



Article

Novel Iron Parameters in Patients with Type 2 Diabetes Mellitus in Relation to Kidney Function

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Abstract: Background/aims: Anemia of chronic disease is a common feature in diabetes and chronic kidney disease. Heparin is the key element involved in iron metabolism; however, studies on new indices of iron status are still ongoing. The aim of the study was to assess novel iron parameters in patients with type 2 diabetes mellitus in relation to kidney function. Methods: The study included 80 type 2 diabetic patients and 23 healthy volunteers. Standard laboratory measurements were used to measure the iron status, complete blood count, creatinine, the estimated glomerular filtration rate (eGFR), serum lipids, and brain natriuretic peptides (BNPs). Commercially available kits were used to measure hepcidin-25, the soluble transferrin receptor (sTfR), growth differentiation factor-15 (GDF-15), and hypoxia-inducible factor-1 alpha. Results: Anemia was present in 65% of the studied patients. The control group was found to have significantly higher hepcidin, sTfR, and GDF-15, and lower hemoglobin and iron. When compared with patients with eGFR values ≥ 60 mL/min/1.73 m² and < 60 mL/min/1.73 m², we found that patients with higher eGFR had higher hemoglobin, ferritin, and HIF-1 alpha, lower BNP, and were younger. We found that levels of HIF-1 alpha are negligible in the studied population and were related to age only in patients with eGFR values ≥ 60 mL/min/1.73 m². Conclusion: A comprehensive assessment of iron status is rarely performed. Novel biomarkers of iron metabolism are not generally related to kidney function. Whether the assessment of HIF-1 alpha would be a marker of efficient anemia therapy with HIF-prolyl hydroxylase inhibitors is still a matter for further study.

Keywords: chronic kidney disease; diabetes mellitus; iron metabolism; hepcidin

1. Introduction

Iron is one of the most common elements on earth and is essential for the proper growth and development of organisms. Imbalance in its quantity can lead to many pathologies. Due to its ability to occur in two forms of oxidation, it participates in the vital process of binding and distributing oxygen, metabolism, cell growth, and proliferation. However, with its ability to accept and transfer electrons, iron can cause severe oxidative stress and tissue damage [1]. Iron homeostasis in the body is maintained by regulators acting at the systemic and cellular levels. The body's ability to excrete iron is limited; therefore, its amount is particularly controlled at the level of its absorption from the gastrointestinal tract [2]. Heparin is the systemic regulator of iron metabolism. It is produced in the liver and depends on the amount of iron in the body [3], the presence of inflammation [4], hypoxia [5,6], and erythropoiesis [7].

Research has disclosed the reciprocal influence between diabetes and iron metabolism. Many of the pathways responsible for iron homeostasis in the body are disrupted in the hyperglycemic environment. On the other hand, excess iron has an effect on insulin action and secretion. Too much iron in the body promotes the development of glucose intolerance, type 2 diabetes [8–14], and gestational diabetes [15,16]. The mechanisms through which iron contributes to the pathogenesis of diabetes are not yet fully understood, but appear to be multifactorial and may vary depending on the cause of iron overload and its tissue distribution [1]. The relationship between excess iron and the pathogenesis of type 2 diabetes has been observed in patients suffering from haemochromatosis and thalassemia [17,18]. On the other hand, lowering iron concentration may be a factor involved in reducing the risk of developing type 2 diabetes [19]. Phlebotomy and chelation improved glycemic control [20–22]. Reducing iron stores through frequent blood donation in healthy volunteers has also increased insulin sensitivity [23].

It is known that the hormone hepcidin is responsible for the regulation of iron metabolism. In addition to iron excess, hepcidin expression is influenced by inflammation. Hyperglycemia occurring in diabetes stimulates the synthesis of pro-inflammatory cytokines, thus leading to the development of chronic inflammation. Clinical trials have shown higher hepcidin levels in diabetic patients, which has correlated with concentration levels of IL-6 and ferritin [24,25]. Pro-inflammatory cytokines increase the expression of hepcidin, which stops the transport of iron from enterocytes to the blood and inhibits its release from macrophages, causing a decrease in its concentration in plasma and its accumulation in cells. The accumulation of iron in adipocytes results in reduced synthesis of adiponectin, thus increasing insulin resistance [26].

Diabetes mellitus is currently a global epidemic. The estimated number of people with diabetes worldwide is 171 million and it will reach 334 million in 2023 [27]. The main chronic complication of diabetes is diabetic kidney disease (DKD). DKD is an important and growing epidemiological and clinical problem. It is the most common cause of chronic kidney disease (CKD) and end-stage renal disease (ESRD) [28–30].

Anemia is one of the most commonly observed complications of chronic kidney disease CKD. The risk of anemia increases with the deterioration of renal function from approximately 27% in CKD stage 1–76% in stage 5 [31–33]. Moreover, 90% of dialysis patients had anemia [34,35]. DKD patients with anemia have a very high risk of cardiovascular diseases (CVD) and a much higher risk of death compared to people with DKD without anemia [36–38]. Anemia in DKD patients develops early, even before GFR is significantly lowered, and is more severe than in patients with non-diabetic kidney disease [39,40].

The pathogenesis of anemia in CKD is a multifactorial process. The most common causes include a decrease in erythropoietin (EPO) synthesis, iron deficiency (IDA), and chronic inflammation. In many patients with CKD, anemia is developed in the form of anemia of chronic disease, resulting from a chronic inflammatory process. The increased level of inflammatory cytokines leads to an increase in hepcidin expression, thus leading to the development of functional iron deficiency—a condition in which the body's iron stores are normal or even increased, but the release of iron from them is not fast enough to ensure normal erythropoiesis. Serum saturation is low, and the concentration of ferritin is normal or high.

As in the published literature, data on iron parameters are scarce; therefore, the aim of the study was to assess novel iron parameters in patients with type 2 diabetes mellitus in relation to kidney function.

2. Materials and Methods

The study group consisted of 80 patients. All patients were hospitalized in the Teaching Hospital of Invasive Cardiology at the Medical University of Białystok in Poland. The median age of patients in the study group was 70 years (min 42; max 88). There were 58 men (72.5%) and 22 women (27.5%) among the study group. All patients in the study group suffered from type 2 diabetes mellitus. Diabetes was defined by the current

guidelines [41] or by the use of insulin or oral hypoglycemic agents. The control group consisted of 23 healthy volunteers (16 females) to obtain normal ranges for biomarkers.

The patients gave written informed consent for this study during hospitalization after receiving comprehensive information and learning about the purpose of the study. Data regarding the course of type 2 diabetes, chronic kidney disease, other chronic diseases, and risk factors were obtained from medical records. Diabetes mellitus in these patients was treated with biguanides, sulfonylurea derivatives, incretin drugs, sodium-glucose cotransporter (SGLT-2) inhibitors, and insulin. The treatment was conducted both in monotherapy as well as in combination therapy consisting of drug groups selected individually to the patient. Serum creatinine concentration was determined in all patients in the study group. Chronic kidney disease recognition was made according to KDIGO 2012 Clinical Practice Guideline for the Evaluation and Management of Chronic Kidney Disease [42]. CKD is defined as the presence of kidney damage or an estimated glomerular filtration rate (eGFR) of less than 60 mL/min/1.73 m², persisting for three months or more. The CKD-EPI (Chronic Kidney Disease Epidemiology Collaboration) formula was also used to estimate the glomerular filtration rate [43] in the study group. The control group consisted of 23 healthy volunteers without type 2 diabetes and chronic kidney disease or other chronic diseases. Basic laboratory results (complete blood count, inflammatory parameters, electrolytes) and hepcidin, HIF1-alpha (Hypoxia-Inducible Factor 1-alpha), GDF-15 (Growth Differentiation Factor 15), sTfR (Soluble Transferrin Receptor), iron, and ferritin concentrations were performed in both the study and control groups. The study patients were divided according to the stage of CKD. eGFR < 60 mL/min/1.73 m² was present in 21 patients, eGFR < 30 mL/min/1.73 m² was present in 2 patients, and eGFR < 15 mL/min/1.73 m² was observed in 1 patient. The patients were divided into two groups depending on the values of eGFR. Group 1 included patients with an eGFR < 60 mL/min/1.73 m², and group 2 included patients with an eGFR ≥ 60 mL/min/1.73 m².

The studied parameters were analyzed in relation to pharmacological treatment, the results of additional examinations, and coexisting diseases. Statistical analysis was performed using the Statistica 13.3 PL program. The aim of this study was to evaluate the relationship of specific factors affecting iron metabolism in diabetic patients (i.e., hepcidin, soluble transferrin receptor- sTfR, growth and differentiation factor 15-GDF-15, hypoxia-inducible factor 1 alpha- HIF-1 alpha with routinely determined iron markers, and kidney function). The approval of the Bioethics Committee of the Medical University of Białystok was obtained (NO: R-I-002/7/2018).

Statistical Analysis

We used the Shapiro–Wilk test to test for normality in the variable distributions. As all continuous variables were not normally distributed, we presented them as median and interquartile range values. Categorical variables were presented as the number of cases and percentages. The Mann–Whitney U tests and T-tests were used to determine the statistical significance of differences between variables. The correlations between variables were determined using multivariable Pearson’s correlations. Data are presented as rank correlations (r) with *p*-values. The threshold of statistical significance for all tests was set at *p* < 0.05. All analyses were performed using MS Excel (Microsoft, 2020, version 16.40, Redmond, WA, USA) and XL Stat (Addinsoft, 2020, version 2020.03.01, New York, NY, USA).

3. Results

Out of 80 patients, 24 patients were diagnosed with reduced renal function (their eGFR value was <60 mL/min/1.73 m²; 30%), whereas the remaining 56 patients (70%) showed normal glomerular filtration rates. Table 1 shows the characteristics of the studied and control populations. Anemia was found in 65% of the studied population according to the WHO definition (hemoglobin < 13 mg/dL for men and <12 mg/dL for women) [44].

Table 1. Characteristics of the studied and control populations.

	Study Participants (N = 80)	Control Cohort (N = 23)	<i>p</i> for the Comparison between Study Participants and the Control Cohort
Age, years; Median (IQR)	70 (11)	51 (8.6)	<0.001
eGFR by CKD-EPI, mL/min/1.73 m ² ; Mean (SD)	70.3 (20.8)	85.9 (15.3)	<0.001
Hemoglobin, g/dL; Mean (SD)	12.9 (1.3)	13.1 (1.4)	<0.001
Hepcidin, mg/mL; Median (IQR)	4.6 (5.3)	2.5 (1.2)	<0.001
sTfR, nmol/L; Mean (SD)	23.7 (7)	8.6 (2)	<0.001
GDF-15, pg/mL; Median (IQR)	1514 (1232)	584 (324)	<0.001
HIF-1 alpha, mg/mL; Median (IQR)	0 (0)	102.6 (112)	0.09
Fe, µg/dL; Mean (SD)	78.7 (24.9)	95.6 (24.7)	0.005
Ferritin, µg/L; Median (IQR)	99.6 (92.5)	104 (90.2)	0.57

The study group was found to have significantly higher hepcidin, sTfR, and GDF-15 levels, and lower hemoglobin and iron levels. When compared with patients with eGFR values ≥ 60 mL/min/1.73 m² and <60 mL/min/1.73 m², we found that patients with higher eGFR levels had higher hemoglobin, ferritin, and HIF-1 alpha levels; lower BNP; and were younger (Table 2).

Table 2. Comparisons between patients with and without chronic kidney disease.

	Study Participants (N = 80)	eGFR < 60 mL/min/1.72 m ² (N = 24)	eGFR > 60 mL/min/1.72 m ² (N = 56)	<i>p</i>
Age, years; Median (IQR)	70 (11)	73.5 (11.3)	69 (9.3)	0.03
Male; % (N)	73 (58)	75.0 (18)	71.4 (40)	0.75
BMI, kg/m ² ; Median (IQR)	30.6 (4.9)	33.3 (5.7)	30.4 (4.2)	0.08
Atrial fibrillation; % (N)	26 (21)	29 (7)	25 (14)	0.70
Hypertension; % (N)	91.3 (73)	92 (22)	91 (51)	0.93
Chronic heart failure; % (N)	34 (27)	33.3 (8)	34 (19)	0.96
Chronic coronary syndrome; % (N)	80 (64)	75 (18)	82.1 (46)	0.46
Acute coronary syndrome; % (N)	40 (32)	45.8 (11)	37.5 (21)	0.44
MAP, mmHg; Mean (SD)	94.3 (18.4)	96.4 (12.9)	93.4 (20.2)	0.49
Heart rate, beats per minute; Mean (SD)	73 (12.6)	71 (11.8)	70 (13)	0.98
Low-density lipoprotein cholesterol, mg/dL; Mean (SD)	86.4 (32.2)	88.1 (38)	82.2 (27.5)	0.46
High-density lipoprotein cholesterol, mg/dL; Mean (SD)	41.8 (11.4)	35 (15.5)	43.5 (11.4)	0.04
TG, mg/dL; Mean (SD)	143 (56.6)	154.5 (53.5)	126 (61)	0.24
eGFR by CKD-EPI, mL/min/1.73 m ² ; Mean (SD)	70.3 (20.8)	50 (18)	81 (21.3)	<0.001
RBC 10 ⁶ /mm ³ ; Mean (SD)	4.3 (0.5)	4.2 (0.63)	4.3 (0.42)	0.09
Hemoglobin, g/dL; Mean (SD)	12.9 (1.3)	12.5 (1.5)	13.1 (1.3)	<0.001
BNP, pg/mL; Median (IQR)	246.3 (406.5)	454 (387.7)	139 (186)	<0.001
Hepcidin, mg/mL; Median (IQR)	4.6 (5.3)	4.6 (4.9)	4.6 (5.4)	0.93
sTfR, nmol/L; Mean (SD)	23.7 (7)	26.3 (5.3)	23 (8.7)	0.08
GDF-15, pg/mL; Median (IQR)	1514 (1232)	1906 (1485)	1351 (1013)	0.1
HIF-1 alpha, mg/mL; Median (IQR)	0 (0)	0 (11.7)	0 (0)	0.02
Fe, µg/dL; Mean (SD)	78.7 (24.9)	75.5 (29.5)	78 (33.8)	0.28
Ferritin, µg/L; Median (IQR)	99.6 (92.5)	78 (49.6)	117 (108)	0.03

Table 2. Cont.

	Study Participants (N = 80)	eGFR < 60 mL/min/1.72 m ² (N = 24)	eGFR > 60 mL/min/1.72 m ² (N = 56)	p
Diuretics medication prescribed at discharge; % (N)	77.5 (62)	83.3 (20)	75 (42)	0.36
Calcium channel blockers prescribed at discharge; % (N)	30 (24)	33.3 (8)	32 (18)	0.54
ACE medication prescribed at discharge; % (N)	84 (67)	88 (21)	82 (46)	0.47
BB medication prescribed at discharge; % (N)	88.8 (71)	96 (23)	86 (48)	0.09
Statin medication prescribed at discharge; % (N)	77.5 (62)	83 (20)	75 (42)	0.36

In the studied group, there were statistically significant correlations between age and hemoglobin ($r = -0.23, p < 0.05$), eGFR ($r = -0.36, p < 0.001$), sTfR ($r = -0.29, p < 0.01$), and GDF-15 ($r = -0.35, p < 0.001$); between hemoglobin and eGFR ($r = 0.37, p < 0.001$), BNP ($r = -0.39, p < 0.001$), hepcidin-25 ($r = 0.32, p < 0.01$), and ferritin ($r = 0.30, p < 0.01$); between eGFR and BNP ($r = -0.45, p < 0.001$), sTfR ($r = -0.31, p < 0.01$), and GDF-15 ($r = -0.24, p < 0.05$); between hepcidin-25 and sTfR ($r = -0.22, p < 0.05$), GDF-15 ($r = 0.031, p < 0.01$), and ferritin ($r = 0.47, p < 0.001$); and between sTfR and ferritin ($r = -0.29, p < 0.05$). All the correlations for the studied subgroups are presented in Table 3 (significant correlations are shown in bold). In the linear regression model, hepcidin was related to iron parameters (Table 4).

Table 3. Multivariable Pearson correlations between physical variables.

Hepcidin mg/mL 0.04 (p = 0.74)	-0.03 (p = 0.9)	0.09 (p = 0.69)	0.34 (p = 0.1)	0.25 (p = 0.24)	-0.01 (p = 0.95)	0.3 (p = 0.16)	0.26 (p = 0.22)	-0.29 (p = 0.17)
	Age, years (p = 0.74)	-0.22 (p = 0.31)	-0.02 (p = 0.93)	0.19 (p = 0.37)	-0.03 (p = 0.9)	0.01 (p = 0.99)	0.12 (p = 0.58)	0.22 (p = 0.3)
		eGFR, mL/min/1.73 m ² (p = 0.4)	0.44 (p = 0.03)	-0.11 (p = 0.62)	-0.13 (p = 0.55)	-0.11 (p = 0.61)	-0.09 (p = 0.66)	-0.28 (p = 0.19)
			HGB, g/dL (p = 0.01)	0.13 (p = 0.55)	0.01 (p = 0.99)	-0.12 (p = 0.59)	-0.16 (p = 0.46)	-0.15 (p = 0.47)
			0.31 (p = 0.02)	Ferritin, µg/L (p = 0.46)	-0.08 (p = 0.69)	0.25 (p = 0.24)	0.11 (p = 0.06)	0.05 (p = 0.82)
			0.17 (p = 0.21)	0.27 (p = 0.04)	Fe, µg/dl (p = 0.72)	0.09 (p = 0.66)	-0.07 (p = 0.72)	-0.25 (p = 0.23)
			-0.02 (p = 0.9)	0.12 (p = 0.39)	0.05 (p = 0.7)	HIF-1 alpha pg, mg/mL (p = 0.45)	-0.16 (p = 0.45)	-0.11 (p = 0.61)
			0.01 (p = 0.99)	0.09 (p = 0.53)	0.05 (p = 0.73)	-0.14 (p = 0.31)	GDF-15, pg/mL (p = 0.38)	0.07 (p = 0.74)
			-0.06 (p = 0.65)	-0.29 (p = 0.03)	-0.15 (p = 0.27)	-0.15 (p = 0.28)	0.12 (p = 0.38)	sTfR, nmol/L

Right side r value for group with eGFR < 60 mL/min/1.73 m², left side for group with eGFR ≥ 60 mL/min/1.73 m². Statistically significant correlations are in bold (p < 0.05).

Table 4. Results of linear regression analyses for serum hepcidin concentration.

Variables	β	(95% CI)		<i>p</i>
sTfR	−0.239	−0.431	−0.047	0.015
GDF-15	0.267	0.082	0.453	0.005
HIF-1 alpha	0.109	−0.074	0.292	0.239
Fe	−0.239	−0.416	−0.062	0.009
Ferritin	0.393	0.203	0.582	<0.0001
Hemoglobin	0.335	0.143	0.527	0.001
eGFR by CKD-EPI	−0.159	−0.368	0.050	0.134
Age	0.162	−0.039	0.362	0.113

4. Discussion

In our study, we assessed—for the first time—several iron parameters, including HIF-1 alpha, in relation to routinely determined iron markers. However, we could not find statistically significant differences in iron parameters between patients with eGFR values ≥ 60 mL/min/1.73 m² and <60 mL/min/1.73 m², except for ferritin and HIF-1 alpha. For the study group, hepcidin was significantly related to iron parameters in the linear regression model.

In a recent cross-sectional study, Hayder and Kareem [45] described the possible role of resistin in the iron status pathway in patients with non-insulin-dependent diabetes mellitus and ESRD. In their study, they found that among 50 patients with type 2 diabetes mellitus and a mean eGFR level of 80 mL/min/1.73 m², serum hepcidin and ferritin were higher when compared to the control group (where the mean eGFR was 89 mL/min/1.73 m²), while serum iron was significantly lower. The mean ages of the studied and control groups were not provided; it was only stated that volunteers were apparently normal, healthy subjects who more than 40 years old. We are fully aware that age- and sex-matching are difficult—in particular, when the studied group is almost 70 years old, as patients in the hospital typically are. No detailed clinical characteristics of the studied groups were provided. In many of our previous studies in patients with different types of renal replacement therapies, including kidney transplantation, we showed that the hepcidin was elevated in these patients [46–48]. As it was suggested, hepcidin-25 might be an important regulator of iron homeostasis under erythropoiesis in hemodialyzed patients [49]. In addition, elevated hepcidin-25 not only decreases the release of stored iron but also influences erythropoiesis [50]. Thus, alternatively, several studies indicated that hepcidin-25 was not a reliable marker for iron release monitoring and storage in dialyzed patients [51,52]. In our previous study, we reported higher hepcidin in the early stages of CKD [53]. However, in this study, we focused only on diabetic patients. Wagner et al. [54], in a cohort of 249 diabetic patients with CKD of any stage (with a mean age of 67 years and a mean eGFR level of 51 mL/min/1.73 m²), reported that elevated hepcidin was independently associated with ferritin and worsened kidney function (all $p < 0.05$).

Previously, we also reported that in patients ≥ 65 years, serum concentrations of GDF-15 were significantly higher in comparison with the younger group of patients with early stages of kidney disease [55]. GDF-15 is a member of the transforming growth factor- β cytokine superfamily and has been shown to be one of the regulators of iron status. GDF-15 expression is increased through oxidative stress and inflammation [56]. In the present study, we found that GDF-15 was higher in healthy volunteers but there was no difference between patients with eGFR values ≥ 60 mL/min/1.73 m² and <60 mL/min/1.73 m². In addition, in our previous study on 84 patients with CKD in the early stage (with a mean eGFR level of 67.7 ± 16 mL/min/1.73 m², mean age of 69 years, and with 70% females), we found that sTfR was higher than in the control group [57]. sTfR is a dimeric protein present in serum, which is a part of the transferrin receptor after enzymatic cleavage [58]. In the study of Belo et al. [59], sTfR values were higher in hemodialyzed diabetic patients when compared to hemodialyzed non-diabetic patients and were significantly correlated with circulating iron, ferritin, transferrin saturation, hepcidin, MCH, and MCHC. There

were no statistically significant differences in GDF-15 and hepcidin between diabetic and non-diabetic patients who were on hemodialysis in their study. Similar findings were observed in our study in diabetic patients who were not on dialysis.

sTfR reflects the total amount of transferrin receptors present in the body. The majority of the sTfR is found in the bone marrow. Assessing the level of the sTfR is useful to estimate the gross mass of erythroid bone marrow and may help to distinguish between the anemia of chronic diseases and iron deficiency anemia [60]. However, the lack of assay standardization and the wide variety in the cutoff values limits its interpretation in clinical practice [61].

In the published literature, we could not find a comprehensive study in iron metabolism diabetic patients within the early stages of CKD. We also assessed HIF-1 alpha in addition to other iron metabolism markers. HIF is the main transcription factor of hundreds of genes, including the EPO gene expressed in hypoxic tissues, and it adapts to a hypoxic environment [62,63]. HIF also controls the expression of the proteins playing a role in iron metabolism and utilization (the upregulation of transferrin, soluble transferrin receptor 1 (sTfR1), ceruloplasmin, divalent metal transporter 1 (DMT1), duodenal cytochrome b (Dcytb); and the downregulation of hepcidin). Increases in soluble transferrin receptor 1 improve iron availability, given its role as a carrier protein for transferrin required to import iron into the cell [64]. Researchers suggest that hepcidin suppression most likely results from the stimulation of erythropoiesis, as it seems that hepcidin is not a direct transcriptional target of HIF. Nowadays, hypoxia-inducible factor prolyl hydroxylase inhibitors (HIF-PHIs) represent a novel approach to the treatment of anemia in patients with CKD.

We found that levels of HIF-1 alpha are negligible in the studied population and were related to age only in patients with eGFR values ≥ 60 mL/min/1.73 m². Whether the assessment of HIF-1 alpha would be a marker of efficient anemia therapy with HIF-PHIs is still a matter for further study.

Our study was cross-sectional, was completed using a single center, and included 80 patients that were subsequently admitted to the Department. We are aware that our population is relatively small, but for the assessment of several iron parameters, it appears to be sufficient to show the differences in relation to kidney function as well as to healthy controls. Our real-life data characterize everyday clinical practice with a substantial component of uncertainties and possible confounders.

5. Conclusions

In conclusion, a comprehensive assessment of iron status is rarely performed. Novel biomarkers of iron metabolism are not generally related to kidney function. Whether the assessment of HIF-1 alpha would be a marker of efficient anemia therapy with HIF-PHIs is still a matter for further study.

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Conflicts of Interest: The authors declare no conflict of interest.

References

1. Zager, R.A. Parenteral iron compounds: Potent oxidants but mainstays of anemia management in chronic renal disease. *Clin. J. Am. Soc. Nephrol.* **2006**, *1*, S24–S31. [[CrossRef](#)]
2. Hentze, M.W.; Muckenthaler, M.U.; Galy, B.; Camaschella, C. Two to tango: Regulation of mammalian iron metabolism. *Cell* **2010**, *142*, 24–38. [[CrossRef](#)]
3. Mazur, A.; Feillet-Coudray, C.; Romier, B.; Bayle, D.; Gueux, E.; Ruivard, M.; Coudray, C.; Rayssiguier, Y. Dietary iron regulates hepatic hepcidin 1 and 2 mRNAs in mice. *Metabolism* **2003**, *52*, 1229–1231. [[CrossRef](#)]
4. Nemeth, E.; Valore, E.V.; Territo, M.; Schiller, G.; Lichtenstein, A.; Ganz, T. Hepcidin, a putative mediator of anemia of inflammation, is a type II acute-phase protein. *Blood* **2003**, *101*, 2461–2463. [[CrossRef](#)]
5. Shah, Y.M.; Xie, L. Hypoxia-inducible factors link iron homeostasis and erythropoiesis. *Gastroenterology* **2014**, *146*, 630–642. [[CrossRef](#)]
6. Silvestri, L.; Pagani, A.; Camaschella, C. Furin-mediated release of soluble hemojuvelin: A new link between hypoxia and iron homeostasis. *Blood* **2008**, *111*, 924–931. [[CrossRef](#)] [[PubMed](#)]
7. Nicolas, G.; Chauvet, C.; Viatte, L.; Danan, J.L.; Bigard, X.; Devaux, I.; Beaumont, C.; Kahn, A.; Vaulont, S. The gene encoding the iron regulatory peptide hepcidin is regulated by anemia, hypoxia, and inflammation. *J. Clin. Investig.* **2002**, *110*, 1037–1044. [[CrossRef](#)]
8. Medalie, J.H.; Papier, C.M.; Goldbourt, U.; Herman, J.B. Major factors in the development of diabetes mellitus in 10,000 men. *Arch. Intern. Med.* **1975**, *135*, 811–817. [[CrossRef](#)] [[PubMed](#)]
9. Wilson, P.W.; McGee, D.L.; Kannel, W.B. Obesity, very low density lipoproteins and glucose intolerance over fourteen years: The framingham study. *Am. J. Epidemiol.* **1981**, *114*, 697–704. [[CrossRef](#)] [[PubMed](#)]
10. Catalano, C.; Muscelli, E.; Quiñones, A.; Baldi, S.; Ciociaro, D.; Seghieri, G.; Ferrannini, E. Reciprocal association between insulin sensitivity and the hematocrit in man (Abstract). *Eur. J. Clin. Investig.* **1997**, *27*, 634–637. [[CrossRef](#)] [[PubMed](#)]
11. Wannamethee, S.G.; Perry, I.J.; Shaper, A.G. Hematocrit and risk of NIDDM. *Diabetes* **1996**, *45*, 576–579. [[CrossRef](#)] [[PubMed](#)]
12. Salonen, J.T.; Tuomainen, T.-P.; Nyyssönen, K.; Lakka, H.-M.; Punnonen, K. Relation between iron stores and noninsulin dependent diabetes in men: Case-control study. *BMJ* **1998**, *317*, 727–730. [[CrossRef](#)] [[PubMed](#)]
13. Ford, E.S.; Cogswell, M.E. Diabetes and serum ferritin concentration among U.S. adults. *Diabetes Care* **1999**, *22*, 1978–1983. [[CrossRef](#)] [[PubMed](#)]
14. Barbieri, M.; Ragno, E.; Benvenuti, E.; Zito, G.A.; Corsi, A.; Ferrucci, L.; Paolisso, G. New aspects of the insulin resistance syndrome: Impact on haematological parameters. *Diabetologia* **2001**, *44*, 1232–1237. [[CrossRef](#)]
15. Lao, T.T.; Tam, K.-F.F. Maternal serum ferritin and gestational impaired glucose tolerance. *Diabetes Care* **1997**, *20*, 1368–1369. [[CrossRef](#)]
16. Lao, T.T.; Chan, P.L.; Tam, K.F. Gestational diabetes mellitus in the last trimester: A feature of maternal iron excess? *Diabet. Med.* **2021**, *18*, 218–223. [[CrossRef](#)]
17. Simcox, J.A.; McClain, D.A. Iron and diabetes risk. *Cell Metab.* **2013**, *17*, 329–341. [[CrossRef](#)]
18. Vogiatzi, M.G.; Macklin, E.; Trachtenberg, F.L.; Fung, E.B.; Cheung, A.M.; Vichinsky, E.; Olivieri, N.; Kirby, M.; Kwiatkowski, J.L.; Cunningham, M.; et al. Differences in the prevalence of growth, endocrine and vitamin D abnormalities among the various thalassaemia syndromes in North America. *Br. J. Haematol.* **2009**, *146*, 546–556. [[CrossRef](#)]
19. Ascherio, A.; Rimm, E.B.; Giovannucci, E.; Willett, W.C.; Stampfer, M.J. Blood donations and risk of coronary heart disease in men. *Circulation* **2001**, *103*, 52–57. [[CrossRef](#)] [[PubMed](#)]
20. Dymock, I.; Cassar, J.; Pyke, D.; Oakley, W.; Williams, R. Observations on the pathogenesis, complications and treatment of diabetes in 115 cases of haemochromatosis. *Am. J. Med.* **1972**, *52*, 203–210. [[CrossRef](#)]
21. Platis, O.; Anagnostopoulos, G.; Farmaki, K.; Posantzis, M.; Gotsis, E.; Tolis, G. Glucose metabolism disorders improvement in patients with thalassaemia major after 24–36 months of intensive chelation therapy. *Pediatr. Endocrinol. Rev.* **2004**, *22*, 279–281.
22. Fernandez-Real, J.M.; Penarroja, G.; Castro, A.; Garcia-Bragado, F.; Hernandez-Aguado, I.; Ricart, W. Blood letting in high-ferritin type 2 diabetes: Effects on insulin sensitivity and beta-cell function. *Diabetes* **2002**, *51*, 1000–1004. [[CrossRef](#)]
23. Hua, N.W.; Stoohs, R.A.; Facchini, F.S. Low iron status and enhanced insulin sensitivity in lacto-ovo vegetarians. *Br. J. Nutr.* **2001**, *86*, 515–519. [[CrossRef](#)] [[PubMed](#)]
24. Pradhan, A.D.; Manson, J.E.; Rifai, N.; Buring, J.E.; Ridker, P.M. C-reactive protein, interleukin 6, and risk of developing type 2 diabetes mellitus. *JAMA* **2001**, *286*, 327–334. [[CrossRef](#)]
25. Wish, J.B. Assessing iron status: Beyond serum ferritin and transferrin saturation. *Clin. J. Am. Soc. Nephrol.* **2006**, *1*, S4–S8. [[CrossRef](#)] [[PubMed](#)]
26. Gabrielsen, J.S.; Gao, Y.; Simcox, J.; Huang, J.; Thorup, D.; Jones, D.; Cooksey, R.C.; Gabrielsen, D.; Adams, T.D.; Hunt, S.C.; et al. Adipocyte iron regulates adiponectin and insulin sensitivity. *J. Clin. Investig.* **2012**, *122*, 3529–3540. [[CrossRef](#)] [[PubMed](#)]
27. Wild, S.; Roglic, G.; Green, A.; Sicree, R.; King, H. Global prevalence of diabetes: Estimates for the year 2000 and projections for 2030. *Diabetes Care* **2004**, *27*, 1047–1053. [[CrossRef](#)]
28. Foley, R.N.; Collins, A.J. End-stage renal disease in the United States: An update from the United States renal data system. *J. Am. Soc. Nephrol.* **2007**, *18*, 2644–2648. [[CrossRef](#)]
29. Stengel, B.; Billon, S.; van Dijk, P.C.; Jager, K.J.; Dekker, F.; Simpson, K.; Briggs, J.D. Trends in the incidence of renal replacement therapy for end-stage renal disease in Europe, 1990–1999. *Nephrol. Dial. Transplant.* **2003**, *18*, 1824–1833. [[CrossRef](#)] [[PubMed](#)]

30. Villar, E.; Chang, S.H.; McDonald, S.P. Incidences, treatments, outcomes, and sex effect on survival in patients with end-stage renal disease by diabetes status in Australia and New Zealand (1991–2005). *Diabetes Care* **2007**, *30*, 3070–3076. [[CrossRef](#)]
31. Astor, B.C.; Muntner, P.; Levin, A.; Eustace, J.A.; Coresh, J. Association of kidney function with anemia: The third national health and nutrition examination survey (1988–1994). *Arch. Intern. Med.* **2002**, *162*, 1401–1408. [[CrossRef](#)] [[PubMed](#)]
32. Rossert, J.; Froissart, M. Role of anemia in progression of chronic kidney disease. *Semin. Nephrol.* **2006**, *26*, 283–289. [[CrossRef](#)] [[PubMed](#)]
33. Krikorian, S.; Shafai, G.; Shamim, K. Managing iron deficiency anemia of CKD with IV iron. *US Pharm.* **2013**, *8*, 22–26.
34. McFarlane, S.I.; Chen, S.C.; Whaley-Connell, A.T.; Sowers, J.R.; Vassalotti, J.A.; Salifu, M.O.; Li, S.; Wang, C.; Bakris, G.; McCullough, P.A.; et al. Prevalence and associations of anemia of CKD: Kidney Early Evaluation Program (KEEP) and National Health and Nutrition Examination Survey (NHANES) 1999–2004. *Am. J. Kidney Dis.* **2008**, *51*, S46–S55. [[CrossRef](#)]
35. Stauffer, M.E.; Fan, T. Prevalence of anemia in chronic kidney disease in the United States. *PLoS ONE* **2014**, *9*, e84943. [[CrossRef](#)]
36. Aso, Y.; Suganuma, R.; Wakabayashi, S.; Hara, K.; Nakano, T.; Suetsugu, M.; Matsumoto, S.; Nakamachi, T.; Takebayashi, K.; Morita, K.; et al. Anemia is associated with an elevated serum level of high-molecular-weight adiponectin in patients with type 2 diabetes independently of renal dysfunction. *Transl. Res.* **2009**, *154*, 175–182. [[CrossRef](#)]
37. McFarlane, S.I.; Salifu, M.O.; Makaryus, J.; Sowers, J.R. Anemia and cardiovascular disease in diabetic nephropathy. *Curr. Diabetes Rep.* **2006**, *6*, 213–218. [[CrossRef](#)]
38. van der Meer, I.M.; Ruggerenti, P.; Remuzzi, G. The diabetic CKD patient—A major cardiovascular challenge. *J. Ren. Care* **2010**, *36*, 34–46. [[CrossRef](#)]
39. Gjata, M.; Nelaj, E.; Sadiku, E.; Collaku, E.; Tase, M. Prevalence of anemia in early stage of diabetic nephropathy (chronic kidney disease stage II) and its impact on the progression of renal function: PP.17.123. *J. Hypertens.* **2010**, *28*, e288. [[CrossRef](#)]
40. Pappa, M.; Dounousi, E.; Duni, A.; Katopodis, K.P. Less known pathophysiological mechanisms of anemia in patients with diabetic nephropathy. *Int. Urol. Nephrol.* **2015**, *47*, 1365–1372. [[CrossRef](#)] [[PubMed](#)]
41. Introduction: Standards of medical care in diabetes—2021. *Diabetes Care* **2021**, *44*, S1–S2. [[CrossRef](#)]
42. Kidney Disease: Improving Global Outcomes (KDIGO) Blood Pressure Work Group. KDIGO 2012 clinical practice guideline for the evaluation and management of chronic kidney disease. *Kidney Int. Suppl.* **2013**, *3*, 1–150.
43. Levey, A.S.; Stevens, L.A.; Schmid, C.H.; Zhang, Y.L.; Castro, A.F., 3rd; Feldman, H.I.; Kusek, J.W.; Eggers, P.; van Lente, F.; Greene, T.; et al. A new equation to estimate glomerular filtration rate. *Ann. Intern. Med.* **2009**, *150*, 604–612. [[CrossRef](#)]
44. World Health Organization. Nutritional anemias: Report of a WHO scientific group. *World Health Organ. Tech. Rep. Ser.* **1968**, *405*, 5–37.
45. Hayder, Z.S.; Kareem, Z.S. Resistin hormone in diabetic kidney disease and its relation to iron status and hepcidin. *Int. Urol. Nephrol.* **2020**, *52*, 749–756. [[CrossRef](#)] [[PubMed](#)]
46. Małyżko, J.; Koc-Żórawska, E.; Levin-Iaina, N.; Małyżko, J.; Koźmiński, P.; Kobus, G.; Myśliwiec, M. New parameters in iron metabolism and functional iron deficiency in patients on maintenance hemodialysis. *Pol. Arch. Med. Wewn.* **2012**, *122*, 537–542.
47. Peters, H.P.; Rumjon, A.; Bansal, S.S.; Laarakkers, C.M.; Brand, J.V.D.; Sarafidis, P.; Musto, R.; Małyżko, J.; Swinkels, D.W.; Wetzels, J.F.; et al. Intra-individual variability of serum hepcidin-25 in haemodialysis patients using mass spectrometry and ELISA. *Nephrol. Dial. Transplant.* **2012**, *27*, 3923–3929. [[CrossRef](#)] [[PubMed](#)]
48. Małyżko, J.; Małyżko, J.S.; Pawlak, K.; Myśliwiec, M. Hepcidin, iron status, and renal function in chronic renal failure, kidney transplantation, and hemodialysis. *Am. J. Hematol.* **2006**, *81*, 832–837. [[CrossRef](#)]
49. Zumbrennenbullough, K.B.; Babitt, J.L. The iron cycle in chronic kidney disease (CKD): From genetics and experimental models to CKD patients. *Nephrol. Dial. Transplant.* **2014**, *29*, 263–273. [[CrossRef](#)]
50. Kuragano, T.; Shimonaka, Y.; Kida, A.; Furuta, M.; Nanami, M.; Otaki, Y.; Hasuie, Y.; Nonoguchi, H.; Nakanishi, T. Determinants of hepcidin in patients on maintenance hemodialysis: Role of inflammation. *Am. J. Nephrol.* **2010**, *31*, 534–540. [[CrossRef](#)] [[PubMed](#)]
51. Tessitore, N.; Girelli, D.; Camprostrini, N.; Bedogna, V.; Solero, G.P.; Castagna, A.; Melilli, E.; Mantovani, W.; De Matteis, G.; Olivieri, O.; et al. Hepcidin is not useful as a biomarker for iron needs in haemodialysis patients on maintenance erythropoiesis-stimulating agents. *Nephrol. Dial. Transplant.* **2010**, *25*, 3996–4002. [[CrossRef](#)]
52. van der Weerd, N.C.; Grooteman, M.P.; Bots, M.L.; van den Dorpel, M.A.; den Hoedt, C.H.; Mazairac, A.H.; Nubé, M.J.; Penne, E.L.; Gaillard, C.A.; Wetzels, J.F.M.; et al. Hepcidin-25 in chronic hemodialysis patients is related to residual kidney function and not to treatment with erythropoiesis stimulating agents. *PLoS ONE* **2012**, *7*, e39783. [[CrossRef](#)]
53. Lukaszuk, E.; Lukaszuk, M.; Koc-Żórawska, E.; Bodzenta-Lukaszuk, A.; Małyżko, J. Zonulin, inflammation and iron status in patients with early stages of chronic kidney disease. *Int. Urol. Nephrol.* **2017**, *50*, 121–125. [[CrossRef](#)] [[PubMed](#)]
54. Wagner, M.; Ashby, D.R.; Kurtz, C.; Alam, A.; Busbridge, M.; Raff, U.; Zimmermann, J.; Heuschmann, P.U.; Wanner, C.; Schramm, L. Hepcidin-25 in diabetic chronic kidney disease is predictive for mortality and progression to end stage renal disease. *PLoS ONE* **2015**, *10*, e0123072. [[CrossRef](#)] [[PubMed](#)]
55. Lukaszuk, E.; Lukaszuk, M.; Koc-Zorawska, E.; Bodzenta-Lukaszuk, A.; Małyżko, J. GDF-15, iron, and inflammation in early chronic kidney disease among elderly patients. *Int. Urol. Nephrol.* **2016**, *48*, 839–844. [[CrossRef](#)] [[PubMed](#)]
56. Breit, S.N.; Johnen, H.; Cook, A.; Tsai, V.W.W.; Mohammad, M.; Kuffner, T.; Zhang, H.P.; Marquis, C.; Jiang, L.; Lockwood, G.; et al. The TGF- β superfamily cytokine, MIC-1/GDF15: A pleiotropic cytokine with roles in inflammation, cancer and metabolism. *Growth Factors* **2011**, *29*, 187–195. [[CrossRef](#)]

57. Lukaszyc, E.; Lukaszyc, M.; Koc-Zorawska, E.; Bodzenta-Lukaszyc, A.; Malyszko, J. Fibroblast growth factor 23, iron and inflammation—are they related in early stages of chronic kidney disease? *Arch. Med. Sci.* **2017**, *13*, 845–850. [[CrossRef](#)]
58. Cook, J.D.; Skikne, B.S.; Baynes, R.D. Serum transferrin receptor. *Annu. Rev. Med.* **1993**, *44*, 63–74. [[CrossRef](#)]
59. Belo, L.; Rocha, S.; Valente, M.J.; Coimbra, S.; Catarino, C.; Bronze-Da-Rocha, E.; Rocha-Pereira, P.; Sameiro-Faria, M.D.; Oliveira, J.G.; Madureira, J.; et al. Hepcidin and diabetes are independently related with soluble transferrin receptor levels in chronic dialysis patients. *Ren. Fail.* **2019**, *41*, 662–672. [[CrossRef](#)]
60. Ferguson, B.J.; Skikne, B.S.; Simpson, K.M.; Baynes, R.D.; Cook, J.D. Serum transferrin receptor distinguishes the anemia of chronic disease from iron deficiency anemia. *J. Lab. Clin. Med.* **1992**, *119*, 385–390.
61. Pfeiffer, C.M.; Looker, A.C. Laboratory methodologies for indicators of iron status: Strengths, limitations, and analytical challenges. *Am. J. Clin. Nutr.* **2017**, *106*, 1606S–1614S. [[CrossRef](#)] [[PubMed](#)]
62. Semenza, G.L. Oxygen sensing, homeostasis, and disease. *N. Engl. J. Med.* **2011**, *365*, 537–547. [[CrossRef](#)] [[PubMed](#)]
63. Mastrogiannaki, M.; Matak, P.; Mathieu, J.R.; Delga, S.; Mayeux, P.; Vaulont, S.; Peyssonnaud, C. Hepatic hypoxia-inducible factor-2 down-regulates hepcidin expression in mice through an erythropoietin-mediated increase in erythropoiesis. *Haematologica* **2011**, *97*, 827–834. [[CrossRef](#)] [[PubMed](#)]
64. Shutov, E.; Sułowicz, W.; Esposito, C.; Tataradze, A.; Andric, B.; Reusch, M.; Valluri, U.; Dimkovic, N. Roxadustat for the treatment of anemia in chronic kidney disease patients not on dialysis: A Phase 3, randomized, double-blind, placebo-controlled study (ALPS). *Nephrol. Dial. Transplant.* **2021**. [[CrossRef](#)]