

Letter to the Editor



Novel Sensitive, Two-site ELISA for the Quantification of Der f 1 Using Monoclonal Antibodies

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To the Editor,

The efficacy of allergen immunotherapy depends on the quality of allergen extracts. Currently, biological methods based on intradermal skin tests or skin prick tests are utilized for the preparation and standardization of reference allergen extracts. A competitive inhibition assay is normally used for *in vitro* comparison of allergenic potency and standardization of new preparations. However, quantification of major allergen content is needed for better allergen standardization. The concentration of major allergen in an extract is proportional to its allergenic activity, and maintenance doses (5 to 20 μ g/dose, 50 to 250 μ g/yr) of the major allergen are necessary for effective subcutaneous immunotherapy. The Japanese Society of Allergology determined allergenic potency based on Der p 1 and Der f 1 content, concluding that 38.5 μ g/mL of Der 1 is equivalent to 100,000 JAU/mL.

A monoclonal antibody (mAb)-based sandwich enzyme-linked immunosorbent assay (ELISA) for the quantification of Der f1 and Der p1 was first developed in 1989⁵ and is currently in wide use. More recently, tandem mass spectrometry has begun to be utilized for allergen quantification.⁶ However, its use is limited due to the need for expensive instruments and highly trained staffs for routine use.

The study presented here was undertaken to develop a new sensitive, two-site ELISA for quantification of Der f1. Hybridomas producing mAbs were prepared by immunization with recombinant (r) Der f1, which was produced in *Pichia* as previously described. First, 17 clones that strongly recognized rDer f1 were selected by ELISA. Five clones (1C9, 1G8, 3D7, 13C5, and 13F7) that showed good reactivity both in ELISA and western blotting were selected for further cloning (**Figure A and B**). However, western blotting showed that clones 1C9 and 13F7 did not retain the ability to recognize Der f1. The combination of 3D7 as a capture antibody and biotinylated 13F7 as a detection antibody showed optimal results for the detection of Der f1 in the extract. The detection limits were as low as 50 ng/mL of extract and 2 pg/mL of rDer f1 (**Figure C and D**).

Der f 1 content in the 4 commercial extracts was 3.47– $19.16 \,\mu g/mL$ when using the Indoor Biotechnologies antibodies and 2.79– $18.59 \,\mu g/mL$ with the investigatory antibodies (**Supplementary Fig. S1**). No significant difference was shown between the results obtained by the 2 kits (P = 0.1 to 0.7). Der f 1 concentrations in the 21 dust samples were 18– $363.2 \, ng/g$ dust with Indoor Biotechnologies antibodies and 14– $322.5 \, ng/g$ dust with the investigatory

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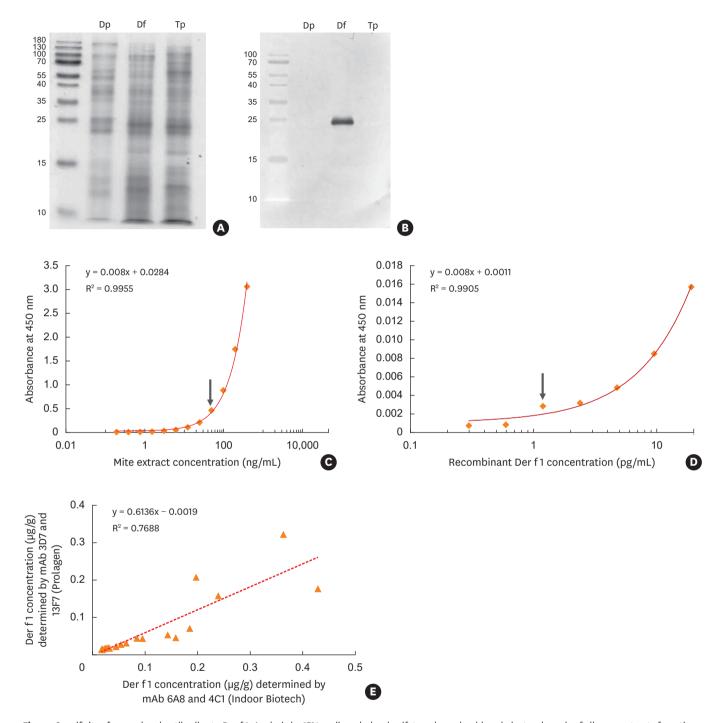


Figure. Specificity of monoclonal antibodies to Der f 1. Analysis by 15% sodium dodecyl sulfate polyacrylamide gel electrophoresis of allergen extracts from three species of dust mite: Dermatophagoides pteronyssinus (Dp), D. farinae (Df), and Tyrophagus putrescentiae (Tp) (A). Detection of native Der f 1 by monoclonal antibodies 3D7 (B). Detection of native Der f 1 in allergen extract by a two-site ELISA system. The two-site ELISA was quantified with doubling dilutions of the extract from 390 ng/mL of the extract (C). The two-site ELISA was quantified with doubling dilutions from 20 pg/mL of recombinant Der f 1 (D). Arrows indicate the detection limits. Comparison of Der f 1 concentration in house dust samples quantified by two ELISA systems (E). mAb, monoclonal antibody; ELISA, enzyme-linked immunosorbent assay.

antibodies (**Figure E**). The Pearson's correlation coefficient between the 2 assays was 0.876824 (P < 0.0001). As expected, Der f1 content in the commercial extracts and dust samples was not significantly different between the two-site ELISA systems.



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Disclosure

JD Kim, KY Jeong, KH Park, JH Lee, and JW Park share equivalents of Prolagen, Ltd. JW Park reports serving as an unpaid chief technology officer for Prolagen. KY Jeong is a Technical Advisor of Prolagen. JT Kim and JD Kim are co-chief executive officers. HH Kim, SH Kim, DJ Kim, and YJ Shin are employees of Prolagen. The interests of these authors did not influence academic fairness in conducting this study, analyzing results, and writing a paper. Other authors have no potential conflicts of interest to disclose.

In conclusion, we developed novel mAbs against rDer f1 useful for both qualitative and quantitative analyses. The antibodies should also be useful for the assessment of allergen exposure.

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SUPPLEMENTARY MATERIAL

Supplementary Fig. S1

Comparison of Der f1 content in commercial *Dermatophagoides farinae* extracts quantified by two enzyme-linked immunosorbent assay systems.

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