GENOME SEQUENCES





Complete Genome Sequence of *Streptococcus mutans* Strain MD, Which Produces Highly Potent Mutacins

Saswati Biswas,^a DIndranil Biswas^a

^aDepartment of Microbiology, Molecular Genetics, and Immunology, University of Kansas Medical Center, Kansas City, Kansas, USA

ABSTRACT Here, we report the complete genome sequence of *Streptococcus mutans* strain MD, which produces potent mutacins capable of inhibiting streptococci. MD is a relatively uncharacterized strain whose genome information was unavailable. This study provides useful information for comparative genomic study and for understanding the repertoire of mutacins in *S. mutans*.

S(1). The human oral microbiome is composed of >700 different bacterial species, and >30% of them belong to the genus *Streptococcus* (2, 3). *S. mutans* produces numerous small antibacterial peptide bacteriocins, called mutacins; they inhibit the growth of competing bacteria, including streptococci. This suppression of other bacteria allows *S. mutans* to become one of the dominant species in the oral cavity to form dental caries. Bacteriocins are broadly classified into two classes, namely, lantibiotics (class I), which are peptides containing dehydrated amino acids and unusual amino acids such as lanthionine, and nonlantibiotics (class II), which are linear unchanged peptides. The nature and potency of mutacins produced by *S. mutans* are highly strain dependent and vary significantly (4–6).

Our group has been working with a prolific mutacin-producing strain that was previously referred to as T8 (7) (M. Duncan, personal communication). However, this T8 strain is not the same as the original T8 strain that produces mutacin II (8). We also observed that this particular isolate displayed a very different inhibitory spectrum and produced a two-peptide lantibiotic, Smb, similar to that produced by the GS-5 strain (7, 9, 10); however, its inhibitory potency is greater than that of the Smb produced by the GS-5 strain. Therefore, we renamed this isolate MD to distinguish it from the original T8 strain (8). To study this isolate further, we determined its complete genome sequence.

The bacterial culture was cultivated at 37°C in Todd-Hewitt medium (BBL) supplemented with 0.2% yeast extract, under microaerophilic conditions. Genomic DNA was isolated using a MasterPure DNA purification kit (Lucigen) as described previously (11–13). The quality and quantity of the isolated genomic DNA were verified by using gel electrophoresis and a NanoDrop spectrophotometer (Thermo Fisher Scientific), respectively. DNA was sheared using a g-TUBE (Covaris), and SMRTbell DNA libraries were prepared using the Express template preparation kit v2.0 (Pacific Biosciences) according to the manufacturer's protocol. Samples were pooled into a single multiplexed library, size selected using BluePippin (Sage Sciences), and sequenced on a Sequel II system with Sequel II chemistry v1.0, at SNPsaurus. Raw reads were converted to the fasta format using SAMtools (14) and were quality controlled using FastQC v0.10.1 with default parameters. Canu v1.7 (15) was used with default settings to assemble the genome from 29,320 raw reads (N_{50} value of 9,099 bp) with a total of 221,612,731 bases. The assembled genome had 113-fold genome coverage. The genome harbors a circular chromosome of 2,009,428 bp (GC content of 36.89%) and a plasmid of 13,108 bp (GC content of 43%) with homology to pVA838 (16). The genome

Citation Biswas S, Biswas I. 2020. Complete genome sequence of *Streptococcus mutans* strain MD, which produces highly potent mutacins. Microbiol Resour Announc 9:e00616-20. https://doi.org/10.1128/MRA.00616-20.

Editor David Rasko, University of Maryland School of Medicine

Copyright © 2020 Biswas and Biswas. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Indranil Biswas, ibiswas@kumc.edu.

Received 26 May 2020 Accepted 16 July 2020 Published 13 August 2020 annotation was carried out using the IGS Prokaryotic Annotation Pipeline at the Institute for Genome Sciences at the University of Maryland (17) and set *oriC/dnaA* as the start.

Genome analysis by CRISPRCasFinder (18) suggests that the MD genome harbors three CRISPR/Cas loci, which is similar to the *S. mutans* GS-5 strain. We analyzed the genome to predict putative biosynthetic gene clusters (BGCs), including those for bacteriocins (mutacin), using the antiSMASH (19) and BAGEL4 (20) servers with default search options. As expected, the MD genome contains genes for the lantibiotic Smb (7, 10) and several nonlantibiotics, as well as several BGC loci. The genome sequencing also identified three new methylation motifs in *S. mutans* in addition to the GATC motif. The new motifs are ATGCAT (m6A), CNGCGNCGGNTNNNG, and CTNNNNTATANCNNNAC. This study will add valuable information regarding the evolution and production of the lantibiotic Smb among various *S. mutans* isolates.

Data availability. The complete genome sequence of the *S. mutans* MD strain has been deposited in GenBank under the accession numbers CP044493 (chromosome) and CP044494 (plasmid). The GenBank assembly number for the genome is GCA_008831325.1. Raw reads have been deposited in the SRA under the accession number SRR11818585.

ACKNOWLEDGMENTS

We thank Margaret Duncan for providing the MD strain. We also thank the Institute for Genome Sciences Analysis Engine service at the University of Maryland School of Medicine for functional annotation of the sequences and assistance with the submission of the sequences to GenBank.

This work was supported by National Institute of Dental and Craniofacial Research grant DE026955 awarded to I.B. and S.B. and grant DE026937 awarded to I.B.

REFERENCES

- Loesche WJ. 1986. Role of *Streptococcus mutans* in human dental decay. Microbiol Rev 50:353–380. https://doi.org/10.1128/MMBR.50.4.353-380 .1986.
- Dewhirst FE, Chen T, Izard J, Paster BJ, Tanner AC, Yu WH, Lakshmanan A, Wade WG. 2010. The human oral microbiome. J Bacteriol 192: 5002–5017. https://doi.org/10.1128/JB.00542-10.
- Costalonga M, Herzberg MC. 2014. The oral microbiome and the immunobiology of periodontal disease and caries. Immunol Lett 162:22–38. https://doi.org/10.1016/j.imlet.2014.08.017.
- Kamiya RU, Hofling JF, Goncalves RB. 2008. Frequency and expression of mutacin biosynthesis genes in isolates of *Streptococcus mutans* with different mutacin-producing phenotypes. J Med Microbiol 57:626–635. https://doi.org/10.1099/jmm.0.47749-0.
- Kamiya RU, Taiete T, Goncalves RB. 2011. Mutacins of *Streptococcus mutans*. Braz J Microbiol 42:1248–1258. https://doi.org/10.1590/S1517 -83822011000400001.
- Merritt J, Qi F. 2012. The mutacins of *Streptococcus mutans*: regulation and ecology. Mol Oral Microbiol 27:57–69. https://doi.org/10.1111/j .2041-1014.2011.00634.x.
- Biswas S, Biswas I. 2013. SmbFT, a putative ABC transporter complex, confers protection against the lantibiotic Smb in streptococci. J Bacteriol 195:5592–5601. https://doi.org/10.1128/JB.01060-13.
- Novak J, Caufield PW, Miller EJ. 1994. Isolation and biochemical characterization of a novel lantibiotic mutacin from *Streptococcus mutans*. J Bacteriol 176:4316–4320. https://doi.org/10.1128/jb.176.14.4316-4320.1994.
- Parrot M, Charest M, Lavoie MC. 1989. Production of mutacin-like substances by *Streptococcus mutans*. Can J Microbiol 35:366–372. https:// doi.org/10.1139/m89-056.
- Yonezawa H, Kuramitsu HK. 2005. Genetic analysis of a unique bacteriocin, Smb, produced by *Streptococcus mutans* GS5. Antimicrob Agents Chemother 49:541–548. https://doi.org/10.1128/AAC.49.2.541-548.2005.
- 11. Biswas S, Biswas I. 2016. Complete genome sequence of *Lactobacillus rhamnosus* strain LRB. Genome Announc 4:e01208-16. https://doi.org/10 .1128/genomeA.01208-16.

- Biswas S, Biswas I. 2014. A conserved streptococcal membrane protein, LsrS, exhibits a receptor-like function for lantibiotics. J Bacteriol 196: 1578–1587. https://doi.org/10.1128/JB.00028-14.
- Biswas S, Biswas I. 2012. Complete genome sequence of Streptococcus mutans GS-5, a serotype c strain. J Bacteriol 194:4787–4788. https://doi .org/10.1128/JB.01106-12.
- Ramirez-Gonzalez RH, Bonnal R, Caccamo M, Maclean D. 2012. Biosamtools: Ruby bindings for SAMtools, a library for accessing BAM files containing high-throughput sequence alignments. Source Code Biol Med 7:6. https://doi.org/10.1186/1751-0473-7-6.
- Koren S, Walenz BP, Berlin K, Miller JR, Bergman NH, Phillippy AM. 2017. Canu: scalable and accurate long-read assembly via adaptive k-mer weighting and repeat separation. Genome Res 27:722–736. https://doi .org/10.1101/gr.215087.116.
- Macrina FL, Tobian JA, Jones KR, Evans RP, Clewell DB. 1982. A cloning vector able to replicate in *Escherichia coli* and *Streptococcus sanguis*. Gene 19:345–353. https://doi.org/10.1016/0378-1119(82)90025-7.
- Galens K, Orvis J, Daugherty S, Creasy HH, Angiuoli S, White O, Wortman J, Mahurkar A, Giglio MG. 2011. The IGS standard operating procedure for automated prokaryotic annotation. Stand Genomic Sci 4:244–251. https://doi.org/10.4056/sigs.1223234.
- Couvin D, Bernheim A, Toffano-Nioche C, Touchon M, Michalik J, Neron B, Rocha EPC, Vergnaud G, Gautheret D, Pourcel C. 2018. CRISPRCas-Finder, an update of CRISRFinder, includes a portable version, enhanced performance and integrates search for Cas proteins. Nucleic Acids Res 46:W246–W251. https://doi.org/10.1093/nar/gky425.
- Weber T, Blin K, Duddela S, Krug D, Kim HU, Bruccoleri R, Lee SY, Fischbach MA, Muller R, Wohlleben W, Breitling R, Takano E, Medema MH. 2015. antiSMASH 3.0: a comprehensive resource for the genome mining of biosynthetic gene clusters. Nucleic Acids Res 43:W237–W243. https://doi.org/10.1093/nar/gkv437.
- van Heel AJ, de Jong A, Song C, Viel JH, Kok J, Kuipers OP. 2018. BAGEL4: a user-friendly Web server to thoroughly mine RiPPs and bacteriocins. Nucleic Acids Res 46:W278–W281. https://doi.org/10.1093/nar/gky383.