

Draft Genome Sequence of a Thermophilic Desulfurization Bacterium, *Geobacillus thermoglucosidasius* Strain W-2

Lin Zhu,^a Mingchang Li,^a Shuyi Guo,^a Wei Wang^{a,b}

Key Laboratory of Molecular Microbiology and Technology, Ministry of Education, TEDA Institute of Biological Sciences and Biotechnology, Nankai University, TEDA, Tianjin, People's Republic of China^a; Tianjin Key Laboratory of Microbial Functional Genomics, TEDA, Tianjin, People's Republic of China^b

***Geobacillus thermoglucosidasius* strain W-2 is a thermophilic bacterium isolated from a deep-subsurface oil reservoir in northern China, which is capable of degrading organosulfur compounds. Here, we report the draft genome sequence of *G. thermoglucosidasius* strain W-2, which may help to elucidate the genetic basis of biodegradation of organosulfur pollutants under heated conditions.**

Received 13 June 2016 Accepted 15 June 2016 Published 4 August 2016

Citation Zhu L, Li M, Guo S, Wang W. 2016. Draft genome sequence of a thermophilic desulfurization bacterium, *Geobacillus thermoglucosidasius* strain W-2. *Genome Announc* 4(4):e00793-16. doi:10.1128/genomeA.00793-16.

Copyright © 2016 Zhu et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Wei Wang, nkweiwang@nankai.edu.cn.

Geobacillus thermoglucosidasius strain W-2 was isolated from a deep-subsurface oil reservoir in northern China, which can degrade organosulfur compounds. Sulfur presenting in fuels leads to SO₂ emission during combustion, which causes not only serious air pollution, but also metal catalysts poisoning. Benzothio-phenene (BT) and dibenzothiophene (DBT) are the most widely studied heterocyclic sulfur compounds with respect to its susceptibility to microbial desulfurization (1, 2). To date, two major DBT biodesulfurization pathways (“Kodama” and “4S”) have been widely characterized (3–5), and a BT biodesulfurization pathway has been newly identified in a *Gordonia terrae* strain C-6 (6). In addition, alkanesulfonates are the major alkyl sulfur-containing compounds and the desulfonation mechanism has been investigated (7). However, the previously reported pathways for degrading BT, DBT and alkanesulfonates were all found in mesophilic bacteria. We report the draft genome sequence of strain W-2, which may provide further insights into genetic information of a thermophilic mechanism of biodesulfurization.

The genome sequencing of strain W-2 was carried out using the Illumina HiSeq 2500 platform at the Majorbio Bio-Pharm Technology Co., Ltd. (Shanghai, China), and Illumina paired-end (PE) libraries were constructed. *de novo* assembly was performed by using SOAPdenovo (version 2.04) and GapCloser (version 1.12). The final genome draft of strain W-2 contains 51 contigs, with a total size of 3,894,555 bp and an average G+C content of 43.34%. The average contig length was 76,664 bp, with the largest contig being 560,848 bp. Gene prediction and annotation were performed as described previously (8). As a result, 3,935 protein-encoding genes, one rRNA operon, and 76 tRNA genes for all 20 amino acids were predicted.

Genes encoding for three putative alkanesulfonate monooxygenases, seven putative sulfonate ABC transporters, and two putative sulfate permeases were identified in the draft genome. They were hypothesized to be responsible for organosulfur compounds degradation (9–11). In addition, we also found some genes that encode nitronate monooxygenases, which catalyze oxidative deni-

trification of nitroalkanes to carbonyl compounds and nitrites (12). It may be the first time that the two groups of enzymes responsible for biodesulfurization and bidenitrification pathways have been found in thermophilic bacteria. The functions and mechanisms of the two biodegradation pathways in strain W-2 are under investigation. As thermophilic enzymes offer major biotechnological advantages over mesophilic enzymes (13), the draft genome of strain W-2 may provide an excellent platform for further improvement of this organism for bioremediation and other biotechnological applications at elevated temperature.

Accession number(s). The whole-genome shotgun project of *G. thermoglucosidasius* strain W-2 has been deposited at DDBJ/EMBL/GenBank under the accession number **LXMA00000000**. The version described in this paper is the first version, LXMA01000000.

ACKNOWLEDGMENTS

This work was supported by the National Natural Science Foundation of China (grants 31570119, 31370121, and 31070078) and by the Tianjin Municipal Science and Technology Committee (grants 15JCZDJC32400 and 11JCZDJC16100).

REFERENCES

- McFarland BL, Boron DJ, Deever W, Meyer JA, Johnson AR, Atlas RM. 1998. Biocatalytic sulfur removal from fuels: applicability for producing low sulfur gasoline. *Crit Rev Microbiol* 24:99–147. <http://dx.doi.org/10.1080/10408419891294208>.
- Kurita S, Endo T, Nakamura H, Yagi T, Tamiya N. 1971. Decomposition of some organic sulfur compounds in petroleum by anaerobic bacteria. *J Gen Appl Microbiol* 17:185–198. <http://dx.doi.org/10.2323/jgam.17.185>.
- Denome SA, Olson ES, Young KD. 1993. Identification and cloning of genes involved in specific desulfurization of dibenzothiophene by *Rhodococcus* sp. strain IGTS8. *Appl Environ Microbiol* 59:2837–2843.
- Denome SA, Oldfield C, Nash LJ, Young KD. 1994. Characterization of the desulfurization genes from *Rhodococcus* sp. strain IGTS8. *J Bacteriol* 176:6707–6716.
- Kodama K, Umehara K, Shimizu K, Nakatani S, Minoda Y, Yamada K.

1973. Identification of microbial products from dibenzothiophene and its proposed oxidation pathway. *Agric Biol Chem* 37:45–50.
6. Wang W, Ma T, Lian K, Zhang Y, Tian H, Ji K, Li G. 2013. Genetic analysis of benzothiophene biodesulfurization pathway of *Gordonia terrae* strain C-6. *PLoS One* 8:e84386. <http://dx.doi.org/10.1371/journal.pone.0084386>.
 7. Van der Ploeg JR, Iwanicka-Nowicka R, Bykowski T, Hryniewicz MM, Leisinger T. 1999. The *Escherichia coli* *ssuEADCB* gene cluster is required for the utilization of sulfur from aliphatic sulfonates and is regulated by the transcriptional activator Cbl. *J Biol Chem* 274:29358–29365. <http://dx.doi.org/10.1074/jbc.274.41.29358>.
 8. Yao N, Ren Y, Wang W. 2013. Genome sequence of a thermophilic Bacillus, *Geobacillus thermodenitrificans* DSM465. *Genome Announc* 1(6):e01046-13. <http://dx.doi.org/10.1128/genomeA.01046-13>.
 9. Van Hamme JD, Bottos EM, Bilbey NJ, Brewer SE. 2013. Genomic and proteomic characterization of *Gordonia* sp. NB4-1Y in relation to 6:2 fluorotelomer sulfonate biodegradation. *Microbiology* 159:1618–1628. <http://dx.doi.org/10.1099/mic.0.068932-0>.
 10. Erwin KN, Nakano S, Zuber P. 2005. Sulfate-dependent repression of genes that function in organosulfur metabolism in *Bacillus subtilis* requires Spx. *J Bacteriol* 187:4042–4049. <http://dx.doi.org/10.1128/JB.187.12.4042-4049.2005>.
 11. Ellis HR. 2011. Mechanism for sulfur acquisition by the alkanesulfonate monooxygenase system. *Bioorg Chem* 39:178–184. <http://dx.doi.org/10.1016/j.bioorg.2011.08.001>.
 12. Ha JY, Min JY, Lee SK, Kim HS, Kim DJ, Kim KH, Lee HH, Kim HK, Yoon HJ, Suh SW. 2006. Crystal structure of 2-nitropropane dioxygenase complexed with FMN and substrate. Identification of the catalytic base. *J Biol Chem* 281:18660–18667. <http://dx.doi.org/10.1074/jbc.M601658200>.
 13. Feng L, Wang W, Cheng J, Ren Y, Zhao G, Gao C, Tang Y, Liu X, Han W, Peng X, Liu R, Wang L. 2007. Genome and proteome of long-chain alkane degrading *Geobacillus thermodenitrificans* NG80-2 isolated from a deep-subsurface oil reservoir. *Proc Natl Acad Sci U S A* 104:5602–5607. <http://dx.doi.org/10.1073/pnas.0609650104>.