

Original Article



OPEN ACCESS

Received: Sep 14, 2017

Revised: Nov 20, 2017

Accepted: Nov 21, 2017

*Correspondence to

Yong Woo Jung

Department of Pharmacy, Korea University,
2511 Sejong-ro, Sejong 30019, Korea.
E-mail: yjung@korea.ac.kr

Soo An Choi

Department of Pharmacy, Korea University,
2511 Sejong-ro, Sejong 30019, Korea.
E-mail: sachoi@korea.ac.kr

[†]Hyun Gyung Kim and Sang Hoon Kim
contributed equally to this work.

Copyright © 2017. The Korean Association of
Immunologists

This is an Open Access article distributed
under the terms of the Creative Commons
Attribution Non-Commercial License ([https://
creativecommons.org/licenses/by-nc/4.0/](https://creativecommons.org/licenses/by-nc/4.0/))
which permits unrestricted non-commercial
use, distribution, and reproduction in any
medium, provided the original work is properly
cited.

Conflict of Interest

The authors declared no potential conflicts of
interest.

Abbreviations

CNT, carbon nanotube; DMEM, Dulbecco's
modified Eagle's high glucose medium; LPS,
lipopolysaccharide; MWCNT, multi-walled
carbon nanotube; NO, nitric oxide; PP,
polypropylene; PVDF, polyvinylidene fluoride;
SWCNT, single-walled carbon nanotube

Polyvinylidene Fluoride Alters Inflammatory Responses by Activation-induced Cell Death in Macrophages

Hyun Gyung Kim^{1,†}, Sang Hoon Kim^{1,†}, Taek-Seung Kim², Tae Won Park³, Ran Won⁴,
Hee-Deung Park², Soo An Choi^{1,*}, Yong Woo Jung^{1,*}

¹Department of Pharmacy, Korea University, Sejong 30019, Korea

²School of Civil, Environmental and Architectural Engineering, Korea University, Seoul 02841, Korea

³Department of Psychiatry, Chonbuk National University Medical School, Jeonju 54907, Korea

⁴Department of Biomedical Laboratory Science, Division of Health Sciences, Dongseo University, Busan
47011, Korea

ABSTRACT

Carbon nanotubes (CNTs) are nanomaterials that have been employed in generating diverse materials. We previously reported that CNTs induce cell death in macrophages, possibly via asbestosis. Therefore, we generated CNT-attached polyvinylidene fluoride (PVDF), which is an established polymer in membrane technology, and then examined whether CNT-attached PVDF is immunologically safe for medical purposes compared to CNT alone. To test this, we treated RAW 264.7 murine macrophages (RAW cells) with CNT-attached PVDF and analyzed the production of nitric oxide (NO), a potent proinflammatory mediator, in these cells. RAW cells treated with CNT-attached PVDF showed reduced NO production in response to lipopolysaccharide. However, the same treatment also decreased the cell number suggesting that this treatment can alter the homeostasis of RAW cells. Although cell cycle of RAW cells was increased by PVDF treatment with or without CNTs, apoptosis was enhanced in these cells. Taken together, these results indicate that PVDF with or without CNTs modulates inflammatory responses possibly due to activation-induced cell death in macrophages.

Keywords: Carbon nanotubes; Polyvinylidene fluoride; Macrophages; Nitric oxide; Cell death; Inflammation

INTRODUCTION

In the last 2 decades, membrane technology has been greatly improved and is widely used for multiple applications such as water treatment, removal of toxic materials, protein filtration, attachment of molecules, and so on. For these applications, attempts have been made to produce various materials in the membrane form. One of these materials, polyvinylidene fluoride (PVDF), has received intense consideration because of its exceptional properties including mechanical durability, thermal stability, chemical endurance, and so on (1-4). Therefore, PVDF membranes have been intensively studied in chemical and biological filtration for separation purposes. Although the potential of PVDF in the filtration industry is unavoidably high, its effect on the immune system is incompletely studied.

Author Contributions

Conceptualization: Kim HG, Kim SH, Choi SA, Jung YW; Data curation: Kim HG, Kim SH; Formal analysis: Kim HG, Kim SH; Funding acquisition: Jung YW; Resources: Kim TS, Park TW, Won R, Park HD; Supervision: Jung YW; Writing - original draft: Kim HG, Kim SH, Choi SA, Jung YW.

The immune system comprises 2 arms: innate and adaptive immunity (5). Innate immunity acts as the first line of defense, whereas adaptive immunity works specifically to fight infections (6). These 2 arms usually cooperate to mount proper immune responses against pathogenic molecules or organisms, even though they use different pathways (7). We focused on innate immune cells because these are the first cells to identify foreign materials and decide whether an immune response needs to be initiated (8). Therefore, stimulation of innate immune cells may induce improper responses to infectious diseases.

Macrophages have been studied over the last few decades for their roles in response to pathogens (9). These cells can produce proinflammatory molecules such as nitric oxide (NO) and proinflammatory cytokines when stimulated by bacterial endotoxins such as lipopolysaccharide (LPS), CpG, or others (8,10). Production of these molecules can initiate innate immune responses to possible harmful materials. Thus, macrophage activation may determine whether or not a particular material can induce inflammatory responses (11).

Using this system, we previously reported that single-walled carbon nanotubes (SWCNTs) induce cell death and tumor necrosis factor α production in macrophages without affecting NO production (12). Since carbon nanotubes (CNTs) are free floating materials, we hypothesized that CNTs-attached PVDF do not affect macrophage function. Here, we examined if PVDF and CNTs could modulate the immune response mounted by LPS-stimulated macrophages. We found that PVDF membrane with or without CNTs induced apoptosis in macrophages. These materials also inhibited NO production by LPS-activated macrophages, possibly due to cell death. In addition, macrophages stimulated by PVDF showed increased number of cells in the G2 and S phase of the cell cycle. These data suggest that PVDF can modulate inflammatory responses.

MATERIALS AND METHODS

Cells

The murine macrophage cell line RAW 264.7 (RAW), was cultured in DMEM containing 10% Hyclone™ fetal bovine serum (GE Healthcare Life Sciences, Logan, UT, USA) and 1% Hyclone™ penicillin/streptomycin (GE Healthcare Life Sciences) at 37°C under 5% CO₂.

Reagents

LPS from *Escherichia coli* 0111:B4 was purchased from Sigma-Aldrich Korea (Yongin, Korea). Two types of CNT filters were fabricated using multi-walled carbon nanotube (MWCNT) and SWCNT inks (Applied Carbon Nano Technology, Pohang, Korea). In the inks, CNTs were suspended in a water-based solvent containing dispersants. The average width and length of the CNTs was 18 nm and 0.1 μ m, respectively. Before preparing the CNT filters, the MWCNT (1.0 wt.%) ink was diluted 10-fold in deionized (DI) water, whereas the SWCNT (0.1 wt.%) ink was not diluted. The diluted MWCNT ink (10 ml) or SWCNT ink (10 ml) was passed through a 47 mm PVDF filter (0.1 μ m pore size; Merck Millipore, Billerica, MA, USA) using a vacuum filtration unit. The residual dispersants in these prepared CNT filters were removed by passing 200 ml of DI water through the filters using the same filtration unit. The fabricated CNT filters were kept in a desiccator before the toxicity test.

NO assay

The Griess reagent measures nitrite (NO₂⁻) concentration, which is used as a standard of inflammation. Griess reagent was prepared with 1% sulfanilamide, 0.1%

naphthylethylenediamine dihydrochloride, and 2% phosphoric acid, which were dissolved in water and the bottle was wrapped the bottle in aluminum foil until use. Following the pretreatment of cells for 18 to 24 h with PVDF, SWCNT, or MWCNT, and treatment with LPS at 3 h after cell seeding, an equal volume of supernatant and Griess reagent was mixed in a 96-well plate as described previously (13). Absorbance was measured using a spectrophotometer at 540 nm. DMEM treated cells was used as the negative control. The nitrite concentration was determined based on a standard curve generated using sodium hydroxide nitrite.

MTT assay

RAW cells grown in a 96-well plate (1×10^6 cells/ml) were treated with PVDF, SWCNT, MWCNT, or LPS (1 ng/ml) for 18 to 24 h. Cells treated with DMEM were used as the negative control. Cell viability was measured using the CellTiter 96[®] Non-Radioactive Cell Proliferation Assay (Promega Corp., Madison, WI, USA) according to the manufacturer's instructions. Cell death greater than 20% was considered significant.

Cell cycle analysis

Cells were stimulated with LPS and each membrane with SWCNT, or MWCNT for 24 h. The incubated cells were washed with PBS and fixed in ice-cold 70% ethanol at -20°C for 1 h. Cells were stained with Krishan buffer at 37°C for 30 min after washing in PBS (14). The stained cells were examined by flow cytometry and analyzed using FlowJo software (Tree Star Inc., San Carlos, CA, USA).

Apoptosis/necrosis detection

Cells were treated with LPS and each membrane was treated with SWCNT, or MWCNT. The treated cells were stained with annexin V and propidium iodide (BD Biosciences, San Jose, CA, USA) using a BD Pharmingen[™] FITC Annexin V Apoptosis Detection Kit II (BD Biosciences). Stained cells were detected by flow cytometry (BD LSRFortessa[™] cell analyzer; BD Biosciences) and the acquired data was analyzed using FlowJo software version 9.3.3 (Tree Star Inc.).

Statistical analysis

Experimental data were analyzed using the Student's t-test, and $p < 0.05$ was considered statistically significant.

RESULTS

PVDF treatment decreased NO production in LPS-activated macrophages

To test our hypothesis that CNTs bound to PVDF membranes do not modulate macrophage function, we treated RAW cells with 3 different PVDF membranes: 1) PVDF membrane alone, 2) SWCNT-attached PVDF membrane, and 3) MWCNT-attached PVDF membrane. NO production in these cells was measured after 24 h of treatment. To our surprise, LPS-stimulated RAW cells treated with any of these materials produced reduced amounts of NO compared to the negative control (Fig. 1). These data suggest that PVDF or CNT-attached PVDF can inhibit macrophage activation by LPS.

The number of macrophages was reduced following treatment with CNT-attached PVDF

As NO production in LPS-stimulated macrophages was inhibited by PVDF with or without CNT attachment, we examined whether there were any changes in cell number. RAW cells

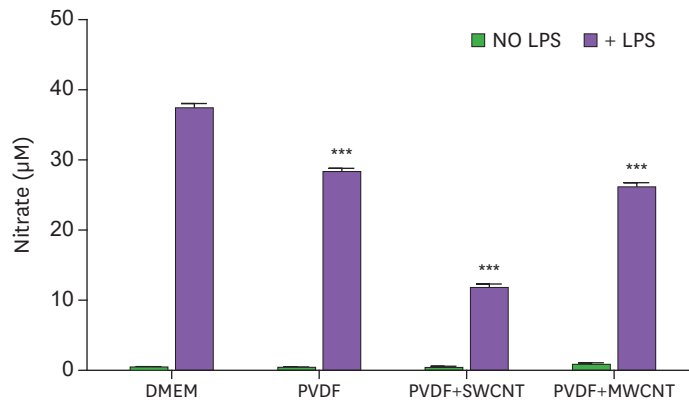


Figure 1. Effect of PVDF membrane on NO production in LPS-stimulated RAW cells. RAW cells were cultured and stimulated with PBS or LPS. These cells were then treated with one of the membranes for 24 h, as indicated. Cells treated with DMEM served as negative controls. Culture supernatants were harvested and tested for NO production. Data are shown as mean±standard deviation values of triplicates in each group. These data are representative of 2 independent experiments. ***p<0.001.

were seeded in a 96-well plate and treated with or without LPS. After 24 h of treatment, the cell number was examined by an MTT assay. Surprisingly, treatment with PVDF alone decreased the number of macrophages (**Fig. 2**). Moreover, the number of macrophages was also reduced by treatment with SWCNT- or MWCNT-bound PVDF. These findings suggest that PVDF with or without SWCNT affects macrophage homeostasis, resulting in reduced expression of NO.

PVDF increased cell cycle of macrophages

Since we found reduced number of RAW cells after PVDF treatment, we hypothesized that these cells underwent cell cycle arrest. To test this hypothesis, we treated RAW cells with PVDF, SWCNT-bound PVDF, or MWCNT-bound PVDF for 24 h and performed cell cycle analysis. In response to PVDF alone, RAW cells showed increased frequency in the S/G2 phase regardless of LPS stimulation, suggesting that PVDF induced activation and proliferation of RAW cells. Although RAW cells without LPS stimulation were not affected

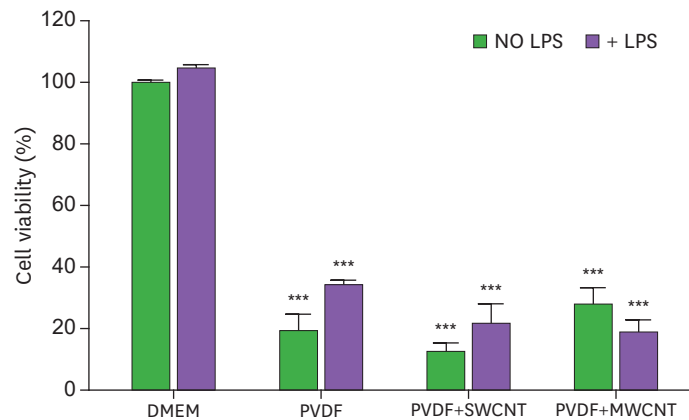


Figure 2. Influence of PVDF membrane on RAW cell survival with or without LPS stimulation. PBS or LPS-stimulated RAW cells were treated with the indicated membrane for 24 h. DMEM treated RAW cells were used as negative controls. After removing the culture supernatants, the cells were examined for survival compared with the negative controls. The plotted values represent survival rate±standard deviation. These data are representative of 2 independent experiments. ***p<0.001.

by treatment with PVDF+SWCNT or PVDF+MWCNT, LPS-stimulated RAW cells showed a substantially increased ratio of cells in the S/G2 phase with the same treatments (Fig. 3). We thus concluded that treatment with PVDF does not induce cell cycle arrest in macrophages. Instead, PVDF increased the proportion of macrophages in the cell cycle probably due to activation of these cells, especially upon stimulation with LPS.

PVDF treatment induced apoptosis in murine macrophages

Considering that the frequency of PVDF-treated macrophages in the S/G2 phase was increased compared to controls, we examined whether these cells underwent activation-induced cell death. RAW cells were stimulated either with PBS or LPS, and then treated with PBS, PVDF alone, SWCNT-bound PVDF, or MWCNT-bound PVDF for 24 h. These cells were then analyzed for annexin V expression by flow cytometry. As shown in Fig. 4, the ratio of apoptotic RAW cells in the LPS-stimulated group was 3 times greater than the negative controls. When these cells were treated with PVDF alone, the ratio of apoptotic cells was increased irrespective of the absence (5.83→19.2) or presence (17.4→35.3) of LPS. Although the percentage of apoptotic cells upon treatment with CNT-bound PVDF was unchanged in

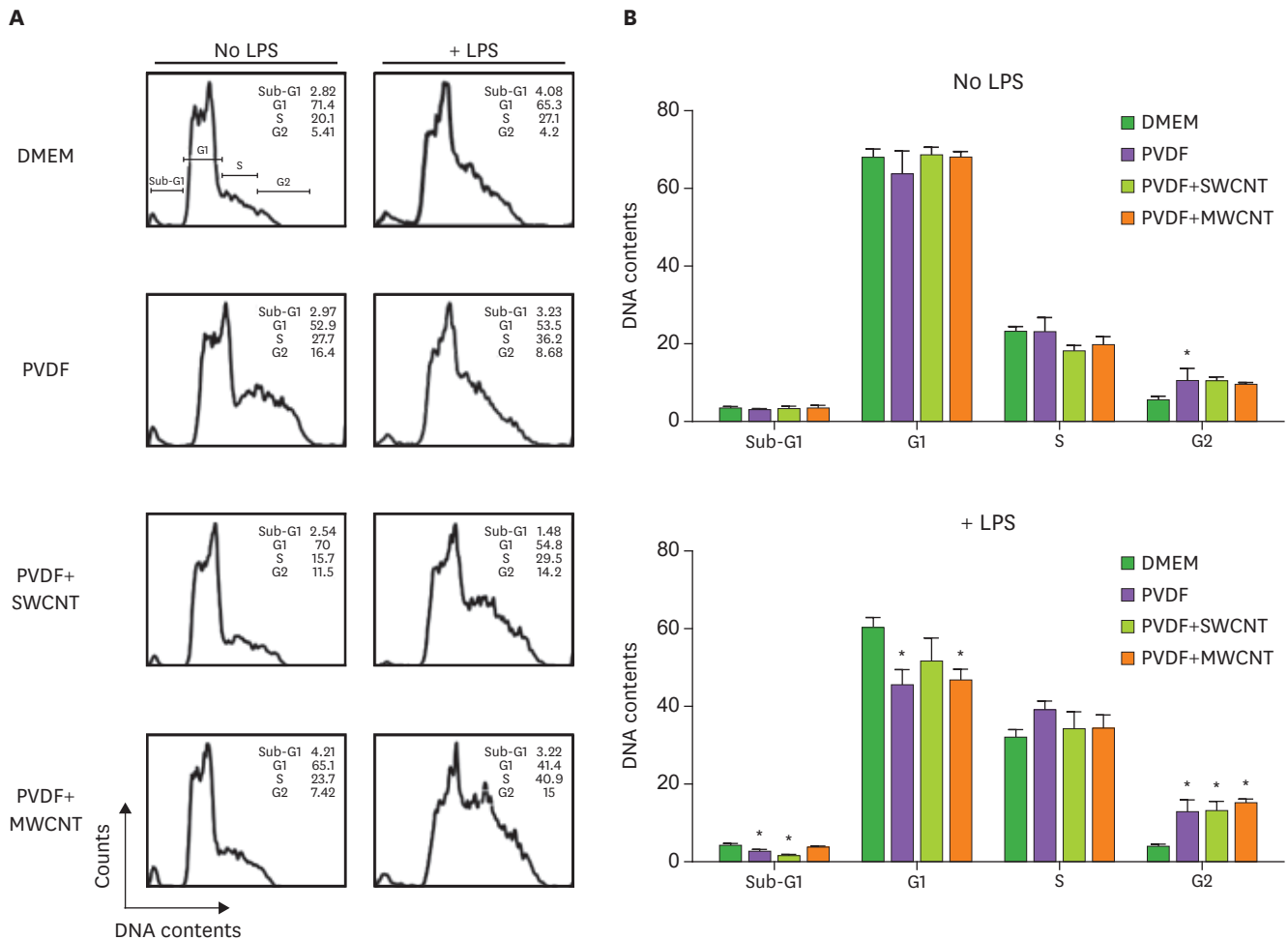


Figure 3. Altered cell cycle in RAW cells after exposure to PVDF membrane. Cells were treated with PVDF, PVDF+SWCNT, or PVDF+MWCNT. The cells were then exposed to LPS or LPS-free conditions for 21 h, and then stained using the Krishan staining buffer. (A) Stained cells were analyzed by flow cytometry and a histogram was obtained. The cell cycle of these cells was examined based on the DNA content of RAW cells. (B) The plotted values represent mean±standard deviation. The data are representative of 3 independent experiments. *p<0.05.

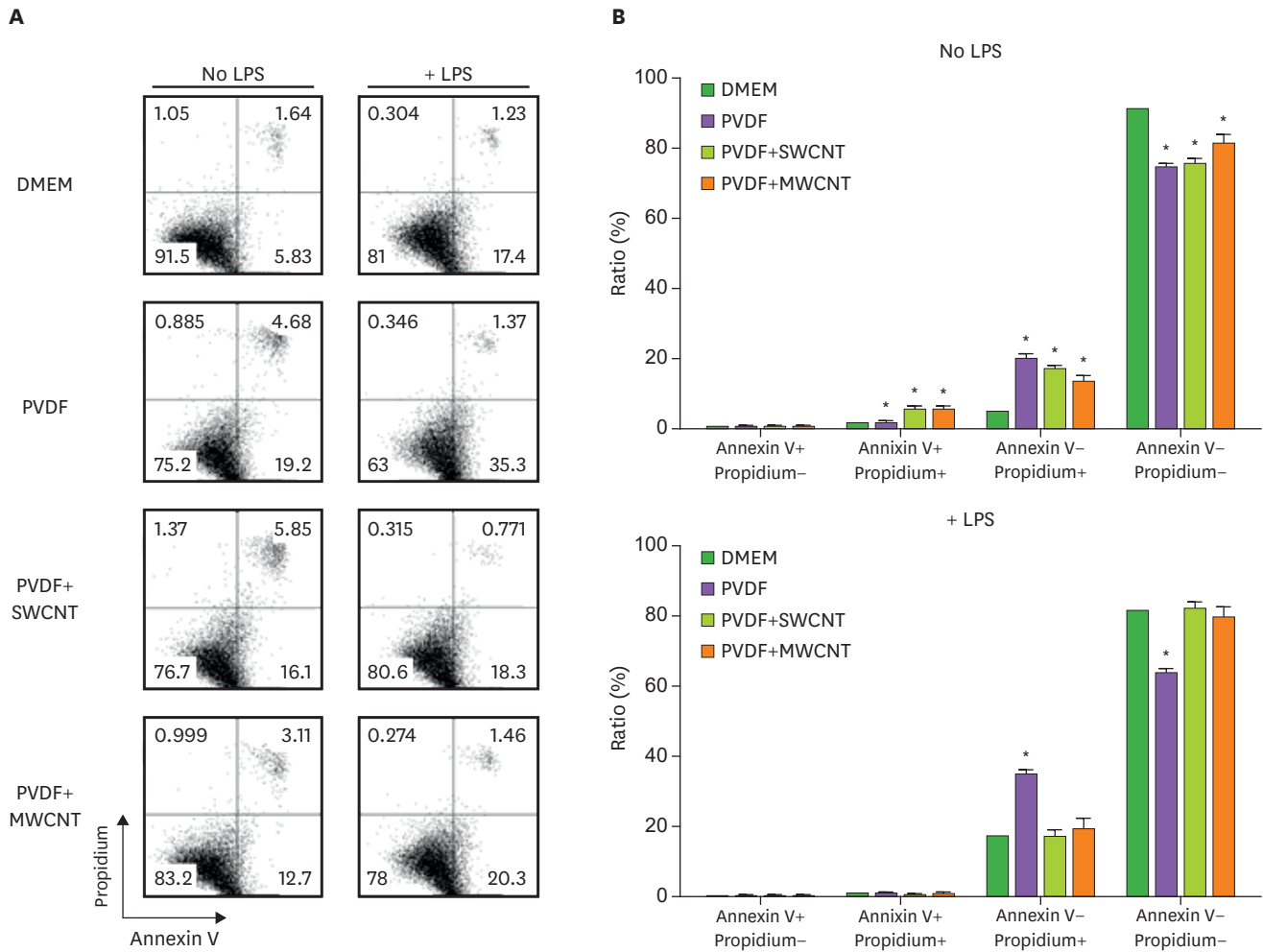


Figure 4. Increased apoptosis in RAW cells after treatment with PVDF. Cells were treated with the 3 types of membrane: PVDF, PVDF+SWCNT, and PVDF+MWCNT. The cells were then exposed to LPS or LPS-free conditions for 21 h and then stained with annexin V and propidium iodide. (A) Stained cells were examined by flow cytometry. (B) The data shown are means of triplicates±standard deviation. Similar results were obtained in 3 independent experiments. *p<0.05.

the LPS treated RAW cells, the ratio of annexin V+ cells was increased by 2-fold compared to the DMEM control in the absence of LPS. Overall, these data suggest that PVDF induces activation-induced cell death in macrophages irrespective of the presence of CNT, resulting in decreased production of NO.

DISCUSSION

PVDF has been widely used in the membrane industry but its biological effects remain to be addressed. In this study, we examined PVDF for its ability to modulate inflammatory responses in murine macrophages. When macrophages were treated with PVDF, NO production in response to LPS was inhibited. We also observed reduced numbers of these cells after PVDF treatment for 24 h, probably due to activation-induced cell death.

The proinflammatory effects of PVDF have been reported previously in animal models. PVDF was found to induce inflammatory and fibrotic reactions, even though the level of these

reactions was lower than those induced by polypropylene (PP) (15). In other reports, PVDF and PP induced comparable foreign body reactions such as granuloma formation (16,17). PVDF could induce inflammation in these animal models, involving various leukocytes including T cells, B cells, macrophages, and possibly others. Since it is difficult to dissect the roles of each cell type, we focused on the effects of PVDF on LPS-stimulated macrophages. Although PVDF showed a reduced level of inflammation compared to PP in previous reports, we found that PVDF showed toxicity in macrophages resulting in cell death. Altogether, these data suggest that PVDF modulates inflammatory reactions by causing cell death in macrophages.

Fig. 4 shows that treatment with PVDF induced apoptosis in macrophages; however, the cause of this cell death is not clear. One possible explanation is that these cells underwent activation-induced cell death (18). Macrophages treated with PVDF appeared to be activated as observed by their increased proportion in the S/G2 phase of the cell cycle. Notably, treatment with PVDF alone showed more cells in the G2 phase than the PVDF+CNT treatment in normal macrophages, whereas treatment with PVDF alone showed fewer cells in the G2 phase compared to treatment with CNT-attached PVDF in LPS-stimulated macrophages. These data may indicate that the effects of PVDF combined with CNT may vary depending on the activation status of macrophages.

We previously demonstrated that CNT could induce necrosis in macrophages (12). This is probably due to the asbestosis caused by “frustrated macrophages” (19). We therefore hypothesized that membrane-bound CNT does not induce cell death and could facilitate the use of CNT in health purposes. However, PVDF itself altered the survival of macrophages. Therefore, other mesh materials need to be explored for carrying CNT with minimal inflammatory responses.

In this study, we examined the function of CNT-attached PVDF on inflammatory responses. Macrophages treated with CNT-attached PVDF showed reduced NO production probably due to cell death. PVDF could also activate macrophages resulting in their proliferation. Finally, PVDF-treated macrophages underwent cell death probably through activation-induced cell death. Taken together, these data indicate that PVDF needs to be investigated for medical applications.

ACKNOWLEDGEMENTS

We appreciate Kyong Hoon Kim, Seung-min Yeon, and Ji Hwa Hong for their critical comments. This study is supported by Basic Science Research Program (Grant No. NRF-2014R1A4A1007304 and NRF-2017R1A2B4003376) of the National Research Foundation of Korea funded by the Korea government.

REFERENCES

1. Oh SJ, Kim N, Lee YT. Preparation and characterization of PVDF/TiO₂ organic-inorganic composite membranes for fouling resistance improvement. *J Membr Sci* 2009;345:13-20.
CROSSREF
2. Deng B, Yu M, Yang X, Zhang B, Li L, Xie L, Li J, Lu X. Antifouling microfiltration membranes prepared from acrylic acid or methacrylic acid grafted poly(vinylidene fluoride) powder synthesized via pre-irradiation induced graft polymerization. *J Membr Sci* 2010;350:252-258.
CROSSREF

3. Chiang YC, Chang Y, Higuchi A, Chen WY, Ruaan RC. Sulfobetaine-grafted poly(vinylidene fluoride) ultrafiltration membranes exhibit excellent antifouling property. *J Membr Sci* 2009;339:151-159.
[CROSSREF](#)
4. Lang WZ, Xu ZL, Yang H, Tong W. Preparation and characterization of PVDF-PFSA blend hollow fiber UF membrane. *J Membr Sci* 2007;288:123-131.
[CROSSREF](#)
5. Flajnik MF, Du Pasquier L. Evolution of innate and adaptive immunity: can we draw a line? *Trends Immunol* 2004;25:640-644.
[PUBMED](#) | [CROSSREF](#)
6. Janeway CA Jr, Medzhitov R. Introduction: the role of innate immunity in the adaptive immune response. *Semin Immunol* 1998;10:349-350.
[PUBMED](#) | [CROSSREF](#)
7. Hoebe K, Janssen E, Beutler B. The interface between innate and adaptive immunity. *Nat Immunol* 2004;5:971-974.
[PUBMED](#) | [CROSSREF](#)
8. Ulevitch RJ, Tobias PS. Recognition of gram-negative bacteria and endotoxin by the innate immune system. *Curr Opin Immunol* 1999;11:19-22.
[PUBMED](#) | [CROSSREF](#)
9. MacMicking J, Xie QW, Nathan C. Nitric oxide and macrophage function. *Annu Rev Immunol* 1997;15:323-350.
[PUBMED](#) | [CROSSREF](#)
10. Boscá L, Zeini M, Través PG, Hortelano S. Nitric oxide and cell viability in inflammatory cells: a role for NO in macrophage function and fate. *Toxicology* 2005;208:249-258.
[PUBMED](#) | [CROSSREF](#)
11. Aderem A, Underhill DM. Mechanisms of phagocytosis in macrophages. *Annu Rev Immunol* 1999;17:593-623.
[PUBMED](#) | [CROSSREF](#)
12. Kim KH, Yeon SM, Kim HG, Lee H, Kim SK, Han SH, Min KJ, Byun Y, Lee EH, Lee KS, et al. Single-walled carbon nanotubes induce cell death and transcription of TNF- α in macrophages without affecting nitric oxide production. *Inflammation* 2014;37:44-54.
[PUBMED](#) | [CROSSREF](#)
13. Kim HG, Yeon SM, Kim KH, Kim H, Park JI, Kang HJ, Cha EJ, Park HD, Kang HJ, Park TW, et al. Estrogenic endocrine-disrupting chemicals modulate the production of inflammatory mediators and cell viability of lipopolysaccharide-stimulated macrophages. *Inflammation* 2015;38:595-605.
[PUBMED](#) | [CROSSREF](#)
14. Krishan A. Rapid flow cytofluorometric analysis of mammalian cell cycle by propidium iodide staining. *J Cell Biol* 1975;66:188-193.
[PUBMED](#) | [CROSSREF](#)
15. Conze J, Junge K, Weiss C, Anurov M, Oettinger A, Klinge U, Schumpelick V. New polymer for intra-abdominal meshes--PVDF copolymer. *J Biomed Mater Res B Appl Biomater* 2008;87:321-328.
[PUBMED](#) | [CROSSREF](#)
16. Binnebösel M, Ricken C, Klink CD, Junge K, Jansen M, Schumpelick V, Lynen Jansen P. Impact of gentamicin-supplemented polyvinylidene fluoride mesh materials on MMP-2 expression and tissue integration in a transgenic mice model. *Langenbecks Arch Surg* 2010;395:413-420.
[PUBMED](#) | [CROSSREF](#)
17. Klosterhalfen B, Junge K, Hermanns B, Klinge U. Influence of implantation interval on the long-term biocompatibility of surgical mesh. *Br J Surg* 2002;89:1043-1048.
[PUBMED](#) | [CROSSREF](#)
18. Munn DH, Beall AC, Song D, Wrenn RW, Throckmorton DC. Activation-induced apoptosis in human macrophages: developmental regulation of a novel cell death pathway by macrophage colony-stimulating factor and interferon gamma. *J Exp Med* 1995;181:127-136.
[PUBMED](#) | [CROSSREF](#)
19. Donaldson K, Murphy FA, Duffin R, Poland CA. Asbestos, carbon nanotubes and the pleural mesothelium: a review of the hypothesis regarding the role of long fibre retention in the parietal pleura, inflammation and mesothelioma. *Part Fibre Toxicol* 2010;7:5.
[PUBMED](#) | [CROSSREF](#)