

Complete Genome Sequence of *Simiduia agarivorans* SA1^T, a Marine Bacterium Able To Degrade a Variety of Polysaccharides

Silk Yu Lin, a Wung Yang Shieh, a Jwo-Sheng Chen, b Sen-Lin Tanga,c

Institute of Oceanography, National Taiwan University, Taipei, Taiwana; College of Health Care, China Medical University, Taichung, Taiwana; Biodiversity Research Center, Academia Sinica, Taipei, Taiwana

Simiduia agarivorans strain SA1^T is able to degrade a variety of polysaccharides found in marine algae, plants, and animals. The genome of *S. agarivorans* SA1^T consists of a single chromosome (4,309,711 bp), and its information may provide insights into the polysaccharide-degrading capability, cell division, flagellar motility, and chemotaxis of this bacterium.

Received 14 October 2012 Accepted 25 October 2012 Published 15 January 2013

Citation Lin SY, Shieh WY, Chen J-S, Tang S-L. 2013. Complete genome sequence of *Simiduia agarivorans* SA1^T, a marine bacterium able to degrade a variety of polysaccharides. Genome Announc. 1(1):e00039-12. doi:10.1128/genomeA.00039-12.

Copyright © 2013 Lin et al. This is an open-access article distributed under the terms of the Attribution 3.0 Unported Creative Commons License.

Address correspondence to Sen-Lin Tang, sltang@gate.sinica.edu.tw.

S.Y.L. and W.Y.S. contributed equally to this article.

Simiduia agarivorans SA1^T is a marine bacterium isolated from shallow coastal seawater and is able to degrade a variety of refractory polysaccharides, such as agar, alginate, cellulose, and chitin (1). The phylogeny analysis based on the 16S rRNA gene showed that *Simiduia* species formed a distinct lineage within other bacterial neighbors of the class *Gammaproteobacteria* (1).

S. agarivorans SA1^T is a type strain in this clade. Moreover, this bacterium reproduces itself by a unique multiple offspring formation. The finishing and annotation of the *S. agarivorans* SA1^T genome are useful to elucidate its genomic features, including those relevant to its versatile metabolic capabilities and cell division manner.

The sequencing technologies used included the Illumina GA2 (Solexa), HiSeq 2000 (Illumina), and 454 GS Junior sequencing (Roche) systems (2). The Illumina paired-end sequencing was performed at the Biodiversity Research Center of Academia Sinica (Taiwan) and yielded ~21,945,490 reads of 63 bp in length. The output data were processed and assembled using Off-Line Basecaller (OLB) software, producing 135 contigs covering approximately 4.2 Mb. The data were combined with the results of another Illumina mate-pair sequencing that was performed at Yourgene BioScience (Taiwan), and 22,750,330 mate-end reads of 40 bp in length were obtained. The data sets were assembled using Velvet software (3). The resulting assembly consisted of 17 large contigs of over 71 kb (maximum contig size of 950 kb). The 454 GS Junior sequencing was performed at the Microarray and Sequencing Core of Academia Sinica, generating 143,752 reads with a modal length of 446 bp. The output data were processed and assembled using Newbler software (4), producing 87 contigs covering approximately 4.3 Mb. After combining all of the data, a genome scaffold of 4.309 Mb was generated. The remaining gaps and unassertive assembled regions were verified by PCR/Applied Biosystems (ABI) sequencing to obtain a single contig.

An optical map of the *S. agarivorans* SA1^T genome was constructed at Yourgene BioScience with the HindIII restriction enzyme, yielding 504 ordered restriction fragments. The genome

size was estimated to be approximately 4.29 Mb, which was determined from the sum of all restriction fragments of the map. The assembly was finished by scaffolding the contigs on the optical map using the MapSolver software (OpGen). The correct sequence mapping relied upon the optical map. The genome was annotated using the NCBI Prokaryotic Genomes Automatic Annotation Pipeline (PGAAP) employed for submission (http://www.ncbi.nlm.nih.gov/genomes/static/Pipeline.html).

The *S. agarivorans* SA1^T genome encodes a 4,309,711-bp circular molecule with a G+C content of 55.9% and no plasmid. We compiled 3,786 (The original gene number is the sum of cds + rRNA + tRNA.) coding sequences (CDSs), 3 rRNA operons, and 41 tRNAs from the nucleotide sequence. The genome contains more than five polysaccharide-hydrolyzing enzyme systems, with a total of 45 CDSs involved in the hydrolysis of agar, alginate, cellulose, chitin, and xylan. It contains more than 47 CDSs involved in the cell division process, including those for cell division regulation, murein and shape determination, chromosome partition, Z-ring formation, the membrane-embedding Tol-Pal system, and amidase.

Nucleotide sequence accession number. The complete genome sequence of *S. agarivorans* SA1^T has been deposited in GenBank under accession number CP003746.

ACKNOWLEDGMENTS

This work was supported by the Biodiversity Research Center of Academia Sinica (Taiwan).

We thank the Biofuel High Throughput Sequencing Core of the Biodiversity Research Center, the Microarray Core and DNA Sequencing Core of the Institute of Molecular Biology of Academia Sinica, and Yourgene BioScience (http://www.yourgene.com.tw) for providing the sequencing service.

REFERENCES

 Kislyuk AO, Katz LS, Agrawal S, Hagen MS, Conley AB, Jayaraman P, Nelakuditi V, Humphrey JC, Sammons SA, Govil D, Mair RD, Tatti KM, Tondella ML, Harcourt BH, Mayer LW, Jordan IK. 2010. A computa-

- tional genomics pipeline for prokaryotic sequencing projects. Bioinformatics ${\bf 26}:1819-1826$.
- Zerbino DR, Birney E. 2008. Velvet: algorithms for de novo short read assembly using de Bruijn graphs. Genome Res. 18:821–829.
- 3. Margulies M, Egholm M, Altman WE, Attiya S, Bader JS, Bemben LA, Berka J, Braverman MS, Chen YJ, Chen Z, Dewell SB, Du L, Fierro JM, Gomes XV, Godwin BC, He W, Helgesen S, Ho CH, Ho CH, Irzyk GP, Jando SC, Alenquer ML, Jarvie TP, Jirage KB, Kim JB, Knight JR, Lanza JR, Leamon JH, Lefkowitz SM, Lei M, Li J, Lohman KL, Lu H, Makhijani
- VB, McDade KE, McKenna MP, Myers EW, Nickerson E, Nobile JR, Plant R, Puc BP, Ronan MT, Roth GT, Sarkis GJ, Simons JF, Simpson JW, Srinivasan M, Tartaro KR, Tomasz A, Vogt KA, Volkmer GA, Wang SH, Wang Y, Weiner MP, Yu P, Begley RF, Rothberg JM. 2005. Genome sequencing in microfabricated high-density picolitre reactors. Nature 437: 376–380.
- 4. Shieh WY, Liu TY, Lin SY, Jean WD, Chen JS. 2008. *Simiduia agarivorans* gen. nov., sp. nov., a marine, agarolytic bacterium isolated from shallow coastal water from Keelung, Taiwan. Int. J. Syst. Evol. Microbiol. **58**:895–900.