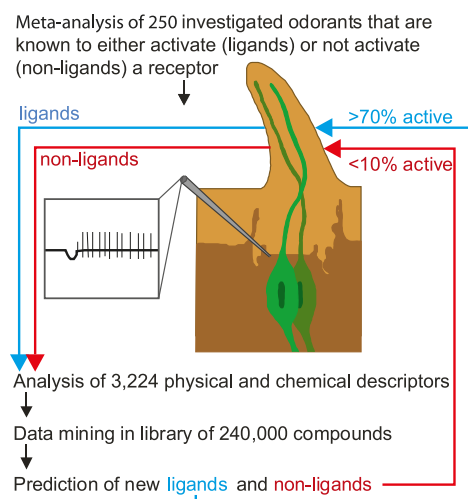




activate at least one receptor. Although this is a tiny number compared with the number of odorants that flies are usually exposed to, Boyle, McInally and Ray were able to gain fresh insights into the receptor-odorant interactions by performing a highly detailed meta-analysis on these 251 odorants to identify the properties that cause an odorant to target a particular receptor (**Figure 1**). In addition to the 'usual suspects' of molecular properties (e.g., whether the odorant is an alcohol, an ester or an aldehyde), they took into account some 3,224 physical and/or chemical properties of the odorants, including obvious properties like molecular weight and three-dimensional structure, and less obvious properties like the 'eigenvalue sum from electronegativity weighted distance matrix'.



**Figure 1.** Predicting odorant-receptor interactions.

Boyle et al. performed a meta-analysis of 250 odorants and 51 receptors and developed an algorithm (based on some 3,224 physical and chemical properties of the odorants) to predict whether a given odorant will interact with a given receptor. This algorithm was then used to 'mine' a library of 240,000 compounds and identify ligands (blue line) and non-ligands (red line) for nine receptors. Experiments were performed with 141 compounds (11–23 per receptor): 71% of the compounds that were predicted to be ligands were found to interact with the relevant receptor, and less than 10% of the compounds that were predicted to be non-ligands were found to interact. The illustration shows an insect sensillum housing two olfactory receptor neurons (one pale green, the other dark green), each with a cell body and a nucleus, and a dendrite that extends into the tip of the sensillum. The tip is filled with a fluid called the sensillum lymph (pale brown) that is excreted by trichogen cells (dark brown). The expanded detail shows the neuronal response to a ligand as measured in the single sensillum recordings performed by Boyle et al.

This approach was pioneered by groups at Goethe University in Frankfurt (**Schmucker et al., 2007**) and the Weizmann Institute (**Haddad et al., 2008**). However, instead of analysing all the receptors and all the physical and chemical properties, the Riverside researchers used an algorithm that allowed the most critical properties for each receptor to be identified. Next they screened a list of more of 240,000 odorants to find those that they expected to interact with nine different receptors. Finally, they tested these predictions in experiments: Their predictions were correct more than 70% of the time, compared with a success rate of just 10% for odorants chosen at random. Hence, although odorants do not follow any linear rules like light and sound, we can still use their physical and chemical properties to predict whether an odorant interacts with a specific receptor and later, we hope, be able to understand why it interacts.

These results will be of interest beyond a narrow group of specialists. According to the United Nations Food and Agriculture Organization, insects and insect-spread diseases are responsible for an estimated 20–40% of world-wide crop production being lost every year. Furthermore, malaria and dengue fever, which are both spread by mosquitoes, kill more than 1 million people every year (and infect another 250 million). As insects typically use olfactory cues to find new hosts, a better understanding of odorant-receptor interactions promises substantial improvements for human food supply and health.

**Markus Knaden** is at the Max Planck Institute for Chemical Ecology, Jena, Germany  
mknaden@ice.mpg.de

**Bill Hansson** is at the Max Planck Institute for Chemical Ecology, Jena, Germany  
hansson@ice.mpg.de

**Competing interests:** The authors declare that no competing interests exist.

**Published** 15 October 2013

## References

- Boyle S, McInally S, Ray A.** 2013. Expanding the olfactory code by in silico decoding of odor-receptor chemical space. *eLife* **2**:e01120. doi: [10.7554/eLife.01120](https://doi.org/10.7554/eLife.01120).
- Buck L, Axel R.** 1991. A novel multigene family may encode odorant receptors. *Cell* **91**:175–87. doi: [10.1016/0092-8674\(91\)90418-X](https://doi.org/10.1016/0092-8674(91)90418-X).
- Haddad RR, Khan R, Takahashi YK, Mori K, Harel D, Sobel N.** 2008. A metric for odorant comparison. *Nat Meth* **5**:425–9. doi: [10.1038/nmeth.1197](https://doi.org/10.1038/nmeth.1197).
- Hallem EA, Carlson JR.** 2006. Coding of odors by a receptor repertoire. *Cell* **125**:143–60. doi: [10.1016/j.cell.2006.01.050](https://doi.org/10.1016/j.cell.2006.01.050).
- Kauer JS, White J.** 2001. Imaging and coding in the olfactory system. *Annu Rev Neurosci* **24**:963–79. doi: [10.1146/annurev.neuro.24.1.963](https://doi.org/10.1146/annurev.neuro.24.1.963).

- Masterto B**, Heffner H, Ravizza R. 1969. The evolution of human hearing. *J Acoust Soc Am* **45**:966–75. doi: [10.1121/1.1911574](https://doi.org/10.1121/1.1911574).
- Nakagawa T**, Sakurai T, Nishioka T, Touhara K. 2005. Insect sex-pheromone signals mediated by specific combinations of olfactory receptors. *Science* **307**:1638–42. doi: [10.1126/science.1106267](https://doi.org/10.1126/science.1106267).
- Schmuker M**, de Bruyne M, Haehnel M, Schneider G. 2007. Predicting olfactory receptor neuron responses from odorant structure. *Chem Cent Jour* **1**:11. doi: [10.1186/1752-153X-1-11](https://doi.org/10.1186/1752-153X-1-11).
- Schnapf JL**, Kraft TW, Baylor DA. 1987. Spectral sensitivity of human cone photoreceptors. *Nature* **325**:439–41. doi: [10.1038/325439a0](https://doi.org/10.1038/325439a0).
- Stensmyr MC**, Dweck HKM, Farhan A, Ibba I, Strutz A, Mukunda L, et al. 2012. A conserved dedicated olfactory circuit for detecting harmful microbes in *Drosophila*. *Cell* **151**:1345–57. doi: [10.1016/j.cell.2012.09.046](https://doi.org/10.1016/j.cell.2012.09.046).
- Vosshall LB**. 2000. Olfaction in *Drosophila*. *Curr Opin Neurobiol* **10**:498–503. doi: [10.1016/S0959-4388\(00\)00111-2](https://doi.org/10.1016/S0959-4388(00)00111-2).