### REVIEW



# Metabolic reprogramming: a bridge between aging and tumorigenesis

### Stanislav Drapela (D, Didem Ilter and Ana P. Gomes (D)

Department of Molecular Oncology, H. Lee Moffit Cancer Center & Research Institute, Tampa, FL, USA

#### Keywords

aging; cellular energetics; immune response; metabolic reprogramming; tumorigenesis

#### Correspondence

A. P. Gomes, Department of Molecular Oncology, H. Lee Moffit Cancer Center & Research Institute, 12902 Magnolia Drive, Tampa, FL, 33612, USA E-mail: ana.gomes@moffitt.org

Stanislav Drapela and Didem Ilter contributed equally to this article

(Received 1 March 2022, revised 7 May 2022, accepted 23 May 2022, available online 19 June 2022)

doi:10.1002/1878-0261.13261

Aging is the most robust risk factor for cancer development, with more than 60% of cancers occurring in those aged 60 and above. However, how aging and tumorigenesis are intertwined is poorly understood and a matter of significant debate. Metabolic changes are hallmarks of both aging and tumorigenesis. The deleterious consequences of aging include dysfunctional cellular processes, the build-up of metabolic byproducts and waste molecules in circulation and within tissues, and stiffer connective tissues that impede blood flow and oxygenation. Collectively, these age-driven changes lead to metabolic reprogramming in different cell types of a given tissue that significantly affects their cellular functions. Here, we put forward the idea that metabolic changes that happen during aging help create a favorable environment for tumorigenesis. We review parallels in metabolic changes that happen during aging and how these changes function both as adaptive mechanisms that enable the development of malignant phenotypes in a cell-autonomous manner and as mechanisms that suppress immune surveillance, collectively creating the perfect environment for cancers to thrive. Hence, antiaging therapeutic strategies that target the metabolic reprogramming that occurs as we age might provide new opportunities to prevent cancer initiation and/or improve responses to standard-of-care anticancer therapies.

### 1. Introduction

Aging is defined as a series of progressive physiological changes that lead to a decline in biological functions, which in turn lead to multiple diseases, collectively known as age-related diseases [1]. For many years,

#### Abbreviations

nuclear and mitochondrial DNA (mtDNA) damage that had accumulated over time was thought to be the root of aging [2,3]. However, this paradigm has shifted in recent years with the discovery that chronological and biological age do not necessarily match [4–8] and with the observation that lifespan can be prolonged

a-KG, α-ketoglutarate; AMP, adenosine monophosphate; APC, antigen-presenting cells; Arg1, arginase 1; ATP, adenosine triphosphate; CAR-T, chimeric antigen receptor T; CR, caloric restriction; ERK, extracellular signal-regulated kinase; ETC, electron transport chain; GAPDH, glyceraldehyde-3-phosphate dehydrogenase; HIF-1 α, hypoxia-inducible factor 1α; IGF-1, insulin-like growth factor 1; IL-4, interleukin 4; INF-γ, interferon-gamma; JNK, c-Jun N-terminal kinase; LDH, lactate dehydrogenase; LKB1, liver kinase 1; MAPK, mitogen-activated protein kinases; MCC, multistage model of carcinogenesis; MMA, methylmalonic acid; mtDNA, mitochondrial DNA; mTOR, mechanistic target of rapamycin; NAD, nicotinamide adenine dinucleotide; NADPH, reduced nicotinamide adenine dinucleotide phosphate; NF- κB, nuclear factorkappa B; NFAT, nuclear factor of activated T cells; NK, natural killer; OXPHOS, oxidative phosphorylation; PARP, poly (ADP-ribose) polymerase; PBMC, peripheral blood mononuclear cells; PD1, programmed cell death protein 1; PET, positron emission tomography; Polg, DNA polymerase G; PTEN, phosphatase and tensin homolog; PTP1b, protein tyrosine phosphatase 1b; REDOX, reduction–oxidation; ROS, reactive oxygen species; TCA, tricarboxylic acid; TCR, T-cell receptor; TFAM, mitochondrial transcription factor A; TFEB, transcription factor EB; T<sub>h</sub>, T helper; TIL, tumor-infiltrating lymphocyte; T<sub>reg</sub>, regulatory T; VEGF, vascular endothelial growth factor.

Molecular Oncology **16** (2022) 3295–3318 © 2022 The Authors. *Molecular Oncology* published by John Wiley & Sons Ltd on behalf of Federation of European Biochemical Societies.

This is an open access article under the terms of the Creative Commons Attribution License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

and that certain traits of aging can be readily reversed with antiaging interventions [9-26].

Among age-related diseases, cancer is becoming a major public health and economic issue with its incidence dramatically on the rise [27]. The rise in cancer incidence is inextricably linked to an increase in lifespan and to the consequent increase in the aging population. The World Health Organization projects that the proportion of the world's population aged over 60 will double to 22% by 2050 [28]. More than 60% of cancers are diagnosed in those aged 60 and above, underscoring aging as a major, well-recognized, risk factor for cancer development [29]. Furthermore, aging predicts both cancer incidence and prognosis [30-33]: older patients have worse outcomes with unfavorable progression-free and overall survival rates [34-36], suggesting that the aging and tumorigenic processes are highly intertwined.

Knudson's hypothesis states that due to increased exposure to mutagens, which allow cells to accumulate cancer-causing genetic mutations, cancer incidence increases with age [37]. However, observations that life choices like diet and physical activity can have drastic consequences for cancer susceptibility and outcome [38–41] have challenged this hypothesis. In contrast, Armitage and Doll [42] described the multi-stage model of carcinogenesis (MMC), which was further developed by Nowell [43]. This model explains carcinogenesis as a Darwinian somatic selection process (Box 1), during which both cell-extrinsic (selection) and cell-intrinsic (mutations) factors act as drivers and are more aligned with how the aging process is currently perceived. Additionally, a recent study underscored the part aging plays in the tumorigenic paradigm by indicating that MCC needs to include aging-dependent somatic selection to be able to explain cancer incidence across tissues and species [44]. Collectively, recent findings suggest that the aging process is both a selective force and a driver for tumorigenesis emphasizing the importance of the host environment, in addition to genetic hits, for cancer cell fate and patient outcomes.

One of the most striking distinctions between tumors and nontransformed tissues is the differences in their metabolism, such as deregulated uptake of glucose and amino acids, use of glycolysis and tricarboxylic acid (TCA) cycle intermediates for biosynthesis, and increased demand for reductive power to counteract the buildup of toxic byproducts [45,46]. These differences are considered to be a hallmark of cancer [47] and have been used to diagnose tumors as in the case of <sup>18</sup>F-fluorodeoxyglucose positron emission tomography (PET) imaging [48], which detects glucose uptake. Much of the metabolism occurring in cancer cells serves to support anabolic and reduction–oxidation (redox) reactions. These reactions generate the building blocks needed to maintain high rates of proliferation without inducing cell death caused by the accumulation of reactive oxygen species (ROS) [49–51].

Metabolic alterations, ranging from deregulated nutrient sensing to profound alterations in central carbon metabolism and mitochondrial dysfunction, are also at the center of the aging process [52]. Highlighting the importance of metabolism in the aging process, many of the well-established interventions that extend lifespan and healthspan, such as caloric restriction (CR), rapamycin (Box 1), and metformin (Box 1), either target metabolism or have major metabolic effects, such as improvements in insulin sensitivity, glucose homeostasis, and changes in body weight [15,53-58]. Conversely, hypercaloric nutrition and sedentary lifestyles, which also have major metabolic consequences, can accelerate the aging process [59–62]. Moreover, inborn errors of metabolism (Box 1) accelerate aging: for example, adult polyglucosan body disease (APBD) causes neurogenic bladder dysfunction and dementia, mucolipidosis type II/III causes joint stiffness, while Gaucher disease type 1/3 and Niemann Pick A/B lead to osteoporosis and arthritis, respectively [63–66]. Interestingly, patients with Werner syndrome, a premature-aging syndrome described as progeria (Box 1) of adults, display an increased risk of cancer incidence [67,68]. However, patients with Hutchinson Gilford progeria syndrome (HGPS), a premature-aging syndrome that has a childhood onset, do not develop tumors even though they have high levels of DNA damage; instead, cells collected from these patients are resistant to neoplastic transformations [69]. The reasons for this discrepancy in the two progeria syndromes are not known, but the very short life expectancy (on average 14.5 years) of HGPS patients might partially explain the low incidence of cancer diagnosis in these patients. On the other hand, loci associated with extreme longevity also influence metabolism [70,71], and centenarians (Box 1) display a very low incidence of cancer, only 4% of them die from cancer [72,73]. Considering this stark anticorrelation between extreme longevity and cancer-associated mortality it is reasonable to conceive that the mechanisms that promote increased healthspan and lifespan must also play a role in suppressing tumor initiation or progression. Therefore, an older individual is a different metabolic entity than a younger one, suggesting that the metabolic alterations that occur with aging have important consequences for tumorigenesis. The

### Box 1. Glossary.

Adenosine monophosphate (AMP)-activated protein kinase (AMPK): AMPK is a kinase that senses the energy status of the cell, and it is highly conserved from yeast to human. Increased AMP/ADP ratio indicates that the cell is in low energy status and the AMPK is activated. Activated AMPK promotes changes such as increased fatty acid oxidation and glucose uptake, and inhibition of lipogenesis and cholesterol synthesis.

**Anaplerosis**: Anaplerosis refers to the replenishing of tricarboxylic acid (TCA) cycle intermediates, which can be used as substrates for various biosynthetic pathways. Since the TCA cycle is a hub of energy production, replenishing these intermediates is crucial and the levels of these intermediates in the mitochondria are therefore highly regulated.

Antigen-presenting cells (APCs): APCs are a heterogeneous group of immune cells including dendritic cells, macrophages, and B cells that mediate the cellular immune response by processing and presenting antigens for recognition by certain lymphocytes such as T cells.

 $CD8^+$  T cells:  $CD8^+$  T cells are critical mediators of adaptive immunity. They include cytotoxic T cells, which are important for killing cancerous or virally infected cells, and CD8-positive suppressor T cells, which restrain certain types of the immune response.

**Centenarian**: Centenarian is an individual who lived beyond their 100<sup>th</sup> birthday. Since these people live beyond the current life expectancy, they are associated with longevity. The aging field has been studying these people to understand which physiological characteristics of these people lead to their extreme longevity.

**Chimeric antigen receptor T (CAR-T) cells:** CAR-T cells are immunotherapy tools: T cells that are genetically engineered to express a T-cell receptor. These cells can be derived from the patient directly or from another health donor. The receptors are called chimeric because they combine into a single receptor both T-cell activating and antigenbinding functions, and they give the CAR-T cells the ability to target a specific protein.

**Conplastic mouse**: Conplastic mice are strains that share the nuclear genome but differ in their mitochondrial genome. They are derived by backcrossing the nuclear genome from one inbred strain into the cytoplasm of another. These mice make it possible to study differences in mitochondrial haplotypes and various phenotypes.

**Glycolytic flux**: The glycolytic flux refers to the flux between fructose 6-phosphate (F6P) and fructose-1,6-bisphosphate (FBP), and simply describes the rate of glucose breakdown.

Host defenses: The term host defenses refers to two complementary, frequently interacting systems: (a) innate immunity, which protects against microorganisms in general; and (b) adaptive immunity, which protects against a particular microorganism.

Hypoxia-inducible factor  $1\alpha$  (HIF- $1\alpha$ ): HIF- $1\alpha$  is a transcription factor that acts as a sensor of available O<sub>2</sub> levels and also plays an important role in glucose metabolism. The stability of HIF1 is dependent on molecular O<sub>2</sub> levels in the cell, and it is stabilized when oxygen is limited. It induces the expression of glycolysis and glucose transporter genes while inhibiting the glucose carbon flow into the mitochondria. In proliferating cells, HIF1 signaling can be activated even at normal O<sub>2</sub> levels. For example, HIF1 can be induced transcriptionally and translationally downstream of growth factor signaling.

**Immune checkpoint blockade**: Immune checkpoints are pathways that regulate the immune system and are crucial for selftolerance. However, cancer cells can hijack these pathways to protect themselves from immune surveillance. A class of anticancer immunotherapies blocks inhibitory checkpoint receptors that are expressed in tumor cells. In some cancers, such as Hodgkin lymphoma and natural killer T-cell lymphoma, these immune checkpoint blockade therapies have led to high response rates. However, in other tumor types, such as breast and prostate, they have not proved to be successful.

**Immune surveillance**: Immune surveillance is a monitoring process of the immune system, including natural killer cells, cytotoxic T cells, or macrophages, to detect and destroy virally infected and neoplastically transformed cells in the body. This phenomenon is mediated by tumor-specific antigens (found exclusively on tumor cells) or tumor-associated antigens (found on both tumor and normal cells but overexpressed on tumor cells).

**Inborn errors of metabolism**: Inborn errors of metabolism refer to a group of genetic diseases that involve a defective metabolic enzyme. These defective enzymes lead to the accumulation of toxic substances and interfere with normal function. Some of these disorders are detected during newborn screening tests while others are diagnosed as symptoms

### Box 1. (Continued)

appear. Depending on the condition, in addition to treating the symptoms and complications, dietary interventions, dialysis, bone marrow or organ transplantation can be used to treat these conditions.

**Kynurenine**: Kynurenine is a tryptophan metabolite and synthesized by both indoleamine 2,3-dioxygenase (IDO-1) and tryptophan-2,3-dioxygenase-2 (TDO-2). It binds, among other endogenous molecules, to the aryl hydrocarbon receptor (AHR) in multiple immune cell types, leading to immune suppression.

Lactate dehydrogenase (LDH) A/B ratio: Lactate dehydrogenase is an enzyme that catalyzes the conversion of lactate to pyruvate and the reverse reaction using NAD(H) as a co-factor. The direction of the reaction is governed by the specific lactate isoforms: LDHA preferentially converts pyruvate into lactate; LDHB preferentially converts lactate into pyruvate. The LDHA/B subunit ratio determines the overall direction of the reaction.

**Lactylation**: Lactylation, also known as histone lysine lactylation, is a new lactate-induced histone post-translational modification proposed and identified in 2019 by mass spectrometry. It is prevalently found in fungi and mammalian cells where it directly stimulates gene transcription and regulates the glycolytic flux.

**Macrophage polarization**: Macrophage polarization refers to the process by which macrophages produce distinct functional phenotypes as a reaction to specific microenvironmental stimuli and signals. Macrophages can be polarized into classically activated (M1) and alternatively activated (M2) macrophages. While M1 macrophages produce proinflammatory cytokines, mediate resistance to pathogens, and stimulate antitumor activity, M2 macrophages mainly secrete antiinflammatory cytokines, which reduce inflammation and contribute to immune suppression and tumor growth.

**Mechanistic target of rapamycin (mTOR)**: mTOR is a protein kinase that is a key regulator of metabolic homeostasis and combined with other proteins forms two distinct protein complexes, mTORC1/2. In response to growth factors, nutrients, energy levels, and stress, it regulates various cellular functions including cell growth, proliferation, survival, motility, protein synthesis, transcription, and autophagy.

**Metformin**: Metformin is the first-line therapy for patients with prediabetes or type 2 diabetes. It is mechanism of action is not fully known.

**Microbiome**: Microbiome is the collection of all microorganisms (including bacteria, fungi, and viruses) that naturally lives in symbiosis inside another organism including humans.

**Mitochondrial transcription factor A (TFAM)**: TFAM is a mitochondrial transcription factor that is a central activator of mitochondrial transcription and participates in mitochondrial DNA replication.

**Nuclear factor of activated T cells (NFAT)**: NFAT is a family of transcription factors critical for the regulation of early gene transcription in response to T-cell receptor-mediated signals in lymphocytosis. When phosphorylated, it is confined to the cell cytoplasm in its inactive state. After T-cell activation, the ensuing calcium influx activates the phosphatase calcineurin that activates NFAT by dephosphorylation.

**Polyamines:** Polyamines are organic compounds that have two or more amino groups. They can occur naturally (spermine, spermidine) but may be also prepared synthetically. Polyamines stimulate cell proliferation, have the ability to modulate the immune response, and are essential components of T-cell and B-cell activation, where they act as effectors.

**Progeria**: Progeria in this context is used to refer to progeroid syndromes, a group of rare genetic disorders. They cause premature aging, and the affected individuals physiologically appear older than they are.

**Rapamycin**: Rapamycin is a small molecule inhibitor that is isolated from the bacterium *Streptomyces hygroscopicus*, and it is an inhibitor of the mechanistic target of rapamycin kinase (mTOR). It has been used as an immunosuppressant to prevent organ rejection after transplants, to treat a rare lung disease called lymphangioleiomyomatosis (LAM), and has been tested for its antiaging and anticancer effects. Later similar molecules, called rapalogs, were designed to have more favorable pharmacokinetics.

**Regulatory T cells (T<sub>regs</sub>):**  $T_{regs}$  are a specialized subpopulation of T cells that act to suppress the immune response, thereby maintaining homeostasis and self-tolerance. It has been shown that  $T_{regs}$  are able to inhibit T-cell proliferation and cytokine production and play a critical role in preventing autoimmunity.

### Box 1. (Continued)

**Senescence**: Cellular senescence or biological aging is a phenomenon characterized by the cessation of cell division. It refers to irreversible gradual deterioration of functional characteristics in living organisms accompanied by a stable growth arrest and other phenotypic alterations such as proinflammatory secretome known as senescence-associated secretory phenotype (SASP).

Short-chain fatty acids (SCFAs): SCFAs are fatty acids with fewer than six carbon atoms derived from intestinal microbial fermentation of indigestible foods. Two common SCFAs are acetate and butyrate. SCFAs serve as the main energy source of colonocytes, making them crucial for gastrointestinal health.

Sirtuins: Sirtuins are a family of signaling molecules that have roles in metabolism and implicated in aging. They are highly conserved from bacteria to humans. They can act as deacetylases and ADP-ribosyl transferases, and play roles in various cellular processes including metabolism, inflammation, cell cycle, DNA repair, and tumorigenesis.

Somatic selection: Somatic mutations are alterations to DNA that occur after conception. They can happen in any cell type except the germline; therefore, they cannot be passed on to children. Somatic selection increases the abundance of cells that have favorable mutations and therefore works in favor of cancer.

 $T_h1$  cell-associated transcription factor (T-bet): T-bet is an immune cell transcription factor expressed in CD4<sup>+</sup> T lymphocytes committed to  $T_h1$  T-cell differentiation and development. However, it has been also recognized to have a role in both the adaptive and innate immune systems. T-bet also directs T-cell homing to proinflammatory sites by the regulation of CXCR3 expression.

**Tumor-infiltrating lymphocytes (TILs)**: TILs are all lymphocytic cell populations that have moved from the blood into a tumor where they can recognize and kill cancer cells. TILs have been described in several solid tumors, including breast cancer, and are emerging as an important biomarker in predicting the efficacy and outcome of treatment. In cancer therapy, tumor-infiltrating lymphocytes are removed from a patient's tumor, grown in large numbers in a laboratory, and then given back to the patient to help the immune system kill the cancer cells.

association of cancer with metabolic reprogramming and increased age might also explain why interventions that maintain metabolic health also have cancerprotective effects.

In the sections below, we discuss the main metabolic pathways and processes that change with aging and how these produce an environment conducive to tumorigenesis. On the one hand, we discuss the metabolic reprogramming that occurs in cells with age that mirrors the metabolic alterations that occur in tumor cells. On the other hand, we also discuss how the metabolic environment of the old host affects the immune compartment, rendering it less prone to immune surveillance (Box 1). We propose that collectively age-induced metabolic reprogramming endows premalignant cells with the means and opportunities to thrive as cancers.

# 2. Age-induced metabolic rewiring in transformation

To maintain growth, cancer cells employ a variety of metabolic adaptations, the nature of which is

collectively determined by the physiology of their cell of origin, the identity of transforming lesions, and the tissue in which cancer cells arise from. It is, therefore, reasonable to suggest that an individual's metabolic state, which changes considerably with age, might enable tumorigenesis and explain why the biggest risk factor for tumorigenesis is old age (Fig. 1 and Table 1).

### 2.1. Nutrient and energy sensors

A key hallmark of aging is the deregulation of nutrient-sensing pathways [1]. Nutrient-sensing pathways ensure that a correct balance is achieved between the availability of nutrients and the cellular capacity to use them in order to maintain homeostasis [74]. Strikingly, of the hundreds of genes that influence lifespan in animal models, a large proportion of them function in nutrient-sensing pathways, including components of insulin-like growth factor 1 (IGF-1)/Insulin signaling, mechanistic target of rapamycin (mTOR; Box 1), adenosine monophosphate (AMP)-activated protein

	Table '	1. Evidence	for agin	g-related	changes as	contributors o	r deterrents c	of tumorigenesis.
--	---------	-------------	----------	-----------	------------	----------------	----------------	-------------------

Aging-related changes	Favors tumorigenesis	Does not favor tumorigenesis	
Progeria	<ul> <li>Patients with Werner Syndrome, called adult progeria, display increased cancer risk [67,68]</li> </ul>	<ul> <li>Patients with Hutchinson Gilford progeria syndrome do not develop tumors and cells derived from these patients are resistant to transformation [60]</li> </ul>	
Deregulation of	Caloric restriction extends lifespan and protects from		
nutrient sensing	tumorigenesis [77]		
	<ul> <li>PISN/ARI/ITTO pattway is hyperactivated in various tumor types [172]</li> <li>Inhibition of mTOR has significant benefits as an anticancer therapy [104]</li> </ul>	<ul> <li>No strong evidence</li> </ul>	
	<ul> <li>Sirtuins act as tumor suppressors [81,87–91]</li> <li>Sirtuins are mutated or deleted in various cancers [89,92]</li> </ul>		
Warburg-like metabolism	<ul> <li>Warburg-like metabolism is sufficient to drive tumorigenesis [94]</li> <li>Extracellular acidification suppresses the proper</li> </ul>		
	function of CD8 <sup>+</sup> T cells [197]	<ul> <li>No strong evidence</li> </ul>	
	• Elevated lactate levels trigger the polarization of CD4 <sup>+</sup>		
	<ul> <li>Lactate promotes T<sub>reg</sub> phenotype in CD4<sup>+</sup> T cells</li> </ul>		
	through the activation of NF- $\kappa$ B and FoxP3 [201]		
	<ul> <li>Lactate induces the protumorigenic M2-like polarization of macrophages [203 204]</li> </ul>		
Alterations in mitochondrial fitness	<ul> <li>mtDNA copy numbers and specific mutant mtDNA alleles have functional and clinical consequences for tumor cells [116 117]</li> </ul>		
	<ul> <li>Conplastic mice with different mtDNAs and levels of mitochondrial fitness have different tumor incidence</li> </ul>		
$N\Delta D^+$ Decline	<ul> <li>Depletion of NAD<sup>+</sup> leads to the inhibition of PARP-</li> </ul>		
NAD Decime	dependent DNA damage repair creating genomic instability and DNA damage [132]	No strong evidence	
	<ul> <li>NAD+ supplementation can enhance the tumor-killing afficiency of tumor infiltrating T cells [234]</li> </ul>		
	<ul> <li>NAD+ supplementation in rodents showed varying</li> </ul>		
	antitumor effects depending on dosage and organ [129]		
ROS	<ul> <li>ROS-mediated genomic instability promotes cancer development [163]</li> </ul>	<ul> <li>Antioxidant treatments promote tumor progression [166]</li> </ul>	
	<ul> <li>ROS leads to hyperactivation of mTOR [173,174]</li> </ul>	<ul> <li>ROS production inhibits melanoma</li> </ul>	
	ROS stabilizes protumorigenic transcription factors: HIE-19 [177] NEE2 2/NEE2 [178] NE- vB [179]	metastasis [166]	
Glutamine and TCA			
cycle intermediates			
	Cancers are addicted to glutamine [255]	<ul> <li>Glutamine promotes the development of proinflammatory T<sub>h</sub>1 and T<sub>h</sub>17 cells in vitro and in vivo [211]</li> </ul>	
		• a-KG acts as a metabolic regulator of	
		differentiation into $T_b1$ cells [213,214]	
Polyamines		<ul> <li>Spermidine promotes the homeostatic differentiation of CD4<sup>+</sup> T<sub>h</sub> and T<sub>reg</sub> cells</li> </ul>	
	Spermine favors macrophage polarization towards a protumoriganic M2 phageture [210]	<ul> <li>Spermidine contributes to the</li> </ul>	
	<ul> <li>Spermine mediates loss of cytotoxic activity in lymphokine-activated killer cells [219]</li> </ul>	rejuvenation of old T cells via the eIF5A- mediated regulation of TFEB and autophagy [226,227]	
Kynurenine	<ul> <li>Increased kynurenine levels are observed in old nation to [247, 249]</li> </ul>		
	<ul> <li>Kynurenine inhibits the proliferation of CD4<sup>+</sup>, CD8<sup>+</sup> T cells, and NK cells, and thereby restricts appropriate</li> </ul>	• No strong evidence	
	<ul> <li>Immune responses [242]</li> <li>Kynurenine reprograms CD4<sup>+</sup> T<sub>h</sub>17 cells into immune suppressive T<sub>reg</sub> cells [245]</li> </ul>		

Table	1.	(Continued).
-------	----	--------------

Aging-related changes	Favors tumorigenesis	Does not favor tumorigenesis	
Microbiome	<ul> <li>Pharmacological degradation of kynurenine increases the proliferation of CD8<sup>+</sup> lymphocytes <i>in vivo</i> [246]</li> <li>Dysbiosis promotes inflammation and tumorigenesis [256]</li> </ul>	<ul> <li>SCFAs enhance polarization effects set by the cytokine milieus present at the time of T-cell priming and differentiation [253]</li> <li>Fecal transplants from PD-1 responder patients decrease tumor burden and size in combination with anti-PD1 therapy [255]</li> <li><i>Faecalibacterium</i> promotes cytotoxic CD8+ T-cell recruitment to tumors [255]</li> <li>SCFAs decrease proinflammatory cytokine secretion by macrophages and dendritic cells [253]</li> </ul>	



**Fig. 1.** Protumorigenic effects of aging-associated metabolism in the epithelium. Aging promotes metabolic and redox rewiring, including an increase in ROS and the induction of a Warburg-like metabolism, which leads to the activation of protumorigenic and proliferation controlling signaling pathways PI3K/AKT/mTOR, p38, JNK, and ERK, and the upregulation of protumorigenic transcription factors such as HIF-1 $\alpha$ , NRF2 or NF-kB. On the other hand, aging also leads to the suppression of antitumorigenic pathways, including the inhibition of sirtuins via a decrease in NAD<sup>+</sup>. Together these age-induced alterations create a metabolic environment in aged epithelial cells that empowers carcinogenesis. ERK, extracellular signal-regulated kinase; HIF-1  $\alpha$ , hypoxia-inducible factor 1 $\alpha$ ; IGF-1, insulin-like growth factor 1; JNK, c-Jun N-terminal kinase; NAD, nicotinamide adenine dinucleotide; NF-kB, nuclear factor-kappa B; NRF2, nuclear factor erythroid 2-related factor 2; OXPHOS, oxidative phosphorylation; p38, p38 mitogen-activated protein kinase; PI3K/Akt/mTOR, phosphoinositide 3-kinases/protein kinase B/mechanistic target of rapamycin; ROS, reactive oxygen species.

kinase (AMPK; Box 1), and the sirtuins (Box 1) (reviewed in [1]).

High rates of cell proliferation and growth, as occur in cancer, require the increased production of proteins, nucleic acids, and lipids from available nutrients and energy. The ability of cancer cells to sense and adapt to ever-changing nutrient and energetic conditions is key to their survival [75]. In fact, a key feature of tumorigenesis is the abnormal activity of the nutrientsensing pathways. Often oncogenic mutations occur in nutrient-sensing pathways, such as activating mutations in *PIK3CA*, a major component of the insulin/IGF-1 pathway, or inactivating mutations in liver kinase 1 (*LKB1*), a major regulator of the energetic sensor, AMPK [76]. Several other oncogenic drivers, including oncogenic *KRAS* and the inactivation of *TP53*, result in deregulated nutrient-sensing pathways [75]. Together, these findings suggest that the deregulation of nutrient-sensing mechanisms is at the nexus of aging and cancer.

Direct evidence for this paradigm was provided by a landmark study in the late 1980s in which caloric restriction (CR), the most well-established antiaging intervention that functions largely by regulating nutrient-sensing pathways, was shown to block tumor formation in a mouse model of liver cancer [77]. CR increases cellular levels of NADPH and delays age-dependent downregulation of many cellular processes [73]; and broadly induces the expression of genes known to inhibit oxidative stress (e.g., Mt1, Mt2), inflammation (e.g., Nfkbia, Timp3) and tumorigenesis (e.g., Txnip, Zbtb16) [78]. At least partially, the efficacy of CR and fasting on cancer might be due to the indirect effects on body mass and the consequent metabolic changes. Moreover, rapamycin and rapalogs, inhibitors of the nutrient sensor mTOR and the only pharmacological agents that are generally agreed to modulate lifespan extension, are important therapeutic tools for the treatment of several types of cancer, such as mantle cell lymphoma, endometrial cancer, and renal cell carcinoma [79].

Another important energetic sensor is the sirtuin family of NAD<sup>+</sup>- dependent deacetylases. The decreased activity of sirtuins, particularly of SIRT1, SIRT3, and SIRT6 due to decreased NAD<sup>+</sup> availability, has been implicated in the aging process [10,80-83]. The reasons for decreased NAD<sup>+</sup> levels during aging are not fully established; however, defects in synthesis, increased consumption through NAD<sup>+</sup>-dependent enzymes such as CD38, and changes in the composition of cells or tissues with age have been suggested and tested as potential mechanistic explanations [84,85]. Sirtuins have also been shown to play important roles in suppressing tumorigenesis due to their ability to inhibit cell cycle progression, inactivate oncogenic pathways, promote DNA repair, and due to their key role as regulators of metabolism [80,86-90]. Highlighting how important the inactivation of sirtuins is for tumorigenesis, SIRT3 is deleted in 40% of human breast and ovarian tumors [88], and point mutations that inactivate SIRT6 are also a feature of many types of human cancer [91]. Thus, it is likely that as organisms age, the deregulation of nutrient- and energy-sensing pathways promotes a cellular environment that is conducive to tumorigenesis and that contributes, at least in part, to the increased incidence of cancer in the elderly.

### 2.2. Cellular energetics

Cancer cells have high energetic demands to maintain the growth rates that are necessary for tumors to thrive. Most mammalian cells use glucose as a fuel source. Glucose is metabolized through glycolysis in a multistep process, resulting in the creation of pyruvate. In healthy cells, under normal oxygen levels, much of this pyruvate enters the mitochondria, where it is oxidized in the TCA cycle to generate reducing equivalents to fuel oxidative phosphorylation (OXPHOS) and adenosine triphosphate (ATP) production. However, in cancer cells, much of the pyruvate from glycolysis is directed away from mitochondria and is used in lactate production, a phenomenon known as the Warburg effect [45]. In this metabolic process, glucose carbons are diverted to adjacent pathways, such as the pentose phosphate pathway, to fuel the generation of the building blocks needed for biomass production [45].

The prevailing view is that Warburg-like metabolic changes in cancer cells are caused by oncogenic mutations [45]. However, recent evidence shows that Warburg-like metabolism is sufficient to drive tumorigenesis in the absence of oncogenic driver mutations [92]. This suggests that the conditions that promote such metabolic shifts can function as tumor drivers even in the absence of oncogenic mutations. Strikingly, aging is associated with a reduction in OXPHOS and a concomitant increase in aerobic glycolysis in many tissues, including the brain, liver, and muscle [10,93-95]. Moreover, old animals have increased lactate levels in their serum and tissues, which is a hallmark of Warburg-like metabolism [96]. This phenomenon is thought to occur via age-induced epigenetic changes in major regulators of this switch, including the activation of mTOR and the stabilization under normoxic conditions of the hypoxia-inducible factor  $1\alpha$  (HIF-1 $\alpha$ ; Box 1) [10,52,97]. Interestingly and as mentioned above, inhibition of mTOR signaling is one of the few pharmacological agents that is well accepted to extend lifespan [98–101], while also having significant benefits as an anticancer therapy [102]. Furthermore, old age causes a shift in the lactate dehydrogenase (LDH) A/B ratio (Box 1) to favor the production of lactate, thereby diverting pyruvate away from the TCA cycle and OXPHOS [96].

Another important contributor to energetic regulation and the shift towards Warburg-like metabolism resides in the mtDNA, which consists of multiple copies of a small circular DNA molecule that encodes several components of the electron transport chain (ETC). Changes in mtDNA abundance and integrity have long been associated with aging [103–105]. Further supporting the role of mitochondrial fitness in the aging process, mice that express error-prone mitochondrial DNA polymerase (*Polg<sup>mut/mut</sup>*) highlight the importance of mtDNA alterations for the aging process. These mutant mice exhibit severe ETC deficiencies along with a

Age-induced metabolic reprogramming and cancer

premature-aging phenotype [106]. On the other hand, Polg<sup>mut/wt</sup>, which accumulate fewer point mutations than the *Polg<sup>mut/mut</sup>* mice show no alterations in lifespan [107]. Although mitochondrial fitness was not evaluated in the *Polg<sup>mut/wt</sup>* mice, when combined with the evidence provided by the  $Polg^{mut/mut}$  mice [106], this discrepancy suggests that a threshold exists for the phenotypic effects of mtDNA mutation loads. Moreover, a recent study showed reduced expression of a mitochondrial complex I subunit (NDUFS2) in mice has no effect on either lifespan or healthspan [108]. However, it is important to note that the reduced expression of NDUFS2 was not shown to significantly affect the function of complex I [108]. Notably in contrast with what occurs in normal mice, in all three of these mouse models, mitochondrial alterations occur early in life, with not many additional changes at older ages. Thus, it is possible that they simply reflect adaptation mechanisms to cope with the genetic alterations or the failure to hit the right cell types at the right time.

When considering the effects of age-induced mitochondrial changes on tumorigenesis, it is important to note that tumors rarely occur as a response to acute changes in OXPHOS. However, gradually accumulating age-induced mtDNA damage, such as that which occurs during the aging process, creates a persistent metabolically favorable environment for tumor growth [109]. This is supported by observations that mtDNA mutations are detected and shown to be involved in human ovarian, gastric, prostate [110], and pancreatic cancers [111]. Moreover, whole-genome sequencing analysis of tumors across a large array of tumor types has shown that specific mutant mtDNA alleles and mtDNA copy numbers have functional and clinical consequences for tumor cells [112,113]. As a result of evolutionary adaptations to the environment, specific mutations have become fixed in mtDNA, giving rise to divergent mtDNA haplotypes [114]. These mutationssuch as ones in the regulatory regions in the mtDNA control region and the established protein binding sites like for mitochondrial transcription factor A (TFAM; Box 1) [114], and mutations in the mitochondrialencoded subunits of the ETC, which regulate the proton pump part of the ETC and thereby affect mitochondrial ETC activity [115,116]—create diversity in mitochondrial fitness, which in turn create energetic traits that are better suited to specific environmental conditions (reviewed in [117]). Hence, different haplotypes give rise to different mitochondrial energetics and metabolic landscapes and have been shown to affect the rate of aging in conplastic mouse models (Box 1) [118]. Strikingly, different mtDNA haplotypes have a significant impact on tumor formation in both age-induced

[118] and forced *in vivo* mouse models of tumorigenesis via the induction of oncogenic drivers [119].

Carbon metabolism via glycolysis and the TCA cycle requires NAD<sup>+</sup> as an electron acceptor. Enhanced glycolvtic activity in cancer cells renders them dependent on the constant regeneration of NAD<sup>+</sup>, which is achieved through the LDH-mediated conversion of pyruvate into lactate [45]. Paradoxically, NAD<sup>+</sup> levels decline with age in both humans and mice in various tissues, including the brain, muscle, liver, skin, pancreas, and adipose tissue [10,120,121]. Not surprisingly NAD<sup>+</sup> supplementation has been widely tested as a therapeutic agent against aging disorders and to increase lifespan, with some success [122-124]. However, its role as an anticancer agent remains highly controversial. Cancer cells have a heightened need for NAD<sup>+</sup> to support their metabolism; therefore, providing cancer cells with extra NAD<sup>+</sup> supplementation can have devastating effects. On the other hand, there can be beneficial effects of  $NAD^+$ supplementation against tumor cells; for instance, Surjana et al. summarized the effects of niacin and nicotinamide treatments of rodents from different studies to show that the effects depended on dosage and organ [125]. This study suggests that there is an optimum level of NAD<sup>+</sup> levels in cells and deviation from these levels in either direction can aid in the tumorigenic process.

While the question of how tumor cells cope with the lower levels of NAD<sup>+</sup> available in an aged host remains largely unanswered, it is important to point out that NAD<sup>+</sup> biosynthesis is highly compartmentalized and tightly regulated [126,127]. Furthermore, LDH activity and lactate production increase with age, suggesting that even though total NAD<sup>+</sup> levels decline with age,  $NAD^+$  availability for LDH is retained [96]. In fact, the age-induced decline in NAD<sup>+</sup> levels in mice has been shown to directly cause a Warburg-like metabolic phenotype through the induction of HIF-1 $\alpha$  under normal oxygen tension [10]. NAD<sup>+</sup> is also an essential co-factor for poly (ADP-ribose) polymerases (PARPs), a family of enzymes involved in DNA repair, and cellular survival. Consequently, a decline in NAD<sup>+</sup> levels leads to the inhibition of PARP-dependent DNA damage repair and thereby might favor tumorigenesis by creating genomic instability and DNA damage [128]. Deficiency in PARP1 has also been shown to accelerate both aging and spontaneous tumorigenesis in mice [129], while high PARP expression has been associated with improved survival in pancreatic cancer patients [130].

### 2.3. ROS

ROS are a natural consequence of oxidative metabolism, during which highly unstable and reactive oxygen can oxidize many molecules and form reactive oxygen species [131]. ROS can be generated in various cellular compartments and via different mechanisms (reviewed in [132]). ROS are generally considered to be damaging agents that can structurally and/or functionally compromise macromolecules, such as nucleic acids, proteins, and lipids, and as such, they are essential mediators of oxidative stress. In order to ensure homeostasis, cells are equipped with a network of detoxification systems, which include enzymatic (e.g., superoxide dismutase, catalase, and thioredoxins) and nonenzymatic (e.g., glutathione, vitamin C, and vitamin E) antioxidants [132]. However, an environment that is conducive to damage is created when the balance between ROS and antioxidant systems is disturbed. enabling the development of several pathological conditions. Thus, it comes as no surprise that one of the oldest theories of aging lies within this paradigm. First stated in 1950s by Denham Harman, the free radical theory of aging hypothesizes that oxidative stress creates cumulative damage that acts as a key driver of aging. Evidence for this comes from observed elevations in ROS in aged tissues [59,133-137], which likely occur as a result of elevated ROS production due to defective OXPHOS, combined with a decline in antioxidant capacity. For example, two of the major antioxidants in cells, glutathione and reduced nicotinamide adenine dinucleotide phosphate (NADPH, which is a necessary co-factor for glutathione regeneration and which powers enzymatic antioxidant systems), become depleted as we age [138– 149]). Despite this, several studies have disproved this theory. These studies report that neither treating animal models with antioxidant products nor overexpressing antioxidant enzymes protects them from aging or from age-related diseases [150–155]. Thus, the relationship between oxidative stress and aging is complex and not completely understood [156]. In the context of cancer, high levels of ROS have also been hypothesized to be carcinogenic due to its ability to cause DNA damage, thereby acting as a mutagen, or by promoting genomic instability upon activation of topoisomerase II [132,157,158]. Moreover, mtDNA is even more susceptible to DNA damage than is nuclear DNA, as mtDNA lacks histones and its DNA repair mechanisms are more limited [159]. In fact, it has been shown that in 80 years of life, the frequency of mtDNA mutations increased about 5-fold [160]. As discussed above, mtDNA mutations have been detected and implicated in human ovarian [161], gastric [162], prostate [110], and pancreatic cancers [111], and were recently found to cause metabolic reprograming of human intestinal tumor cells and accelerate intestinal tumorigenesis in mice [109], highlighting the importance of mtDNA mutation frequency for tumorigenesis. Strikingly, as in the aging context, antioxidant treatments do not prevent tumorigenesis and in certain contexts have tumor-promoting effects [132,163]. This is thought to be partly due to the ability of ROS to drive cell cycle arrest, senescence (Box 1), and apoptosis when in excess.

In addition to their DNA damaging capability, ROS have important roles as signaling molecules with important consequences for tumorigenesis [164]. A classic example of a protumorigenic signaling pathway that is regulated by ROS is the PI3K/AKT/mTOR pathway-a key mediator of growth factor signaling that enables uncontrolled growth, proliferation, and survival [165-168]. These are essential features of malignant cells that allow them to thrive as cancers. One of the ways hyperactivation of the PI3K/AKT/ mTOR pathway is achieved in various tumor types [169] is by the ROS-mediated oxidation of key cysteine residues in negative regulators (phosphatase and tensin homolog (PTEN) and protein tyrosine phosphatase 1b (PTP1b)), which renders both proteins inactive [170,171]. Another major signaling node regulated by ROS via the inhibition of phosphatases is the mitogenactivated protein kinases (MAPKs) p38, c-Jun Nkinase (JNK), and extracellular signalterminal regulated kinase (ERK) [172]. Similar to the PI3K/ AKT/mTOR pathway, the MAPKs play a key role in the regulation of cellular growth and survival, and their abnormal activation is a known driver of uncontrolled cell proliferation and resistance to apoptosis [173]. ROS also regulate key protumorigenic transcription factors; they stabilize HIF-1a [174] and NFE2L2/NRF2 [175] and induce the transcriptional activity of nuclear factorkappa B (NF-  $\kappa$ B) [176], among others [177].

Cancer cells maintain a delicate balance of ROS since increased levels help promote many aspects of tumor initiation and progression, but the cells also express increased levels of ROS detoxifying systems to assure the ROS levels do not become deleterious [178,179]. Considering the parallels between the roles of ROS in aging and tumorigenesis, it is conceivable that a mild agedriven and noncytotoxic elevation of ROS might contribute to tumorigenesis by inducing cell growthpromoting pathways, and by evoking genetic abnormalities that can function as cancer drivers. However, it is likely that once ROS levels increase beyond a certain threshold, cell death mechanisms would be triggered that function as an antitumorigenic mechanism [180].

Together, the evidence discussed in this section draws the parallels between age-induced metabolic reprogramming and the metabolic changes that empower tumorigenesis and discuss the potential role of the age-induced metabolic reprogramming as a potential promoter of tumorigenesis. Proof of principle for this concept emerged from a recent study demonstrating that changes in circulatory metabolite levels due to global metabolic deregulation of the aged host contribute to the tumorigenic process. Specifically, methylmalonic acid (MMA), a byproduct of propionate metabolism, accumulates in circulation during aging and was shown to be sufficient to drive tumor progression *in vivo* [144].

# 3. Metabolic suppression of host defenses by the aging process

As with everything in evolution, a combination of the right traits and opportunities is needed to enable tumor initiation. Thus, the ability of a host to mount anticancer defenses (Box 1) is an essential determinant of cancer incidence. While innate immunity, the first line of defense, provides fast and effective immune responses, it lacks long-term memory. By comparison, adaptive immunity has a high degree of antigen specificity and allows for memory formation. The aging process severely alters both innate and adaptive immunity, leading to a decline in immune cell activation and in proper immune responses [181,182].

Cytotoxic CD8<sup>+</sup> T cells (Box 1) of the adaptive immune system are the most powerful anticancer immune response effectors and form the backbone of cancer immunotherapies [183]. Importantly, not only do T-cell numbers decline with age [184,185] but also the landscape of T cells changes with age, giving rise to T cells with reduced functionality [186]. Aged T cells in general also contribute to the general inflammatory state of old hosts by secreting a multitude of cytokines in their defective state [185–187]. A possible explanation for the decrease in T-cell numbers with age is thymic atrophy, which can hinder the development and maturation of T cells [188]. However, the exact mechanisms by which aging has such pronounced effects in the T-cell compartment remain unknown. Over the past decade, metabolism has emerged as a key regulator of both innate and adaptive immune functions. This seems to be particularly important in the T-cell compartment, where a significant degree of metabolic reprogramming is required for T-cell activation (reviewed in [189]). In an additional layer of complexity, functionally distinct T-cell subsets require distinct biosynthetic and energetic pathways to support their specific functional needs (reviewed in [190]). Interestingly, the metabolism of young and old immune cells has been shown to be

fundamentally different (reviewed in [182]). Thus, it is conceivable that the effects of aging on immune cell fate and function are partly mediated by metabolic alterations (Fig. 2 and Table 1).

### 3.1. Lactate

As described above, high lactate levels are a feature of the aging process. The root of many of lactate's functions lies in its ability to disrupt the pH balance of the extracellular environment in a phenomenon known as lactic acidosis [191]. Extracellular acidification caused by elevated lactate levels suppresses the proper functioning of CD8<sup>+</sup> T cells and thus reduces antitumor immunity [192]. This partly happens via the acidity-induced repression of a family of transcription factors, the nuclear factor of activated T-cell (NFAT; Box 1) family, in CD8<sup>+</sup> T and natural killer (NK) cells (Fig. 2). The repression of NFAT in these cells reduces their cytotoxicity by inhibiting their production of interferon-gamma (IFN- $\gamma$ ) production [193]. Lactate also shapes immune cell function beyond its ability to acidify the environment. Much like cancer cells, cytotoxic CD8<sup>+</sup> T cells depend on high rates of glycolysis and on the efficient secretion of lactate. High levels of serum lactate, as occur in aging, disrupt the concentration gradient that the lactate exporter, MCT-1, depends on, resulting in the accumulation of intracellular lactate [194]. High levels of intracellular lactate promote the reduction in NAD<sup>+</sup> to NADH, which disrupts glycolytic flux (Box 1) via glyceraldehyde-3phosphate dehydrogenase (GAPDH). This in turn leads to a failure to sustain the metabolic reprogramming that is necessary for CD8<sup>+</sup> effector T-cell activation and expansion [194,195]. Lactate also exerts immune suppressive effects in the CD4<sup>+</sup> T-cell compartment. Elevated lactate levels can trigger the polarization of CD4<sup>+</sup> T cells and can cause a reduction in the numbers of antitumor T helper (Th) 1 cell by inducing the SIRT1mediated deacetylation/degradation of the Th1 cellassociated transcription factor T-bet (Box 1) [196]. Lactate further contributes to immunosuppression by promoting a regulatory T-cell (Treg; Box 1) phenotype in  $CD4^+$  T cells through the activation of NF- $\kappa$ B and FoxP3; FoxP3 is a master regulator of immunosuppressive  $T_{reg}$  cell gene expression (Fig. 2) [196].

In addition to T cells, lactate can alter the function of other components of the immune system. For instance, high lactate levels impair the differentiation and activation of dendritic cells, which are antigenpresenting cells (APCs; Box 1) that incite the mobilization of other immune cells [197]. Through the stabilization of HIF-1 $\alpha$ , lactate induces the protumorigenic M2-like polarization of macrophages (Box 1; Fig. 2)



Fig. 2. Aging-induced metabolic suppression of host defenses. Age-induced metabolic alterations shape both innate and adaptive immunity resulting in a decline in immune cell activation and proper immune responses. Aging promotes a global decline in NAD<sup>+</sup> within the immune compartment due to an increase in the expression of NADase CD38 leading to a decline in cytotoxic effector activity and the induction of an inflammatory state. The decline in glutamine and a-KG availability with age further negatively impacts the function and differentiation of T cells by limiting substrate availability for TCA anaplerosis and redox reactions. The age-induced decline in spermidine inhibits autophagy in T cells and disrupts the homeostatic differentiation of CD4<sup>+</sup> T cells into specific subsets. On the other hand, aging also induces lactate production and promotes acidification; this acidification reduces the activity of NFAT and IFN-y in T cells and NK cells inhibiting their cytotoxic ability and induces protumorigenic M2-like polarization of macrophages via stabilization of HIF-1a and induction of Arg1 expression. Similarly, ageinduced elevation in spermine levels favors polarization towards a protumorigenic M2 macrophage phenotype, both dampening the immune response. In addition, lactate contributes to immunosuppression by promoting an activation of T<sub>reg</sub> phenotype of CD4<sup>+</sup> T cells through NFkB and FoxP3 activity. This phenomenon can also be controlled by the age-induced increase in kynurenine, which blocks the cytotoxic activity of T and NK cells. Advanced age might also cause a shift in microbiome composition affecting SCFAs in old hosts and thereby influence antitumor immunity. SCFAs have been shown to regulate innate immune cells via decreasing the secretion of proinflammatory cytokines by macrophages. On the other hand, SCFAs tend to enhance the polarization effects set by the cytokine milieus present at the time of T-cell priming and differentiation. Green and red arrows indicate increased or decreased level, respectively. a-KG, α-ketoglutarate; Arg1, arginase 1; CD38, cluster of differentiation 38; FoxP3, forkhead box P3; HIF-1 α, hypoxia-inducible factor 1α; INF-γ, interferon-gamma; M1MΦ, M1polarized macrophages; M2MΦ, M2-polarized macrophages; mTORC1, mechanistic target of rapamycin complex 1; NAD, nicotinamide adenine dinucleotide; NFAT, nuclear factor of activated T cells; NF-kB, nuclear factor-kappa B; NK cell, natural killer cell; p300, histone acetyltransferase p300; SCFAs, short-chain fatty acids; TCA, tricarboxylic acid; TFEB, transcription factor EB; T<sub>rea</sub>, regulatory T.

[198,199]. M2-polarized macrophages contribute to tumor progression by inducing angiogenesis through the secretion of vascular endothelial growth factor (VEGF) [199] and by inhibiting antitumor T- and NKcell activity. High expression levels of Arginase 1 (Arg1), a characteristic of M2-like macrophages, lead to the depletion of arginine, a metabolite that is essential for T- and NK-cell proliferation, thereby impairing T- and NK-cell-mediated antitumor immune response [200,201]. The ability of lactate to induce Arg1 in macrophages has recently been attributed to a novel epigenetic, post-translational modification called lactylation (Box 1) [202]. Histone lactylation, similarly to histone acetylation and methylation, which are wellestablished regulators of epigenetic transcription, has been shown to regulate the transcription of a metabolic gene set by regulating 28 different lysine residues in histones [202].

Lactate is now widely accepted to be a key messenger for immunosuppression that acts on multiple different immune cell types and at each stage of the immune cell response. As such, the increase in lactate as a function of age is likely to be a key mediator of the decrease in immune surveillance that helps tumors to thrive.

### 3.2. Glutamine and TCA cycle intermediates

In addition to increased glycolysis, T cells rely on continued TCA flux to power lipid synthesis, a key component of the metabolic reprogramming that underlies T-cell activation (reviewed in [190]). This is enabled not only by glucose-derived carbon but also by glutamine-mediated  $\alpha$ -ketoglutarate ( $\alpha$ -KG) production and consequent TCA cycle anaplerosis (Box 1) [203]. To achieve this, T cells induce the expression of the glutamine transporter, ASCT2 [204]. In addition to sustaining TCA anaplerosis, ASCT2 elevation is required for T-cell receptor (TCR)-stimulated activation of mTORC1 [205]—a major regulator of anabolic reprogramming and cell growth in CD4<sup>+</sup> T cells. The uptake of glutamine by CD4<sup>+</sup> T cells also influences the development of proinflammatory  $T_h1$  and  $T_h17$ cells in vitro and in vivo [206]. Interestingly, circulatory glutamine levels decline significantly during the aging process in both mice and humans [144,207], suggesting that a decline in glutamine availability might also contribute to age-induced T-cell dysfunction (Fig. 2). Serum levels of a-KG also significantly decrease as a function of age, further supporting the role of the agemediated decline in T-cell activation via the reduction in TCA cycle anaplerosis [144]. a-KG acts as a metabolic regulator of CD4<sup>+</sup> T-cell differentiation to T<sub>h</sub>1 cells by promoting the expression of T-bet, and by promoting the activation of CD4<sup>+</sup> T-cell proliferation via mTORC1 [208,209]. In addition, both glutamine and a-KG provide synergistic support for interleukin 4 (IL4)-induced M2 macrophage activation via Jmjd3dependent metabolic and epigenetic reprogramming [210,211]. Thus, glutamine and a-KG deficiency in old age can both impair cytotoxic adaptive immune function and shape the innate compartment away from antitumor to tumor-promoting activities.

### 3.3. Polyamines

Like lactate, polyamines (Box 1) can modulate several aspects of the immune response. Polyamine metabolism is a fundamental process that governs the ability of CD4<sup>+</sup> T<sub>h</sub> cells to polarize into different functional fates [212], highlighting the importance of cell-intrinsic polyamines for immune regulation. By contrast, the cell-extrinsic roles of polyamines in immune function are more complex and depend on the particular type of polyamine. For example, the polyamine spermine appears to have both proinflammatory and protumorigenic properties (Fig. 2). Macrophages are particularly susceptible to modulation by spermine due to their expression of fetuin, which binds spermine and facilitates its import [213]. Exposure to spermine favors macrophage polarization towards a protumorigenic M2 phenotype, dampening the immune response (Fig. 2) [214]. Additionally, exposure of peripheral

blood mononuclear cells (PBMCs) to spermine results in the loss of the adhesion molecules, CD11a and CD56, and in the loss of cytotoxic activity in lymphokine-activated killer cells. Conversely, polyamine spermidine has broad anti-inflammatory properties and supports the function of several types of immune cells. Dietary supplementation of spermidine promotes the homeostatic differentiation of CD4<sup>+</sup> T<sub>h</sub> and T<sub>reg</sub> cells, protecting them from the intestinal inflammatory disease [212], attenuating the development of colitis [215], and delaying senescence in mice [216]. This is especially relevant when considering the effects of aging in tumorigenesis as spermidine levels have been shown to decline with age [144], and its enrichment is observed in extremely long-living human populations [217]. Moreover, dietary supplementation of spermidine has been reported to extend lifespan [218,219]. This is associated with the ability of spermidine to restore the proper function of immune cells, as shown in vitro using human T cells from old donors [220]. Mechanistically, spermidine contributes to the rejuvenation of old B and T cells via the eIF5Amediated regulation of transcription factor EB (TFEB) and via the induction of autophagy (Fig. 2) [220,221]. Furthermore, spermidine contributes to autophagy in immune cells by inhibiting the acetyltransferase p300, which results in the deacetylation of multiple autophagy-related proteins [222]. Taken together, these reports demonstrate a central role in the agedependent decline in spermidine in deregulating immune function, suggesting that its consequent reduction in immune surveillance might be a key component of age-induced tumorigenesis. In support of this idea, dietary supplementation of spermidine in mice protects against hepatocellular carcinoma formation induced by chemical insults [223].

### 3.4. NAD<sup>+</sup>

NAD<sup>+</sup> is a key metabolite for cellular function and homeostasis. NAD<sup>+</sup> regulates the development, reprogramming, and differentiation of immune cells via its pivotal role in metabolic and redox reactions, and through signaling mechanisms regulated by sirtuins (reviewed in [224]). The inhibition of NAD<sup>+</sup> production severely hinders both innate and adaptive immunity [225–227] and was recently identified as an important modulator of tumor-infiltrating lymphocytes (TILs; Box 1) [228]. Interestingly, NAD<sup>+</sup> degradation is heightened upon the activation of various immune cells, including macrophages, T cells, and NK cells, through the induction of CD38, a NADase (Fig. 2) [229,230]. Importantly, CD38 is an established marker of T-cell exhaustion and is highly expressed in TILs that can be reinvigorated by antiprogrammed cell death protein 1 (PD1) immune checkpoint blockade (Box 1) as shown in vitro in human T cells isolated from non-small-cell lung cancer tumors [231]. This finding suggests that CD38-mediated NAD<sup>+</sup> depletion is an important regulator of T-cell function in TILs. In support of this notion, the in vivo suppression of CD38 in murine T cells increased NAD<sup>+</sup> levels and inhibited tumor growth [232]. The enrichment of CD38<sup>+</sup> immune cells is seen in different mouse tissues during aging [233] and is induced by signals secreted by senescent cells [230,233]. These reports suggest that CD38 induction is responsible for the age-induced decline in NAD<sup>+</sup> in immune cells, which consequently leads to immunosuppression and to the creation of an environment that is conducive to tumorigenesis. In support of this notion, NAD<sup>+</sup> supplementation has been found to enhance the tumor-killing efficacy of T cells and the adoptive chimeric antigen receptor T (CAR-T; Box 1) cell and anti-PD1 immune checkpoint blockade response in mouse models [228,234].

### 3.5. Kynurenine

Kynurenine (Box 1) is an intermediary in tryptophan catabolism that has well-established immunosuppressive functions. It inhibits the proliferation of CD4<sup>+</sup>, CD8<sup>+</sup> T cells, and NK cells and thereby regulates the availability of adaptive immune cells to mount appropriate immune responses (reviewed in [235]). In addition, kynurenine plays important roles in modulating the function of different immune cells. When human NK cells are isolated from healthy donors and treated with kynurenine in vitro, it blocks their cytotoxic activity through the cytokine-mediated upregulation of specific triggering receptors that are responsible for inducing NK-cellmediated killing (Fig. 2) [236]. Moreover, through its ability to activate the aryl hydrocarbon receptor, kynurenine reprograms CD4<sup>+</sup> T<sub>h</sub>17 cells into immune suppressive  $T_{reg}$  cells [237,238]. In addition, the pharmacological degradation of kynurenine using a pharmacologically optimized enzyme (PEGylated kynureninase), increased the proliferation of CD8<sup>+</sup> lymphocytes in vivo and their infiltration into mouse tumors [239], further highlighting the importance of kynurenine as a mechanism of immune suppression with important consequences for tumorigenesis. Importantly, kynurenine levels have been shown to increase with age and are linked to higher mortality in humans [240-242]. This suggests that targeting kynurenine production in older individuals might help to reinvigorate their immune systems and boost their antitumor immune responses.

### 3.6. Microbiome-derived short-chain fatty acids

Mounting evidence indicates that small molecules and metabolites produced by our gut commensals can have beneficial or detrimental effects on many human diseases, including cancer and aging. The relationship between a healthy microbiome (Box 1) and the host is generally a symbiotic one, with gut flora providing essential metabolic and immunomodulatory contributions [243]. When this balance is disrupted, however, the microbiome can be corrupted to aid the carcinogenic process, particularly in cancers of the gastrointestinal tract. Fascinatingly, bacteria also exist in human tumors themselves, largely localized intracellularly inside tumor cells or inside immune cells in the tumor microenvironment [244]. The main way by which the commensal microbiome shapes the host is through the secretion of short-chain fatty acids (SCFAs; Box 1), the end products of nondigestible carbohydrate fermentation, of which acetate, propionate, and butyrate are the most predominant [244]. Acetate is the most abundant SCFA, at about three times the levels of butyrate and propionate. It is formed by most enteric bacteria through fermentation or can be produced by a diverse group of acetogenic bacteria from hydrogen and carbon dioxide [245]. Propionate is largely formed through succinate metabolism, while most butyric acid is produced by butyryl-CoA:acetate CoA-transferase [245]. Microbial-derived SCFAs serve important regulatory functions in both the innate and adaptive components of the immune system (reviewed in [246]). At the innate level, SCFAs have a general anti-inflammatory function by decreasing proinflammatory cytokines secreted by macrophages and dendritic cells. On the other hand, even though SCFAs have different effects on the different T-cell subtypes, overall SCFAs tend to enhance the polarization effects set by the cytokine milieus present at the time of T-cell priming and differentiation (reviewed in [246]). Highlighting the importance of the host-microbiome interactions in cancer, some commensal bacteria have been shown to contribute to the onset and progression of cancer through the modulation of immune responses to tumors [247,248]. Moreover, tumor-bearing mice that were given fecal transplants from PD1 responder patients exhibited decreased tumor burden and tumor size when receiving anti-PD1 therapy [248]. Accordingly, Faecalibacterium promoted cytotoxic CD8+ T-cell recruitment to tumors, which may boost anti-PD1 responses [248]. Thus, microbiota-dependent immune system shaping may be one of the critical modes of altering host response to cancer therapy. Interestingly, changes in

the commensal bacteria that populate the human body have been shown to occur during the aging process [249,250]. Advanced age has been shown to be associated with a higher prevalence of opportunistic bacteria such as Pseudomonadota and Enterobacteriaceae and a depletion of Bifidobacterium and Clostridiale in the gut when compared to young individuals [251,252]. Interestingly, enrichment of enterobacteria has been shown to correlate with lower levels of fecal SCFAs [253]. Moreover, metformin, which as mentioned above has been shown to have longevity-promoting effects [53], was shown to significantly influence bacteria by suppressing bacterial folate metabolism and thereby mediating longevity in C. elegans [254]. Although no direct evidence exists to support the idea that the different composition of the microbiome directly contributes to the decline in immune surveillance with old age, it is possible that the shift in microbiome composition with age affects the levels of SCFAs in old hosts and thereby influence antitumor immunity.

# 4. Conclusions and future perspectives

Metabolic reprogramming is a hallmark of cell fate decisions; in order for a cell to commit to a specific fate and remain viable, it needs to adjust its metabolism to maintain homeostasis. This is especially important in the context of complex processes, such as tumorigenesis and immune cell activation. Therefore, both carcinogenesis and immune cell function are tightly linked to the metabolic status of an organism. Here, we propose that the metabolic changes that occur with aging promote an environment that is conducive to tumor initiation. When put into context, a significant body of published data supports this idea and clearly demonstrates the similarities between ageinduced metabolic reprogramming, and that occurs in cancer cells or suppresses protective mechanisms of anticancer immunity. The concepts highlighted in this review only skim the surface of our emerging understanding of how the metabolism of an aging host contributes to tumor initiation, and a significant amount of work is needed to illuminate this age-induced metabolic crosstalk and its bona fide importance for tumorigenesis. For instance, is age-induced metabolic reprogramming sufficient to drive tumorigenesis? What are the relative contributions of the metabolic alterations in the premalignant cells versus the metabolicdriven suppression of immune surveillance to tumor formation? Does adaptation to the aged host require different pathways than the ones employed in a young host? Are there differences in microbiome-derived

SCFAs that modulate tumorigenesis in young and old hosts? Can we take advantage of these potential differences to tailor anticancer therapies to the age of the host and thereby increase positive outcomes for cancer patients? Do antiaging strategies also have beneficial effects in the treatment of cancer? The answers to these questions remain largely unknown, in part due to the complexity and time requirements of modeling the aging process within a cancer context. However, if this hypothesis holds true, it opens the door to the possibility of utilizing antiaging strategies (such as intermittent fasting, rapamycin, NAD<sup>+</sup> precursors, and metformin, which target the metabolic changes that occur with age), to 'rejuvenate' the host, as a strategy to prevent age-induced tumor initiation and to enhance the effectiveness of anticancer therapies in the most vulnerable cancer patients: the elderly.

### Acknowledgements

We are grateful to members of the Gomes and DeNicola laboratories for critical discussions on this topic. The Gomes Lab is supported by a Pathway to Independence Award to A.P.G. from NCI (R00CA218686; R00CA218686-04S1), a New Innovator Award from OD/NIH (DP2AG0776980), the American Lung Association, the Florida Health Department Bankhead-Coley Research Program, the Florida Breast Cancer Foundation, The George Edgecomb Society of Moffitt Cancer Center and the Evolutionary Therapy Center of Excellence of Moffitt Cancer Center. Figures were made with BioRender.

# **Conflict of interest**

The authors declare no conflict of interest.

# **Author contributions**

APG conceived, wrote, and edited the manuscript. SD and DI wrote and edited the manuscript. SD conceived and APG edited the figures in the manuscript.

# References

- Lopez-Otin C, Blasco MA, Partridge L, Serrano M, Kroemer G. The hallmarks of aging. *Cell*. 2013;**153**:1194–217.
- 2 Gensler HL, Bernstein H. DNA damage as the primary cause of aging. Q Rev Biol. 1981;56:279–303.
- 3 Pinto M, Moraes CT. Mechanisms linking mtDNA damage and aging. *Free Radic Biol Med.* 2015;85:250–8.

- 4 Levine ME. Modeling the rate of senescence: can estimated biological age predict mortality more accurately than chronological age? *J Gerontol A Biol Sci Med Sci.* 2013;**68**:667–74.
- 5 Petkovich DA, Podolskiy DI, Lobanov AV, Lee SG, Miller RA, Gladyshev VN. Using DNA methylation profiling to evaluate biological age and longevity interventions. *Cell Metab.* 2017;25:954–960.e6.
- 6 Brown PJ, Wall MM, Chen C, Levine ME, Yaffe K, Roose SP, et al. Biological age, not chronological age, is associated with late-life depression. *J Gerontol A Biol Sci Med Sci.* 2018;73:1370–6.
- 7 Chen BH, Marioni RE, Colicino E, Peters MJ, Ward-Caviness CK, Tsai PC, et al. DNA methylation-based measures of biological age: metaanalysis predicting time to death. *Aging (Albany NY)*. 2016;8:1844–65.
- 8 Kresovich JK, Xu Z, O'Brien KM, Weinberg CR, Sandler DP, Taylor JA. Methylation-based biological age and breast cancer risk. *J Natl Cancer Inst.* 2019;**111**:1051–8.
- 9 Conboy IM, Conboy MJ, Wagers AJ, Girma ER, Weissman IL, Rando TA. Rejuvenation of aged progenitor cells by exposure to a young systemic environment. *Nature*. 2005;433:760–4.
- 10 Gomes AP, Price NL, Ling AJ, Moslehi JJ, Montgomery MK, Rajman L, et al. Declining NAD(+) induces a pseudohypoxic state disrupting nuclearmitochondrial communication during aging. *Cell*. 2013;155:1624–38.
- 11 Lu Y, Brommer B, Tian X, Krishnan A, Meer M, Wang C, et al. Reprogramming to recover youthful epigenetic information and restore vision. *Nature*. 2020;**588**:124–9.
- 12 Wang T, Tsui B, Kreisberg JF, Robertson NA, Gross AM, Yu MK, et al. Epigenetic aging signatures in mice livers are slowed by dwarfism, calorie restriction and rapamycin treatment. *Genome Biol.* 2017;18:57.
- 13 Fontana L, Meyer TE, Klein S, Holloszy JO. Longterm calorie restriction is highly effective in reducing the risk for atherosclerosis in humans. *Proc Natl Acad Sci USA*. 2004;**101**:6659–63.
- 14 Fontana L, Villareal DT, Weiss EP, Racette SB, Steger-May K, Klein S, et al. Calorie restriction or exercise: effects on coronary heart disease risk factors. A randomized, controlled trial. *Am J Physiol Endocrinol Metab.* 2007;**293**:E197–202.
- 15 Spadaro O, Youm Y, Shchukina I, Ryu S, Sidorov S, Ravussin A, et al. Caloric restriction in humans reveals immunometabolic regulators of health span. *Science*. 2022;**375**:671–7.
- 16 Kaeberlein M, Powers RW 3rd, Steffen KK, Westman EA, Hu D, Dang N, et al. Regulation of yeast replicative life span by TOR and Sch9 in response to nutrients. *Science*. 2005;**310**:1193–6.

- 17 Lin SJ, Defossez PA, Guarente L. Requirement of NAD and SIR2 for life-span extension by calorie restriction in Saccharomyces cerevisiae. *Science*. 2000;**289**:2126–8.
- 18 Lee GD, Wilson MA, Zhu M, Wolkow CA, de Cabo R, Ingram DK, et al. Dietary deprivation extends lifespan in Caenorhabditis elegans. *Aging Cell*. 2006;5:515–24.
- 19 Bruce KD, Hoxha S, Carvalho GB, Yamada R, Wang HD, Karayan P, et al. High carbohydrate-low protein consumption maximizes drosophila lifespan. *Exp Gerontol.* 2013;**48**:1129–35.
- 20 Davis TA, Bales CW, Beauchene RE. Differential effects of dietary caloric and protein restriction in the aging rat. *Exp Gerontol.* 1983;**18**:427–35.
- 21 Martens CR, Denman BA, Mazzo MR, Armstrong ML, Reisdorph N, McQueen MB, et al. Chronic nicotinamide riboside supplementation is well-tolerated and elevates NAD(+) in healthy middle-aged and older adults. *Nat Commun.* 2018;9:1286.
- 22 Elhassan YS, Kluckova K, Fletcher RS, Schmidt MS, Garten A, Doig CL, et al. Nicotinamide riboside augments the aged human skeletal muscle NAD(+) metabolome and induces transcriptomic and antiinflammatory signatures. *Cell Rep.* 2019;**28**:1717– 1728.e6.
- 23 Kiss T, Giles CB, Tarantini S, Yabluchanskiy A, Balasubramanian P, Gautam T, et al. Nicotinamide mononucleotide (NMN) supplementation promotes anti-aging miRNA expression profile in the aorta of aged mice, predicting epigenetic rejuvenation and antiatherogenic effects. *Geroscience*. 2019;41:419–39.
- 24 Kiss T, Nyul-Toth A, Balasubramanian P, Tarantini S, Ahire C, Yabluchanskiy A, et al. Nicotinamide mononucleotide (NMN) supplementation promotes neurovascular rejuvenation in aged mice: transcriptional footprint of SIRT1 activation, mitochondrial protection, anti-inflammatory, and antiapoptotic effects. *Geroscience*. 2020;42:527–46.
- 25 Ma S, Sun S, Geng L, Song M, Wang W, Ye Y, et al. Caloric restriction reprograms the single-cell transcriptional landscape of Rattus Norvegicus aging. *Cell*. 2020;**180**:984–1001.e22.
- 26 Ocampo A, Reddy P, Martinez-Redondo P, Platero-Luengo A, Hatanaka F, Hishida T, et al. In vivo amelioration of age-associated hallmarks by partial reprogramming. *Cell.* 2016;167:1719– 1733.e12.
- 27 Cancer statistics by the National Cancer Institute. September 25, 2020. Available from: https://www. cancer.gov/about-cancer/understanding/statistics
- 28 Valastyan S, Benaich N, Chang A, Reinhardt F, Weinberg RA. Concomitant suppression of three target genes can explain the impact of a microRNA on metastasis. *Genes Dev.* 2009;23:2592–7.

- 29 White MC, Holman DM, Goodman RA, Richardson LC. Cancer risk among older adults: time for cancer prevention to go silver. *Gerontologist*. 2019;**59**:S1–6.
- 30 Ecker BL, Kaur A, Douglass SM, Webster MR, Almeida FV, Marino GE, et al. Age-related changes in HAPLN1 increase lymphatic permeability and affect routes of melanoma metastasis. *Cancer Discov.* 2019;9:82–95.
- 31 Kaur A, Ecker BL, Douglass SM, Kugel CH, Webster MR, Almeida FV, et al. Remodeling of the collagen matrix in aging skin promotes melanoma metastasis and affects immune cell motility. *Cancer Discov*. 2019;9:64–81.
- 32 Han Z, Brown JR, Niederkorn JY. Growth and metastasis of intraocular tumors in aged mice. *Invest Ophthalmol Vis Sci.* 2016;**57**:2366–76.
- 33 Loughran EA, Leonard AK, Hilliard TS, Phan RC, Yemc MG, Harper E, et al. Aging increases susceptibility to ovarian cancer metastasis in murine allograft models and alters immune composition of peritoneal adipose tissue. *Neoplasia*. 2018;20:621–31.
- 34 Migkou M, Kastritis E, Roussou M, Gkotzamanidou M, Gavriatopoulou M, Nikitas N, et al. Short progression-free survival predicts for poor overall survival in older patients with multiple myeloma treated upfront with novel agent-based therapy. *Eur J Haematol.* 2011;87:323–9.
- 35 Thigpen T, Brady MF, Omura GA, Creasman WT, McGuire WP, Hoskins WJ, et al. Age as a prognostic factor in ovarian carcinoma. The Gynecologic Oncology Group experience. *Cancer.* 1993;71:606–14.
- 36 Deng F, Xu X, Lv M, Ren B, Wang Y, Guo W, et al. Age is associated with prognosis in serous ovarian carcinoma. J Ovarian Res. 2017;10:36.
- 37 Knudson AG. Mutation and cancer: statistical study of retinoblastoma. *Proc Natl Acad Sci USA*. 1971;68:820–3.
- 38 De Lorenzo MS, Baljinnyam E, Vatner DE, Abarzúa P, Vatner SF, Rabson AB. Caloric restriction reduces growth of mammary tumors and metastases. *Carcinogenesis*. 2011;**32**:1381–7.
- 39 Simone BA, Dan T, Palagani A, Jin L, Han SY, Wright C, et al. Caloric restriction coupled with radiation decreases metastatic burden in triple negative breast cancer. *Cell Cycle*. 2016;15:2265–74.
- 40 Alvarado A, Gil da Costa RM, Faustino-Rocha AI, Ferreira R, Lopes C, Oliveira PA, et al. Effects of exercise training on breast cancer metastasis in a rat model. *Int J Exp Pathol.* 2017;**98**:40–6.
- 41 Ligibel J. Lifestyle factors in cancer survivorship. J Clin Oncol. 2012;30:3697–704.
- 42 Armitage P, Doll R. The age distribution of cancer and a multi-stage theory of carcinogenesis. 1954. Int J Epidemiol. 2004;33:1174–9.

- 43 Nowell PC. The clonal evolution of tumor cell populations. *Science*. 1976;**194**:23–8.
- 44 Rozhok A, DeGregori J. A generalized theory of agedependent carcinogenesis. *Elife*. 2019;8:e39950.
- 45 Vander Heiden MG, Cantley LC, Thompson CB. Understanding the Warburg effect: the metabolic requirements of cell proliferation. *Science*. 2009;**324**:1029–33.
- 46 Pavlova NN, Thompson CB. The emerging hallmarks of cancer metabolism. *Cell Metab.* 2016;**23**:27–47.
- 47 Hanahan D, Weinberg RA. Hallmarks of cancer: the next generation. *Cell*. 2011;**144**:646–74.
- 48 Zhu A, Lee D, Shim H. Metabolic positron emission tomography imaging in cancer detection and therapy response. *Semin Oncol.* 2011;**38**:55–69.
- 49 Vander Heiden MG, DeBerardinis RJ. Understanding the intersections between metabolism and cancer biology. *Cell*. 2017;168:657–69.
- 50 Pavlova NN, Zhu J, Thompson CB. The hallmarks of cancer metabolism: still emerging. *Cell Metab.* 2022;**34**:355–77.
- 51 Schulze A, Harris AL. How cancer metabolism is tuned for proliferation and vulnerable to disruption. *Nature*. 2012;491:364–73.
- 52 Lopez-Otin C, Galluzzi L, Freije JMP, Madeo F, Kroemer G. Metabolic control of longevity. *Cell*. 2016;**166**:802–21.
- 53 Martin-Montalvo A, Mercken EM, Mitchell SJ, Palacios HH, Mote PL, Scheibye-Knudsen M, et al. Metformin improves healthspan and lifespan in mice. *Nat Commun.* 2013;4:2192.
- 54 Harrison DE, Strong R, Sharp ZD, Nelson JF, Astle CM, Flurkey K, et al. Rapamycin fed late in life extends lifespan in genetically heterogeneous mice. *Nature*. 2009;460:392–5.
- 55 Zhang Y, Bokov A, Gelfond J, Soto V, Ikeno Y, Hubbard G, et al. Rapamycin extends life and health in C57BL/6 mice. *J Gerontol A Biol Sci Med Sci.* 2014;69:119–30.
- 56 Bjedov I, Toivonen JM, Kerr F, Slack C, Jacobson J, Foley A, et al. Mechanisms of life span extension by rapamycin in the fruit fly *Drosophila melanogaster*. *Cell Metab.* 2010;11:35–46.
- 57 Anisimov VN, Zabezhinski MA, Popovich IG, Piskunova TS, Semenchenko AV, Tyndyk ML, et al. Rapamycin increases lifespan and inhibits spontaneous tumorigenesis in inbred female mice. *Cell Cycle*. 2011;10:4230–6.
- 58 Colman RJ, Anderson RM, Johnson SC, Kastman EK, Kosmatka KJ, Beasley TM, et al. Caloric restriction delays disease onset and mortality in rhesus monkeys. *Science*. 2009;**325**:201–4.
- 59 Hegab AE, Ozaki M, Meligy FY, Kagawa S, Ishii M, Betsuyaku T. High fat diet activates adult mouse lung stem cells and accelerates several aging-induced effects. *Stem Cell Res.* 2018;33:25–35.

- 60 Shadyab AH, Macera CA, Shaffer RA, Jain S, Gallo LC, LaMonte MJ, et al. Associations of accelerometer-measured and self-reported sedentary time with leukocyte telomere length in older women. *Am J Epidemiol.* 2017;**185**:172–84.
- 61 Gao Y, Zhang W, Zeng LQ, Bai H, Li J, Zhou J, et al. Exercise and dietary intervention ameliorate high-fat diet-induced NAFLD and liver aging by inducing lipophagy. *Redox Biol.* 2020;**36**:101635.
- 62 Tarumi T, Gonzales MM, Fallow B, Nualnim N, Pyron M, Tanaka H, et al. Central artery stiffness, neuropsychological function, and cerebral perfusion in sedentary and endurance-trained middle-aged adults. J Hypertens. 2013;31:2400–9.
- 63 Barzilai N, Gabriely I, Atzmon G, Suh Y, Rothenberg D, Bergman A. Genetic studies reveal the role of the endocrine and metabolic systems in aging. *J Clin Endocrinol Metab.* 2010;95:4493–500.
- 64 Langeveld M, Hollak CEM. Bone health in patients with inborn errors of metabolism. *Rev Endocr Metab Disord*. 2018;**19**:81–92.
- 65 van Karnebeek CD, Stockler S. Treatable inborn errors of metabolism causing intellectual disability: a systematic literature review. *Mol Genet Metab.* 2012;105:368–81.
- 66 Zebhauser PT, Cordts I, Hengel H, Haslinger B, Lingor P, Akman HO, et al. Characterization of cognitive impairment in adult polyglucosan body disease. *J Neurol.* 2022;269:2854–61.
- 67 Lauper JM, Krause A, Vaughan TL, Monnat RJ Jr. Spectrum and risk of neoplasia in Werner syndrome: a systematic review. *PLoS One.* 2013;8:e59709.
- 68 Tokita M, Kennedy SR, Risques RA, Chun SG, Pritchard C, Oshima J, et al. Werner syndrome through the lens of tissue and tumour genomics. *Sci Rep.* 2016;6:32038.
- 69 Fernandez P, Scaffidi P, Markert E, Lee JH, Rane S, Misteli T. Transformation resistance in a premature aging disorder identifies a tumor-protective function of BRD4. *Cell Rep.* 2014;9:248–60.
- 70 Atzmon G, Schechter C, Greiner W, Davidson D, Rennert G, Barzilai N. Clinical phenotype of families with longevity. J Am Geriatr Soc. 2004;52:274–7.
- 71 Atzmon G, Rincon M, Rabizadeh P, Barzilai N. Biological evidence for inheritance of exceptional longevity. *Mech Ageing Dev.* 2005;**126**:341–5.
- 72 Joseph SC, Delcastilo E, Loukas M, Osiro S. Common cancers in centenarians. *Med Sci Monit*. 2014;20:18–23.
- 73 Pavlidis N, Stanta G, Audisio RA. Cancer prevalence and mortality in centenarians: a systematic review. *Crit Rev Oncol Hematol.* 2012;83:145–52.
- 74 Gomes AP, Blenis J. A nexus for cellular homeostasis: the interplay between metabolic and signal transduction pathways. *Curr Opin Biotechnol.* 2015;**34**:110–7.

- 75 Torrence ME, Manning BD. Nutrient sensing in cancer. *Annu Rev Cancer Biol.* 2018;**2**:251–69.
- 76 Shaw RJ, Kosmatka M, Bardeesy N, Hurley RL, Witters LA, DePinho RA, et al. The tumor suppressor LKB1 kinase directly activates AMP-activated kinase and regulates apoptosis in response to energy stress. *Proc Natl Acad Sci USA*. 2004;101:3329–35.
- 77 Lagopoulos L, Stalder R. The influence of food intake on the development of diethylnitrosamine-induced liver tumours in mice. *Carcinogenesis*. 1987;**8**:33–7.
- 78 Swindell WR. Genes and gene expression modules associated with caloric restriction and aging in the laboratory mouse. *BMC Genomics*. 2009;10:585.
- 79 Guertin DA, Sabatini DM. Defining the role of mTOR in cancer. *Cancer Cell*. 2007;12:9–22.
- 80 Oberdoerffer P, Michan S, McVay M, Mostoslavsky R, Vann J, Park SK, et al. SIRT1 redistribution on chromatin promotes genomic stability but alters gene expression during aging. *Cell*. 2008;135:907–18.
- 81 Kanfi Y, Naiman S, Amir G, Peshti V, Zinman G, Nahum L, et al. The sirtuin SIRT6 regulates lifespan in male mice. *Nature*. 2012;**483**:218–21.
- 82 Someya S, Yu W, Hallows WC, Xu J, Vann JM, Leeuwenburgh C, et al. Sirt3 mediates reduction of oxidative damage and prevention of age-related hearing loss under caloric restriction. *Cell*. 2010;**143**:802–12.
- 83 Brown K, Xie S, Qiu X, Mohrin M, Shin J, Liu Y, et al. SIRT3 reverses aging-associated degeneration. *Cell Rep.* 2013;3:319–27.
- 84 McReynolds MR, Chellappa K, Baur JA. Age-related NAD(+) decline. *Exp Gerontol.* 2020;**134**:110888.
- 85 Schultz MB, Sinclair DA. Why NAD(+) declines during aging: it's destroyed. *Cell Metab.* 2016;23:965– 6.
- 86 Sebastian C, Zwaans BM, Silberman DM, Gymrek M, Goren A, Zhong L, et al. The histone deacetylase SIRT6 is a tumor suppressor that controls cancer metabolism. *Cell*. 2012;**151**:1185–99.
- 87 Bell EL, Emerling BM, Ricoult SJ, Guarente L. SirT3 suppresses hypoxia inducible factor 1alpha and tumor growth by inhibiting mitochondrial ROS production. *Oncogene*. 2011;**30**:2986–96.
- 88 Finley LW, Carracedo A, Lee J, Souza A, Egia A, Zhang J, et al. SIRT3 opposes reprogramming of cancer cell metabolism through HIF1alpha destabilization. *Cancer Cell*. 2011;19:416–28.
- 89 Jeong SM, Xiao C, Finley LW, Lahusen T, Souza AL, Pierce K, et al. SIRT4 has tumor-suppressive activity and regulates the cellular metabolic response to DNA damage by inhibiting mitochondrial glutamine metabolism. *Cancer Cell*. 2013;23:450–63.
- 90 Serrano L, Martinez-Redondo P, Marazuela-Duque A, Vazquez BN, Dooley SJ, Voigt P, et al. The tumor suppressor SirT2 regulates cell cycle progression and

genome stability by modulating the mitotic deposition of H4K20 methylation. *Genes Dev.* 2013;**27**:639–53.

- 91 Kugel S, Feldman JL, Klein MA, Silberman DM, Sebastian C, Mermel C, et al. Identification of and molecular basis for SIRT6 loss-of-function point mutations in cancer. *Cell Rep.* 2015;13:479–88.
- 92 Zhang Y, Xu Y, Lu W, Ghergurovich JM, Guo L, Blair IA, et al. Upregulation of antioxidant capacity and nucleotide precursor availability suffices for oncogenic transformation. *Cell Metab.* 2021;33:94– 109.e8.
- 93 Bowling AC, Mutisya EM, Walker LC, Price DL, Cork LC, Beal MF. Age-dependent impairment of mitochondrial function in primate brain. *J Neurochem*. 1993;60:1964–7.
- 94 Hagen TM, Yowe DL, Bartholomew JC, Wehr CM, Do KL, Park JY, et al. Mitochondrial decay in hepatocytes from old rats: membrane potential declines, heterogeneity and oxidants increase. *Proc Natl Acad Sci USA*. 1997;94:3064–9.
- 95 Trounce I, Byrne E, Marzuki S. Decline in skeletal muscle mitochondrial respiratory chain function: possible factor in ageing. *Lancet*. 1989;1:637–9.
- 96 Ross JM, Oberg J, Brene S, Coppotelli G, Terzioglu M, Pernold K, et al. High brain lactate is a hallmark of aging and caused by a shift in the lactate dehydrogenase A/B ratio. *Proc Natl Acad Sci USA*. 2010;107:20087–92.
- 97 Wu LE, Gomes AP, Sinclair DA. Geroncogenesis: metabolic changes during aging as a driver of tumorigenesis. *Cancer Cell*. 2014;**25**:12–9.
- 98 Vellai T, Takacs-Vellai K, Zhang Y, Kovacs AL, Orosz L, Muller F. Genetics: influence of TOR kinase on lifespan in *C. elegans. Nature*. 2003;**426**:620.
- 99 Kapahi P, Zid BM, Harper T, Koslover D, Sapin V, Benzer S. Regulation of lifespan in drosophila by modulation of genes in the TOR signaling pathway. *Curr Biol.* 2004;14:885–90.
- 100 Lamming DW, Ye L, Katajisto P, Goncalves MD, Saitoh M, Stevens DM, et al. Rapamycin-induced insulin resistance is mediated by mTORC2 loss and uncoupled from longevity. *Science*. 2012;335:1638–43.
- 101 Wu JJ, Liu J, Chen EB, Wang JJ, Cao L, Narayan N, et al. Increased mammalian lifespan and a segmental and tissue-specific slowing of aging after genetic reduction of mTOR expression. *Cell Rep.* 2013;4:913–20.
- 102 Li J, Kim SG, Blenis J. Rapamycin: one drug, many effects. *Cell Metab.* 2014;19:373–9.
- 103 Cortopassi GA, Arnheim N. Detection of a specific mitochondrial DNA deletion in tissues of older humans. *Nucleic Acids Res.* 1990;18:6927–33.
- 104 Gadaleta MN, Rainaldi G, Lezza AM, Milella F, Fracasso F, Cantatore P. Mitochondrial DNA copy number and mitochondrial DNA deletion in adult and senescent rats. *Mutat Res.* 1992;275:181–93.

- 105 Arnheim N, Cortopassi G. Deleterious mitochondrial DNA mutations accumulate in aging human tissues. *Mutat Res.* 1992;275:157–67.
- 106 Trifunovic A, Wredenberg A, Falkenberg M, Spelbrink JN, Rovio AT, Bruder CE, et al. Premature ageing in mice expressing defective mitochondrial DNA polymerase. *Nature*. 2004;**429**:417–23.
- 107 Vermulst M, Bielas JH, Kujoth GC, Ladiges WC, Rabinovitch PS, Prolla TA, et al. Mitochondrial point mutations do not limit the natural lifespan of mice. *Nat Genet*. 2007;**39**:540–3.
- 108 McElroy GS, Chakrabarty RP, D'Alessandro KB, Vasan K, Tan J, Stoolman JS, et al. Reduced expression of mitochondrial complex I subunit Ndufs2 does not impact healthspan in mice. *Sci Rep.* 2022;**12**:5196.
- 109 Smith AL, Whitehall JC, Bradshaw C, Gay D, Robertson F, Blain AP, et al. Age-associated mitochondrial DNA mutations cause metabolic remodelling that contributes to accelerated intestinal tumorigenesis. *Nat Cancer*. 2020;1:976–89.
- 110 Petros JA, Baumann AK, Ruiz-Pesini E, Amin MB, Sun CQ, Hall J, et al. mtDNA mutations increase tumorigenicity in prostate cancer. *Proc Natl Acad Sci* USA. 2005;**102**:719–24.
- 111 Schopf B, Weissensteiner H, Schafer G, Fazzini F, Charoentong P, Naschberger A, et al. OXPHOS remodeling in high-grade prostate cancer involves mtDNA mutations and increased succinate oxidation. *Nat Commun.* 2020;**11**:1487.
- 112 Gorelick AN, Kim M, Chatila WK, La K, Hakimi AA, Berger MF, et al. Respiratory complex and tissue lineage drive recurrent mutations in tumour mtDNA. *Nat Metab.* 2021;3:558–70.
- 113 Yuan Y, Ju YS, Kim Y, Li J, Wang Y, Yoon CJ, et al. Comprehensive molecular characterization of mitochondrial genomes in human cancers. *Nat Genet*. 2020;**52**:342–52.
- 114 Suissa S, Wang Z, Poole J, Wittkopp S, Feder J, Shutt TE, et al. Ancient mtDNA genetic variants modulate mtDNA transcription and replication. *PLoS Genet*. 2009;5:e1000474.
- 115 Zhang J, Zhao F, Fu Q, Liang M, Tong Y, Liu X, et al. Mitochondrial haplotypes may modulate the phenotypic manifestation of the LHON-associated m.14484T>C (MT-ND6) mutation in Chinese families. *Mitochondrion*. 2013;13:772–81.
- 116 Uittenbogaard M, Brantner CA, Fang Z, Wong LC, Gropman A, Chiaramello A. Novel insights into the functional metabolic impact of an apparent de novo m.8993T>G variant in the MT-ATP6 gene associated with maternally inherited form of Leigh syndrome. *Mol Genet Metab.* 2018;**124**:71–81.
- 117 Wallace DC. A mitochondrial paradigm of metabolic and degenerative diseases, aging, and cancer: a dawn

for evolutionary medicine. *Annu Rev Genet*. 2005;**39**:359–407.

- 118 Latorre-Pellicer A, Moreno-Loshuertos R, Lechuga-Vieco AV, Sanchez-Cabo F, Torroja C, Acin-Perez R, et al. Mitochondrial and nuclear DNA matching shapes metabolism and healthy ageing. *Nature*. 2016;**535**:561–5.
- 119 Chattopadhyay M, Jenkins EC, Lechuga-Vieco AV, Nie K, Fiel MI, Rialdi A, et al. The portrait of liver cancer is shaped by mitochondrial genetics. *Cell Rep.* 2022;**38**:110254.
- 120 Zhu XH, Lu M, Lee BY, Ugurbil K, Chen W. In vivo NAD assay reveals the intracellular NAD contents and redox state in healthy human brain and their age dependences. *Proc Natl Acad Sci USA*. 2015;**112**:2876–81.
- 121 Fang EF, Lautrup S, Hou Y, Demarest TG, Croteau DL, Mattson MP, et al. NAD(+) in aging: molecular mechanisms and translational implications. *Trends Mol Med.* 2017;23:899–916.
- 122 Fang EF, Kassahun H, Croteau DL, Scheibye-Knudsen M, Marosi K, Lu H, et al. NAD(+) replenishment improves lifespan and Healthspan in ataxia telangiectasia models via mitophagy and DNA repair. *Cell Metab.* 2016;24:566–81.
- 123 Hou Y, Lautrup S, Cordonnier S, Wang Y, Croteau DL, Zavala E, et al. NAD(+) supplementation normalizes key Alzheimer's features and DNA damage responses in a new AD mouse model with introduced DNA repair deficiency. *Proc Natl Acad Sci USA*. 2018;115:E1876–85.
- 124 Scheibye-Knudsen M, Mitchell SJ, Fang EF, Iyama T, Ward T, Wang J, et al. A high-fat diet and NAD(+) activate Sirt1 to rescue premature aging in cockayne syndrome. *Cell Metab.* 2014;20:840–55.
- 125 Surjana D, Halliday GM, Damian DL. Role of nicotinamide in DNA damage, mutagenesis, and DNA repair. J Nucleic Acids. 2010;2010:1–13.
- 126 Nikiforov A, Kulikova V, Ziegler M. The human NAD metabolome: functions, metabolism and compartmentalization. *Crit Rev Biochem Mol Biol.* 2015;**50**:284–97.
- 127 Johnson S, Imai SI. NAD (+) biosynthesis, aging, and disease. *F1000Res*. 2018;7:132.
- 128 Li J, Bonkowski MS, Moniot S, Zhang D, Hubbard BP, Ling AJ, et al. A conserved NAD(+) binding pocket that regulates protein-protein interactions during aging. *Science*. 2017;355:1312–7.
- 129 Piskunova TS, Yurova MN, Ovsyannikov AI, Semenchenko AV, Zabezhinski MA, Popovich IG, et al. Deficiency in poly(ADP-ribose) Polymerase-1 (PARP-1) accelerates aging and spontaneous carcinogenesis in mice. *Curr Gerontol Geriatr Res.* 2008;**2008**:754190.
- 130 Klauschen F, von Winterfeld M, Stenzinger A, Sinn BV, Budczies J, Kamphues C, et al. High nuclear

poly-(ADP-ribose)-polymerase expression is prognostic of improved survival in pancreatic cancer. *Histopathology*. 2012;**61**:409–16.

- 131 Kudryavtseva AV, Krasnov GS, Dmitriev AA, Alekseev BY, Kardymon OL, Sadritdinova AF, et al. Mitochondrial dysfunction and oxidative stress in aging and cancer. *Oncotarget*. 2016;7:44879–905.
- 132 Chio IIC, Tuveson DA. ROS in cancer: the burning question. *Trends Mol Med.* 2017;**23**:411–29.
- 133 Jacinto TA, Meireles GS, Dias AT, Aires R, Porto ML, Gava AL, et al. Increased ROS production and DNA damage in monocytes are biomarkers of aging and atherosclerosis. *Biol Res.* 2018;51:33.
- 134 Luceri C, Bigagli E, Femia AP, Caderni G, Giovannelli L, Lodovici M. Aging related changes in circulating reactive oxygen species (ROS) and protein carbonyls are indicative of liver oxidative injury. *Toxicol Rep.* 2018;**5**:141–5.
- 135 Kuka S, Tatarkova Z, Racay P, Lehotsky J, Dobrota D, Kaplan P. Effect of aging on formation of reactive oxygen species by mitochondria of rat heart. *Gen Physiol Biophys.* 2013;**32**:415–20.
- 136 Brawek B, Loffler M, Wagner K, Huppertz HJ, Wendling AS, Weyerbrock A, et al. Reactive oxygen species (ROS) in the human neocortex: role of aging and cognition. *Brain Res Bull.* 2010;81:484–90.
- 137 Petrosillo G, Matera M, Casanova G, Ruggiero FM, Paradies G. Mitochondrial dysfunction in rat brain with aging involvement of complex I, reactive oxygen species and cardiolipin. *Neurochem Int.* 2008;53:126– 31.
- 138 Dietzmann K. [Histochemical demonstration of enzyme activity changes in the regio postcentralis of the mouse brain under the effect of the aging process]. *J Hirnforsch.* 1981;22:405–8.
- 139 Kaliman PA, Konovalova EO. [Age-dependent characteristics of the regulation of cytoplasmic NADP+-dehydrogenases in the liver of rats on different diets]. Ukr Biokhim Zh (1978). 1985;57:38–42.
- 140 Lemeshko VV, Nikitchenko IV, Kaliman PA.
   [Enzymes of the antioxidant system of rat liver during aging]. Ukr Biokhim Zh (1978). 1983;55:523–8.
- 141 Maurya PK, Kumar P, Chandra P. Age-dependent detection of erythrocytes glucose-6-phosphate dehydrogenase and its correlation with oxidative stress. *Arch Physiol Biochem*. 2016;**122**:61–6.
- 142 Kil IS, Lee YS, Bae YS, Huh TL, Park JW. Modulation of NADP(+)-dependent isocitrate dehydrogenase in aging. *Redox Rep.* 2004;9:271–7.
- 143 Machado A, Ayala A, Gordillo E, Revilla E, Santa Maria C. Relationship between enzymatic activity loss and post-translational protein modification in aging. *Arch Gerontol Geriatr.* 1991;12:187–97.
- 144 Gomes AP, Ilter D, Low V, Endress JE, Fernandez-Garcia J, Rosenzweig A, et al. Age-induced

accumulation of methylmalonic acid promotes tumour progression. *Nature*. 2020;**585**:283-7.

- 145 Hazelton GA, Lang CA. Glutathione contents of tissues in the aging mouse. *Biochem J.* 1980;188:25–30.
- 146 Liu RM, Dickinson DA. Decreased synthetic capacity underlies the age-associated decline in glutathione content in fisher 344 rats. *Antioxid Redox Signal*. 2003;5:529–36.
- 147 Lang CA, Naryshkin S, Schneider DL, Mills BJ, Lindeman RD. Low blood glutathione levels in healthy aging adults. J Lab Clin Med. 1992;120:720–5.
- 148 Stohs SJ, Lawson T, Al-Turk WA. Changes in glutathione and glutathione metabolizing enzymes in erythrocytes and lymphocytes of mice as a function of age. *Gen Pharmacol.* 1984;15:267–70.
- 149 Sekhar RV, Patel SG, Guthikonda AP, Reid M, Balasubramanyam A, Taffet GE, et al. Deficient synthesis of glutathione underlies oxidative stress in aging and can be corrected by dietary cysteine and glycine supplementation. *Am J Clin Nutr.* 2011;94:847– 53.
- 150 Flores LC, Roman MG, Cunningham GM, Cheng C, Dube S, Allen C, et al. Continuous overexpression of thioredoxin 1 enhances cancer development and does not extend maximum lifespan in male C57BL/6 mice. *Pathobiol Aging Age Relat Dis.* 2018;8:1533754.
- 151 Perez VI, Van Remmen H, Bokov A, Epstein CJ, Vijg J, Richardson A. The overexpression of major antioxidant enzymes does not extend the lifespan of mice. *Aging Cell*. 2009;8:73–5.
- 152 Bjelakovic G, Nikolova D, Gluud LL, Simonetti RG, Gluud C. Mortality in randomized trials of antioxidant supplements for primary and secondary prevention: systematic review and meta-analysis. *JAMA*. 2007;297:842–57.
- 153 Bjelakovic G, Nikolova D, Gluud LL, Simonetti RG, Gluud C. Antioxidant supplements for prevention of mortality in healthy participants and patients with various diseases. *Cochrane Database Syst Rev.* 2012;**2012**:CD007176.
- 154 Sesso HD, Christen WG, Bubes V, Smith JP, MacFadyen J, Schvartz M, et al. Multivitamins in the prevention of cardiovascular disease in men: the Physicians' health study II randomized controlled trial. *JAMA*. 2012;**308**:1751–60.
- 155 Macpherson H, Pipingas A, Pase MP. Multivitaminmultimineral supplementation and mortality: a metaanalysis of randomized controlled trials. *Am J Clin Nutr.* 2013;97:437–44.
- 156 Gladyshev VN. The free radical theory of aging is dead. Long live the damage theory! Antioxid Redox Signal. 2014;20:727–31.
- 157 Cheung EC, Vousden KH. The role of ROS in tumour development and progression. *Nat Rev Cancer*. 2022;**22**:280–97.

- 158 Radisky DC, Levy DD, Littlepage LE, Liu H, Nelson CM, Fata JE, et al. Rac1b and reactive oxygen species mediate MMP-3-induced EMT and genomic instability. *Nature*. 2005;**436**:123–7.
- 159 Alexeyev M, Shokolenko I, Wilson G, LeDoux S. The maintenance of mitochondrial DNA integrity–critical analysis and update. *Cold Spring Harb Perspect Biol.* 2013;5:a012641.
- 160 Kennedy SR, Salk JJ, Schmitt MW, Loeb LA. Ultrasensitive sequencing reveals an age-related increase in somatic mitochondrial mutations that are inconsistent with oxidative damage. *PLoS Genet*. 2013;9:e1003794.
- 161 Liu VW, Shi HH, Cheung AN, Chiu PM, Leung TW, Nagley P, et al. High incidence of somatic mitochondrial DNA mutations in human ovarian carcinomas. *Cancer Res.* 2001;61:5998–6001.
- 162 Wu CW, Yin PH, Hung WY, Li AF, Li SH, Chi CW, et al. Mitochondrial DNA mutations and mitochondrial DNA depletion in gastric cancer. *Genes Chromosomes Cancer*. 2005;44:19–28.
- 163 Piskounova E, Agathocleous M, Murphy MM, Hu Z, Huddlestun SE, Zhao Z, et al. Oxidative stress inhibits distant metastasis by human melanoma cells. *Nature*. 2015;**527**:186–91.
- 164 Schieber M, Chandel NS. ROS function in redox signaling and oxidative stress. *Curr Biol.* 2014;24: R453–62.
- 165 Kim JH, Choi TG, Park S, Yun HR, Nguyen NNY, Jo YH, et al. Mitochondrial ROS-derived PTEN oxidation activates PI3K pathway for mTOR-induced myogenic autophagy. *Cell Death Differ*. 2018;25:1921–37.
- 166 Yalcin S, Marinkovic D, Mungamuri SK, Zhang X, Tong W, Sellers R, et al. ROS-mediated amplification of AKT/mTOR signalling pathway leads to myeloproliferative syndrome in Foxo3(-/-) mice. *EMBO J.* 2010;**29**:4118–31.
- 167 Zhao Y, Hu X, Liu Y, Dong S, Wen Z, He W, et al. ROS signaling under metabolic stress: cross-talk between AMPK and AKT pathway. *Mol Cancer*. 2017;16:79.
- 168 Koundouros N, Poulogiannis G. Phosphoinositide 3kinase/Akt signaling and redox metabolism in cancer. *Front Oncol.* 2018;8:160.
- 169 Liu GY, Sabatini DM. mTOR at the nexus of nutrition, growth, ageing and disease. *Nat Rev Mol Cell Biol.* 2020;21:183–203.
- 170 Lee SR, Yang KS, Kwon J, Lee C, Jeong W, Rhee SG. Reversible inactivation of the tumor suppressor PTEN by H2O2. J Biol Chem. 2002;277:20336–42.
- 171 Salmeen A, Andersen JN, Myers MP, Meng TC, Hinks JA, Tonks NK, et al. Redox regulation of protein tyrosine phosphatase 1B involves a sulphenylamide intermediate. *Nature*. 2003;423:769–73.
- 172 Son Y, Cheong YK, Kim NH, Chung HT, Kang DG, Pae HO. Mitogen-activated protein kinases and

- 173 Dhillon AS, Hagan S, Rath O, Kolch W. MAP kinase signalling pathways in cancer. *Oncogene*. 2007;**26**:3279–90.
- 174 Qutub AA, Popel AS. Reactive oxygen species regulate hypoxia-inducible factor 1alpha differentially in cancer and ischemia. *Mol Cell Biol.* 2008;**28**:5106–19.
- 175 Kansanen E, Kuosmanen SM, Leinonen H, Levonen AL. The Keap1-Nrf2 pathway: mechanisms of activation and dysregulation in cancer. *Redox Biol.* 2013;1:45–9.
- 176 Morgan MJ, Liu ZG. Crosstalk of reactive oxygen species and NF-kappaB signaling. *Cell Res.* 2011;21:103–15.
- 177 Turpaev KT. Reactive oxygen species and regulation of gene expression. *Biochemistry (Mosc)*. 2002;67:281–92.
- 178 Reczek CR, Chandel NS. The two faces of reactive oxygen species in cancer. *Annu Rev Cancer Biol.* 2017;1(1):79–98.
- 179 Liou GY, Storz P. Reactive oxygen species in cancer. *Free Radic Res.* 2010;**44**:479–96.
- 180 Wang Y, Qi H, Liu Y, Duan C, Liu X, Xia T, et al. The double-edged roles of ROS in cancer prevention and therapy. *Theranostics*. 2021;11:4839–57.
- 181 Montecino-Rodriguez E, Berent-Maoz B, Dorshkind K. Causes, consequences, and reversal of immune system aging. J Clin Invest. 2013;123:958–65.
- 182 Weyand CM, Goronzy JJ. Aging of the immune system. Mechanisms and therapeutic targets. Ann Am Thorac Soc. 2016;13(Suppl 5):S422–8.
- 183 Raskov H, Orhan A, Christensen JP, Gogenur I. Cytotoxic CD8(+) T cells in cancer and cancer immunotherapy. *Br J Cancer*. 2021;**124**:359–67.
- 184 Li M, Yao D, Zeng X, Kasakovski D, Zhang Y, Chen S, et al. Age related human T cell subset evolution and senescence. *Immun Ageing*. 2019;16:24.
- 185 Elyahu Y, Hekselman I, Eizenberg-Magar I, Berner O, Strominger I, Schiller M, et al. Aging promotes reorganization of the CD4 T cell landscape toward extreme regulatory and effector phenotypes. *Sci Adv.* 2019;5:eaaw8330.
- 186 Egorov ES, Kasatskaya SA, Zubov VN, Izraelson M, Nakonechnaya TO, Staroverov DB, et al. The changing landscape of naive T cell receptor repertoire with human aging. *Front Immunol.* 2018;9:1618.
- 187 Pieren DKJ, Smits NAM, van de Garde MDB, Guichelaar T. Response kinetics reveal novel features of ageing in murine T cells. Sci Rep. 2019;9:5587.
- 188 Palmer S, Albergante L, Blackburn CC, Newman TJ. Thymic involution and rising disease incidence with age. *Proc Natl Acad Sci USA*. 2018;115:1883–8.
- 189 Slack M, Wang T, Wang R. T cell metabolic reprogramming and plasticity. *Mol Immunol*. 2015;68:507–12.

- 190 MacIver NJ, Michalek RD, Rathmell JC. Metabolic regulation of T lymphocytes. *Annu Rev Immunol*. 2013;**31**:259–83.
- 191 de la Cruz-Lopez KG, Castro-Munoz LJ, Reyes-Hernandez DO, Garcia-Carranca A, Manzo-Merino J. Lactate in the regulation of tumor microenvironment and therapeutic approaches. *Front Oncol.* 2019;9:1143.
- 192 Ippolito L, Morandi A, Giannoni E, Chiarugi P. Lactate: a metabolic driver in the tumour landscape. *Trends Biochem Sci.* 2019;44:153–66.
- 193 Brand A, Singer K, Koehl GE, Kolitzus M, Schoenhammer G, Thiel A, et al. LDHA-associated lactic acid production blunts tumor immunosurveillance by T and NK cells. *Cell Metab.* 2016;24:657–71.
- 194 Fischer K, Hoffmann P, Voelkl S, Meidenbauer N, Ammer J, Edinger M, et al. Inhibitory effect of tumor cell-derived lactic acid on human T cells. *Blood*. 2007;**109**:3812–9.
- 195 Quinn WJ 3rd, Jiao J, TeSlaa T, Stadanlick J, Wang Z, Wang L, et al. Lactate limits T cell proliferation via the NAD(H) redox state. *Cell Rep.* 2020;**33**:108500.
- 196 Comito G, Iscaro A, Bacci M, Morandi A, Ippolito L, Parri M, et al. Lactate modulates CD4(+) T-cell polarization and induces an immunosuppressive environment, which sustains prostate carcinoma progression via TLR8/miR21 axis. Oncogene. 2019;**38**:3681–95.
- 197 He Z, Zhu X, Shi Z, Wu T, Wu L. Metabolic regulation of dendritic cell differentiation. *Front Immunol.* 2019;10:410.
- 198 Zhang L, Li S. Lactic acid promotes macrophage polarization through MCT-HIF1alpha signaling in gastric cancer. *Exp Cell Res.* 2020;**388**:111846.
- 199 Colegio OR, Chu NQ, Szabo AL, Chu T, Rhebergen AM, Jairam V, et al. Functional polarization of tumour-associated macrophages by tumour-derived lactic acid. *Nature*. 2014;**513**:559–63.
- 200 Geiger R, Rieckmann JC, Wolf T, Basso C, Feng Y, Fuhrer T, et al. L-arginine modulates T cell metabolism and enhances survival and anti-tumor activity. *Cell.* 2016;**167**:829–842.e13.
- 201 Oberlies J, Watzl C, Giese T, Luckner C, Kropf P, Muller I, et al. Regulation of NK cell function by human granulocyte arginase. *J Immunol.* 2009;**182**:5259–67.
- 202 Zhang D, Tang Z, Huang H, Zhou G, Cui C, Weng Y, et al. Metabolic regulation of gene expression by histone lactylation. *Nature*. 2019;**574**:575–80.
- 203 Chapman NM, Boothby MR, Chi H. Metabolic coordination of T cell quiescence and activation. *Nat Rev Immunol.* 2020;**20**:55–70.
- 204 Carr EL, Kelman A, Wu GS, Gopaul R, Senkevitch E, Aghvanyan A, et al. Glutamine uptake and metabolism are coordinately regulated by ERK/

MAPK during T lymphocyte activation. *J Immunol*. 2010;**185**:1037–44.

- 205 Nakaya M, Xiao Y, Zhou X, Chang JH, Chang M, Cheng X, et al. Inflammatory T cell responses rely on amino acid transporter ASCT2 facilitation of glutamine uptake and mTORC1 kinase activation. *Immunity*. 2014;40:692–705.
- 206 Johnson MO, Wolf MM, Madden MZ, Andrejeva G, Sugiura A, Contreras DC, et al. Distinct regulation of Th17 and Th1 cell differentiation by glutaminasedependent metabolism. *Cell*. 2018;**175**:1780–1795.e19.
- 207 Tomas-Loba A, Bernardes de Jesus B, Mato JM, Blasco MA. A metabolic signature predicts biological age in mice. *Aging Cell*. 2013;**12**:93–101.
- 208 Klysz D, Tai X, Robert PA, Craveiro M, Cretenet G, Oburoglu L, et al. Glutamine-dependent alphaketoglutarate production regulates the balance between T helper 1 cell and regulatory T cell generation. *Sci Signal.* 2015;8:ra97.
- 209 Sener Z, Cederkvist FH, Volchenkov R, Holen HL, Skalhegg BS. T helper cell activation and expansion is sensitive to glutaminase inhibition under both hypoxic and normoxic conditions. *PLoS One.* 2016;11: e0160291.
- 210 Liu PS, Wang H, Li X, Chao T, Teav T, Christen S, et al. Alpha-ketoglutarate orchestrates macrophage activation through metabolic and epigenetic reprogramming. *Nat Immunol.* 2017;**18**:985–94.
- 211 Liu M, Chen Y, Wang S, Zhou H, Feng D, Wei J, et al. Alpha-ketoglutarate modulates macrophage polarization through regulation of PPARgamma transcription and mTORC1/p70S6K pathway to ameliorate ALI/ARDS. *Shock*. 2020;**53**:103–13.
- 212 Puleston DJ, Baixauli F, Sanin DE, Edwards-Hicks J, Villa M, Kabat AM, et al. Polyamine metabolism is a central determinant of helper T cell lineage fidelity. *Cell*. 2021;**184**:4186–4202.e20.
- 213 Zhang M, Wang H, Tracey KJ. Regulation of macrophage activation and inflammation by spermine: a new chapter in an old story. *Crit Care Med.* 2000;28: N60–6.
- 214 Latour YL, Gobert AP, Wilson KT. The role of polyamines in the regulation of macrophage polarization and function. *Amino Acids*. 2020;**52**:151– 60.
- 215 Carriche GM, Almeida L, Stuve P, Velasquez L, Dhillon-LaBrooy A, Roy U, et al. Regulating T-cell differentiation through the polyamine spermidine. J Allergy Clin Immunol. 2021;147:335–348.e11.
- 216 Kibe R, Kurihara S, Sakai Y, Suzuki H, Ooga T, Sawaki E, et al. Upregulation of colonic luminal polyamines produced by intestinal microbiota delays senescence in mice. *Sci Rep.* 2014;**4**:4548.
- 217 Pucciarelli S, Moreschini B, Micozzi D, De Fronzo GS, Carpi FM, Polzonetti V, et al. Spermidine and

spermine are enriched in whole blood of nona/centenarians. *Rejuvenation Res.* 2012;**15**:590–5.

- 218 Eisenberg T, Knauer H, Schauer A, Buttner S, Ruckenstuhl C, Carmona-Gutierrez D, et al. Induction of autophagy by spermidine promotes longevity. *Nat Cell Biol.* 2009;**11**:1305–14.
- 219 Eisenberg T, Abdellatif M, Schroeder S, Primessnig U, Stekovic S, Pendl T, et al. Cardioprotection and lifespan extension by the natural polyamine spermidine. *Nat Med.* 2016;**22**:1428–38.
- 220 Alsaleh G, Panse I, Swadling L, Zhang H, Richter FC, Meyer A, et al. Autophagy in T cells from aged donors is maintained by spermidine and correlates with function and vaccine responses. *Elife*. 2020;**9**:e57950.
- 221 Zhang H, Alsaleh G, Feltham J, Sun Y, Napolitano G, Riffelmacher T, et al. Polyamines control eIF5A hypusination, TFEB translation, and autophagy to reverse B cell senescence. *Mol Cell*. 2019;76:110–125.e9.
- 222 Pietrocola F, Lachkar S, Enot DP, Niso-Santano M, Bravo-San Pedro JM, Sica V, et al. Spermidine induces autophagy by inhibiting the acetyltransferase EP300. *Cell Death Differ*. 2015;**22**:509–16.
- 223 Yue F, Li W, Zou J, Jiang X, Xu G, Huang H, et al. Spermidine prolongs lifespan and prevents liver fibrosis and hepatocellular carcinoma by activating MAP1S-mediated autophagy. *Cancer Res.* 2017;77:2938–51.
- 224 Grahnert A, Grahnert A, Klein C, Schilling E, Wehrhahn J, Hauschildt S. Review: NAD +: a modulator of immune functions. *Innate Immun.* 2011;**17**:212–33.
- 225 Skokowa J, Lan D, Thakur BK, Wang F, Gupta K, Cario G, et al. NAMPT is essential for the G-CSFinduced myeloid differentiation via a NAD(+)-sirtuin-1-dependent pathway. *Nat Med.* 2009;**15**:151–8.
- 226 Bruzzone S, Fruscione F, Morando S, Ferrando T, Poggi A, Garuti A, et al. Catastrophic NAD+ depletion in activated T lymphocytes through Nampt inhibition reduces demyelination and disability in EAE. *PLoS One.* 2009;**4**:e7897.
- 227 Pittelli M, Cavone L, Lapucci A, Oteri C, Felici R, Niccolai E, et al. Nicotinamide phosphoribosyltransferase (NAMPT) activity is essential for survival of resting lymphocytes. *Immunol Cell Biol.* 2014;92:191–9.
- 228 Wang Y, Wang F, Wang L, Qiu S, Yao Y, Yan C, et al. NAD(+) supplement potentiates tumor-killing function by rescuing defective TUB-mediated NAMPT transcription in tumor-infiltrated T cells. *Cell Rep.* 2021;36:109516.
- 229 Piedra-Quintero ZL, Wilson Z, Nava P, Guerau-de-Arellano M. CD38: an immunomodulatory molecule in inflammation and autoimmunity. *Front Immunol.* 2020;11:597959.

- 230 Covarrubias AJ, Kale A, Perrone R, Lopez-Dominguez JA, Pisco AO, Kasler HG, et al. Senescent cells promote tissue NAD(+) decline during ageing via the activation of CD38(+) macrophages. *Nat Metab.* 2020;2:1265–83.
- 231 Wu P, Zhao L, Chen Y, Xin Z, Lin M, Hao Z, et al. CD38 identifies pre-activated CD8+ T cells which can be reinvigorated by anti-PD-1 blockade in human lung cancer. *Cancer Immunol Immunother*. 2021;70:3603–16.
- 232 Chatterjee S, Daenthanasanmak A, Chakraborty P, Wyatt MW, Dhar P, Selvam SP, et al. CD38-NAD(+) Axis regulates immunotherapeutic anti-tumor T cell response. *Cell Metab.* 2018;27:85–100.e8.
- 233 Chini CCS, Peclat TR, Warner GM, Kashyap S, Espindola-Netto JM, de Oliveira GC, et al. CD38 ecto-enzyme in immune cells is induced during aging and regulates NAD(+) and NMN levels. *Nat Metab.* 2020;2:1284–304.
- 234 Yu YR, Imrichova H, Wang H, Chao T, Xiao Z, Gao M, et al. Disturbed mitochondrial dynamics in CD8(+) TILs reinforce T cell exhaustion. *Nat Immunol.* 2020;21:1540–51.
- 235 Proietti E, Rossini S, Grohmann U, Mondanelli G. Polyamines and kynurenines at the intersection of immune modulation. *Trends Immunol.* 2020;**41**:1037– 50.
- 236 Della Chiesa M, Carlomagno S, Frumento G, Balsamo M, Cantoni C, Conte R, et al. The tryptophan catabolite L-kynurenine inhibits the surface expression of NKp46- and NKG2D-activating receptors and regulates NK-cell function. *Blood.* 2006;**108**:4118–25.
- 237 Mezrich JD, Fechner JH, Zhang X, Johnson BP, Burlingham WJ, Bradfield CA. An interaction between kynurenine and the aryl hydrocarbon receptor can generate regulatory T cells. *J Immunol.* 2010;**185**:3190– 8.
- 238 Siska PJ, Jiao J, Matos C, Singer K, Berger RS, Dettmer K, et al. Kynurenine induces T cell fat catabolism and has limited suppressive effects in vivo. *EBioMedicine*. 2021;74:103734.
- 239 Triplett TA, Garrison KC, Marshall N, Donkor M, Blazeck J, Lamb C, et al. Reversal of indoleamine 2,3dioxygenase-mediated cancer immune suppression by systemic kynurenine depletion with a therapeutic enzyme. *Nat Biotechnol.* 2018;**36**:758–64.
- 240 van der Goot AT, Zhu W, Vazquez-Manrique RP, Seinstra RI, Dettmer K, Michels H, et al. Delaying aging and the aging-associated decline in protein homeostasis by inhibition of tryptophan degradation. *Proc Natl Acad Sci USA*. 2012;109:14912–7.
- 241 de Bie J, Guest J, Guillemin GJ, Grant R. Central kynurenine pathway shift with age in women. J Neurochem. 2016;136:995–1003.

- 242 Kim BJ, Lee SH, Koh JM. Clinical insights into the kynurenine pathway in age-related diseases. *Exp Gerontol.* 2020;**130**:110793.
- 243 Clemente JC, Ursell LK, Parfrey LW, Knight R. The impact of the gut microbiota on human health: an integrative view. *Cell*. 2012;**148**:1258–70.
- 244 Nejman D, Livyatan I, Fuks G, Gavert N, Zwang Y, Geller LT, et al. The human tumor microbiome is composed of tumor type-specific intracellular bacteria. *Science*. 2020;368:973–80.
- 245 Louis P, Hold GL, Flint HJ. The gut microbiota, bacterial metabolites and colorectal cancer. *Nat Rev Microbiol.* 2014;**12**:661–72.
- 246 Kim CH. Immune regulation by microbiome metabolites. *Immunology*. 2018;**154**:220–9.
- 247 Kostic AD, Chun E, Robertson L, Glickman JN, Gallini CA, Michaud M, et al. Fusobacterium nucleatum potentiates intestinal tumorigenesis and modulates the tumor-immune microenvironment. *Cell Host Microbe*. 2013;14:207–15.
- 248 Gopalakrishnan V, Spencer CN, Nezi L, Reuben A, Andrews MC, Karpinets TV, et al. Gut microbiome modulates response to anti-PD-1 immunotherapy in melanoma patients. *Science*. 2018;**359**:97–103.
- 249 Cedergren R, Gray MW, Abel Y, Sankoff D. The evolutionary relationships among known life forms. J Mol Evol. 1988;28:98–112.
- 250 Kim M, Benayoun BA. The microbiome: an emerging key player in aging and longevity. *Transl Med Aging*. 2020;4:103–16.
- 251 Odamaki T, Kato K, Sugahara H, Hashikura N, Takahashi S, Xiao JZ, et al. Age-related changes in gut microbiota composition from newborn to centenarian: a cross-sectional study. *BMC Microbiol.* 2016;16:90.
- 252 Hopkins MJ, Sharp R, Macfarlane GT. Age and disease related changes in intestinal bacterial populations assessed by cell culture, 16S rRNA abundance, and community cellular fatty acid profiles. *Gut.* 2001;48:198–205.
- 253 Dostal A, Chassard C, Hilty FM, Zimmermann MB, Jaeggi T, Rossi S, et al. Iron depletion and repletion with ferrous sulfate or electrolytic iron modifies the composition and metabolic activity of the gut microbiota in rats. J Nutr. 2012;142:271–7.
- 254 Cabreiro F, Au C, Leung KY, Vergara-Irigaray N, Cocheme HM, Noori T, et al. Metformin retards aging in *C. elegans* by altering microbial folate and methionine metabolism. *Cell.* 2013;**153**:228–39.
- 255 Wise DR, Thompson CB. Glutamine addiction: a new therapeutic target in cancer. *Trends Biochem Sci.* 2010;**35**:427–33.
- 256 Biragyn A, Ferrucci L. Gut dysbiosis: a potential link between increased cancer risk in ageing and inflammaging. *Lancet Oncol.* 2018;19:e295–304.