# $\Box$ CASE REPORT $\Box$

# Transient Worsening of Photosensitivity due to Cholelithiasis in a Variegate Porphyria Patient

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## Abstract

Variegate porphyria (VP) is an autosomal dominant disease caused by mutations of the protoporphyrinogen oxidase (*PPOX*) gene. This porphyria has unique characteristics which can induce acute neurovisceral attacks and cutaneous lesions that may occur separately or together. We herin report a 58-years-old VP patient complicated with cholelithiasis. A sequencing analysis indicated a novel c.40G>C mutation (p.G14R) in the *PPOX* gene. His cutaneous photosensitivity had been worsening for 3 years before the emergence of chole-cystitis and it then gradually improved after cholecystectomy and ursodeoxycholic acid treatment with a slight decline in the porphyrin levels in his blood, urine and stool. In VP patients, a worsening of photosensitivity can thus be induced due to complications associated with some other disease, thereby affecting their porphyrin-heme biosynthesis.

Key words: variegate porphyria, photosensitivity, cholelithiasis, PPOX gene, protoporphyrin

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# Introduction

Variegate porphyria (VP; OMIM 176200) is an autosomal-dominant disease caused by mutations of the protoporphyrinogen oxidase (PPOX; E.C.1.3.3.4) gene. PPOX catalyses oxidation of protoporphyrinogen to protoporphyrin in the heme biosynthetic pathway, and the PPOX activity is reduced to approximately half of its normal level in heterozygous patients (1). VP has the unique, characteristic presentation of a combination of acute neurovisceral attacks and cutaneous symptoms that can occur either separately or together. The cutaneous lesions have increased photosensitivity that results in skin fragility and a tendency for blistering, and this condition typically leads to milia, scarring, thickening, and pigmentation in the chronic phase (2). Based on clinical experience, it is supposed that the presence of cutaneous lesions occurs in adulthood and it is rare before puberty. Recent genetic evidence has suggested the prevalence of cutaneous symptoms in VP patients to be approximately 30-40% (3). There appears to be no correlation between the mutation type and clinical manifestations (4). Fraunberg et al. showed that the occurrence of skin symptoms was related to a more than fourfold increase in urinary coproporphyrin excretion, but normal faecal protoporphyrin excretion predicted freedom from skin symptoms (5). However, the clinical factors that determine skin symptoms in VP are not well known.

We herein report a patient who showed chronic blistering due to photosensitivity along with the complication of cholelithiasis. We identified a novel c.40G>C mutation in the *PPOX* gene in this patient. His skin condition improved after cholecystectomy and ursodeoxycholic acid (UDCA) treatment, so we assumed that the emergence of cutaneous symptoms may thus have been induced by cholelithiasis in addition to an abnormal metabolism of porphyrin.

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Figure 1. Chronic cutaneous symptoms before (a) and after (b) cholecystectomy. (a) Blisters, erosions, crusts and milia on the back of the hands are observed. (b) The cutaneous symptoms improved except for pigmentation at 3 years after cholecystectomy.

 Table 2.
 Porphyrin Concentrations before and 3 Years after Cholecystectomy.

porphyrins	before	after	reference
Urine (µg/g creatinine)			
Uroporphyrin	29	18	<36
Coproporphyrin	367	128	<170
Blood (µg/dL RBC)			
Coproporphyrin	<1	<1	<1
Protoporphyrin	90	75	30-86
Faeces (µg/24h)			
Uroporphyrin	<1	<1	<170
Coproporphyrin I	799	497	<500
Coproporphyrin III	6,550	4,575	<400
Protoporphyrin	12,087	11,204	<1,500
Copro III/Copro I	8.19	9.21	<1.20

#### **Case Report**

A 58-year-old male was referred to our hospital for surgical treatment of cholecystolithiasis. He had been diagnosed as having acute porphyria because of an acute attack after the administration of an antifungal agent 7 years previously (though the diagnosis was not made at that time). Until recently, there had been no cutaneous symptoms except for two episodes of the transient formation of blisters over his entire body during sunbathing when he was in his thirties.

He presented with a 3-year history of chronic blistering

Table 1. Laboratory Data.

WBC	5,090	/µL	ALP	258	U/L
Neut	2.890	/μL	γ-GTP	45	U/L
RBC	$464 \times 10^{4}$	/µL	BUN	11	mg/dL
Hb	14.0	g/dL	Cr	0.79	mg/dL
Ht	41.9	%	Sodium	142	mmol/L
Plt	13.9×10 <sup>4</sup>	/µL	Potassium	4.1	mmol/L
ТР	7.4	g/dL	Chloride	105	mmol/L
Alb	4.0	g/dL	Ferrum	79	µg/dL
T.Bil	0.7	mg/dL	UIBC	209	µg/dL
AST	18	U/L	TIBC	288	µg/dL
ALT	22	U/L	Ferritin	68.1	ng/mL
LDH	157	U/L	CRP	0.1	mg/dL

WBC: white blood cell, Neut: neutrophil, RBC: red blood cell, Ht: hematocrit, Plt: platelet, TP: total protein, Alb: albumin, T.Bil: total bilirubin, AST: asparate aminotransferase, ALT: alanine aminotransferase, LDH: lactate dehydrogenase,  $\gamma$ -GTP:  $\gamma$ -glutamyl transferase, BUN: blood ureanitrogen, Cr: creatinine, TIBC: total iron binding capacity, UIBC: unsaturated iron binding capacity, CRP: C-reactive protein

on the backs of his hands due to photosensitivity, and he had developed skin fragility, blisters and scarring (Fig. 1a). Facial hyperpigmentation was also present. Liver function and inflammatory reaction tests did not show abnormal findings (Table 1). The serum levels of iron and ferritin were also within the normal limits. A computed tomography (CT) scan of the abdomen demonstrated 30 mm and 4 mm calcified gallstones in the neck of the gallbladder and distal bile duct, respectively. Despite three episodes of acute cholecystitis in the previous 6 months, no elevations in urinary levels of  $\delta$ -Aminolevulinic acid (ALA) or porphobilinogen (PBG) had been observed. A stool examination suggested the excessive excretion of protoporphyrin consistent with a biochemical diagnosis of VP (Table 2). Sequencing analyses using polymerase chain reaction primers (4), identified a heterogeneous novel c.40G>C mutation in exon 2 of the PPOX gene (Fig. 2a).

Owing to the characteristics of the acute attack and photosensitivity, cholecystectomy was done avoiding use of porphyrinogenic agents for anaesthesia and protection the patient from phototoxic injury due to surgical luminaires. Lights in the operating room were covered with orange filters (Lumicool 1905; Yamahira, Saitama, Japan), and a yellow filter (Dichroic Filter-Y; Koshin, Kanagawa, Japan) was used for the headlight, and emitted light at wavelengths < 500 nm and 460 nm, respectively (6). There was no clinical evidence of any exacerbation of the patient's illness throughout the perioperative period.

Postoperatively, he was given UDCA. The urinary excretion of uroporphyrin and coproporphyrin and protoporphyrin in the erythrocytes fell to normal limits after 3 years (Table 2). In the patient's faeces, porphyrin excretion also decreased slightly. Chronic blistering on the backs of his hands due to photosensitivity had also completely improved at this time (Fig. 1b). Skin pigmentation remained but there was no fragility, blistering or pronounced scarring.



Figure 2. Analyses of the p.G14R mutation of the PPOX gene. (a) Missense mutation of a heterozygous G→C transition in exon 2 of the PPOX gene. (b) Sequence alignment of the PPOX enzyme from *Homo sapiens* (HS), mouse (MM), *Rattus norvegicus* (RN), *Xenopus laevis* (XL), *Drosophila melanogaster* (DM) and *Nicotiana tabacum* (NT). Residue numbers and secondary structures are based on human PPOX. Conserved residues are highlighted in red. The arrow indicates the amino-acid change found in our patient. (c) Predicted three-dimensional (3D) structures of the wild-type (blue) and p. G14R mutated (green) type of PPOX at the N-terminal  $\beta\alpha\beta$  domain and their superimposition. Predicted amino-acid sequences were modelled to the 3D structure using I-TASSER software (17), and figure drawings were carried out using Swiss-PdbViewer (18). The region of amino acid 14 is shown in red. Hydrogen bonds are shown as yellow dashed lines. In the merge image, side chains of amino acids are represented by the same colors with their structures. This model shows that a p.G14R mutation would loosen an  $\alpha$ 1-helix and warp  $\beta$ 1-sheet, which thus causes an extension of the loop between them (black dashed line). The side chain of mutational amino acid arginine is sticking out over  $\alpha$ 1 helix of the PPOX (arrow), while the side chain of wild type amino acid glycine has no projection.

### Discussion

In VP patients, excess porphyrins accumulate in the skin and dermal vessels. They produce reactive singlet oxygen molecules when they are activated by excessive exposure to sunlight, resulting in skin fragility and a tendency to develop blister formation (7). However, not all VP patients show these cutaneous symptoms. The number of skin lesions in VP deteriorates with sunlight exposure, but it is not known which factors determine the emergence of chronic cutaneous symptoms. Our patient experienced two episodes of transient, acute photosensitive dermatitis after excessive exposure to sunlight while in his thirties, but did not present with chronic cutaneous lesions until 3 years previously. Since a 30 mm calcified gallstone was observed in the cystic duct at the first visit, he would have been complicated with cholelithiasis for several years. Therefore, it is supposed that his cutaneous symptoms became apparent with the progression of his gallstone. Moreover, His cutaneous lesions improved with no specific therapy except for cholecystectomy and UDCA treatment, so we hypothesised that cholelithiasis induced these symptoms.

The prevalence of cholelithiasis has been reported to increase in VP patients, and some patients exhibit chronic skin symptoms before cholelithiasis becomes apparent (8). It has been suggested that a disturbance of porphyrin excretion to bile due to cholestasis might cause increases in porphyrin accumulation. However, there was no sign of cholestasis or liver dysfunction in laboratory data of this patient (Table 1). Several VP cases presented with cutaneous symptoms before the definitive complications of another disease became apparent (8-14). Most of these cases had liver-related disease, but some did not (Table 3). Therefore, the exacerbation of

Pat. no/ Sex	Age at diag- nosis (years)	Duration of CS (years)	Complications	Mutation in the PPOX gene	Reference
1/F	70	1	HCC	Unknown	9
2/F	79	1	HCC	Unknown	10
3/F	82	0.5	HCC	1,082-1,083 ins C	11
4/F	26	2	Coeliac disease	Unknown	12
5/M	55	2.5	metastatic colon cancer	1,082-1,083 ins C	13
6/F	44	0.5	Cervical cancer	557-558 del GT	14
7/F	59	9	Cholelithiasis	Unknown	8
8/M	28	11	Cholelithiasis	Unknown	8
9/M	58	3	Cholelithiasis	c.40G>C	Present case

 Table 3.
 Overview of Cutaneous Symptom of Variegate Porphyria Patients Associated with Chronic Disease.

CS: cutaneous symptom, PPOX: protoporphyrinogen oxidase, HCC: hepatocellular carcinoma

skin lesions is due not only to liver dysfunction, but is also dependent upon systemic stress, which may alter heme biosynthesis. Furthermore, in patients with homozygous VP, severe cutaneous symptoms were reported from childhood even though they had experienced no acute attacks (15), suggesting that a severe deficiency of protoporphyrinogen oxidase (PPOX) can cause chronic cutaneous lesions rather than acute attacks. In most cases, increases in the levels of protoporphyrins in erythrocytes are observed, but not for patients with heterozygous cases. Therefore, increased levels of protoporphyrin in the blood may predict the emergence of chronic cutaneous symptoms in VP patients. During chronic stress (as seen in our patient), the suppression of PPOX activity and increases in protoporphyrinogen levels (including other porphyrin metabolites) may be induced. Then, autooxidation of water-soluble protoporphyrinogen to insoluble protoporphyrin may occur in erythrocytes and peripheral tissues, resulting in their accumulation in the skin and thus leading to the onset of cutaneous symptoms.

In our case, we identified a c.40G>C mutation in exon 2 of the PPOX gene, which resulted in a substitution of a nonpolar glycine by a polar arginine (p.G14R). This glycine residue is evolutionarily highly conserved in humans, mice, Rattus norvegicus, Xenopus laevis, Drosophila melanogaster and Nicotiana tabacum, attesting to its importance (Fig. 2b), and lies in the flavin adenine dinucleotide (FAD)-binding domain in the amino-terminal  $\alpha 1$  helix of the PPOX (16). Modelling of the p.G14R mutation reveals conformational changes at the canonical FAD binding site (Fig. 2c). Consequently, this mutant seems to have a reduced activity. Moreover, 28 amino acids in the amino terminus of PPOX contain a functioning signal for mitochondrial targeting (19). A positively charged and hydrophilic arginine substitution disrupts the hydrophobic face in this lesion, and this affects the interaction with the mitochondrial outer membrane receptor Tom20 (translocase of outer mitochondrial membrane 20). Therefore, a p.G14R mutation may disrupt the correct transport of PPOX into mitochondria.

The use of protective light filters during surgical procedures for VP patients is not commonly recommended. However, phototoxic injury due to operating-room lights in a Japanese VP patient was reported recently (14). That patient presented with chronic blistering due to photosensitivity and increased levels of protoporphyrins in erythrocytes with a complication of cervical cancer. Thus, in our patients, we could not exclude the possibility of phototoxic injury due to surgical luminaires, and therefore used filters to avoid that risk. Photosensitivity may be dependent upon the extent of accumulation of cutaneous porphyrins, light quality or duration of irradiation. After cholecystectomy and the dosage of UDCA, the porphyrin levels in his blood, urine and stool decreased and cutaneous photosensitivity improved. Therefore, the onset of photosensitivity should be considered to be a potential and transient symptom in some VP patients who have complications.

#### The authors state that they have no Conflict of Interest (COI).

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#### References

- 1. Puy H, Gouya L, Deybach JC. Porphyrias. Lancet 375: 924-937, 2010.
- Timonen K, Niemi KM, Mustajoki P, Tenhunen R. Skin changes in variegate porphyria. Clinical, histopathological, and ultrastructural study. Arch Dermatol Res 282: 108-114, 1990.
- Hift RJ, Meissner D, Meissner PN. A systematic study of the clinical and biochemical expression of variegate porphyria in a large South African family. Br J Dermatol 151: 465-471, 2004.
- **4.** Whatley SD, Puy H, Morgan RR, et al. Variegate porphyria in Western Europe: identification of PPOX gene mutations in 104 families, extent of allelic heterogeneity, and absence of correlation between phenotype and type of mutation. Am J Hum Genet **65**: 984-994, 1999.
- **5.** von und zu Fraunberg M, Timonen K, Mustajoki P, Kauppinen R. Clinical and biochemical characteristics and genotype-phenotype correlation in Finnish variegate porphyria patients. Eur J Hum Genet **10**: 649-657, 2002.
- Takahashi R, Hirai I, Murayama S, et al. Experience of cholecystectomy in a porphyria patient with photosensitivity. Tando 27: 817-821, 2013 (in Japanese, Abstract in English).
- 7. Poh-Fitzpatrick MB. Molecular and cellular mechanisms of porphyrin photosensitization. Photodermatol 3: 148-157, 1986.
- 8. Herrick AL, Moore MR, Thompson GG, Ford GP, McColl KE.

Cholelithiasis in patients with variegate porphyria. J Hepatol 12: 50-53, 1991.

- Tidman MJ, Higgins EM, Elder GH, MacDonald DM. Variegate porphyria associated with hepatocellular carcinoma. Br J Dermatol 121: 503-505, 1989.
- **10.** Grabczynska SA, McGregor JM, Hawk JL. Late onset variegate porphyria. Clin Exp Dermatol **21**: 353-356, 1996.
- Schneider-Yin X, van Tuyll van Serooskerken AM, Went P, et al. Hepatocellular carcinoma in variegate porphyria: a serious complication. Acta Derm Venereol 90: 512-515, 2010.
- 12. Dal Sacco D, Parodi A, Cozzani E, Biolcati G, Griso D, Rebora A. A case of variegate porphyria with coeliac disease and beta-thalassaemia minor. Dermatology 209: 161-162, 2004.
- Hanneken S, Kuerten V, Hoernke M, Neumann NJ. Metastatic colon cancer triggering an acute attack of variegate porphyria. Int J Colorectal Dis 24: 127-128, 2009.
- Masuoka E, Bito T, Oka M, Nakano H, Nishigori C. A case of variegae porphyria diagnosed by genetic analysis. Hifurinshou 53: 277-282, 2011 (in Japanese).

- 15. Palmer RA, Elder GH, Barrett DF, Keohane SG. Homozygous variegate porphyria: a compound heterozygote with novel mutations in the protoporphyrinogen oxidase gene. Br J Dermatol 144: 866-869, 2001.
- 16. Qin X, Tan Y, Wang L, et al. Structural insight into human variegate porphyria disease. FASEB J 25: 653-664, 2011.
- **17.** Zhang Y. I-TASSER server for protein 3D structure prediction. BMC Bioinformatics 23 **9**: 40, 2008.
- Guex N, Peitsch MC. SWISS-MODEL and the Swiss-PdbViewer: An environment for comparative protein modeling. Electrophoresis 18: 2714-2723, 1997.
- von und zu Fraunberg M, Nyröen T, Kauppinen R. Mitochondrial targeting of normal and mutant protoporphyrinogen oxidase. J Biol Chem 278: 13376-13381, 2003.

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