DOI: 10.1111/1759-7714.13939

ORIGINAL ARTICLE

WILEY

miRNA-218-5p increases cell sensitivity by inhibiting PRKDC activity in radiation-resistant lung carcinoma cells

Xiaoke Chen[†]

Yuanyuan Xu[†] | Long Jiang 💿 | Qiang Tan 💿

Shanghai Lung Cancer Center, Shanghai Chest Hospital, Shanghai Jiao Tong University, Shanghai, China

Correspondence

Qiang Tan, Shanghai Lung Cancer Center, Shanghai Chest Hospital, Shanghai Jiao Tong University, No.241 Huaihai West Road, Xuhui District, Shanghai, 200030, China. Email: tttqiang323@163.com

Funding information National Natural Science Fundation of China,

Grant/Award Number: 8187070347

Abstract

Background: Non-small cell lung carcinoma (NSCLC) is a malignancy with the highest mortality rate. Currently, surgery combined with radiotherapy is the first choice in the clinical treatment of lung carcinoma (LC); however, long-term radiotherapy leads to radiation resistance in patients, resulting in treatment failure.

Methods: In this study, a new microRNA-218-5p (miRNA-218-5p) was identified, and its function in LC was investigated.

Results: Reverse transcription quantitative polymerase chain reaction (RT-qPCR) results revealed that miRNA-218-5p was downregulated in LC. Overexpression or inhibition of miRNA-218-5p in LC and targeted binding of protein kinase, DNAactivated, catalytic polypeptide (PRKDC) to miRNA-218-5p were confirmed by comprehensive bioinformatic analysis. Exosomes from A549 and H1299 cells were cocultured with miRNA-218-5p and then cotransfected into radiation-resistant A549R and H1299R cells; the proliferation of radiation-resistant LC cells was found to be effectively inhibited and apoptosis was induced. Overexpression of miRNA-218-5p and X-irradiation could enhance the radiosensitivity of LC cells. Exogenous miRNA-218-5p derived from A549 and H1299 cells could be transfected into radiationresistant LC cells and could inhibit PRKDC expression, thus accelerating DNA damage, apoptosis, and radiation sensitization of LC cells.

Conclusions: miRNA-218-5p could induce apoptosis and enhance the radiosensitivity of LC cells through regulatory activities, thus suggesting its application as a potential target for LC treatment.

KEYWORDS

exosomes, lung carcinoma, miRNA-218-5p, PRKDC, radiation resistance

INTRODUCTION

Lung carcinoma (LC) has become an increasingly serious public health concern worldwide.¹ Statistics indicate that in 2018 there were more than 2 million new LC patients globally² and 1.7 million deaths. LC can be grouped into small cell LC (SCLC) and non-small cell lung carcinoma (NSCLC); NSCLC accounts for over 80% of LCs. Radiotherapy is an important treatment option for malignant thoracic tumors.³⁻⁵ At present, radiotherapy combined with surgery

[†]Both authors contributed equally to this work and should be considered as equal first

is known to exert potent synergistic effects,^{6, 7} which can induce cell death by breaking the DNA of tumor cells. However, carcinoma treatment is a lengthy process. Patients often show adverse reactions during long-term radiotherapy, and an increasing number of studies have revealed that NSCLC can acquire resistance to radiotherapy, reducing its efficacy, and tumors can recur and metastasize.⁸ In addition, for epidermal growth factor receptor-mutant lung adenocarcinoma, the radiotherapeutic effects of tyrosine kinase inhibitors are also not ideal.⁹ Therefore, exploring radiotherapy tolerance is crucial.

microRNAs (miRNAs), which have emerged as a subject of extensive research, are short-chain non-coding RNAs

© 2021 The Authors. Thoracic Cancer published by China Lung Oncology Group and John Wiley & Sons Australia, Ltd.

coauthors.

This is an open access article under the terms of the Creative Commons Attribution-NonCommercial License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited and is not used for commercial purposes

comprising 19–25 nucleotide bases. Recent studies have confirmed that miRNAs can target downstream genes through mRNA 3'-untranslated regions (UTRs) and participate in tumorigenesis.^{10–12} The key role of miRNAs in the pathogenesis and response to treatment has been confirmed in different carcinomas.¹³ For example, Meng et al.¹⁴ reported that miR-183-5p plays a tumor-inhibitory role in LC by inhibiting PIK3CA. The levels of miRNA-218-5p, an miRNA that was discovered early on, were found to be low in LC¹⁵; studies have revealed that exosomes containing miRNA-218-5p play a role in osteoblast differentiation as well as oxaliplatin resistance in colon carcinoma.^{16, 17} However, there are few studies on the role of miRNA-218-5p in tumor tolerance to radiotherapy in LC.

Cells can communicate directly via cell-to-cell contacts and synapses through chemical receptors.¹⁸ Recent studies have shown that exosomes also act in cell-to-cell communication.¹⁹ Exosomes can carry lipids, proteins, mRNA, and various miRNAs.²⁰ It has been reported that exosomes engage recipient cells by fusion with target cell membranes, suggesting that exosomes may act in transporting radiationinduced serum miRNAs and thus affect the proliferation and radio resistance of cells.²¹

In this study, we found that miRNA-218-5p reduced the tolerance of LC cells to radiotherapy, thus suggesting a potential therapeutic strategy for tumor tolerance to radio-therapy in LC.

METHODS

Clinical data

A total of 42 patients with NSCLC who received radiotherapy between January 2013 and January 2015 were selected as participants. The patient received 60 Gy/30 min of radiotherapy. Peripheral venous blood was obtained 24 h before treatment, and peripheral venous blood was collected again after the patient completed the course of treatment. In addition, 30 healthy individuals during the same period were included in a control group. Peripheral blood samples were collected from both groups, held at room temperature for 60 min, and centrifuged at $3000 \times \text{g}$ for 10 min. The supernatant was then obtained and stored at -80° C. The patients had not received chemotherapy before this study but were diagnosed with NSCLC. All subjects agreed to participate in the study and signed consent forms voluntarily. This

CHEN ET AL.

research was approved by the Shanghai Chest Hospital, Shanghai Jiao Tong University and was carried out in compliance with the Declaration of Helsinki.²²

Cell culture and irradiation

HEK-293 T, A549, H1299, HCC827, and BEAS-2B cell lines were procured from the American Type Culture Collection and cultured in Dulbecco's modified Eagle's medium. A549 and H1299 cell lines were irradiated for radiation-resistance analysis. When cell confluence reached 50%, A549 and H1299 cells were subjected to X-ray irradiation (2 Gy), cultured, digested with trypsin at a 90% convergence rate, and then reirradiated at a 50% convergence rate, with a 2 Gy daily fraction size being administered 30 times for a total dose of 60 Gy. Parental cells were cultured without radiation under the same conditions. In addition, there was an interval of four or five months between the last two partial exposures. Cells were replated for colony formation after recovering for 2-3 weeks after the last irradiation. After 2-3 weeks, the progenies of the clonogenic survivors were obtained.

Cell transfection

miRNA-218-5p mimic, miRNA-218-5p inhibitor, inhibitor-NC, and protein kinase, DNA-activated, catalytic polypeptide (PRKDC) small interfering RNA (si-PRKDC) were synthesized by GenePharma Co. Ltd. The synthesized RNA was transfected using Lipofect 3000.

Reverse transcription quantitative polymerase chain reaction (RT-qPCR)

Total RNA was obtained using TRIzol reagent, and reverse transcription was performed to obtain cDNA. TransStart SYBR Green qPCR Supermix (TransGen Biotech) was used for PCR amplification using an ABI 7500 system, and the reaction systems were configured according to the manufacturer's instructions. Three replicate wells were employed, and the test was conducted three times. *U6* and *GAPDH* were used as internal references for miRNAs and mRNAs, respectively, with $2^{-\triangle \triangle qct}$ for data analysis.²³ The PCR primer sequences are listed in Table 1.

TABLE 1 PCR primer sequences

Gene	Upstream primer	Downstream primer
miRNA-218-5p	5'-TTGCGGATGGTTCCGTCA AGCA-3'	5'-ATCCAGTGCAGGGTCCGAGG-3'
PRKDC	5'-GAAAGGAGGTTCTAAACTAC-3'	5'-GCTACGTGAATATAGACCATA-3'
U6	5'-CTCGCTTCGGCAGCACA-3'	5'-AACGCTTCACGAATTTGCGT-3'
GAPDH	5'-CCATCTTCCAGGAGCGAGAT-3'	5'-TGCTGATGATCTTGAGGCTG-3'

Colony formation

After 48 h of transfection, the cells were treated with X-rays at 0, 2, 4, 6, or 8 Gy. Cells were cultured and evaluated for survival using a colony forming assay. Cells were reinoculated into six-well plates, cultured overnight to allow cell adhesion, treated with 0–8 Gy X-rays, cultured at 37°C for two weeks, fixed with methanol for 20 min, and stained with 0.1% crystal violet.

3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyl-2H4tetrazolium bromide (MTT) assay

MTT was used to evaluate cell proliferation. After 48 h of transfection, the cells were resuspended and the densityadjusted cells (2×10^3 cells/well) were seeded into 96-well plates. After 24 h of culture, 20 µl MTT solution (5 mg/ml) was added at 24, 48, and 72 h, followed by incubation at 37°C for another 4 h. Next, the medium was aspirated and 150 µl of dimethyl sulfoxide (DMSO) was added. The optical density (OD) was determined at 492 nm.

Cells in DNA synthesis assay

Cell-Light EdU DNA cell proliferation kit (Ribo Bio) was used to determine the proliferation rate of A549R and H1299R cells according to the manufacturer's instructions. Briefly, cells were incubated with 50 μ M EdU for 2 h, then fixed, permeabilized, and stained with EdU. The nuclei were stained with Hoechst 33342 at a concentration of 5 μ g/ml for 30 min, and the cells were then inspected using a fluorescence microscope (Olympus). The viability index was calculated as follows: Viability index = experimental OD value/control OD value. In addition, 5-ethynyl-20-deoxyuridine (EdU) was used to evaluate cell proliferation.

Apoptosis

Cells were obtained after 48 h of transfection, and apoptosis was evaluated using annexin V-fluorescein isothiocyanate (FITC). Transfected cells were harvested, rinsed, and resuspended. Next, the cells were stained with FITC and propidium iodide (PI) for 15 min. Cytek Aurora and FlowJo were used for the analysis.

Western blotting (WB)

Total protein was extracted, and exosome protein levels were determined using BCA (Bicinchoninic acid, Beyotime), followed by quantification. The proteins were transferred to a polyvinylidene fluoride membrane, which was sealed with phosphate buffer and mixed with diluted anticleaved PARP, anticleaved caspase 3, anti-CD63, and anti- β -actin antibodies. Secondary antibody labeled with horseradish peroxidase was added, followed by incubation for 1 h. Enhanced chemiluminescence was employed to determine protein levels using Bio-Rad ChemiDoc XRS densitometry and Image Lab version 4.1.

Dual-luciferase reporter gene constructs

Potential targets of miRNA-216-5p were identified, and PRKDC was the target of miRNA-216-5p. To verify that PRKDC was a target of miRNA-216-5p, pCMV-REPORT-PRKDC-3'UTR-WT was established by inserting the PCMV-reporter vector into the DNA sequence of PRKDC 3'UTR. The binding site of

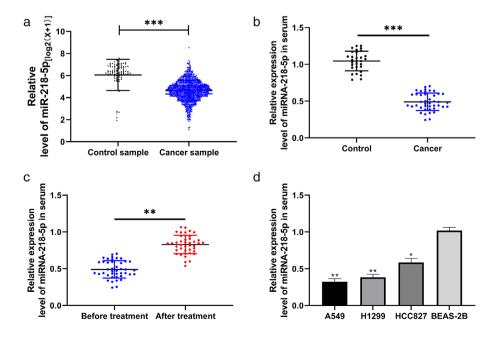


FIGURE 1 miRNA-218-5p expression in lung carcinoma patients after radiotherapy. (a) Relative expression of miRNA-218-5p in lung carcinoma in TCGA. (b) Reverse transcription quantitative polymerase chain reaction (RT-qPCR) was used to determine miRNA-218-5p expression in the serum of control and patient groups. ***p < 0.001, lung carcinoma groups compared with the control. (c) RT-qPCR was used to determine miRNA-218-5p expression in serum before and after radiotherapy. ***p < 0.001 after radiotherapy compared with before radiotherapy. (d) RTqPCR was used to determine miRNA-218-5p expression in lung carcinoma cell lines. **p < 0.01 and *p < 0.05 carcinoma cell lines compared with the BEAS-2B cell line

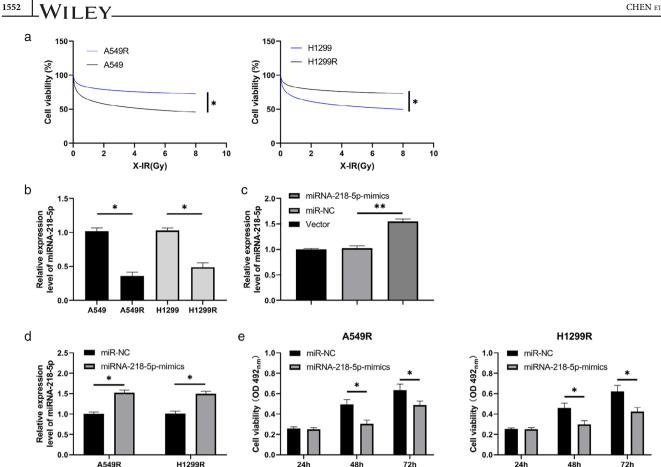


FIGURE 2 Role of miRNA-218-5p in the growth of radiation-induced lung carcinoma cells. (a) MTT assay was used to evaluate metabolic survival of parental cells and radiation-resistant cell lines under different doses of X-rays. (b) RT-qPCR was used to determine miRNA-218-5p expression in radiation-resistant cell lines. (c) RTqPCR was used to determine miRNA-218-5p expression in miRNA-218-5p-mimic plasmid. (d) RT-qPCR was used to determine miRNA-218-5p expression after transfection of miRNA-218-5p-mimic into radiation-resistant cell lines. (e) MTT assay was used to evaluate the proliferation of irradiated cell lines after transfection of the miRNA-218-5p-mimic. (b), p < 0.05, radiation-resistant cell lines compared with the control cell lines. (c–e), *p < 0.05, miRNA-218-5p-mimic compared with the NC

miRNA-216-5p was confirmed. A pCMV-REPORT-PRKDC-3'UTR-mut with a mutant miRNA-216-5p binding site was also constructed. miRNA-216-5p mimic and control were transfected into HEK-293 T cells, and the luciferase activity was tested.

Tumor formation in nude mice

A total of 12 male BALB/c nude mice (5-weeks-old) were procured from Charles River Laboratories. A normal diet was fed to the mice. This study conformed to the Animal Ethics Committee and the Laboratory Animal Guidelines. Overexpressed miRNA-216-5p (A549R cells) were injected into the mice, and X-ray irradiation (12 Gy) was conducted when tumor diameters reached 5 mm. The mice were grouped into an miRNA-NC group, miRNA-216-5p-mimics group, miRNA-NC + 12 Gy group, and miRNA-216-5pmimics+2 Gy group, with three mice in each group. Two days after X-ray irradiation, the tumor volume (V) was measured using a caliper. The spherical volume was calculated as follows: spherical volume = $4/3 \pi$ r³ or $(4/3) \pi$ (length/ 2) × (width/2) × (depth/2).

Statistical analysis

SPSS v.24.0 was applied for analysis, and GraphPad 8 was used for visualization. Counting data were represented in (%) and compared using the Chi-squared test. Measurement data were represented as means ± SEM and compared using Student's t-test. Single-factor analysis of variance (ANOVA) and least significant difference (LSD) post-hoc test were applied for multiple groups. Repeated measures ANOVA and Bonferroni post-hoc test were applied for multiple time points. The diagnostic predictive value was tested using the receiver operating characteristic curve. At p < 0.05, the difference was considered to be statistically significant.

RESULTS

Role of radiotherapy in improving miRNA-218-5p levels in LC

First, miRNA-218-5p levels in LC from The Cancer Genome Atlas (TCGA) data were determined, and RT-qPCR results revealed that the miRNA-218-5p level in the serum of LC

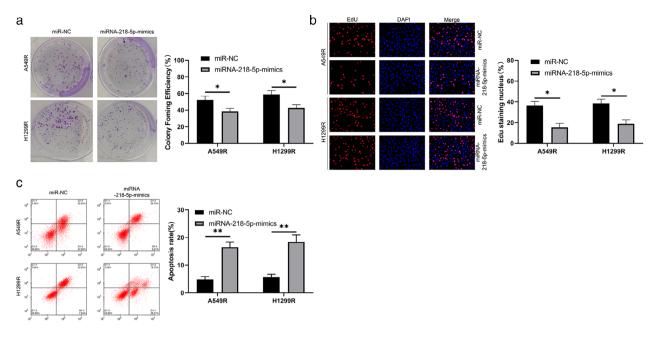


FIGURE 3 Role of miRNA-218-5p in the apoptosis of radiation-induced lung carcinoma cells. (a) Colony formation was used to evaluate changes in the cloning ability of irradiated cell lines after transfection of the miRNA-218-5p-mimic. (b) EdU was used to evaluate the proliferation of irradiated cell lines after the transfection of the miRNA-218-5p-mimic. (c) Flow cytometry was used to evaluate changes in the apoptosis rate of irradiated cell lines after transfection of the miRNA-218-5p-mimic. *p < 0.05; **p < 0.01

patients was evidently lower than that in the normal control group (Figure 1a,b). Further comparison showed that the miRNA-218-5p level in the serum of LC patients after radio-therapy was evidently higher than that before intervention (Figure 1c). In addition, the miRNA-218-5p level in LC cell lines was evaluated using RT-qPCR, and the results revealed that the miRNA-218-5p levels in the LC cell lines after X irradiation declined (Figure 1d). This suggests that miRNA-218-5p may be a latent observation index for radiotherapy in LC.

Elevation of miRNA-218-5p levels on proliferation and apoptosis of radiationresistant LC cells

A549 and H1299 cells were exposed to X-rays to establish radiation-resistant LC cells. Cloning experiments showed that the radiation-resistant cell lines (A549R and H1299R) had enhanced resistance to X-rays compared with the parental cells (Figure 2a). RT-qPCR revealed that the miRNA-218-5p abundance in A549R and H1299R cells was evidently lower than that in parental cells, suggesting that miRNA-218-5p may play a role in the radiation resistance of LC cells (Figure 2b,c). Therefore, we established an miRNA-218-5p-mimic plasmid and transferred it into A549R and H1299R cells to further observe any changes in cellular biological functions (Figure 2d). MTT (Figure 2e) assays, cloning assays (Figure 3a), and EdU experiments (Figure 3b) revealed that the metabolic survival (Figure 2e), clonogenic survival (Figure 3a), and DNA synthesis activity (Figure 3b), respectively, of A549R and H1299R cells

declined after transfection with the miRNA-218-5p-mimic, whereas apoptosis of A549R and H1299R cells was elevated after transfection of the miRNA-218-5p-mimic, as revealed by flow cytometry (Figure 3c).

miRNA-218-5p exhibits targeted binding to PRKDC

It has been proven that miRNAs regulate tumor growth by combining with downstream target genes. PRKDC and miRNA-218-5p have latent binding targets, identified through online website prediction analysis (Figure 4a). To verify the correlations among them, we first tested PRKDC in LC in TCGA database and found that PRKDC expression in LC is elevated (Figure 4b). We then tested PRKDC expression in LC patients using RT-qPCR, and the results were consistent with those obtained from the database (Figure 4c). Through correlation analysis, PRKDC expression was negatively correlated with miRNA-218-5p levels in the serum of LC patients (Figure 4d), suggesting that miRNA-218-5p may exhibit targeted binding to PRKDC. To verify this speculation, we conducted dual-luciferase reporter gene construct experiments, and the results revealed that the miRNA-218-5pmimic could hinder the fluorescence activity of PRKDC-WT (Figure 4e), whereas the miRNA-218-5p-inhibitor enhanced the fluorescence activity of PRKDC-WT. In addition, RT-qPCR and WB revealed that PRKDC mRNA and protein levels in cells transfected with the miRNA-218-5p-mimic or inhibitor were obviously hindered and accelerated, respectively (Figure 4f,g). This suggests that miRNA-218-5p exhibits targeted binding to PRKDC.

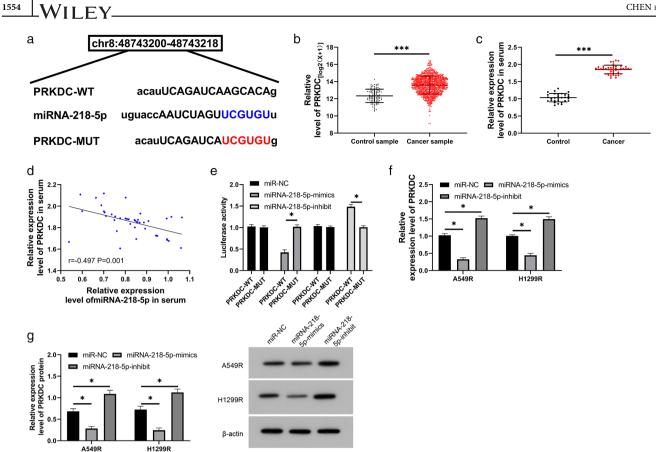


FIGURE 4 Targeted binding of miRNA-218-5p and protein kinase, DNA-activated, catalytic polypeptide (PRKDC). (a) StarBase predictions were used to identify the binding and mutated sites of miRNA-218-5p and PRKDC. (b) Expression of PRKDC in lung carcinoma in TCGA database. (c) Reverse transcription quantitative polymerase chain reaction (RT-qPCR) was used to evaluate the relative expression of PRKDC in the serum of patients with lung carcinoma. (d) Pearson's test was applied to identify any correlation involving miRNA-218-5p and PRKDC in patients with lung carcinoma. (e) Dualluciferase reporter gene analysis of miRNA-218-5p and PRKDC target binding. (f) and (g) RT-qPCR and WB were used to evaluate PRKDC mRNA and protein levels, respectively, in radiation-resistant lung carcinoma cells transfected with miRNA-218-5p-mimic/inhibitor. *p < 0.05; ***p < 0.001

Decreased PRKDC expression can inhibit the growth promotion of miRNA-218-5p-inhibitor on radiation-resistant LC cells

A cotransfection experiment was performed to test the role of miRNA-218-5p and PRKDC in radiation-resistant LC. The metabolic survival of radiation-resistant cells was increased (Figure 5a), the clone survival rate (Figure 5b) was enhanced, and apoptosis was hindered (Figure 6a,b). However, the metabolic and clonal survival of radiation-resistant cells declined after transfection of si-PRKDC. Compared with miRNA-NC + si-NC cells, miRNA-218-5p-inhibitor and si-PRKDC cotransfected cells showed no evident difference in metabolic survival, clonal survival, or apoptosis.

miRNA-218-5p-mimics increases the radiosensitivity of lung cancer nude mice

In the above experiment, we determined the function of miRNA-218-5p in radiation-resistant lung cancer cells. In order to further determine whether miRNA-218-5p can be a potential target in clinical radiotherapy, we established a

xenograft in a nude mouse model. Through measurement, we found that the tumor volume of nude mice after 12 Gy X-ray irradiation was significantly lower than that of miRNA-NC, while the tumor volume in nude mice injected with miRNA-218-5p-mimics combined with 12 Gy X-ray irradiation was significantly lower than that of miRNA-NC (Figure 7a,b). In addition, our research observed that the miRNA-218-5p in the tumor tissue of nude mice after 12 Gy X-ray irradiation was significantly higher than that of miRNA-NC, while miRNA-218-5p in tumors of nude mice after injection of miRNA-218-5p-mimics combined with 12 Gy X-ray irradiation was significantly higher than that of nude mice after miRNA-NC and 12 Gy X-ray irradiation. This suggests that miRNA-218-5p can increase the radio sensitivity in a lung cancer nude mouse model.

DISCUSSION

Radiation exposure increases the levels of free radicals in cells, accompanied by DNA strand breakage and dysfunction of the mitochondria, endoplasmic reticulum, and other organelles, eventually leading to apoptosis.^{24, 25} At present,

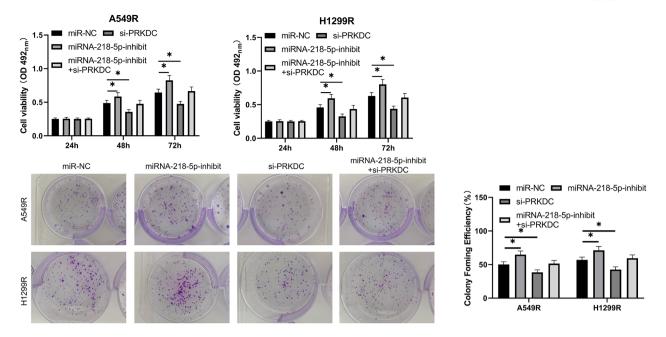


FIGURE 5 Decrease in protein kinase, DNA-activated, catalytic polypeptide (PRKDC) levels can inhibit promotion of miRNA-218-5p-inhibitor on growth of radiation-resistant lung carcinoma cells. (a) MTT assay was used to evaluate changes in the number of cells surviving metabolically after cotransfection of the miRNA-218-5p-inhibitor and si-PRKDC. (b) Colony-forming experiments were performed to evaluate changes in cell survival of reproductive integrity after cotransfection of the miRNA-218-5p-inhibitor and si-PRKDC. *p < 0.05; **p < 0.01

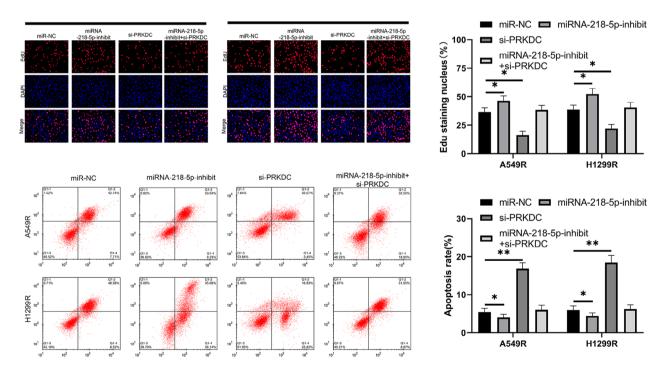


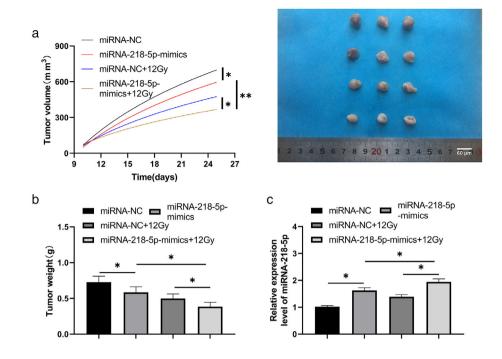
FIGURE 6 Decreasing the level of protein kinase, DNA-activated, catalytic polypeptide (PRKDC) can reverse miRNA-218-5p-inhibitor radiationresistant lung cancer cell apoptosis. (a) An EdU DNA cell proliferation test was used to evaluate changes in the number of cells undergoing DNA synthesis after cotransfection of the miRNA-218-5p-inhibitor and si-PRKDC. (b) Flow cytometry was used to evaluate changes in the apoptosis rates after cotransfection of the miRNA-218-5p-inhibitor and si-PRKDC. *p < 0.05; **p < 0.01

research concerning the mechanism involved in the radiation resistance in LC shows that P53 mutations, an X-linked inhibitor of apoptosis protein, and survivin have positive effects on radiotherapy.²⁶ However, the mechanisms and latent targets of radiotherapy remain unclear. miRNAs play a role in the radiation resistance and radiosensitivity of various tumor cells. For example, Zhu et al.²⁷ reported that miRNA-145 antagonizes the dryness and radiation resistance of colorectal carcinoma mediated by SNAI1; another study showed that miR-99a elevates the

FIGURE 7 miRNA-218-5p-mimics increases the radiosensitivity of lung cancer cells in a nude mice model. (a) The tumor volume changes in the nude mice over 25 days. (b) After 25 days, the nude mice were sacrificed and the tumor tissue mass was compared. *p < 0.05, **p < 0.01

HFY

1556



radiosensitivity of NSCLC via mTOR.²⁸ miRNA-218-5p, an miRNA that was discovered early on, is located on human chromosome 4p15.31. A previous study revealed that miRNA-218-5p levels are low in LC cells,²⁹ as confirmed by our results. In addition, a study has found that miRNA-218-5p plays a role in the sensitivity to ionizing radiation in breast carcinoma; however, there are relatively few studies on the role on miRNA-218-5p in radiation resistance of LC.³⁰ miRNA-218-5p levels in radiation-resistant LC cells were found to be decreased; the metabolic and clonogenic survival of radiation-resistant LC cells were obviously reduced, and apoptosis was induced by elevating miRNA-218-5p levels. In addition, the transfection of the miRNA-218-5p-mimic increased the sensitivity of radiationresistant cells to X-irradiation, suggesting that miRNA-218-5p is a latent therapeutic target for radiation-resistant LC cells.

It has been reported that miRNAs influence downstream target genes to participate in tumor radiation resistance mechanisms.^{31, 32} To further investigate the mechanisms underlying the effect of miRNA-218-5p on the radiation resistance of LC cells, we predicted the downstream target genes of miRNA-218-5p and found that there were targeted binding sites for PRKDC with miRNA-218-5p. This suggests that miRNA-218-5p may be a latent target for radiation resistance. PRKDC belongs to the non-homologous end joining pathway of the DNA double-strand break (DSB) repair response and can be recruited by DNA double-strand breaks to form DNAPK complexes with DNA-bound Ku70/80 heterodimers to promote the repair of DNA DSBs.^{33, 34} PRKDC levels in patients with LC are evidently elevated. Furthermore, miRNA-218-5p levels in the serum of LC patients showed a negative correlation with PRKDC abundance. In addition, miRNA-218-5p exhibited targeted binding to PRKDC. Knockdown of PRKDC could reverse the promotion of the miRNA-218-5p-inhibitor on the proliferation of radiation-resistant LC cells and inhibit apoptotic activity. It is suggested that miRNA-218-5p regulates the growth and apoptosis of radiation-resistant LC cells through PRKDC.

However, this study had certain limitations. First, we did not detect miRNA-218-5p in the serum exosomes of patients with LC. Previous studies have shown that miRNAs in serum exosomes can also be applied as a latent diagnostic marker for tumors. Second, owing to the short collection time of the samples, we did not conduct patient follow-up for a long period. In the future, we aim to perform additional clinical experiments to further verify the significance of miRNA-218-5p in serum exosomes in LC, in order to supplement our experimental results.

This study explored the role of miRNA-218-5p in the radiosensitivity of LC cells. miRNA-218-5p could induce apoptosis and enhance the radiosensitivity of LC cells through regulation, thus suggesting its application as a potential target for LC treatment.

ACKNOWLEDGMENT

This research was funded by National Natural Science Fundation of China Project No. 8187070347.

CONFLICT OF INTEREST

The authors declare that they have no competing interests.

ORCID

Long Jiang b https://orcid.org/0000-0002-6860-755X Qiang Tan b https://orcid.org/0000-0002-3978-7184

REFERENCES

- Watanabe SI, Nakagawa K, Suzuki K, Takamochi K, Ito H, Okami J, et al. Neoadjuvant and adjuvant therapy for stage III non-small cell lung cancer. Jpn J Clin Oncol. 2017;47:1112–8.
- Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin. 2018;68:394–424.
- Romaszko AM, Doboszyńska A. Multiple primary lung cancer: a literature review. Adv Clin Exp Med. 2018;27:725–30.
- Villalobos P, Wistuba II. Lung cancer biomarkers. Hematol Oncol Clin North Am. 2017;31:13–29.
- Steven A, Fisher SA, Robinson BW. Immunotherapy for lung cancer. Respirology. 2016;21:821–33.
- Chua GWY, Chua KLM. Which patients benefit most from stereotactic body radiotherapy or surgery in medically operable non-small cell lung cancer? An in-depth look at patient characteristics on both sides of the debate. Thorac Cancer. 2019;10:1857–67.
- Lemjabbar-Alaoui H, Hassan OU, Yang YW, Buchanan P. Lung cancer: biology and treatment options. Biochim Biophys Acta. 1856;2015: 189–210.
- Rizvi NA, Hellmann MD, Snyder A, Kvistborg P, Makarov V, Havel JJ, et al. Cancer immunology. Mutational landscape determines sensitivity to PD-1 blockade in non-small cell lung cancer. Science. 2015;348:124–8.
- Peng S, Wang R, Zhang X, Ma Y, Zhong L, Li K, et al. EGFR-TKI resistance promotes immune escape in lung cancer via increased PD-L1 expression. Mol Cancer. 2019;18:165.
- Yu Y, Wang Y, Xiao X, Cheng W, Hu L, Yao W, et al. MiR-204 inhibits hepatocellular cancer drug resistance and metastasis through targeting NUAK1. Biochem Cell Biol. 2019;97:563–70.
- 11. Lin YH. MicroRNA networks modulate oxidative stress in cancer. Int J Mol Sci. 2019;20:4497.
- Tuo L, Chu X, Sha S, Zhang X. MicroRNA and lung cancer: a mini review. Zhongguo Fei Ai Za Zhi. 2018;21:727–30.
- 13. Wu L, Hu B, Zhao B, Liu Y, Yang Y, Zhang L, et al. Circulating microRNA-422a is associated with lymphatic metastasis in lung cancer. Oncotarget. 2017;8:42173–88.
- Meng F, Zhang L. miR-183-5p functions as a tumor suppressor in lung cancer through PIK3CA inhibition. Exp Cell Res. 2019;374:315–22.
- Yang Y, Ding L, Hu Q, Xia J, Sun J, Wang X, et al. MicroRNA-218 functions as a tumor suppressor in lung cancer by targeting IL-6/STAT3 and negatively correlates with poor prognosis. Mol Cancer. 2017;16:141.
- Iwamoto N, Fukui S, Takatani A, Shimizu T, Umeda M, Nishino A, et al. Osteogenic differentiation of fibroblast-like synovial cells in rheumatoid arthritis is induced by microRNA-218 through a ROBO/slit pathway. Arthritis Res Ther. 2018;20:189.
- Jin X, Liu X, Zhang Z, Guan Y. IncRNA CCAT1 acts as a microRNA-218 sponge to increase Gefitinib resistance in NSCLC by targeting HOXA1. Mol Ther Nucleic Acids. 2020;19:1266–75.
- Koritzinsky EH, Street JM, Star RA, Yuen PS. Quantification of Exosomes. J Cell Physiol. 2017;232:1587–90.
- Gilligan KE, Dwyer RM. Engineering Exosomes for cancer therapy. Int J Mol Sci. 2017;18:1122.

- Qu Y, Zhang Q, Cai X, Li F, Ma Z, Xu M, et al. Exosomes derived from miR-181-5p-modified adipose-derived mesenchymal stem cells prevent liver fibrosis via autophagy activation. J Cell Mol Med. 2017;21:2491–502.
- Tang Y, Cui Y, Li Z, Jiao Z, Zhang Y, He Y, et al. Radiation-induced miR-208a increases the proliferation and radioresistance by targeting p21 in human lung cancer cells. J Exp Clin Cancer Res. 2016;35:7.
- 22. Issue information-declaration of Helsinki. J Bone Miner Res. 2018;33: BM i–BM ii.
- Livak KJ, Schmittgen TD. Analysis of relative gene expression data using real-time quantitative PCR and the 2(-Delta Delta C [T]) method. Methods. 2001;25:402–8.
- Storch K, Dickreuter E, Artati A, Adamski J, Cordes N. BEMER electromagnetic field therapy reduces cancer cell radioresistance by enhanced ROS formation and induced DNA damage. PLoS One. 2016;11:e0167931.
- Baskar R, Lee KA, Yeo R, Yeoh KW. Cancer and radiation therapy: current advances and future directions. Int J Med Sci. 2012;9:193–9.
- Tommelein J, De Vlieghere E, Verset L, Melsens E, Leenders J, Descamps B, et al. Radiotherapy-activated cancer-associated fibroblasts promote tumor progression through paracrine IGF1R activation. Cancer Res. 2018;78:659–70.
- Zhu Y, Wang C, Becker SA, Hurst K, Nogueira LM, Findlay VJ, et al. miR-145 antagonizes SNAI1-mediated stemness and radiation resistance in colorectal cancer. Mol Ther. 2018;26:744–54.
- Yin H, Ma J, Chen L, Piao S, Zhang Y, Zhang S, et al. MiR-99a enhances the radiation sensitivity of non-small cell lung cancer by targeting mTOR. Cell Physiol Biochem. 2018;46:471–81.
- Yang Q, Li J, Hu Y, Tang X, Yu L, Dong L, et al. MiR-218-5p suppresses the killing effect of natural killer cell to lung adenocarcinoma by targeting SHMT1. Yonsei Med J. 2019;60:500–8.
- Labbé M, Hoey C, Ray J, Potiron V, Supiot S, Liu SK, et al. micro-RNAs identified in prostate cancer: correlative studies on response to ionizing radiation. Mol Cancer. 2020;19:1–18.
- Li Y, Wang P, Wu LL, Yan J, Pang XY, Liu SJ. miR-26a-5p inhibit gastric cancer cell proliferation and invasion through mediated Wnt5a. Onco Targets Ther. 2020;13:2537–50.
- Xu B, Wang C, Wang YL, Chen SQ, Wu JP, Zhu WD, et al. miR-143 inhibits renal cell carcinoma cells metastatic potential by suppressing ABL2. Kaohsiung J Med Sci. 2020;36:592–8.
- Pannunzio NR, Watanabe G, Lieber MR. Nonhomologous DNA endjoining for repair of DNA double-strand breaks. J Biol Chem. 2018; 293:10512–23.
- Brand TM, Iida M, Luthar N, Starr MM, Huppert EJ, Wheeler DL. Nuclear EGFR as a molecular target in cancer. Radiother Oncol. 2013; 108:370–7.

How to cite this article: Chen X, Xu Y, Jiang L, Tan Q. miRNA-218-5p increases cell sensitivity by inhibiting PRKDC activity in radiation-resistant lung carcinoma cells. *Thorac Cancer*. 2021;12:1549–1557. https://doi.org/10.1111/1759-7714.13939