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## Near-Complete Genome Sequence of *Ralstonia solanacearum* T523, a Phylotype I Tomato Phytopathogen Isolated from the Philippines

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**ABSTRACT** *Ralstonia solanacearum* strain T523 is the major phytopathogen causing tomato bacterial wilt in the Philippines. Here, we report the complete chromosome and draft megaplasmid genomes with predicted gene inventories supporting rhizosphere processes, extensive plant virulence effectors, and the production of bioactive signaling metabolites, such as ralstonin, micacocidin, and homoserine lactone.

**R**alstonia solanacearum causes bacterial wilt, one of the most important plant diseases worldwide (1). Bacterial wilt affects 200 species in 50 different families, including tobacco, banana, and solanaceous crops, such as potato and tomato (2). Widespread outbreaks in the Philippines have affected various economically important crops, with severe effects on tomato production (3). Here, we report the genome of *R. solanacearum* strain T523, isolated from wilting tomatoes in the Philippines (3).

Genomic DNA was extracted from R. solanacearum strain T523 cells grown in Kelman's tetrazolium chloride medium (24 h, 28°C) using an MG genomic DNA purification kit (MGmed-Doctor Protein, Republic of Korea), according to the manufacturer's protocol. The whole genome was sequenced at Macrogen, Inc. (Republic of Korea), from 10  $\mu$ g of genomic DNA using a PacBio P6 DNA polymerase binding kit and a PacBio version 4.0 sequencing kit with eight single-molecule real-time (SMRT) cells (C4 chemistry) on the PacBio RS II platform. This generated 139,215 reads from a 20-kb SMRT library (mean subread length, 6,474 bp; N<sub>50</sub>, 9,102 bp). The 9.01-Mb reads were de novo assembled into contigs using the Hierarchical Genome Assembly Process (HGAP version 2.3) (4) to generate a final genome of 5,722,229 bp. One contig is a complete, closed, circular chromosome with a size of 3,652,934 bp, a G+C content of 67%, and a coverage of  $98 \times$ . A second contig is the megaplasmid, with a size of 2,069,295 bp, a G+C content of 67%, and a coverage of  $112\times$ . Gene prediction was performed independently using the NCBI Prokaryotic Genome Annotation Pipeline (PGAP) (5) and the Joint Genome Institute-Integrated Microbial Genomes and Microbiomes (JGI-IMG/M) pipeline (6). Species identity was determined from the genome-wide average nucleotide identity (gANI) and alignment fraction (AF) using the Microbial Species

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Address correspondence to Asuncion K. Raymundo, akraymundo@gmail.com, or Albert Remus R. Rosana, rosana@ualberta.ca. A.D.M. and A.R.R.R. contributed equally to this work. Identifier (MiSI) calculator employed in IMG/M (7). Strain identity was ascertained by the digital DNA-DNA hybridization score using the Genome-to-Genome Distance Calculator (GGDC) version 2.1 (8). Bioactive secondary metabolites and virulence-associated genes were predicted using the antiSMASH version 4 (9) and Ralsto T3E (10) servers, respectively. All programs were run with default parameters unless otherwise noted.

The T523 genome has an ANI of >99% (AF, 0.9) and a dDDH (formula 2) of <70% with *R. solanacearum* GMI1000 and other phylotype I strains, thereby supporting the nomenclature. The genome revealed an extensive repertoire of biosynthetic gene clusters and type III virulence effectors supporting rhizosphere processes and plant symbiotic associations. The chromosome encodes a complete gene cluster for mica-cocidin biosynthesis, a siderophore utilizing a hybrid pathway of nonribosomal peptide synthetase, and a type I iterative polyketide synthase (11). The megaplasmid encodes genes involved in the production of the antibiotic lipopeptide ralstonin/ralsolamycin, with established phytotoxic (12, 13) and antifungal (12, 14) activities, and a putative bacteriocin. Biosynthetic gene clusters for exopolysaccharide, terpene, and homoserine lactone production were detected. Virulence-associated enzyme loci were identified, including pectinase, cellulase, and phospholipase C. Finally, the Ralsto T3E server predicted 37 and 45 *rip 77 (Ralstonia*-injected proteins) genes (10) located in the chromosome and megaplasmid, respectively.

**Data availability.** The sequences were deposited in DDBJ/ENA/GenBank under accession numbers CP022702 and CP022703 for the chromosome and megaplasmid, respectively. The sequencing reads were deposited in the SRA under the accession number SRP159038.

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## REFERENCES

- Yabuuchi E, Kosako Y, Yano I, Hotta H, Nishiuchi Y. 1995. Transfer of two *Burkholderia* and an *Alcaligens* species to *Ralstonia* gen. nov. Microbiol Immunology 39:897–904. https://doi.org/10.1111/j.1348-0421.1995 .tb03275.x.
- Vailleau F, Sartorel E, Jardinaud MF, Chardon F, Genin S, Huguet T, Gentzbittel L, Petitprez M. 2007. Characterization of the interaction between the bacterial wilt pathogen *Ralstonia solanacearum* and the model legume plant *Medicago truncatula*. Mol Plant Microbe Interact 20:159–167. https://doi.org/10.1094/MPMI-20-2-0159.
- Orlina-Villareal ME, Opina NL, Raymundo AK. 2008. A hypervirulent isolate identified from a race 1 *Ralstonia solanacearum* strain from tomato (*Lycopersicon esculentum* mill. cv. L-180). Phil Agric Scientist 91:94–98.
- Chin C-S, Alexander DH, Marks P, Klammer AA, Drake J, Heiner C, Clum A, Copeland A, Huddleston J, Eichler EE, Turner SW, Korlach J. 2013. Nonhybrid, finished microbial genome assemblies from long-read SMRT sequencing data. Nat Methods 10:563–569. https://doi.org/10 .1038/nmeth.2474.
- Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. Nucleic Acids Res 44: 6614–6624. https://doi.org/10.1093/nar/gkw569.
- 6. Markowitz VM, Chen I-MA, Palaniappan K, Chu K, Szeto E, Pillay M, Ratner

A, Huang J, Woyke T, Huntemann M, Anderson I, Billis K, Varghese N, Mavromatis K, Pati A, Ivanova NN, Kyrpides NC. 2014. IMG 4 version of the integrated microbial genomes comparative analysis system. Nucleic Acids Res 42:D560–D567. https://doi.org/10.1093/nar/gkt963.

- Varghese NJ, Mukherjee S, Ivanova N, Konstantinidis KT, Mavrommatis K, Kyrpides NC, Pati A. 2015. Microbial species delineation using whole genome sequences. Nucleic Acids Res 43:6761–6771. https://doi.org/10 .1093/nar/gkv657.
- Meier-Kolthoff JP, Auch AF, Klenk H-P, Göker M. 2013. Genome sequence-based species delimitation with confidence intervals and improved distance functions. BMC Bioinformatics 14:60. https://doi.org/10 .1186/1471-2105-14-60.
- Blin K, Wolf T, Chevrette MG, Lu X, Schwalen CJ, Kautsar SA, Suarez Duran HG, de los Santos ELC, Kim HU, Nave M, Dickschat JS, Mitchell DA, Shelest E, Breitling R, Takano E, Lee SY, Weber T, Medema MH. 2017. antiSMASH 4.0—improvements in chemistry prediction and gene cluster boundary identification. Nucleic Acids Res 45:W36–W41. https://doi .org/10.1093/nar/gkx319.
- Peeters N, Carrère S, Anisimova M, Plener L, Cazalé A-C, Genin S. 2013. Repertoire, unified nomenclature and evolution of the type III effector gene set in the *Ralstonia solanacearum* species complex. BMC Genomics 14:859. https://doi.org/10.1186/1471-2164-14-859.

- Kreutzer MF, Kage H, Gebhardt P, Wackler B, Saluz HP, Hoffmeister D, Nett M. 2011. Biosynthesis of a complex yersiniabactin-like natural product via the *mic* locus in phytopathogen *Ralstonia solanacearum*. Appl Environ Microbiol 77:6117–6124. https://doi.org/10.1128/AEM.05198-11.
- Murai Y, Mori S, Konno H, Hikichi Y, Kai K. 2017. Ralstonins A and B, lipopeptides with chlamydospore-inducing and phytotoxic activities from the plant pathogen *Ralstonia solanacearum*. Org Lett 19: 4175–4178. https://doi.org/10.1021/acs.orglett.7b01685.
- Baldeweg F, Kage H, Schieferdecker S, Allen C, Hoffmeister D, Nett M. 2017. Structure of ralsolamycin, the interkingdom morphogen from the crop plant pathogen *Ralstonia solanacearum*. Org Lett 19:4868–4871. https://doi.org/10.1021/acs.orglett.7b02329.
- Spraker JE, Sanchez LM, Lowe TM, Dorrestein PC, Keller NP. 2016. Ralstonia solanacearum lipopeptide induces chlamydospore development in fungi and facilitates bacterial entry into fungal tissues. ISME J 10:2317–2330. https://doi.org/10.1038/ismej.2016.32.