iCNHHCH3CH3HH7iCNHHCH3CH3HOCH37mCNHBrCH3CH3FOCH37mCNHBrCH3CH3FOCH37mCNHBrCH3CH3FOCH37mCNH6-CH3-3- pridylCH3CH3FOCH37prd⁴CNH6-CH3-3- pry</td><td>VEEV INCompdR¹R²R³R⁴R⁵R⁶R⁷$\hline CPE^{e^{2}} EC_{50}$ (µM)7aNO2HHCH3CH3HOCH30.041 ± 0.97bNO2HHCH3CH3FOCH30.048 ± 0.0047cNO2HHCH3CH3HF0.15 ± 0.017dNO2HHCH3CH3OCH3H0.53 ± 0.037eNO2HHCH3CH3HH0.12 ± 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0.1$7m<</td><td>VEV INF9813LEEV VIOScompdR¹R²R³R⁴R⁵R⁶R⁷CPE⁴ EC₅₀Viral yield redn^bCPE⁴ EC₅₀ (µM)Viral yield redn^b7aNO2HHCH3CH3FOCH30.041 ± 0.95.5 ± 1.20.033 ± 0.0027.9 ± 0.57bNO2HHCH3CH3FOCH30.048 ± 0.0043.7 ± 1.10.027 ± 0.0037.7 ± 0.87cNO2HHCH3CH3HF0.15 ± 0.012.9 ± 0.90.11 ± 0.0046.1 ± 1.87dNO2HHCH3CH3HF0.15 ± 0.012.9 ± 0.90.01 ± 0.0035.7 ± 1.47fNO2HHCH3CH3HH0.12 ± 0.022.6 ± 0.70.059 ± 0.015.7 ± 1.47fNO2HHCH3CYHOCH37.0 ± 0.86.2 ± 0.27gNO2HHCH3CH3HH2.30>307iHNO2HCH3CH3HH2.30>307iFHHCH3CH3HH2.30>307iFHHCH3CH3HH2.30>307iFHHCH3CH3HH2.30>307iFHHCH3CH3HOCH32.5 ± 0.32</td></td></td<>	compd \mathbb{R}^1 \mathbb{R}^2 \mathbb{R}^3 \mathbb{R}^4 7a NO2 H H CH3 7b NO2 H H CH3 7c NO2 H H CH3 7c NO2 H H CH3 7d NO2 H H CH3 7d NO2 H H CH3 7f NO3 H H CH3 7f CN H Br CH3 7m CN H 6-CH3-3- CH3 <td>compd R¹ R² R³ R⁴ R⁵ 7a NO2 H H CH3 CH3 7b NO2 H H CH3 CH3 7b NO2 H H CH3 CH3 7c NO2 H H CH3 CH3 7d NO2 H H CH3 CH3 7d NO2 H H CH3 CH3 7e NO2 H H CH3 CH3 7f NO3 H H CH3 CH3 7t NO4 H H CH3 CH3 7t CN H</td> <td>compdR¹R²R³R⁴R⁵R⁶7aNO2HHCH3CH3H7bNO2HHCH3CH3F7cNO2HHCH3CH3H7dNO2HHCH3CH3H7dNO2HHCH3CH3H7dNO2HHCH3CH3H7fNO2HHCH3CH3H7gNO2HHCH3SHH7gNO2HHCH3HH7gNO2HHCH3H7gFHHCH3H7hCNHHCH3H7hCNHBrCH3H7nCNHBrCH3H7nCNHBrCH3F7nCNHServerlageF7nCNHServerlageCH3F7nCNHServerlageCH3F7nCNHServerlageCH3F7nCNHServerlageCH3F7nCNHServerlageCH3F7nCNHServerlageCH3F7nCNHServerlageCH3F7nCNHServerlageServerlageSer</td> <td>compdR¹R²R³R⁴R⁵R⁶R⁷7aNO2HHCH3CH3HOCH37bNO2HHCH3CH3FOCH37cNO2HHCH3CH3HF7dNO2HHCH3CH3HF7dNO2HHCH3CH3HH7eNO2HHCH3CH3HOCH37gNO2HHCH3CH3HOCH37gNO2HHCH3SnHOCH37gNO2HHCH3CH3HH7fFHHCH3CH3HH7iFHHCH3CH3HH7iCNHHCH3CH3HH7iCNHHCH3CH3HH7iCNHHCH3CH3HH7iCNHHCH3CH3HOCH37mCNHBrCH3CH3FOCH37mCNHBrCH3CH3FOCH37mCNHBrCH3CH3FOCH37mCNH6-CH3-3- pridylCH3CH3FOCH37prd⁴CNH6-CH3-3- pry</td> <td>VEEV INCompdR¹R²R³R⁴R⁵R⁶R⁷$\hline CPE^{e^{2}} EC_{50}$ (µM)7aNO2HHCH3CH3HOCH30.041 ± 0.97bNO2HHCH3CH3FOCH30.048 ± 0.0047cNO2HHCH3CH3HF0.15 ± 0.017dNO2HHCH3CH3OCH3H0.53 ± 0.037eNO2HHCH3CH3HH0.12 ± 0.027fNO2HHCH3CH3HOCH33.0 ± 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7aNO2HHCH3CH3H7bNO2HHCH3CH3F7cNO2HHCH3CH3H7dNO2HHCH3CH3H7dNO2HHCH3CH3H7dNO2HHCH3CH3H7fNO2HHCH3CH3H7gNO2HHCH3SHH7gNO2HHCH3HH7gNO2HHCH3H7gFHHCH3H7hCNHHCH3H7hCNHBrCH3H7nCNHBrCH3H7nCNHBrCH3F7nCNHServerlageF7nCNHServerlageCH3F7nCNHServerlageCH3F7nCNHServerlageCH3F7nCNHServerlageCH3F7nCNHServerlageCH3F7nCNHServerlageCH3F7nCNHServerlageCH3F7nCNHServerlageServerlageSer	compdR ¹ R ² R ³ R ⁴ R ⁵ R ⁶ R ⁷ 7aNO2HHCH3CH3HOCH37bNO2HHCH3CH3FOCH37cNO2HHCH3CH3HF7dNO2HHCH3CH3HF7dNO2HHCH3CH3HH7eNO2HHCH3CH3HOCH37gNO2HHCH3CH3HOCH37gNO2HHCH3SnHOCH37gNO2HHCH3CH3HH7fFHHCH3CH3HH7iFHHCH3CH3HH7iCNHHCH3CH3HH7iCNHHCH3CH3HH7iCNHHCH3CH3HH7iCNHHCH3CH3HOCH37mCNHBrCH3CH3FOCH37mCNHBrCH3CH3FOCH37mCNHBrCH3CH3FOCH37mCNH6-CH3-3- pridylCH3CH3FOCH37prd ⁴ CNH6-CH3-3- pry	VEEV INCompdR ¹ R ² R ³ R ⁴ R ⁵ R ⁶ R ⁷ $\hline CPE^{e^{2}} EC_{50}$ (µM)7aNO2HHCH3CH3HOCH30.041 ± 0.97bNO2HHCH3CH3FOCH30.048 ± 0.0047cNO2HHCH3CH3HF0.15 ± 0.017dNO2HHCH3CH3OCH3H0.53 ± 0.037eNO2HHCH3CH3HH0.12 ± 0.027fNO2HHCH3CH3HOCH33.0 ± 0.17gNO2HHCH3SoHOCH33.0 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^{*a*}CPE assay, V/EEEV MOI = 0.05, Vero 76 cells, assessed at 48 h; data averaged from at least 3 replicates ($n \ge 3$, mean \pm SD). ^{*b*}Viral yield reduction (Vero 76 cells) at 5 μ M compound concentration calculated based on the averaged \log_{10} transformed viral titer of the mock-treated group divided by that of the compound-treated group at 18 h postinfection (n = 4, mean \pm SD). ^{*c*}Vero 76 cells, assessed 72 h post-treatment. ^{*d*}The (*R*)- or (*S*)-stereochemistry is unassigned, but each enantiomer was isolated in >99% ee by chiral HPLC.

mimic the tertiary amine of the cyclic amidine structures, a pharmacophoric element that improved potency for that series.

The scaffold was equipped with an aryl bromide at \mathbb{R}^3 for the purposes of coupling, and then, we explored the effect of installing a morpholine or pyridyl moiety off that position of the diazepinone core. While the morpholine modification did not appreciably improve potency, the augmentation was not deleterious, and for EEEV, it resulted in a promising 4-fold reduction of viral yield (entry 14).

The pyridyl analog **70** gratifyingly exhibited substantially improved CPE activity (EC₅₀ = 0.16–0.24 μ M) and robust reduction of viral replication, especially in the case of EEEV. Cytotoxicity was not observed for compounds tested up to 30 μ M with the exception of racemic **70** which showed some liability at 23 μ M. Nonetheless, the selectivity indices (CC₅₀/ EC₅₀ = 97–146) for **70** reflect a solid margin from which to optimize.

To determine if antiviral activity was influenced by enantiomeric identity, racemic **7o** was subjected to chiral separation, yielding *ent*-**7o** and *ent'*-**7o** in high enantiopurity (>99% ee). No significant difference was observed in the CPE assay between the enantiomers *ent*-**7o** and *ent'*-**7o** for VEEV or EEEV (entries 16 and 17, Table 1), and reduction in viral titer was notably different only for VEEV (entry 17). While the

antiviral activity of the compounds was evaluated in a Vero 76 cell line that is well established for these viruses, antiviral activity and sensitivity to the enantiomeric identity of 70 in primary human neuronal cells were also evaluated. As such, selected compounds, 7a, 7b, 7o, and the corresponding 7o enantiomers, ent- and ent'-70, were assayed for reduction of viral yield for both VEEV and EEEV in human brain primary neuronal cells (HNC001). Following treatment in neuronal cells, infectious viral titers were measured in a tissue culture infectious dose assay (TCID₅₀) using VERO 76 cells (Figure 1). At a compound concentration of 5 μ M, all of the compounds reduced viral titer to the limits of detection of the assay for both VEEV and EEEV compared to virus-only controls after 18 h (Figure 1A). Racemate 70 and its enantiomers were assessed in the same cell line at 1 μ M concentration to see if the antiviral activity could be better differentiated; however, we observed no significant difference in activity among these compounds even at the lower concentration (Figure 1B). Importantly, these results suggest that, barring a racemization event happening during the assay assessment, the stereocenter of 70 does not significantly influence the degree of antiviral activity observed for VEEV or EEEV. Moreover, we also show that substantial reduction in



Figure 1. Selected 70 compounds showing reduction of EEEV and VEEV titers in a TCID₅₀ assay using human brain primary neuronal cells at 5 μ M (panel A) or 1 μ M (panel B) after 18 h. Each experiment was done in triplicate, and averaged data is plotted (n = 3, mean \pm SD). The graphs were generated using GraphPad Prism V.9.3.1.

VEEV and EEEV titers is achieved in a primary human neuronal cell line even at a concentration of 1 μ M.

Selected compounds were also profiled for kinetic aqueous solubility and stability in mouse liver microsomes (MLMs) to gauge prospects for eventual *in vivo* evaluation and optimization (Table 2). The nitro group containing analogs

Table 2. Solubility and Mouse Microsomal Stability Data

entry	compd	solubility ^{<i>a</i>} (μ M)	$\mathrm{MLM}^{b} T_{1/2} \ (\mathrm{min})$
1	7a	0.7	10
2	7b	0.5	10
3	70	2.0	45
^a Kinotic so	lubility in ac	woods DBS buffer pH	1.74 ^b MIM - CD 1

"Kinetic solubility in aqueous PBS buffer, pH 7.4. "MLM = CDmouse liver microsomal stability.

7a,b showed limited aqueous solubility and a short MLM halflife (~10 min), while improved solubility and stability were observed for the nitrile-containing analog 7o. Mouse plasma stability for 7o was found to be robust ($T_{1/2} > 289$ min, CD-1 strain). Nonetheless, future optimization efforts will focus on refining solubility and microsomal stability as the series is advanced.

Given the observed SAR congruency between the older benzamidine series and the piperazinobenzodiazepinone scaffold, we wanted to better visualize shared or differentiated structural features in three-dimensional space, as these pharmacophoric details may guide future optimization. Benzodiazepinone **70**, though not the most potent analog from the survey, retained significant CPE assay potency and robustly attenuated viral replication for both VEEV and EEEV in Vero 76 and human neuronal cells while also bearing an aryl nitrile moiety in place of the potential toxophoric aryl nitro group. This was significant as the benzamidine series did not reflect a similar tolerance, and notably, the presence of the pyridyl appendage of **70** was required to achieve desirable potency. A potent benchmark benzamidine, BDGR-4 (Figure 2A), was independently compared *in silico* to each enantiomer



Figure 2. A. Structure of the benzamidine V/EEEV inhibitor, BDGR-4. B. Superimposed structures of BDGR-4 (gray structure) and (R)-70 (cyan structure) using a flexible align MOE program.

of benzodiazepinone 70 using the program Molecular Operating Environment (MOE).³⁰ Interestingly, alignment of benzamidine BDGR-4 (gray-colored carbon atoms) with either isomer of 70 (shown only for the (R)-isomer of 70 as cyan-colored carbon atoms, Figure 2B) showed exceptional overlap in which the amidine portion of BDGR-4 assumed a similar spatial arrangement as the piperazine of the benzodiazepinone structure. These results, in which the best scored alignments were redundant for each enantiomer of 70, corroborate the lack of differentiation in antiviral activity observed for *ent-*70 and *ent'-*70 in neuronal cells. Further, this model suggests that the required pyridyl appendage of 70 may engage the target with binding interactions that are differentiated from those that are possible for benzamidine BDGR-4.

In the course of exploring a quinazolinone rearrangement that has historically afforded benzamidines or derivatives thereof, we discovered that 2-dichloroquinazolin-4-ones, when treated with N,N'-dialkylethane-1,2-diamines, undergo ring expansion to form a new class of piperazine-fused benzodiazepinones. Furthermore, the analogs generated from this effort show remarkable cellular antiviral activity in two orthogonal antiviral assays against encephalitic alphaviruses, VEEV and EEEV, and without significant cytotoxic liability. The transformation led to modest yields of analogs with nitro-group substituted cores but significantly improved to a suitable range of 49-66% yield when nitrile-group variants were generated. This yield enhancement was also notable given that submicromolar antiviral CPE inhibition and good viral yield reduction were achieved with the nitrile analogs, a milestone not met with the analogously substituted, nitrile-containing benzamidines. Additionally, we observed that several piperazinobenzodiazepinones were effective in significantly reducing both VEEV and EEEV titers in human primary neuronal cells. Preliminary solubility and mouse microsomal stability were determined to be modest, though nitrile compound 70 was comparatively better than two nitro-containing analogs. Last, we analyzed in silico the energy-minimized structures of a leading benzamidine, BDGR-4, and an optimized piperazinobenzodiazepinone 70. This exercise revealed relatively congruent, low-energy conformations that either enantiomer of 70 can adopt a three-dimensional structural arrangement like that of benzamidine BDGR-4 and that the pyridyl appendage of this new benzodiazepinone class may offer an advantage in exploiting new target interactions. Work is ongoing to explore this aspect of the transformation. Ultimately, the discovery and development of these piperazinobenzodiazepinones represent another important milestone in expanding a sparsely stocked pipeline of small molecules that inhibit encephalitic alphaviruses.

ASSOCIATED CONTENT

Supporting Information

The Supporting Information is available free of charge at https://pubs.acs.org/doi/10.1021/acsmedchemlett.1c00539.

Assay and experimental details, synthetic procedures, characterization, chiral separation of 70, including NMR spectra for all new compounds (11a-j, 14a-j, and 7a-q) reported in this work (PDF)

AUTHOR INFORMATION

Corresponding Author

Jennifer E. Golden – Pharmaceutical Sciences Division, School of Pharmacy, University of Wisconsin–Madison, Madison, Wisconsin 53705, United States; Department of Chemistry, University of Wisconsin–Madison, Madison, Wisconsin 53706, United States; orcid.org/0000-0002-6813-3710; Email: jennifer.golden@wisc.edu

Authors

Michael C. Ryan – Pharmaceutical Sciences Division, School of Pharmacy, University of Wisconsin–Madison, Madison, Wisconsin 53705, United States

Eunjung Kim – Department of Microbiology and Immunology, University of Louisville, Louisville, Kentucky 40202, United States

Xufeng Cao – Pharmaceutical Sciences Division, School of Pharmacy, University of Wisconsin–Madison, Madison, Wisconsin 53705, United States

Walter Reichard – Department of Microbiology, Immunology, and Biochemistry, University of Tennessee Health Science Center, Memphis, Tennessee 38163, United States

Tyler J. Ogorek – Department of Chemistry, University of Wisconsin–Madison, Madison, Wisconsin 53706, United States

Pronay Das – Pharmaceutical Sciences Division, School of Pharmacy, University of Wisconsin–Madison, Madison, Wisconsin 53705, United States

Colleen B. Jonsson – Department of Microbiology, Immunology, and Biochemistry, University of Tennessee Health Science Center, Memphis, Tennessee 38163, United States; College of Pharmacy, University of Tennessee Health Science Center, Memphis, Tennessee 38163, United States; orcid.org/0000-0002-2640-7672

Jerome Baudry – Department of Biological Sciences, University of Alabama in Huntsville, Huntsville, Alabama 35899, United States

Donghoon Chung – Department of Microbiology and Immunology, University of Louisville, Louisville, Kentucky 40202, United States; o orcid.org/0000-0003-2626-172X

Complete contact information is available at: https://pubs.acs.org/10.1021/acsmedchemlett.1c00539

Notes

The authors declare no competing financial interest.

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ABBREVIATIONS

BRSM, based on recovered starting material; CPE, cytopathic effect; EEEV, eastern equine encephalitis virus; PMP, *p*-methoxyphenyl; SAR, structure–activity relationships; VEEV, Venezuelan equine encephalitis virus

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