



Ferroptosis contributes to the progression of female-specific neoplasms, from breast cancer to gynecological malignancies in a manner regulated by non-coding RNAs: Mechanistic implications

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ABSTRACT

Ferroptosis, a recently identified type of non-apoptotic cell death, triggers the elimination of cells in the presence of lipid peroxidation and in an iron-dependent manner. Indeed, ferroptosis-stimulating factors have the ability of suppressing antioxidant capacity, leading to the accumulation of reactive oxygen species (ROS) and the subsequent oxidative death of the cells. Ferroptosis is involved in the pathophysiological basis of different maladies, such as multiple cancers, among which female-oriented malignancies have attracted much attention in recent years. In this context, it has also been unveiled that non-coding RNA transcripts, including microRNAs, long non-coding RNAs, and circular RNAs have regulatory interconnections with the ferroptotic flux, which controls the pathogenic development of diseases. Furthermore, the potential of employing these RNA transcripts as therapeutic targets during the onset of female-specific neoplasms to modulate ferroptosis has become a research hotspot; however, the molecular mechanisms and functional alterations of ferroptosis still require further investigation. The current review comprehensively highlights ferroptosis and its association with non-coding RNAs with a focus on how this crosstalk affects the pathogenesis of female-oriented malignancies, from breast cancer to ovarian, cervical, and endometrial neoplasms, suggesting novel therapeutic targets to decelerate and even block the expansion and development of these tumors.

1. Introduction

Female-oriented neoplasms, including breast and gynecological malignancies, are considered one of the leading causes of death in the global female population. Genetics, age, lifestyle and eating habits, menopause state, and history of being exposed to carcinogens are the principal determinants of the onset and frequency of these types of malignancies. Treatment of female-specific cancers, similar to other cancers, is considered a huge challenge, and eliminating tumor cells without hurting non-cancerous cells is a key concept of cancer therapy.

Following the discovery of regulated cell death (RCD) researchers found that the cell death process can potentially be controlled [1–3]. For a long time, apoptosis was considered the only type of RCD, and a broad range of anti-tumor medications were designed to stimulate apoptosis in cancer cells; however, it became clear that tumor cells exhibit, and develop, various degrees of resistance to these drugs and, thus, apoptosis [4]. Accordingly, recent studies have attempted to target non-apoptotic cell death pathways for possible reduction of cancer cell resistance against apoptosis-inducing drugs.

In the context of non-apoptotic RCDs, ferroptosis as an iron-dependent type of RCD, was first characterized in 2012 [5].

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| Abbreviations | | | |
|---------------|---|--------|-------------------------------|
| BC | breast cancer | GSH | glutathione |
| ceRNA | competing endogenous RNA | HCC | hepatocellular carcinoma |
| circRNA | circular RNA | lncRNA | long non-coding RNA |
| DDP | cisplatin | MDA | malondialdehyde |
| DEFerlncRNA | differentially expressed lncRNAs | miRNA | microRNA |
| EC | endometrial cancer | ncRNA | non-coding RNA |
| ecEM | ectopic endometrium cells | OC | ovarian cancer |
| ESC | endometrial stromal cell | ORFs | open reading frames |
| euEM | normal endometrium cells | OS | overall survival |
| FerroScore | ferroptosis score | PI3K | phosphoinositide 3-kinase |
| FIRLs | ferroptosis and iron-metabolism related lncRNAs | RCD | regulated cell death |
| GC | gynecological cancer | ROS | reactive oxygen species |
| | | TNBC | triple-negative breast cancer |
| | | UTR | untranslated region |

Ferroptosis is responsible for the inhibition of RAS mutant tumor cells and it was subsequently shown that this process is strongly related to tumor cell death [5,6]. This RCD pathway is modulated by the canonical *TP53/p53* tumor suppressor gene through blocking the cystine/glutamate antiporter/xCT/system xc⁻ [7,8]. Interestingly, many tumor cells, although resistant to common therapeutic interventions, are sensitive to ferroptosis. Thus, induction of this cell death mechanism might help eradicate those cells [9]. Moreover, ferroptosis is also associated with immunotherapeutic interventions, as it can be triggered in tumor cells by T cells and IFNG/IFN γ (interferon gamma) [10].

According to recent evidence, the ferroptotic flux can be regulated by non-coding RNA (ncRNA) transcripts, among which microRNAs (miRNAs), long non-coding RNAs (lncRNAs), and circular RNAs (circRNAs) play the most important roles [11,12]. In association with ferroptosis, the corresponding ncRNA molecules participate in iron metabolism, as well as ferroptosis-related amino acid metabolism to regulate the ferroptotic flux [13]. Furthermore, reactive oxygen species (ROS) metabolism is also regulated by ncRNAs, and it is accepted that intracellular lipid ROS accumulation is a major stimulant for ferroptosis [14]. Moderate elevation of ROS levels inside cells triggers cell proliferation, survival, and malignant transformation; however, ncRNAs have the ability to modulate ROS levels to maintain redox dynamics and the decrease in ROS blocks ferroptosis [11]. In conclusion, ferroptosis, as a critical process in cancer development, can be controlled by ncRNAs, leading to either the progression or suppression of malignant conditions. In the current review, the modulatory mechanisms related to ferroptosis are first summarized, and then the involvement of this non-apoptotic RCD in female-oriented malignancies, from breast cancer to endometrial neoplasm, under the regulation of ncRNAs is discussed. Current application status and opportunities of targeting ferroptosis in female cancer therapies are then considered.

2. Female-specific neoplasms: from breast cancer to common gynecological malignancies

Breast cancer (BC) is the most common type of female malignancy with more than 2 million cases each year [15]. Breast tumors are often formed and expanded due to ductal hyperproliferation and can represent either a benign or malignant phenotype as the result of the existing relationship between continuous induction and varied tumorigenic parameters. Stromal cells and macrophages residing in the tumor microenvironment significantly affect the progression of this neoplasm [16, 17]. In detail, macrophages provoke the angiogenic processes and also increase the immune resistance of tumor cells through producing a mutagenic inflammatory microenvironment [18]. Moreover, DNA methylation, as well as other epigenetic modifications developed in the tumor microenvironment, elevates the risk of BC tumorigenesis. Consistent with this finding, cancer stem cells also have a role in BC

carcinogenesis, immune evasion, recurrence, and therapeutic resistance (Fig. 1) [19].

In the case of other female-oriented neoplasms, gynecological cancers (GCs) are of great importance among women worldwide. Ovarian cancer (OC), as one of the well-studied GC subtypes, is not as common as other female malignancies but unfortunately has the highest rate of cancer mortality. It been estimated that OCs have an incidence of 11.7–12.1 per 100,000 in European countries and the U.S, with a lower rate among the Middle East and Asian nations [20,21]. Regarding the pathology of OC, a non-homogenous cluster of malignancies are formed and expanded in the germ cells, as well as epithelial cells, fallopian tube, and mesenchyme with different patterns of etiology and molecular biology. Nonetheless, the majority of OCs have been detected with an epithelial origin [22]. Most OC patients are diagnosed when the disease has entered its advanced progression levels, terminating with high mortality rates. Therefore, it is crucial to improve preventive measures and allow a timely diagnosis, as early detection of OC provides a higher survival rate up to 93% [19,23].

Cervical cancer is the other GC in this field, which is categorized as the fourth most common type of malignancy and the fourth prime cause of cancer deaths among the female population (BC, colorectal cancer, and lung neoplasm are the first three leading causes). The Lancet Global Health has reported that approximately 570,000 women experienced CC in 2018, among which 311,000 died [24]. It has been elucidated that human papillomavirus/HPV infections, especially those being developed by carcinogenic species, are the major leading cause of CC onset. Therefore, human papillomavirus screening, along with vaccination against these viral species can improve the preventive approaches [25, 26]. CCs are histologically divided into two principal subtypes, including adenocarcinoma (25% of cases) and squamous cell carcinoma (70% of cases) [27,28]. The conventional Pap smear test can help detect early alterations in the cervical epithelium and thus the early stage of invasive CC [29]. Beyond the ovary and uterine cervix, the endometrium can also be affected by malignancy. In this context, endometrial cancer (EC) is considered as the most common GC with more than 61,000 cases detected yearly in the U.S [30,31]. EC primarily affects the glandular epithelium, lining the uterus, which is typically responsible for releasing substances required for menstruation or even embryonic development [32]. The onset of EC is principally affected by obesity, along with hormonal deregulation, reproductive parameters, and being genetically vulnerable. This GC has a heterogenous genetic pattern except for a subgroup of patients with a cancer predisposition syndrome, Lynch syndrome, stimulated by the germline changes of DNA mismatch repair genes such as *MLH1*, *PMS2*, *MSH2*, and *MSH6*. Furthermore, ECs are represented through a high degree of many other germline mutations in malignancy predisposition genes (Fig. 1) [19,33,34].

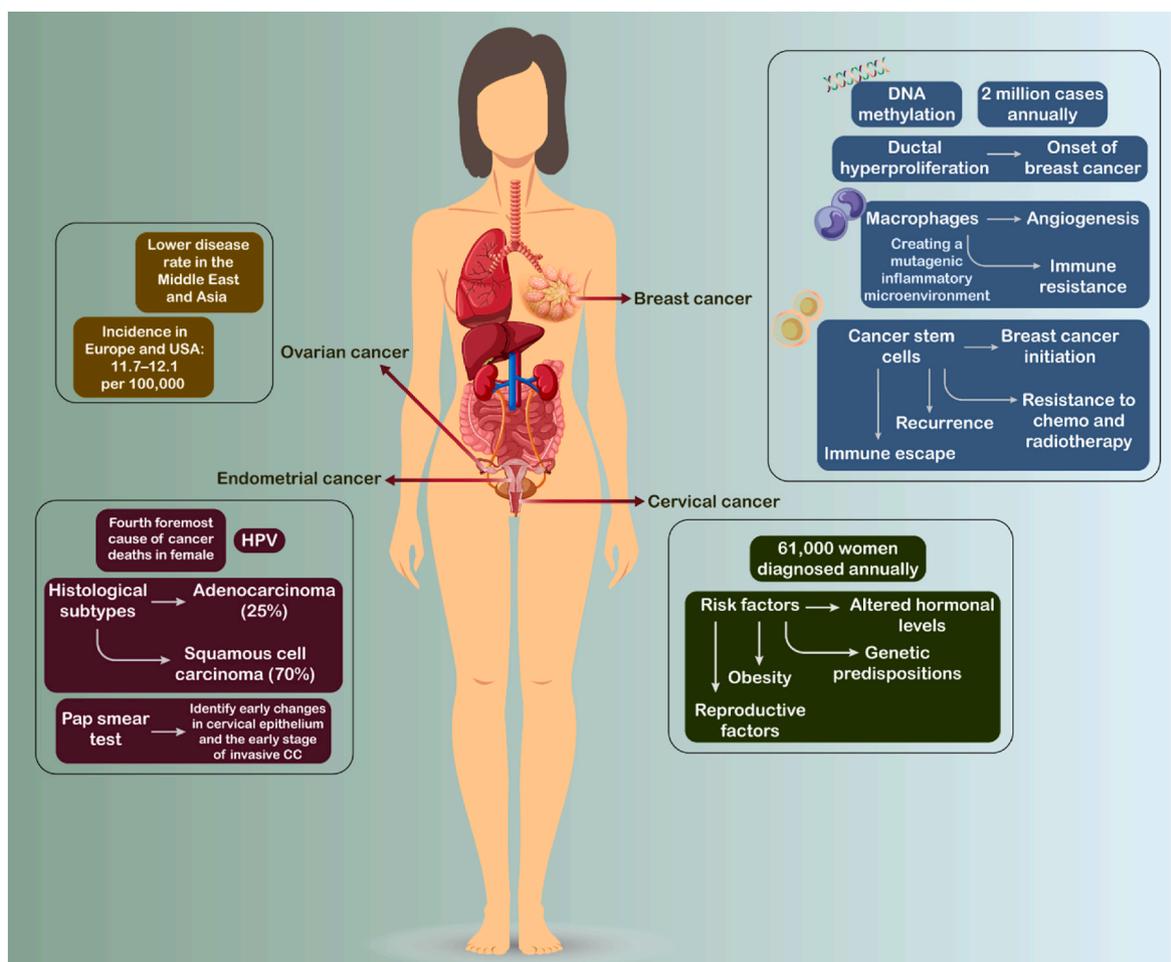


Fig. 1. Female-specific cancers: Breast, ovarian, cervical, and endometrial cancers. (A) Breast cancer: The most common female malignancy (although also occurring in males), BC arises from ductal hyperproliferation and is influenced by the tumor microenvironment. Macrophages and epigenetic modifications in this environment contribute to BC progression. Cancer stem cells also play a role in BC development, immune evasion, recurrence, and resistance to treatment. (B) Ovarian cancer: Though less frequent than other GCs, OC has the highest mortality rate. OC encompasses various types of malignancies with diverse origins. Most cases are of epithelial origin, and early detection is crucial due to high mortality rates in advanced stages; (C) Cervical cancer: The fourth most common female malignancy, CC is primarily caused by human papillomavirus (HPV) infections, particularly carcinogenic strains. Vaccination and screening programs are crucial for prevention. Histologically, CC is divided into adenocarcinoma and squamous cell carcinoma subtypes. (D) Endometrial cancer: The most common GC of the uterus, EC affects the glandular epithelium and is influenced by factors such as obesity, hormonal imbalances, and genetics. Most cases of EC exhibit a heterogeneous genetic pattern, except for those with Lynch syndrome.

3. Ferroptosis: A non-apoptotic type of regulated cell death

Iron (Fe), with a vast array of biological roles in the human body, is critical for cell survival, due to its role in oxygen transport, DNA biosynthesis, and ATP production [35,36]. Moreover, iron is strongly correlated with the onset and expansion of tumors, and thus iron metabolism defects might result in tumor growth [37,38]. Once iron-dependent oxidative phosphorylation has progressed in mitochondria, cells generate ROS and ATP. Increasing ROS levels result in oxidative stress responses, leading to cell injury or death [39]. In this context, ferroptosis primarily results from the aggregation of iron-dependent lipid peroxide (Fig. 2) [40,41].

Although the term ferroptosis was first used in 2012, the inducers of this non-apoptotic RCD were discovered before it was named. During a large-scale analysis conducted in 2003, Stockwell et al., noticed that a newly discovered compound called erastin could provoke death of RAS-mutated tumor cells in a non-apoptotic manner [12]. Later, other chemicals, such as sorafenib, artemisinin, and 1, 2-dioxolane (FINO2), as a cyclic peroxide, were affirmed to stimulate ferroptosis [40,42,43]. Mechanistically, ferroptosis differs from other RCDs and is biochemically identified by intracellular iron and ROS accumulation, lipid

peroxidation, and depletion of glutathione and lipid repair enzymes [44]. Moreover, autophagy, which is a highly conserved eukaryotic cellular recycling process [45–47], has a strong correlation with ferroptosis, as it triggers the removal of proteins related to ferroptosis through ferritinophagy and chaperone-mediated autophagy [48].

In comparison to the other types of regulated cell death processes, it is known that PCD is a critical physiological process in all living organisms, with roles spanning embryonic development, organ function maintenance, aging, and immune response regulation. Phagocytic processes efficiently remove dead cells under normal conditions. Apoptosis, necroptosis, and pyroptosis are the most well-characterized forms of PCD. Apoptosis is a highly conserved cell death pathway across different animal species that has been extensively studied for decades [49]. Initially considered the sole regulated PCD mechanism, apoptosis involves the controlled release of CYCS (cytochrome *c*, somatic) from mitochondria. A delicate balance between pro-apoptotic (BCL2 family) and anti-apoptotic proteins regulates this process. Additionally, initiator caspases (CASP8, CASP9 and CASP10) together with effector caspases (CASP3, CASP6 and CASP7) play crucial roles during apoptosis activation. Apoptosis culminates in a series of distinctive events: nuclear membrane breakdown by CASP6, cleavage of intracellular proteins (e.

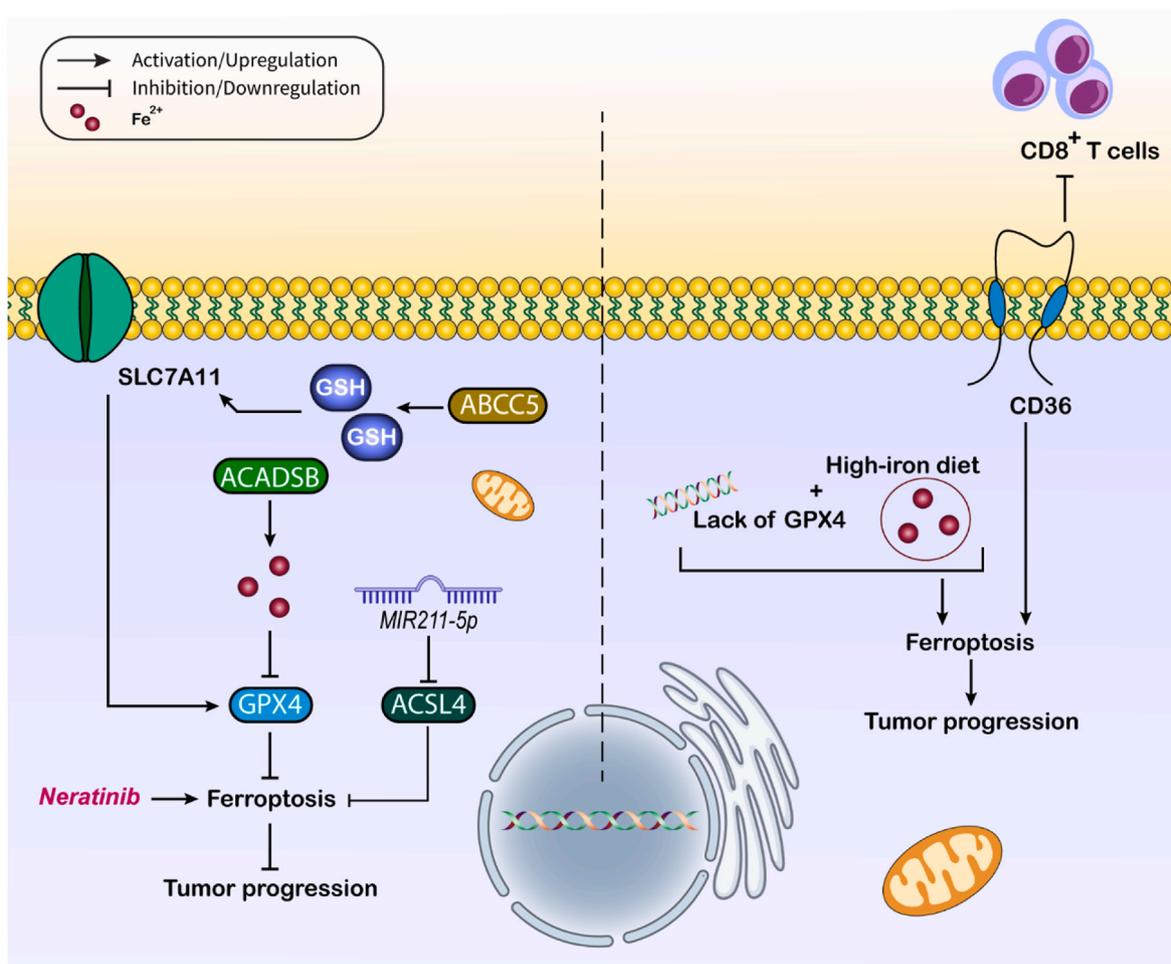


Fig. 2. Ferroptosis: a tumor-suppressor or an oncogenic process? With regard to the tumor suppressive effects of ferroptosis, it disrupts cell proliferation, migration, and invasion in various cancers, including blood malignancies and solid tumors. Studies suggest a negative correlation between ferroptosis and metastasis, with ferroptosis-silenced tumors exhibiting increased metastatic potential. Ferroptosis inducers such as neratinib may block brain metastasis in breast cancer models. Lower expression of ferroptosis inhibitors, including SLC7A11 and ABCC5, and higher expression of ferroptosis modulators such as ACSL4 and GPX4, are associated with better prognosis in some cancers. Conversely, in KRAS-driven pancreatic cancer models, ferroptosis induction via GPX4 knockdown or high-iron diet accelerate tumor progression. CD36 may suppress anti-tumor CD8⁺ T cells by triggering ferroptosis, suggesting a context-dependent role.

g., PARP, lamin, etc.), membrane blebbing, and fragmentation of genomic DNA into nucleosomes [3,50,51]. These hallmarks are employed to identify the specific PCD pathway engaged [3,52].

Previously, apoptosis was contrasted with necrosis, a supposedly uncontrolled process leading to membrane rupture and leakage; however, recent research has identified necroptosis, a non-apoptotic regulated PCD pathway promoting tissue repair and pathogen detection [53, 54]. Triggered by TNF/TNF- α (tumor necrosis factor) or other stimuli such as FAS-FASLG/FasL, TLRs (toll like receptors), and cytosolic nucleic acid sensors, necroptosis relies on TNFRSF1A/TNFR1 (TNF receptor superfamily member 1A) activation [53,55–58]. Whereas these pathways often activate proinflammatory and prosurvival NF κ B/NF- κ B (nuclear factor kappa B) signals, necroptosis is specifically induced when CASP8 is inhibited by microbes or drugs [59].

Furthermore, pyroptosis, as a PCD pathway triggered by inflammasome sensors (NLR family, AIM2 [absent in melanoma 2], MEFV [MEFV innate immunity regulator, pyrin]), culminates in membrane leakage. Inflammasomes detect pathogen-associated molecular patterns/PAMPs and danger/damage-associated molecular patterns/DAMPs, acting as a defense mechanism against pathogens and cellular stress [60]. This lytic cell death prevents microbial spread and alerts the immune system but can lead to pathological inflammation if dysregulated. Initially thought to be CASP3-dependent apoptosis, pyroptosis was distinguished due to

its reliance on CASP1 [60,61]. Parenthetically, the term "pyroptosis" reflects the inflammatory nature ("pyro" meaning fire) and programmed cell death aspect ("ptosis" meaning falling) [62].

Interestingly, cells that experience ferroptosis present different morphological characteristics compared to apoptosis and/or autophagy; for instance, the cell membrane is ruptured during ferroptosis, whereas it is not impaired through apoptotic or autophagic fluxes. In ferroptosis, mitochondrial atrophy, along with the disappearance of the mitochondrial ridge, without any change in nucleus size except for chromatin condensation are considered as the specific characteristics [63]. Recently, ferroptosis has been found to play crucial roles in the pathogenesis of multiple diseases, including a vast array of malignancies. Once the ferroptosis-related regulatory molecular mechanism(s) is deeply explored, the interaction between ferroptosis and cancer development will be better elucidated. As mentioned above, the modulation of ferroptosis is controlled by particular signaling pathways through iron accumulation, lipid peroxidation, and cellular membrane impairments, and thus ferroptosis can be regulated by specific drugs or genetic interventions. Within this context, regulation of homeostasis between oxidative and antioxidant systems is considered the prime mechanism in ferroptosis [64–67]. Thereby, it is expected that cancerous conditions can be ameliorated or even cured by exerting specific interventions into the ferroptotic flux.

4. Ferroptosis plays a dual role in cancer

According to the existing evidence, ferroptosis can serve as a tumor suppressive flux in many blood malignancies, as well as solid tumors to affect multiple steps from the cell cycle and proliferation to tumor expansion [68]. In this context, cytosolic GOT1 (glutamic-oxaloacetic transaminase 1), which is necessary for oxidant/antioxidant balance, blocks cell proliferation and increases cytotoxicity, especially when used in combination with the ferroptosis-inducing agent RSL3 [69,70]. Other than cell proliferation, ferroptosis also blocks the migration of tumor cells, and thus suppresses tumor invasion. As an example, KLF2/Krüppel-like factor 2 (KLF transcription factor 2) can repress tumor cell migration and invasion in clear cell renal carcinoma through GPX4 (glutathione peroxidase 4)-mediated regulation of ferroptosis [71]. ACSL4 (acyl-CoA synthetase long chain family member 4) is another ferroptosis modulator in this regard that can act at downstream of the miRNA *MIR211-5p*, which suppresses cell proliferation, as well as migration and invasion in hepatocellular carcinoma (HCC) [72].

Ferroptosis is also correlated with metastasis in a negative manner, as it has been revealed that metastatic trait is more common in ferroptosis-silenced tumors compared to those with activated ferroptosis [73,74]. Additionally, in HCC, ferroptosis inhibition is closely related to worse tumor expansion, high degrees of metastasis, and poor prognosis [75]. In line with this fact, Liu et al. indicated that resistance against ferroptosis may enhance the probability of metastasis, which can be reversed by ferroptosis inhibitors, such as the aforementioned GPX4 [76]. Consistent with this observation, ferroptosis inducers, such as neratinib as a pan-tyrosine kinase inhibitor, potentially block brain metastasis in a synergistic model of human EGFR⁺ BC metastasis [63, 77].

In clinical settings, ferroptosis has also been proposed to be in negative association with prognosis and overall survival (OS). Within this context, the overexpression of SLC7A11 provides a poor prognosis, specifically due to ferroptosis inhibition [78]. In clinical specimens collected from HCC patients, ABCC5 (ATP binding cassette subfamily C member 5) exhibits an overexpressed pattern with a negative interconnection with ferroptosis through stabilizing SLC7A11 and enhancing reduced glutathione (GSH) levels inside the cells. Sorafenib-resistant HCC cells also exhibit an increase in ABCC5 expression levels, indicating poor prognosis [79,80]. As indicated above, ACSL4 and GPX4 are ferroptosis modulators, and their expression can be utilized as a prognostic factor for disease-free survival. In detail, ACSL4 overexpression results in a better OS, whereas patients with GPX4 overexpression exhibit better metastasis-free survival [81]. Together, these findings propose that ferroptosis can be considered a tumor suppressive process by regulating varied molecular and cellular mechanisms [63].

In contrast, some recent evaluations have suggested that under particular circumstances ferroptosis may act as a tumor activator. This hypothesis can be exemplified by the enhancement of pancreas weight and mortality in KRAS4-driven animals with pancreatic malignancy due to GPX4 knockdown or receiving a high-iron diet. Thus, the absence of GPX4 or the administration of a high-iron diet accelerates the progression of KRAS-induced pancreatic ductal adenocarcinoma [82,83]. CD36 decreases the antitumor activity of CD8⁺ T cells by triggering ferroptosis and lipid peroxidation. Nonetheless, the basic modulatory mechanisms and pathways supporting the carcinogenic role of ferroptosis are still ambiguous [63,84].

5. The interconnection between non-coding RNAs and ferroptosis

As we know, ncRNAs, including miRNAs, lncRNAs, and circRNAs contribute to a vast array of genetic modulatory mechanisms [85,86]. In this era, researchers have tried to elucidate these regulatory pathways that are related to ferroptosis and fortunately have discovered valuable information. Based on their research, they noticed that specific ncRNA

transcripts can control the ferroptosis flux by direct modulation of key factors or indirect regulation of molecular targets located upstream of this process [87–89]. The following sections provide more detailed information on the existing network between ncRNAs and the process of ferroptosis.

5.1. MiRNAs and ferroptosis: regulatory mechanisms

MiRNAs, which are a subclass of the ncRNA superfamily that are 20–25 nucleotides in length [90–92], have recently been shown to control ferroptosis through multiple pathways (Fig. 3) (Table 1) [93]. With a mechanistic view, miRNAs can affect lipid metabolism in tumor cells to regulate ferroptosis. In this regard, MFN2 (mitofusin 2) that is central to the process of mitochondrial fusion, is responsible for the regulation of ROS generation in cellular lipid metabolism [94].

ALOX15 (arachidonate 15-lipoxygenase), which belongs to the LOX family, has the ability of converting arachidonic acid, along with other unsaturated fatty acids residing on the cell membrane, into lipid peroxides, and thus oxidative stress-mediated cell death and inflammation can specifically be regulated by ALOX15 [95]. In gastric neoplasms, exosomal *MIR522* targets ALOX15 further decreasing ROS levels, which in turn blocks ferroptosis and stimulates chemo-resistance in these malignancies [96].

The serine-threonine kinase AURKA (aurora kinase A), as another defined regulator for ferroptosis, aids in controlling mitosis-related spindle formation. Several types of cancer exhibit AURKA overexpression in this context. By siRNA-mediated silencing of AURKA translation, GPX4 expression can be downregulated, implying that GPX4's upstream factors are controlled by AURKA. Upon the conducted bioanalysis, a binding site was found for *MIR4715-3p* in the 3'-untranslated region (3'-UTR) of *AURKA* mRNA. Once *MIR4715-3p* is turned on, AURKA expression declines and a reduction is observed in GPX4 expression in pancreatic cancer cells. Accordingly, the AURKA-GPX4 axis is a prime route by which *MIR4715-3p* may trigger ferroptosis [97,98].

Other than interfering with lipid metabolism, miRNAs can also control ferroptosis through regulating the metabolism of amino acids. For example, the pathway responsible for the breakdown of glutamine is a crucial regulator of intracellular glutamate levels. GLS2 (glutaminase 2) and SLC1A5 (solute carrier family 1 member 5) are significant regulatory factors for the uptake and breakdown of glutamine in this framework. Reduced intracellular glutamate content can decrease ROS accumulation and block ferroptosis, as occurs with lower expression of SLC1A5 or GLS2. Notably, by binding SLC1A5 at the 3'-UTR, *MIR137* overexpression can adversely control ferroptosis in melanoma cells. Niu et al. also suggested that by targeting GLS2 and mitigating intracellular glutamate levels, *MIR103A-3p* can cause ferroptosis resistance in gastric cancer cells. Similarly, it was affirmed that patients suffering from gastric carcinoma have a poor prognosis when *MIR103A-3p* is overexpressed [99–102].

In acute kidney injury, cell damage resulting from ferroptosis is considered a critical mechanism. To enhance the function of the cystine/glutamate antiporter/xCT and raise the intracellular GSH level, ATF4 (activating transcription factor 4) can upregulate the expression of transmembrane transporter SLC7A11 [67]. Moreover, HSPA5 (heat shock protein family A (Hsp70) member 5) is also activated by ATF4 to increase GPX4 expression and activity, thereby shielding cells from ferroptosis. Ferroptosis in HCC cells may be promoted by the overexpression of *MIR214*, which implies that *MIR214* can inhibit *ATF4* transcription and, in turn, the ATF4-HSPA5-GPX4 pathway, leading to the stimulation of ferroptosis [103–105].

Tumor cell ferroptosis can be controlled by miRNAs through interfering with intracellular iron metabolism. Maintaining the balance of Fe²⁺ within the cytoplasm and mediating its entry into mitochondria are the primary functions of TF (transferrin). By controlling TF to lower intracellular Fe²⁺ levels and prevent ferroptosis, *MIR7-5p* causes

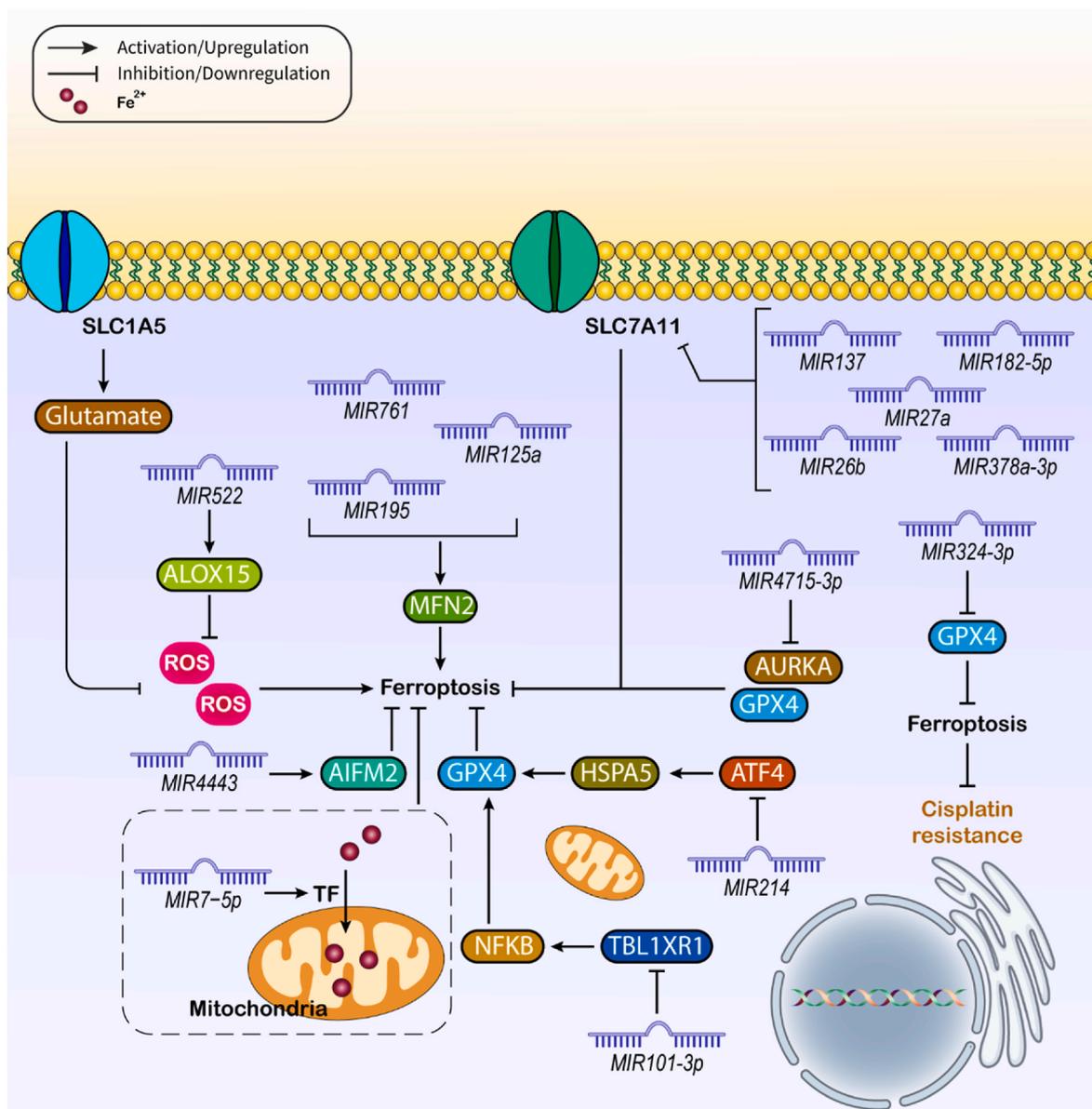


Fig. 3. miRNAs and ferroptosis. miRNAs can influence ferroptosis through various pathways, including lipid and amino acid metabolism, as well as iron homeostasis as illustrated in this figure. *MIR522* in gastric cancer targets *ALOX15*, as an enzyme involved in lipid peroxidation, reducing ROS levels and blocks ferroptosis. *MIR4715-3p* targets *AURKA* in pancreatic cancer, leading to *GPX4* (a ferroptosis inhibitor) downregulation and increased ferroptosis. *MIR137* overexpression in melanoma inhibits *SLC1A5*, reducing glutamate uptake and ferroptosis. *MIR103A-3p* targets *GLS2* in gastric cancer, decreasing glutamate levels and promoting ferroptosis resistance. *MIR214* in HCC inhibits *ATF4*, inducing a decreased *GPX4* expression and promoting ferroptosis. *MIR7-5p* reduces iron uptake by targeting TF, preventing ferroptosis and promoting radiation resistance. *MIR335* targets *FTH1* in Parkinson disease, increasing iron levels and promoting ferroptosis. Several miRNAs (including *MIR124*, *MIR30B-5p*, *MIR20A*, and *MIR485-3p*) have the ability of targeting iron exporters or regulators, influencing ferroptosis susceptibility. Other miRNAs such as *MIR7*, *MIR200A*, *MIR101*, and *MIR455* target negative regulators of *NFE2L2* (a ferroptosis inhibitor), promoting ferroptosis. *MIR28*, *MIR153-3p*, etc. also target *NFE2L2* directly or indirectly, again promoting ferroptosis.

radiation resistance. Besides, in the context of Parkinson disease, *MIR335* targets *FTH1* (ferritin heavy chain 1) to promote ferroptosis by elevating intracellular Fe^{2+} content [106,107]. Mammals have only one cellular iron exporter, *SLC40A1/ferroportin 1*, which is essential for maintaining the cellular iron homeostasis. Underexpression of this iron exporter, achieved by *MIR124*, provokes neuronal death in murine models of intracerebral hemorrhage through increasing ferroptosis [108]. Along these lines, ferroptosis-induced oxidative stress, which has been identified as a major leading cause of pre-eclampsia-related adverse outcomes, is correlated with *MIR30B-5p* that is responsible for downregulation of *SLC40A1/ferroportin 1* to provoke ferroptosis in trophoblasts [109]. In a similar manner, *MIR20A*, along with *MIR485-3p* can also decrease the generation of Fe^{2+} ions through targeting

ferroptosis, which indeed stimulates resistance against ferroptotic flux [36,110].

The *KEAP1* (kelch like ECH associated protein 1)-*NFE2L2* (*NFE2* like bZIP transcription factor 2) regulatory pathway, a key axis that suppresses ferroptosis through blocking ROS generation and decreasing iron load, can be substantially targeted by miRNAs to control ferroptosis. In this regard, overexpressed *MIR7* and *MIR200A* target *KEAP1* to degrade its mRNA and activate *NFE2L2*. In addition, *MIR101* and *MIR455* have the ability to target *CUL3* (cullin 3) to induce *NFE2L2*. In contrast, a vast array of miRNAs, including *MIR28*, *MIR153-3p*, *MIR142-5p*, *MIR27A*, *MIR144*, and *MIR155*, along with others repress *NFE2L2* either directly or indirectly. Thereby, all these miRNAs can be considered as potential molecules in modulating cellular ferroptosis [94,111,112].

Table 1
MiRNAs implicated in ferroptosis regulation during gynecological and breast cancers.

| miRNA | Cancer type | Role in ferroptosis | Target | Reference |
|--------------------|--------------------|---|----------------------------------|-----------|
| <i>MIR1-3p</i> | Ovarian | Enhances the sensitivity of ovarian cancer cells to ferroptosis | <i>FZD7</i> | [156] |
| <i>MIR424-5p</i> | Ovarian | Negatively regulates ferroptosis | <i>ACSL4</i> | [159] |
| <i>MIRNA660-5p</i> | Cervical | Inhibits cancer cell ferroptosis | <i>ALOX15</i> | [161] |
| <i>MIR382-5p</i> | Breast/ ovarian | Promotes ferroptosis | <i>SLC7A11</i> axis | [168] |
| <i>MIR5096</i> | Breast | Promotes ferroptosis | <i>SLC7A11</i> / xCT | [169] |
| <i>MIR499A-5p</i> | Breast | Promotes ferroptosis | <i>PEDS1</i> / <i>TMEM189</i> | [178] |
| <i>MIR324-3p</i> | Breast | Metformin induces ferroptosis by upregulating <i>MIR324-3p</i> | <i>GPX4</i> axis | [173] |

5.2. LncRNAs and ferroptosis

The other subgroup of ncRNAs, i.e., lncRNAs, which are 200 nucleotides or more in length, significantly interfere with gene expression modulation at transcriptional, translational, and post-translational levels by connecting to DNA, mRNA, proteins, or even miRNA transcripts [113,114]. Traditionally, lncRNAs have been defined by the absence of protein-coding capacity. However, recent bioinformatic analyses have identified open reading frames (ORFs) within lncRNA sequences, suggesting potential coding ability [115]. Furthermore, studies have shown a significant correlation between some lncRNAs and ribosomes, hinting at the possibility of lncRNAs harboring coding regions for short peptides [116].

Supporting this notion, analyses of ribosome profiling data revealed that 40% of lncRNAs and pseudogene RNAs in human cells undergo translation. Mass spectrometry data further confirmed the translation of lncRNAs into small peptides [117,118]. For instance, the lncRNA *HOXB-AS3* (*HOXB* cluster antisense RNA 3) encodes a functional 53-amino acid peptide that can suppress colon cancer cell growth [119]. Notably, translated lncRNAs exhibit preferential cytoplasmic localization, whereas untranslated lncRNAs are predominantly nuclear [117]. The translational efficiency of cytoplasmic lncRNAs is similar to that of mRNAs, suggesting active ribosomal engagement. However, the functionality of most lncRNA-derived peptides remains unclear, as they might be unstable byproducts [118]. The evaluation of lncRNA coding potential is inherently challenging due to their structural similarities to mRNAs. Additionally, coding sequences can reside within introns or overlapping exons of other genes, further complicating the analysis. With only a small fraction of lncRNA-encoded products functionally characterized, a vast landscape of potential lncRNA-derived peptides awaits exploration [120]. Ferroptosis is one of the key cellular processes regulated by lncRNAs within cancer cells (Fig. 4) (Table 2). For example, CBS (cystathionine beta-synthase), a potential target for ferroptosis modulation, is regulated by the lncRNA LINC00336 (long intergenic non-protein coding RNA 336) in a positive manner, which in turn triggers cysteine production through trans-sulfuration flux and blocks ferroptosis in lung tumor cells. By binding to *MIR6852*, LINC00336 can also increase the suppressive effects of the CBS axis on ferroptosis [121, 122].

TP53 is a well-known tumor suppressor gene, being responsible for cell cycle arrest induction to repair DNA damage; different *TP53* target genes have interestingly been shown to contribute to ferroptosis modulation. Mao and colleagues noticed that the lncRNA *LINC00472/p53RRA* had the ability of interacting with *G3BP1* (*G3BP* stress granule assembly factor 1) to generate a new complex, i.e., *LINC00472/p53RRA-G3BP1*. In consequence, this newly-formed complex enhances the

intracellular *TP53* gene transcriptome content by disrupting the attachment of *TP53* to *G3BP1* [123,124]. Likewise, *PVT1*, another lncRNA, can also target *TP53* in conjunction with *MIR214*; the targeted *TP53* triggers ferroptosis through repressing the transcription of *SLC7A11*, as well as decreasing the cysteine levels within the cell [125].

LINC00618 is a lncRNA with the potential of increasing ROS and iron accumulation in leukemic patients, thus making cells susceptible to ferroptosis and its inducers. This lncRNA transcript induces ferroptosis in a manner depending on apoptosis [126]. Furthermore, LINC00618 can downregulate *HELLS/LSH* (helicase, lymphoid specific), which otherwise promotes expression of *SLC7A11* and blocks ferroptosis. As a newly recognized lncRNA, *ZFAS1* (*ZNF*X1 antisense RNA 1; located on chromosome 20q13.13) has been associated with varied modulatory roles in an array of diseases [127]. For instance, *ZFAS1* underexpression abolishes pharmacological lipid peroxidation during pneumonic conditions [128]. In detail, *ZFAS1* can serve as a competing endogenous RNA (ceRNA) to potentiate cell susceptibility against ferroptosis through sponging *MIR150-5p* to suppress *SLC38A1* expression; *SLC38A1* is considered a principal mediator of glutamine uptake and lipid peroxidation metabolism [129].

Multiple processes can result in a ferroptosis blockade including exosome-mediated export of iron from the cells. *PROM1* (prominin 1) and *PROM2* are glycoproteins with five transmembrane domains. *PROM1* regulates autophagy through inhibition of *MTOR* (mechanistic target of rapamycin kinase) complex 1 (*MTORC1*), whereas *PROM2* promotes the formation of ferritin-containing exosomes. Recently, it was found that *PROM2* expression in bladder cancer can be induced by the lncRNA *LINC01833/RP11-89* through sponging *MIR129-5p*, resulting in ferroptosis inhibition [130].

5.3. CircRNAs and ferroptosis

CircRNAs, which are well-known for their specific closed loop structure, are synthesized from pre-mRNAs through back splicing or lariat-driven processes [131,132]. This closed ring structure makes circRNAs resistant to exonuclease-mediated degradation, and thus these ncRNA transcripts are expressed in a stable manner [133–135]. Due to their structural stability, multiple binding sites for miRNAs, and the ability to regulate cellular processes, circRNAs have attracted much attention in biological studies and cancer research [136–138].

Within this context, circRNAs are proposed to be involved in the modulation ferroptosis by sponging different miRNAs inside tumor cells (Fig. 4) (Table 3). *CircTTBK2* (tau tubulin kinase 2) is one of these tumor-associated circRNAs that is markedly expressed in gliomas and is responsible for regulating tumor cell proliferation, migration, and invasion [139]. This circRNA also controls cell metabolism by sponging *MIR1283*, *MIR520B*, *MIR217*, and *MIR761*, among which *MIR761* can potentially modulate the ferroptotic flux in HCC via targeting *MFN2* [140–142]. *ITGB8* (integrin subunit beta 8) is a specific target for *circTTBK2* that triggers the suppression of ferroptosis in glioma cells by sponging *MIR761* [143].

According to recently published evidence, ferroptosis can also be considered an autophagy-dependent cell death mode. For example, *ALKBH5* (alkB homolog 5, RNA demethylase) is a major N6-methyladenosine (m6A) demethylase and is a principal autophagy blocker [144,145]. Once *ALKBH5*'s function in autophagy regulation is inhibited by the circRNA *has_circ_0008367*, ferroptosis is triggered in an autophagy-dependent manner [146]. *Circ_0008035* is another ferroptosis-related circRNA that is overexpressed in gastric cancer. Mechanistically, *circ_0008035* underexpression results in the suppression of gastric cancer cell proliferation, which in turn causes an increase in apoptosis, as well as ferroptosis. Li et al. suggest that the inhibition of ferroptosis in gastric cancer is due to direct targeting of *MIR599* by overexpressed *circ_0008035* through the *MIR599-EIF4A1* (eukaryotic translation initiation factor 4A1) regulatory axis [147].

The mutual interaction between different circRNAs and miRNAs in

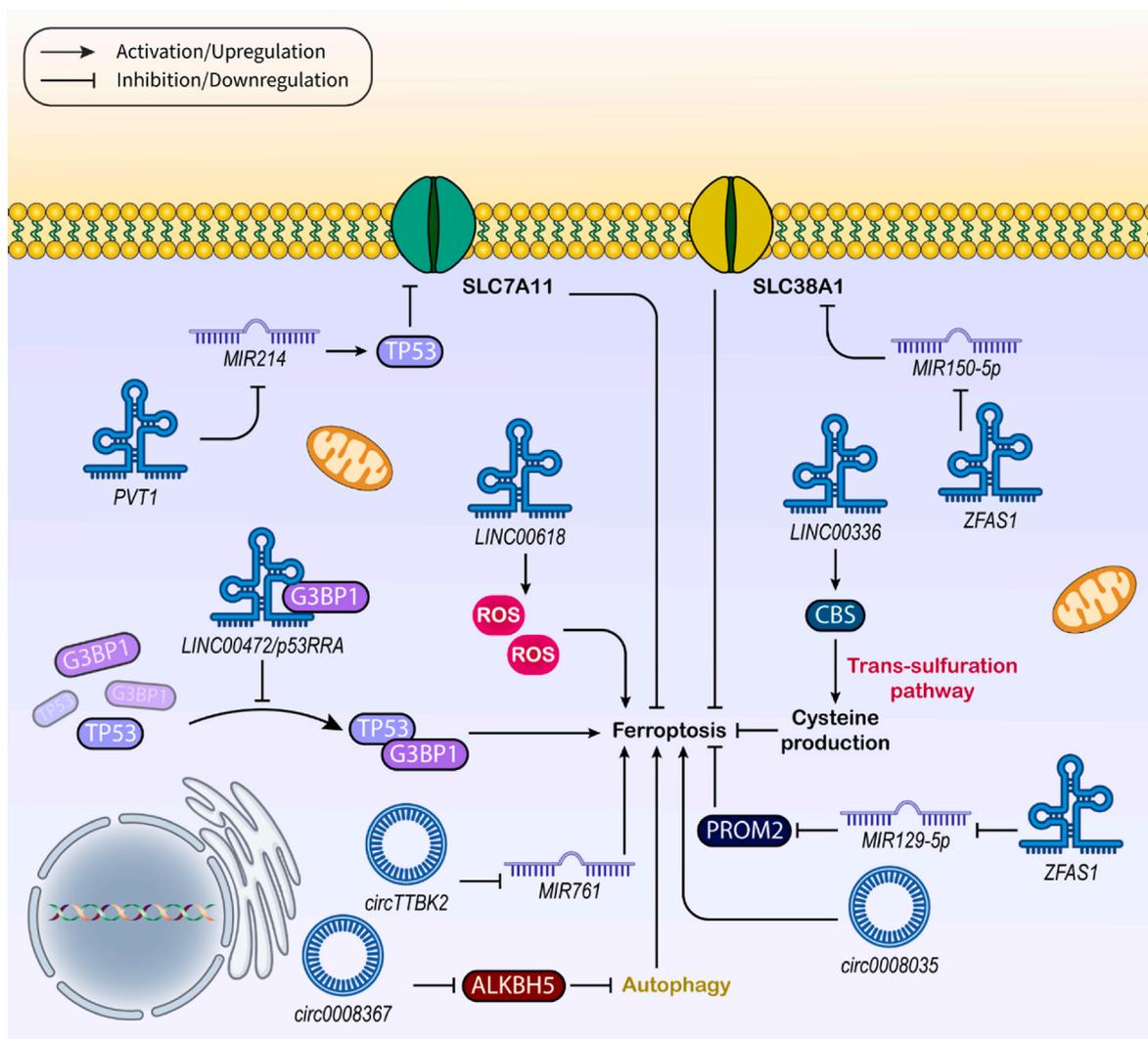


Fig. 4. LncRNAs and circRNAs in association with ferroptosis. LncRNAs and circRNAs have emerged as important regulators of ferroptosis pathways in cancer cells; understanding their specific roles may open doors for novel therapeutic strategies. In the case of lncRNAs, LINC00336 promotes ferroptosis resistance in lung cancer by increasing cysteine production through the CBS axis and suppressing ferroptosis-promoting miRNAs, such as *MIR6852*. *LINC00472/p53RRA* enhances TP53 gene expression, promoting ferroptosis. *PVT1* targets *TP53* in conjunction with *MIR214*, leading to ferroptosis by repressing *SLC7A11* and lowering cysteine levels. *LINC00618* sensitizes leukemic cells to ferroptosis by increasing ROS and iron accumulation and suppressing *SLC7A11* expression. *ZFAS1* acts as a competing endogenous RNA (ceRNA), sponging *MIR150-5p* to increase ferroptosis susceptibility by suppressing *SLC38A1*, which is a key player in glutamine uptake and lipid metabolism. *LINC01833/RP11-89* promotes ferroptosis inhibition in bladder cancer by sponging *MIR129-5p* and inducing *PROM2* expression, which facilitates iron export through ferritin-containing exosomes. Regarding the role of circRNAs, *circTTBK2*, as a glioma-associated circRNA, regulates ferroptosis by sponging multiple miRNAs, including *MIR761*, which can target *MFN2*. *CircTTBK2* sponges *MIR761*, leading to *ITGB8* upregulation and ferroptosis suppression in glioma cells. *Circ_0008367* triggers ferroptosis in an autophagy-dependent manner by inhibiting the autophagy-promoting function of *ALKBH5*. *Circ_0008035* inhibits ferroptosis in gastric cancer by directly targeting *MIR599* and affecting the *MIR599-EIF4A1* regulatory axis. *CircIL4R* and *circEPST11* regulate ferroptosis through the *MIR541-3p-GPX4* and *MIR375-MIR409-3p-MIR515-5p-SLC7A11* axes, respectively. Eventually, *circSNX12* interacts with *MIR224-5p* to target *FTH1*, encoding a regulator of iron homeostasis and ferroptosis susceptibility.

ferroptosis modulation has been demonstrated by several lines of investigation. One recent study shows that *circIL4R* and *circEPST11* can mediate ferroptosis regulation via the *MIR541-3p-GPX4* and *MIR375-MIR409-3p-MIR515-5p-SLC7A11* axes, respectively [148,149]. Another example is seen with *circSNX12* that can interact with *MIR224-5p* to target *FTH1* (ferritin heavy chain 1); *FTH1* is a subunit of the ferritin complex, which negatively correlates with ferroptosis by trapping and then oxidizing Fe^{2+} ions inside cells, as well as decreasing ROS formation. It has been reported that *FTH1* is significantly underexpressed in heart failure conditions; however, there is no evidence for the expression of *circSNX12* in heart tissues, and its function needs to be better elucidated [150,151].

6. The existing crosstalk between ferroptosis and non-coding RNAs regulates the onset and progression of female-oriented cancers: A focus on miRNAs, lncRNAs, and circRNAs

6.1. The regulatory network between microRNAs and ferroptosis in gynecological neoplasms, as well as breast cancer

FZD7 (frizzled class receptor 7) is activated in a wide range of malignancies and was recently found to be overexpressed in OC, which in turn leads to a decrease in platinum-resistant OC cells' sensitivity to the ferroptotic flux, and subsequent tumor cell survival (Fig. 5) [152]. FZD7 has the ability to regulate both canonical and non-canonical WNT signaling mechanisms [153]. When FZD7 is overexpressed the WNT-CTNNB1/ β -catenin pathway is overactivated and a group of

Table 2

LncRNAs in association with ferroptosis regulation, having roles in gynecological and breast neoplasms.

| LncRNA | Cancer type | Role in ferroptosis | Target | Reference |
|--------------------|-------------------------------|-----------------------------|--|-----------|
| <i>ADAMTS9-AS1</i> | Epithelial ovarian cancer | Attenuates ferroptosis | <i>MIR587-SLC7A11</i> axis | [184] |
| <i>CACNA1G-AS1</i> | Ovarian | Inhibits ferroptosis | <i>FTH1</i> | [183] |
| <i>ADAMTS9-AS1</i> | Endometriosis | Represses ferroptosis | <i>MIR6516-5p-GPX4</i> axis | [211] |
| <i>H19</i> | Breast | Inhibits ferroptosis | Downregulation of <i>H19</i> can inhibit autophagy to induce an increase in ferroptosis | |
| <i>RUNX1-IT1</i> | Breast | Inhibits ferroptosis | <i>IGF2BP1-GPX4</i> axis; <i>RUNX1-IT1</i> promotes breast cancer carcinogenesis through blocking ferroptosis via elevating GPX4 | [213] |
| <i>HCP5</i> | Triple-negative breast cancer | Inhibits ferroptosis | Regulating GPX4 expression and lipid ROS level | [214] |
| <i>LINC00460</i> | Breast | Inhibits ferroptosis | <i>MIR320A-MAL2</i> axis | [217] |
| <i>LncFASA</i> | Triple-negative breast cancer | Promotes cancer ferroptosis | Binds to PRDX1 and inhibits its peroxidase activity | [232] |

cancers, such as gastric neoplasm, will be triggered. The upregulated *FZD7* can also provoke the onset and progression of OC depending on the regulation of WNT signaling [154,155].

The above-stated findings are also supported by in vitro analyses, as *FZD7* overexpression enhances cell viability and survival of HO8910 or SKOV3 human OC cell lines that are treated with erastin or RSL3 [156]. Mechanistically, *MIR1-3p*, by binding to the *FZD7* 3'-UTR, directly targets *FZD7* to block its expression. Conversely, the mimics of this miRNA transcript reduce cell viability of the corresponding OC cell lines and elevate the malondialdehyde (MDA; a product of polyunsaturated fatty acid peroxidation) levels inside the erastin- or RSL3-treated cells; *MIR1-3p* blockers exert the opposite effects, as they enhance cell viability and mitigate MDA levels. Thus, erastin or RSL3 induce ferroptosis, and *MIR1-3p* mimics these effects on ferroptosis, and being activated by these treatments, can be suppressed by *FZD7* upregulation. Indeed, *MIR1-3p* increases the sensitivity of OC cells to ferroptosis through targeting *FZD7* [156].

MIR424-5p is another miRNA in association with OC, which is a key player in suppressing ferroptosis [157,158]. *MIR424-5p* achieves this effect by silencing *ACSL4*, a molecule that promotes ferroptosis. When *MIR424-5p* levels decrease, *ACSL4* expression increases, making ovarian

Table 3

CircRNAs that are involved in ferroptosis regulation through the progression of gynecological and breast neoplasms.

| CircRNAs | Cancer type | Role in ferroptosis | Target | Reference |
|-------------------------|--------------------|--|---|-----------|
| <i>circSNX12</i> | Ovarian cancer | Inhibiting ferroptosis | <i>MIR194-5p-SLC7A11</i> axis | [234] |
| <i>hsa_circ_0007615</i> | Ovarian cancer | Knockdown of <i>hsa_circ_0007615</i> in EOC cells leads to the blocking of cell proliferation, migration and invasion, but an increase of cell death presenting as ferroptosis | Sponging <i>MIR874-3p</i> and moderating TUBB3. | [237] |
| <i>circACAP2</i> | Cervical cancer | Suppresses ferroptosis | <i>MIR193A-5p-GPX4</i> | [239] |
| <i>circLMO1</i> | Cervical cancer | Promotes cervical cancer cell ferroptosis | Sponging <i>MIR4192-ACSL4</i> | [243] |
| <i>circEPST11</i> | Cervical cancer | Silencing of <i>circEPST11</i> induces ferroptosis | <i>MIR375-MIR409-3p-MIR515-5p-SLC7A11</i> axis | [242] |
| <i>CircRAPGEF5</i> | Endometrial cancer | <i>circRAPGEF5</i> promotes the formation of TFRC with exon 4 skipping and confers ferroptosis resistance in EC cells | Interaction with RBFOX2 | [244] |
| <i>circ_0000643</i> | Breast cancer | Reduced cell ferroptosis | <i>MIR153-SLC7A11</i> axis | [250] |
| <i>RHOT1</i> | Breast cancer | Inhibits ferroptosis | <i>MIR106A-5p-STAT3</i> axis | [253] |
| <i>CircGFRA1</i> | Breast cancer | Downregulation of <i>circGFRA1</i> promotes ferroptosis | Sponging of <i>MIR1228</i> and enhancing AIFM2 expression | |

cancer cells more susceptible to ferroptosis. This finding suggests that targeting *ACSL4* could be an effective strategy for treating ovarian cancer. *ACSL4* is also involved in other cancers and may be a promising target for broader cancer therapies [159].

Tumor cells and macrophages interact through exosomes. In the case of CC, tumor-associated macrophages may transport miRNAs to cancer cells, reducing their sensitivity to ferroptosis inducers [160]. A macrophage-secreted miRNA, *MIR660-5p*, suppresses ferroptosis by downregulating the lipoxygenase ALOX15, an enzyme involved in lipid metabolism [161]. *MIR660* expression varies across cancer types, with low levels observed in CC and high levels in hepatocellular carcinoma and breast cancer [162,163]. Blocking the transcription factor STAT6 (signal transducer and activator of transcription 6) suppresses *MIR660-5p* levels in tumor-associated macrophages, suggesting its involvement in regulating *MIR660* expression. Further research is needed to fully understand the role of exosomes, miRNAs, and STAT6 in CC [161].

Studies suggest that targeting ferroptosis may be a promising approach for treating BC, as well. Triple-negative breast cancer (TNBC) patients, who are less responsive to traditional therapies, may be particularly sensitive to ferroptosis-inducing agents [164,165]. Lidocaine, a commonly used local anesthetic, affects the development of various cancers, including ovarian and breast cancer. Lidocaine can inhibit the cellular resistance against cytotoxicity, increase apoptosis, and decrease cell proliferation in different types of tumor cells [166,167]. In one study, researchers found that lidocaine induces ferroptosis in both ovarian and breast cancer cells. They also demonstrated that lidocaine augments the expression of *MIR382-5p*, which downregulates *SLC7A11* expression. The inhibition of *MIR382-5p* blocks lidocaine-mediated ferroptosis. These findings suggest that lidocaine may inhibit the malignant progression of ovarian and breast neoplasms by stimulating ferroptosis and that *MIR382-5p* plays a role in this process. However, further research is needed to validate the clinical value of lidocaine in the treatment of these cancers [168]. Additionally in relation to *SLC7A11*, researchers explored how *MIR5096*, a miRNA that is expressed at a low level in human BC cells, affects BC development and spread [169]. They discovered that *MIR5096* lowers the levels of *SLC7A11*; this lowering of *SLC7A11* causes more cell death and less cell growth in BC cells. These researchers also observed that *MIR5096* causes ferroptosis to be more severe in TNBC cells than in other types of BC. Moreover, the study showed that *MIR5096* prevents tumor metastasis from occurring in zebrafish models with transplanted tumors [169]. These results indicate that *MIR5096* might be a useful target for BC therapy, especially TNBC, by causing ferroptosis and stopping tumor development and expansion. More studies are required to fully comprehend how *MIR5096* works and its potential in the clinic [169].

Metformin, a commonly used diabetes medication, also possesses anti-tumor properties; this drug inhibits the proliferation and metastasis of various cancers, including BC [170]. Studies suggest that metformin induces ferroptosis under cancer-forming circumstances. This process

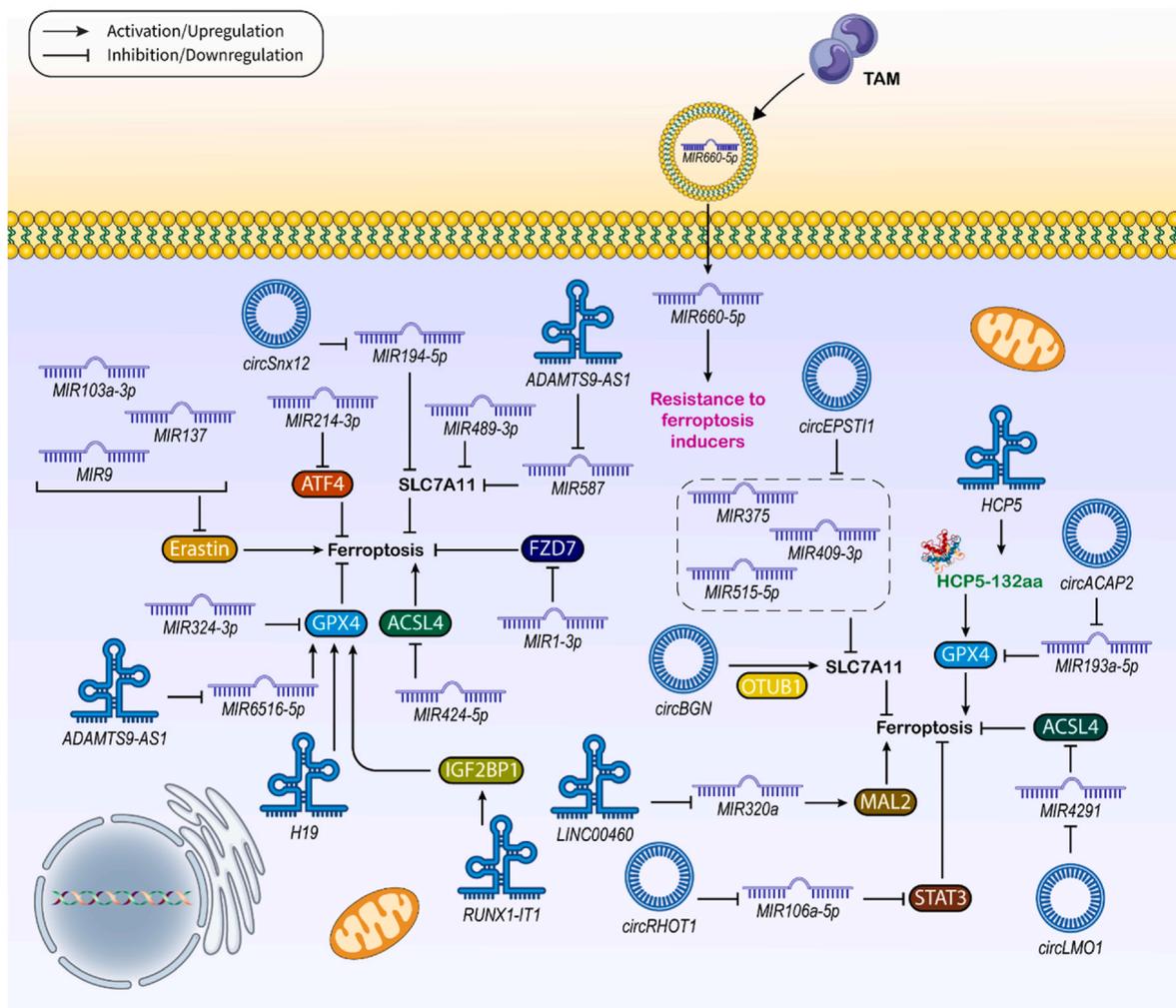


Fig. 5. The interplay between ferroptosis and miRNAs, lncRNAs, and circRNAs relative to the pathogenesis and progression of female-specific neoplasms. TAM, tumor-associated macrophage. See Tables 1–3 for further information.

involves the accumulation of Fe^{2+} and ROS, leading to the downregulation of the antioxidant enzyme GPX4 [171,172]. Metformin also increases the expression of *MIR324-3p*, a miRNA that negatively regulates *GPX4* expression. Based on a luciferase reporter assay, expression of *GPX4* is blocked by a *MIR324-3p* mimic (promoting Fe^{2+} aggregation), whereas a *MIR324-3p* inhibitor enhances the expression of *GPX4* (suppressing Fe^{2+} accumulation) [173]. Both the *MIR324-3p* mimic and the *MIR324-3p* inhibitor have minimal effects on the expression of a *GPX4* mutant with an altered 3'-UTR. These findings highlight the potential of metformin and *MIR324-3p* as novel therapeutic agents for cancer treatment [173].

On another front of investigation, *MIR499A-5p* has been demonstrated to play a dual role in cancer. *MIR499A-5p* is significantly underexpressed in several cancer tissues, including EC, CC, and NSCLC [174–176]. In this regard, once *MIR499A-5p* is downmodulated, tumor growth and metastasis are provoked. However, *MIR499A-5p* also has tumor-suppressing effects by targeting *EIF4E* and *VAV3* (vav guanine nucleotide exchange factor 3) [174,176]. In BC, *MIR499A-5p* is associated with a significant decrease in cancer risk. *PEDS1/TMEM189* (plasmalogen ethanolamine desaturase 1) is involved in the synthesis of ether lipids, which are important for the regulation of ferroptosis [177, 178]. *PEDS1/TMEM189* underexpression sensitizes cells to ferroptosis, and its overexpression contrarily blocks ferroptosis [179,180]. *PEDS1/TMEM189* is highly expressed in various human cancers, and its expression is correlated with the expression of *GPX4* [179]. In BC cells, *MIR499A-5p* directly targets *PEDS1/TMEM189* to decrease its

expression, promoting ferroptosis. This finding suggests that *MIR499A-5p* could be a potential therapeutic target for BC by inducing ferroptosis and inhibiting tumor growth [178].

6.2. Long non-coding RNAs and other ncRNAs associated with ferroptosis in gynecological cancers and breast neoplasm

As mentioned above, the growth and spread of OC is influenced by several molecular factors. One of them is *IGF2BP1* (insulin like growth factor 2 mRNA binding protein 1), a protein that attaches to mRNA and controls its stability and translation, affecting how tumor cells grow and invade [181,182]. Another factor is *CACNA1G-AS1*, a long RNA molecule that does not code for proteins but interacts with *IGF2BP1* and increases the expression of *FTH1*, an enzyme that regulates iron levels. m6A methylation of *FTH1*, which correlates with a poor cancer prognosis, is linked with *CACNA1G-AS1*. High *FTH1* expression makes cells more resistant to ferroptosis, which implies that interfering with the methylation of *FTH1*, especially the *CACNA1G-AS1-IGF2BP1* pathway, could be a potential treatment for OC patients [183]. In addition, researchers also explored if the lncRNA *ADAMTS9-AS1*, can control the expression of *SLC7A11* and prevent ferroptosis in OC cells by competing with *MIR587* [184]. Their in vitro experiments showed that *ADAMTS9-AS1* indeed prevents ferroptosis in OC cells by affecting the *MIR587-SLC7A11* pathway. It was also discovered that *ADAMTS9-AS1* is overexpressed in OC cells, especially in the OVCAR3 and CAOV-3 cell lines. Moreover, reducing *ADAMTS9-AS1* in OC cells stops their growth

and movement by increasing ferroptosis. This result agrees with findings in other cancers, implying that *ADAMTS9-AS1* may have a wider role in controlling cancer cell behavior. The study's findings are backed up by the idea of ceRNAs [185,186]. Fang et al. showed that *ADAMTS9-AS1* can reduce the invasive behavior of BC cells by competing with *MIR513A-5p* and controlling ZFP36 (ZFP36 ring finger protein) expression, which correlates with an improved OS [187]. This finding supports the idea that *ADAMTS9-AS1* can change cellular processes by interfering with small RNA regulation [184].

In OCs, ferroptosis is also enhanced by the interaction of lncRNA *LINC00472/P53RRA* and G3BP1, which prevents the tumor suppressor protein TP53 from exiting the nucleus. In lung cancer, the lncRNA *LINC00336* prevents the formation of ferritin [188]. We know that the HIF1A (hypoxia inducible factor 1 subunit alpha) signaling pathway stimulates tumor growth and spread by controlling various signaling molecules. The response of OC cells that are resistant to cisplatin can be improved by reducing HIF1A [189,190]. The lncRNAs *TLR8-AS1* and *LBX2-AS1* are linked to OC spread and resistance to chemotherapy drugs, and they could be used as targets for therapy or diagnosis [191–193]. A relevant study found 11 lncRNAs related to ferroptosis that are increased in OC tissues, indicating their role in OC growth and expansion. These lncRNAs could be potential targets for OV therapy [194].

In order to help patients with OC make treatment decisions and predict their prognosis, a research team discovered a signature consisting of eight ferroptosis and iron-metabolism related lncRNAs (FIRLs) [195]. Included in the signature are the following FIRLs: *AC083880.1*, *LINC01558*, *AL023583.1*, *AP005205.2*, *AC007114.1*, *LINC00665*, and *AC138904.1*. To visualize the FIRLs and make their interpretation easier, the researchers created two nomograms in addition to the signature [195]. The potential of a multi-label fusion collaborative matrix factorization/MLFCMF approach with an area under the curve/AUC value of 0.8612 has been shown in previous research for predicting lncRNA-disease associations [196]. Furthermore, studies on OC cell lines have looked into the function of a number of FIRLs, including *TONSL-AS1* [197], *SNHG20* (small nucleolar RNA host gene 20) [198], and *MSC-AS1* [199]. Still, not much research has been done on the function of FIRLs in the creation of OC risk signatures. This study lays the groundwork for upcoming in vivo and in vitro assays and offers a significant step toward comprehending the prognostic significance of FIRLs in OC [195].

Beyond the OC, CCs are often diagnosed at an advanced stage, making it difficult to treat effectively [200]. Ferroptosis-related lncRNAs have been studied in other cancers, but their role in CC prognosis is poorly understood. Researchers constructed a co-expression network of ferroptosis-related genes and lncRNAs, identifying 1393 lncRNAs with ferroptosis-related functions. Using these lncRNAs, they developed a prognostic model for CC [201]. The model showed that *AC099568.2* was the most consistently associated lncRNA with CC prognosis across different stages of the disease. This suggests that *AC099568.2* may be a valuable prognostic biomarker and may play a role in CC development. The CC prognostic model developed in this study also shows superior predictive accuracy compared to other prognostic models [202,203], highlighting its potential as a valuable tool for improving CC patient outcomes [201].

Classifying ferroptosis-related gene expression in CC patients was essential due to the importance of ferroptosis in CC immune modulation and the heterogeneity of ferroptosis phenotype in individual CC cells. The low-ferroptosis score (FerroScore) group shows significantly higher expression of immune checkpoints CTLA4 (cytotoxic T-lymphocyte associated protein 4) and PDCD1 (programmed cell death 1) compared to the high-FerroScore group [204]. This observation suggests that low-FerroScore patients may benefit more from checkpoint blockade therapy, which is consistent with the predictions of the submap algorithm. It has been proposed that combining immune checkpoint inhibitors with ferroptosis inducers could be a promising therapeutic

strategy, leading to the development of novel combination therapies and immunotherapeutic agents [205].

Three other ferroptosis-associated lncRNAs were also affirmed to be expressed in CC samples [206–208]. Of these lncRNAs, *AC026790.1* underwent additional validation in multiple ferroptosis-related experiments. Ferroptosis-related markers such as MDA, Fe^{2+} , and ROS levels are found to increase in cells overexpressing *AC026790.1* in comparison to the control group, indicating that *AC026790.1* promotes erastin-induced ferroptosis [205]. The results show that *AC026790.1* may be an important molecule in controlling ferroptosis in CC and may represent a viable target for CC treatment. For CC patients, combining lncRNA analysis with FerroScore may enhance patient prognosis and enable individualized treatment [205].

In the case of EC, the levels of *ADAMTS9-AS1*, as a subtype of lncRNA family, are much higher in EC than in normal endometrium cells [209]. *ADAMTS9-AS1* not only prevents ferroptosis and enhances endometrial stromal cell (ESC) movement and growth, but also acts as a ceRNA that absorbs *MIR6516-5p* [210,211]. By doing so, *ADAMTS9-AS1* boosts the production of GPX4, an essential enzyme that protects cells from ferroptosis. These findings highlight the significance of the *ADAMTS9-AS1-MIR6516-5p-GPX4*-ferroptosis pathway in regulating the survival and migration of embryonic stem cells [211]. This pathway could also be a potential therapeutic target for ectopic endometrium treatment. The pathophysiology of ectopic pregnancy has been demonstrated to involve the lncRNA *ADAMTS9-AS1*. In contrast to normal endometrium cells (euEM), ectopic endometrium cells (ecEM) exhibit significantly higher expression of *ADAMTS9-AS1* [209]. Functional studies were carried out in both human and mouse models to examine the biological role of *ADAMTS9-AS1* in ectopic pregnancy. The researchers discovered that whereas overexpressing *ADAMTS9-AS1* enhances cell viability and migration, inhibiting its expression in endometrial stem cells decreases these properties. *ADAMTS9-AS1* may activate ecEMs, according to these results [211]. Also, by sponging *MIR6516-5p*, *ADAMTS9-AS1* functions as a ceRNA. Furthermore, GPX4 is negatively regulated by the miRNA *MIR6516-5p*. *ADAMTS9-AS1* protects ecEMs from ferroptosis by sponging *MIR6516-5p*, which delays the degradation of GPX4, resulting in elevated GPX4 expression. According to these results, the miRNA-GPX4 axis is the mechanism by which *ADAMTS9-AS1* controls ferroptosis resistance [211]. Through the regulation of *MIR6516-5p-GPX4*-dependent ferroptosis, *ADAMTS9-AS1* also expedites the proliferation and migration of ESCs. The development of novel treatment approaches for ectopic pregnancy may result from these findings, which offer fresh perspectives on the function of *ADAMTS9-AS1* [211].

Researchers found that in ecEM cells as opposed to euEM cells, nine differentially expressed lncRNAs (DEFerlncRNAs) are present. They then employed Cox regression analysis to create a prognostic model for ectopic pregnancy based on these nine DEFerlncRNAs. Based on the analysis, it was found that the DEFerlncRNA prognostic model is a more effective, sensitive, and specific prognostic index in EC [212]. Furthermore, the researchers employed Gene Set Enrichment Analysis/GSEA to detect differentially expressed genes between the two groups (ecEM and euEM). They discovered that these differentially expressed genes are functionally enriched in a set of pathways, such as the Hedgehog signaling pathway, apoptotic flux, extracellular matrix receptor interaction, and natural killer cell-mediated cytotoxicity, along with others. For the purpose of predicting the prognosis of EC and immune-infiltrating conditions, the researchers also discovered a 9-lncRNA model, which includes *CFAP58-DT* (*CFAP58* divergent transcript) [212]. The lncRNA in the model with the highest coefficient is *CFAP58-DT*, and poor EC outcomes are associated with high *CFAP58-DT* expression. Researchers knocked down *CFAP58-DT* expression in HEC-1A and Ishikawa cells to further explore the function of *CFAP58-DT* in EC. They discovered that this significantly decreases cell viability, invasion, and migration abilities. These results imply that *CFAP58-DT* might contribute to EC carcinogenesis. Nonetheless, to clarify the

mechanisms behind this lncRNA's effects on EC cells and to confirm its role in EC, more clinical research is necessary [212].

For another female-oriented malignancy, i.e. BC, the GEO, TCGA, and related cohorts reveal that *RUNX1-IT1* (*RUNX1* intronic transcript 1) is overexpressed in BC tissues [213]. It was also discovered that *RUNX1-IT1* inhibition blocks tumor growth in vivo, as well as BC cell invasion and proliferation in vitro. Further research shows that *RUNX1-IT1* directly binds to IGF2BP1 and promotes the formation of ribonucleoprotein condensates. Because of this binding, IGF2BP1 is able to bind to *GPX4* mRNA with greater tenacity, preventing *GPX4* mRNA degradation. Consequently, the elevated *GPX4* protein promotes BC tumorigenesis by obstructing ferroptosis and lipid peroxidation. These results point to *RUNX1-IT1* as a novel oncogenic lncRNA [213]. In addition, the research suggests that the development and progression of breast cancer are caused by dysregulation of the *RUNX1-IT1*-IGF2BP1-*GPX4* axis. Furthermore, it was demonstrated that by controlling the IGF2BP1-*GPX4* axis, *RUNX1-IT1* inhibits ferroptosis and thus promotes BC tumorigenesis. This discovery offers a possible therapeutic target and prognostic marker for BC [213].

According to the findings of a relevant study, the 132-amino acid protein HCP5-132aa, which is encoded by the lncRNA *HCP5*, regulates the ferroptosis pathway and contributes to the advancement of TNBC [214,215]. Scientists discovered that *GPX4* is expressed at a lower level when the HCP5-132aa ORF is knocked down. Elevated levels of ROS, a characteristic of ferroptosis, are caused by this decrease in *GPX4* activity [214]. They also discovered that when the HCP5-132aa ORF is knocked down, the effects are akin to those of the drug erastin, which causes ferroptosis, by increasing mitochondrial membrane density and decreasing mitochondrial crest. Moreover, they demonstrated that the ferroptosis activators RSL3 and erastin stimulate cells, which results in the highest ROS levels that are directly elevated by knocking down the HCP5-132aa ORF. Using ferroptosis inhibitors and overexpressing HCP5-132aa can counteract this effect. The results of their research point to HCP5-132aa as a potential therapeutic target for TNBC, as well as a novel prognostic factor for the disease [214].

In BC tissues, the lncRNA *LINC00460* has a higher expression level than in normal breast tissues [216]. This lncRNA enhances the growth and survival of BC cells by preventing ferroptosis. *LINC00460* increases the expression of *MAL2* (*mal*, T cell differentiation protein 2), a protein that belongs to the *MAL* family and is associated with cancer development, by binding to *MIR320A* [217]. The effects of *LINC00460* inhibition on BC cell proliferation and ferroptosis are reversed by *MAL2* overexpression. These results indicate that the *LINC00460*-*MAL2* pathway could be a new indicator and treatment option for BC [217].

Another group of researchers revealed that four lncRNAs *LINC01152*, *AC004585.1*, *MAPT-IT1*, and *AC026401.3* have a significant relation to BC prognosis [218]. Low expression of *LINC01152* is linked to poor OS in patients. The study also showed that *MAPT-IT1* has a significant association with BC OS. BC has a higher expression of *AC026401.3*, which suggests that *AC026401.3* could be a prognostic marker for BC [216,219,220]. These results imply that these four lncRNAs could be potential indicators of BC prognosis [218].

Largely found in the cytoplasm, the lncRNA *H19* (*H19* imprinted maternally expressed transcript) is responsible for regulating a number of different biological functions. For example, *H19* is necessary for embryonic development and growth and is involved in controlling the expression of other genes via a variety of processes [221]. Many human cancers have been related to elevated *H19* expression, indicating that *H19* may be a promising target for treatment. *H19* can boost ferroptosis by preventing autophagy [222]. This revelation emphasizes the function of *H19* in controlling cellular fate and clarifies the intricate relationship between autophagy and ferroptosis. In human tissues and plasma, *H19* also shows stability, which makes it a viable biomarker and therapeutic target for the treatment of cancer. Taken together, these data offer a theoretical framework for comprehending the function of *H19* in the genesis of cancer and the medicinal possibilities of metformin, a

medication that causes ferroptosis [223].

A specific evaluation performed by Fan et al. identified the lncRNA *LncFASA* as a tumor suppressor in triple-negative breast cancer (TNBC) [224]. They reported that *LncFASA* increases TNBC susceptibility to ferroptosis. Mechanistically, *LncFASA* directly binds to the Ahpc-TSA domain of *PRDX1* (peroxiredoxin 1), a peroxidase enzyme. This binding drives the formation of *PRDX1*-containing droplets and disrupts its peroxidase activity through liquid-liquid phase separation. Consequently, *LncFASA* disrupts intracellular ROS homeostasis, leading to lipid peroxidation accumulation via the *SLC7A11*-*GPX4* axis. Notably, high *LncFASA* expression correlates with improved overall survival in breast cancer patients. Furthermore, *LncFASA* impedes the growth of breast xenograft tumors by promoting ferroptosis, highlighting its potential as a therapeutic target [224].

Twenty-one ferroptosis-related lncRNAs were found to be correlated with recurrence-free survival (RFS) in BC patients. To predict the recurrence of BC, these lncRNAs may combine to form a new signature. Among the 21 lncRNAs, *LINC01235* is particularly significant because aggressive BC cells express a high level of this gene [225]. Another lncRNA in the signature, *LINC02166*, is also implicated in autophagy and has the potential to enhance the prognostic significance of BC [84]. It is still unclear what other lncRNAs do in BC. In a different study, researchers looked at the TCGA-BCRA cohort's expression of ferroptosis-related lncRNAs and selected lncRNAs linked to OS in BC patients [226]. *CYTOR* (cytoskeleton regulator RNA) is overexpressed in colorectal cancer samples and is linked to a poorer prognosis. This suggests that *CYTOR* may have an impact on proliferation and metastasis [227]. Another lncRNA that was studied, *USP30-AS1*, is also linked to a longer overall survival in patients with cervical cancer. The study's findings add to our understanding of lncRNA involvement in BC [228]. A notable upsurge is observed in *LINC01235* and *LINC02166* in breast cancer cell lines [229,230]. More investigation is required to elucidate the precise biological roles of these lncRNAs, which may play a significant role in the development of tumors. Ferroptosis-related lncRNA models developed in this work also provide hints regarding the molecular mechanisms underlying ferroptosis. The lack of stratification analysis based on BC molecular subtypes and the limited validation using external databases, however, could introduce biases into the results and therefore call for more research [225].

Together, it is possible to determine the prognosis of BC and possibly uncover the underlying mechanisms of lncRNAs in ferroptosis by examining the ferroptosis-associated lncRNAs found in different studies. With respect to accurately predicting how BC patients will react to immunotherapy, the developed predictive model shows great promise. To verify the model using separate datasets and determine whether it applies to various BC molecular subtypes, more research is necessary [231].

6.3. Circular RNAs also interfere with ferroptosis to control the onset and progression of female-oriented cancers

As substantial regulators of chemosensitivity in a variety of cancers, circRNAs have come to light [233]. Researchers examined whether the ferroptosis-regulating circRNA *circSNX12* aids in cisplatin resistance in OC in light of the increasing significance of ferroptosis in cancer treatment [234]. It was observed that OC tissues and cisplatin (DDP)-resistant cells have considerably higher levels of *circSNX12*. Ferroptosis is improved and DDP sensitivity is regained in DDP-resistant OC cells by reducing *circSNX12* expression [234]. Additionally, DDP's anti-tumor efficacy in vivo is improved by pharmacologically suppressing *circSNX12* via viral shRNA delivery. These results show that chemo-resistance in OC can be effectively prevented in vitro and in vivo by downregulating *circSNX12*. Researchers determined that *MIR194-5p* might be *circSNX12*'s target by using bioinformatic analysis [234]. Several malignancies have been linked to *MIR194-5p* as a mechanism of chemo-resistance. The researchers found that *MIR194-5p* regulates ZEB1

(zinc finger E-box binding homeobox 1) and MDM2 (MDM2 proto-oncogene) expression in OC cells, which supports earlier findings [235,236]. Additionally, they noticed that through sequestering *MIR194-5p*, *circSNX12* increases the expression of *SLC7A11* in OC. *MIR194-5p*'s exact function in OC DDP resistance is still unknown. The researchers did note, however, that in DDP-resistant OC tissues, *MIR194-5p* expression is repressed. To fully understand the mechanisms by which circRNA-miRNA interactions affect cancer will require additional research [234].

To determine which circRNAs are overexpressed in OC, researchers looked at the publicly accessible dataset GSE192410. Using 113 OC tissues, along with the corresponding normal tissues as a patient cohort, they assessed the expression levels of a particular circRNA, *hsa_circ_0007615*. They verified that *hsa_circ_0007615* has a prognostic value for predicting the overall survival and recurrence of OC patients by examining clinical parameters, Kaplan-Meier curves, and Cox proportional hazards models [237]. With OC cell lines, the researchers performed cell-based experiments to evaluate the functional implications of *hsa_circ_0007615*. Assays for cell proliferation, transwell migration, and cell death were used to examine the consequences of *hsa_circ_0007615* knockdown [237]. Based on their research, they discovered that *hsa_circ_0007615* knockdown promotes ferroptosis while suppressing invasion, migration, and proliferation of cells. Notably, blocking the regulatory microRNA *MIR874-3p* can counteract the tumor-suppressive effect of *hsa_circ_0007615* knockdown. *MIR874-3p* targets *TUBB3* (tubulin beta 3 class III), a gene involved in cell proliferation and microtubule stability. All things considered, *hsa_circ_0007615* may be a useful biomarker for OC prognosis, and its suppression can halt the growth of tumors. By controlling *TUBB3* and sponging *MIR874-3p*, *hsa_circ_0007615* may mechanistically aid in the advancement of OC. *Hsa_circ_0007615* may thus prove to be the basis for a useful treatment [237].

In the development of some cancers, the circRNA *circACAP2* is essential. Research has demonstrated that *MIR143-3p* regulates HK2 (hexokinase 2) expression, thereby promoting invasion and migration of neuroblastoma cells [238]. Through its interaction with *MIR193A-5p* and impact on GPX4 expression, *circACAP2* modulates cancer cell ferroptosis. Scavenging ROS is a key function of GPX4, which helps shield cells from ferroptosis [239]. Reduction of cell viability and elevation of ROS, Fe²⁺, and iron levels are observed in CC cells upon downregulating *circACAP2* with siRNAs. By inhibiting ferroptosis, these results imply that *circACAP2* aids in the advancement of CC. Additionally, *circACAP2* functions as a *MIR193A-5p* ceRNA, targeting and downregulating GPX4 expression. *circACAP2* indirectly promotes GPX4 expression and shields cells from ferroptosis by sequestering *MIR193A-5p* [239]. *CircACAP2* knockdown-induced inhibition of cell viability may be mitigated by overexpressing GPX4 or inhibiting *MIR193A-5p*. GPX4 and *circACAP2* have elevated expression in CC tissues, whereas the expression of *MIR193A-5p* is downregulated. These data imply that a potential therapeutic target for CC treatment may be the *circACAP2-MIR193A-5p-GPX4* axis [239].

Prior studies have demonstrated that *circEPSTI1* stimulates the growth of CC cells by controlling the ferroptosis mediated by *SLC7A11*. Additionally, *circEPSTI1* is essential for the invasion and metastasis of cancer [240]. Research has shown that *circEPSTI1* controls the expression of *EPSTI1* and influences the progression of OC through *MIR942*, and it also modulates the apoptosis and proliferation of BC through the ceRNA mechanism of *MIR6809-MIR4753* [241]. Nevertheless, the precise function of *circEPSTI1* in CC remains incompletely understood. A recent study looked into *circEPSTI1*'s function in CC. In comparison to normal cells, the researchers observed that CC cell lines have higher levels of *circEPSTI1* expression. They also showed that the growth of CC cell lines is markedly repressed by *circEPSTI1* knockdown using siRNAs [242]. Furthermore, in mouse xenograft models of HeLa cell lines, *circEPSTI1* silencing inhibits the growth of tumors. According to these results, *circEPSTI1* functions as an oncogene and may present a therapeutic

target worth pursuing in the management of CC. The mechanism by which *circEPSTI1* works was further examined. It was discovered that *MIR375*, *MIR409-3p*, and *MIR515-5p* are sequestered by *circEPSTI1*, which functions as a ceRNA [242]. These microRNAs target and suppress the expression of the ferroptosis-related gene *SLC7A11*. Through the sequestration of these miRNAs, *circEPSTI1* suppresses ferroptosis and indirectly increases the expression of *SLC7A11*, both of which are involved in CC cell proliferation [242].

Subsequent research revealed that ferrostatin-1/Fer-1 significantly blocks *circLMO1*-induced cell death, suggesting that *circLMO1* causes ferroptosis to trigger cell death in CC cells [243]. It was discovered that *circLMO1* does not control *SLC7A11* expression, in contrast to an earlier study. Rather, *circLMO1* sequesters *MIR4291* to cause ferroptosis, which in turn causes CC cells to express *ACSL4* more abundantly. As anticipated, the promotion of ferroptosis by *circLMO1* is effectively countered by overexpressing *MIR4291* or downregulating *ACSL4* [243]. The results indicate that *circLMO1* is downregulated in cancerous cells, and its overexpression suppresses the growth and metastasis of CC by encouraging ferroptosis mediated by *MIR4291-ACSL4*. For the purpose of creating therapeutic approaches for the treatment of CC, this mechanism offers a fresh target [243].

With regard to the gynecological malignancy endometrial cancer, tumor cells have significantly higher levels of the circRNA *circRAPGEF5*, which is primarily concentrated in the nucleus [244]. Researchers found that *circRAPGEF5* increases EC cell proliferation and resistance to ferroptosis through gain-of-function and loss-of-function experiments. By controlling the RNA binding protein RBFOX2 (RNA binding fox-1 homolog 2) splicing activity toward the *TFRC* (transferrin receptor) pre-mRNA, *circRAPGEF5* exhibits its pro-tumor effects. *circRAPGEF5* can directly bind to the RBFOX2 protein's Fox-1 C-terminal domain, significantly lowering RBFOX2's binding to downstream genes' pre-mRNAs [244]. Prior work conducted by Hilmar and colleagues showed that MALT1 (MALT1 paracaspase), a protease involved in the BCR (BCR activator of RhoGEF and GTPase)-NFKB signaling pathway, has its splicing regulation controlled by RBFOX2. This discovery was expanded upon in this study by demonstrating that RBFOX2 is also essential for the splicing of the transcripts of multiple other genes, such as *TFRC*, *ANXA2*, *EIF5A*, *ITGAE*, *SIKE1*, and *TSPO*, in EC cells. Furthermore, it was found that RBFOX2's ability to splice these target genes is diminished by a direct binding of the circular RNA *circRAPGEF5*. The formation of alternative splicing isoforms that facilitate tumor progression is a result of *circRAPGEF5* binding to RBFOX2. More research is necessary to determine the exact molecular mechanism by which *circRAPGEF5* blocks RBFOX2 splicing activity [244].

Furthermore, the phosphoinositide 3-kinase (PI3K) signaling pathway and the inactivation of the PTEN (phosphatase and tensin homolog) tumor suppressor gene, which are common characteristics of many cancers, have been demonstrated to make human cancer cells resistant to ferroptosis [245]. Furthermore, when the PI3K-AKT (AKT serine/threonine kinase)-MTOR signaling pathway is inhibited, cancer cells are generally more vulnerable to ferroptosis [245]. According to these findings, endometrial cancer often exhibits dysregulation of the PI3K-AKT-MTOR pathway, and the majority of ECs have *PTEN* gene inactivating mutations, which are present in up to 83% of endometrioid tumors [246]. These data lead us to hypothesize that *circRAPGEF5* could sequester RBFOX2 and thereby desensitize EC cells to ferroptosis. This novel mechanism may contribute to the resistance of EC cells to ferroptosis, suggesting that *circRAPGEF5* may be a viable therapeutic target for the treatment of this disease [244].

Other than the genital tract malignancies, to find better ways for treating ERBB2/HER2-positive BC patients, it is crucial to understand why some of them become resistant to trastuzumab, a drug that is often used for this type of cancer [247]. Researchers found that a circular RNA molecule called *circBGN* is very abundant in trastuzumab-resistant breast cancer tissues and is associated with poor prognosis [248]. *CircBGN* promotes ferroptosis by increasing the interaction between

OTUB1 (OTU deubiquitinase, ubiquitin aldehyde binding 1) and SLC7A11 and the level of SLC7A11 protein. This ferroptosis is not affected by inhibitors of other cell death pathways, such as apoptosis, necroptosis, or autophagy, suggesting that it is specifically related to trastuzumab resistance. Moreover, the ferroptosis inducer erastin can make breast cancer cells more sensitive to trastuzumab, and this effect is enhanced when *circBGN* is knocked down. These findings suggest that a new strategy to overcome trastuzumab resistance in ERBB2/HER2-positive breast cancer patients could be to target the *circBGN*-OTUB1-SLC7A11 pathway and use ferroptosis inducers together with trastuzumab [248].

One transcription factor involved in the development of BC is FOXQ1 (forkhead box Q1) [249]. Prior research has demonstrated that FOXQ1 can accelerate the growth of tumors by upregulating the expression of ferroptosis-inhibiting genes *SLC7A11* and *GPX4*. Nevertheless, it is still unknown how FOXQ1 controls SLC7A11. The authors of this study examined the function of FOXQ1 in controlling SLC7A11 and discovered that FOXQ1 can upregulate the expression of *circ_0000643*, a circular RNA that interacts with *MIR153*, to control the expression of SLC7A11 [250]. Additionally, the authors demonstrated that FOXQ1 can directly bind to the *circ_0000643* host gene's promoter to boost transcription of that gene. The development of novel therapeutic approaches for breast cancer may aim to target this mechanism [250].

BC progression has also been demonstrated to be influenced by *circ0052112*, *circ0001982*, and *circ0072309*. The targets of these circular RNAs increase invasion and migration of BC cells, inhibit *MIR143* to enhance carcinogenesis, and regulate *MIR492* to suppress invasion and proliferation, respectively, in BC cells [104,251,252]. Not only does *circRHOT1* suppress apoptosis and ferroptosis in BC cells, but it also stimulates the invasion, migration, and multiplication of these cells by controlling SLC7A11. Thus, *circRHOT1* may be a target for breast cancer treatment [253]. Mechanistically, by enclosing *MIR106A-5p*, *circRHOT1* suppresses ferroptosis in BC cells. In these cells, *MIR106A-5p* induces ferroptosis by targeting *STAT3* (signal transducer and activator of transcription 3). The inhibition of proliferation and enhancement of apoptosis caused by *circRHOT1* depletion in BC cells are reversed by overexpression of *STAT3* and inhibition of *MIR106A-5p*. These results provide new evidence that the *circRHOT1*-*MIR106A-5p*-*STAT3* signaling pathway is important in controlling the progression of BC and uncover a correlation between *circRHOT1*, *MIR106A-5p*, and *STAT3* [253].

In order to help BC cells withstand the cell death brought on by ferroptosis, researchers have identified a new system against ferroptosis involving AIFM2/FSP1 (apoptosis inducing factor mitochondria associated 2). Due to its sponging of *MIR1228*, *circGFRA1* can control the expression of AIFM2. The increase in AIFM2 levels prevents ERBB2/HER2-positive BC cells from undergoing ferroptosis [254]. Through the reduction of CoQ10 (coenzyme Q10), an endogenous antioxidant that inhibits ferroptosis, by NAD(P)H, AIFM2 has an anti-ferroptosis effect [255,256]. Another route that keeps cells safe from ferroptosis is the GSH-GPX4 system. Researchers discovered that ERBB2/HER2-positive BC cells have higher expression of *GPX4*, AIFM2, and the glutathione-level indicator GSH:GSSG ratio. ERBB2/HER2-positive breast cancer treatment may benefit from targeting these pathways as a therapeutic approach [254].

7. Ferroptosis and ncRNAs: exploring therapeutic strategies, limitations, and directions of improvement that could be further investigated

Ferroptosis has been implicated in various pathologies including neurodegeneration, organ fibrosis, and ischemia-reperfusion injuries [65,257,258]. Notably, cancer cells are particularly susceptible to ferroptosis [259,260]. Mesenchymal and dedifferentiated cancer cells, often resistant to traditional therapies and apoptosis, are highly susceptible to ferroptosis inducers, suggesting its potential to overcome therapeutic resistance.

Several strategies are being explored to exploit ferroptosis for cancer therapy. Targeting key ferroptosis enzymes in cancer cells is one approach. Pharmacological and genetic inhibition of the cystine/glutamate antiporter/xCT, achieved by blocking SLC3A2 and SLC7A11, has yielded promising results in preclinical models with minimal toxicity [14,261,262]. Similarly, targeting AIFM2/FSP1 is a promising avenue due to its dispensability in normal development, suggesting a potentially broad therapeutic window [263,264].

However, *GPX4*, another ferroptosis target, is crucial for healthy tissues including renal cells and neurons [265–267]. Clearly, *GPX4* inhibitors (e.g., RSL3) require specific delivery to cancer cells to minimize side effects. Indirect ferroptosis inducers such as erastin may suffer from low solubility and rapid metabolic breakdown [268]. Encapsulation of ferroptosis inducers within protective delivery systems, such as nanoparticles, is being explored to address this issue.

Nanoparticle-based delivery of iron, peroxides, and ncRNAs targeting inhibitors of ferroptosis are actively being investigated in vitro and in vivo. ncRNAs, in particular, offer several advantages. They are naturally occurring cellular molecules, potentially leveraging existing metabolic pathways. Additionally, ncRNAs often target multiple genes across interconnected pathways, leading to a broader yet specific anti-cancer response — exemplified by the *MIR15*-*MIR16* cluster, which regulates multiple anti-apoptotic and cell cycle proteins [269]. Eventually, ncRNA therapeutics hold promise for cost-effective production through chemical synthesis.

Despite these advantages, ncRNA-based ferroptosis therapies face potential limitations. First, the regulation of tumorigenesis through ncRNA-mediated ferroptosis may have restricted efficacy. Second, individual variations in ncRNA expression and response to therapeutic interventions pose a challenge for predictability. Third, achieving a balance between promoting ferroptosis for tumor suppression and preventing chemoresistance using ncRNAs requires further investigation. Consequently, further research is necessary to explore the clinical potential of targeting ferroptosis-related ncRNAs.

Our understanding of ferroptosis remains incomplete, with several key questions unanswered. The precise relationship between ferroptosis and other regulated cell death pathways, such as TP53-mediated apoptosis, with share some upstream mechanisms, requires further elucidation. While iron is a key player in ferroptosis, the possibility of redox-independent roles for iron and the involvement of other metals such as copper necessitate further investigation. Additionally, a comprehensive understanding of the molecular events leading to ferroptosis execution is lacking. This knowledge gap is particularly pronounced concerning the downstream events following lipid peroxidation, especially the critical point(s) beyond which ferroptosis becomes irreversible.

Furthermore, the lack of specific markers for identifying ferroptosis in live cells and intact tissues presents a significant challenge. ncRNAs, a diverse group of non-coding transcripts with remarkable regulatory and biomarker potential, remain largely unexplored in the context of ferroptosis and cancer. While current evidence suggests that dysregulation of tightly controlled ncRNA networks in cancer frequently suppresses ferroptosis, which promotes tumor cell survival and progression, further research is necessary. Nonetheless, the ability to artificially induce ferroptosis holds significant therapeutic promise for cancer treatment.

8. Conclusion

Ferroptosis is known as a newly identified regulated cell death mechanism, involving iron-dependent lipid peroxidation and oxidative stress. This type of regulated cell death has been implicated in various diseases, particularly cancers. This review explores the role of ferroptosis in the progression of female-specific cancers, such as breast cancer and gynecological malignancies, and how it is regulated by ncRNAs. The review summarizes the current understanding of the molecular mechanisms, biomarker potential, and therapeutic implications of ferroptosis

and ncRNAs in these cancers. Additionally, it highlights the challenges and promising directions for future research in this area. The authors conclude that ferroptosis and ncRNAs hold promise as targets for the diagnosis and treatment of female-specific cancers, and that further studies are essential to fully comprehend their intricate interactions and functions.

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References

- [1] Q. Nie, et al., Induction and application of ferroptosis in cancer therapy, *Cancer Cell Int.* 22 (1) (2022) 1–19.
- [2] T. Hirschhorn, B.R. Stockwell, The development of the concept of ferroptosis, *Free Radic. Biol. Med.* 133 (2019) 130–143.
- [3] L. Galluzzi, et al., Molecular mechanisms of cell death: recommendations of the Nomenclature Committee on cell death 2018, *Cell Death Differ.* 25 (3) (2018) 486–541.
- [4] B.A. Carneiro, W.S. El-Deiry, Targeting apoptosis in cancer therapy, *Nat. Rev. Clin. Oncol.* 17 (7) (2020) 395–417.
- [5] S.J. Dixon, et al., Ferroptosis: an iron-dependent form of nonapoptotic cell death, *Cell* 149 (5) (2012) 1060–1072.
- [6] C.M. Bebbler, et al., Ferroptosis in cancer cell biology, *Cancers* 12 (1) (2020) 164.
- [7] D. Tang, et al., The molecular machinery of regulated cell death, *Cell Res.* 29 (5) (2019) 347–364.
- [8] B. Chu, et al., ALOX12 is required for p53-mediated tumour suppression through a distinct ferroptosis pathway, *Nat. Cell Biol.* 21 (5) (2019) 579–591.
- [9] J. Tsoi, et al., Multi-stage differentiation defines melanoma subtypes with differential vulnerability to drug-induced iron-dependent oxidative stress, *Cancer Cell* 33 (5) (2018) 890–904. e5.
- [10] W. Wang, et al., CD8+ T cells regulate tumour ferroptosis during cancer immunotherapy, *Nature* 569 (7755) (2019) 270–274.
- [11] Y. Luo, et al., Regulation of ferroptosis by non-coding RNAs in the development and treatment of cancer, *Oncol. Rep.* 45 (1) (2021) 29–48.
- [12] Y. Mou, et al., Ferroptosis, a new form of cell death: opportunities and challenges in cancer, *J. Hematol. Oncol.* 12 (1) (2019) 1–16.
- [13] H.O. Fearnhead, P. Vandenabeele, T. Vanden Berghe, How do we fit ferroptosis in the family of regulated cell death? *Cell Death Differ.* 24 (12) (2017) 1991–1998.
- [14] M.A. Badgley, et al., Cysteine depletion induces pancreatic tumor ferroptosis in mice, *Science* 368 (6486) (2020) 85–89.
- [15] S. Lukasiewicz, et al., Breast cancer—epidemiology, risk factors, classification, prognostic markers, and current treatment strategies—an updated review, *Cancers* 13 (17) (2021) 4287.
- [16] Y. Feng, et al., Breast cancer development and progression: risk factors, cancer stem cells, signaling pathways, genomics, and molecular pathogenesis, *Genes & diseases* 5 (2) (2018) 77–106.
- [17] C. Sonnenschein, A.M. Soto, Carcinogenesis explained within the context of a theory of organisms, *Prog. Biophys. Mol. Biol.* 122 (1) (2016) 70–76.
- [18] C. Dumars, et al., Dysregulation of macrophage polarization is associated with the metastatic process in osteosarcoma, *Oncotarget* 7 (48) (2016) 78343.
- [19] F. Naz, et al., The role of long non-coding RNAs (lncRNAs) in female oriented cancers, *Cancers* 13 (23) (2021).
- [20] P. Jessmon, et al., Epidemiology and treatment patterns of epithelial ovarian cancer, *Expert Rev. Anticancer Ther.* 17 (5) (2017) 427–437.
- [21] F. Naz, et al., The role of long non-coding RNAs (lncRNAs) in female oriented cancers, *Cancers* 13 (23) (2021) 6102.
- [22] J. Huang, et al., Worldwide burden, risk factors, and temporal trends of ovarian cancer: a global study, *Cancers* 14 (9) (2022) 2230.
- [23] Z. Nash, U. Menon, Ovarian cancer screening: current status and future directions, *Best Pract. Res. Clin. Obstet. Gynaecol.* 65 (2020) 32–45.
- [24] D. Singh, et al., Global estimates of incidence and mortality of cervical cancer in 2020: a baseline analysis of the WHO Global Cervical Cancer Elimination Initiative, *Lancet Global Health* 11 (2) (2023) e197–e206.
- [25] J. Lei, et al., HPV vaccination and the risk of invasive cervical cancer, *N. Engl. J. Med.* 383 (14) (2020) 1340–1348.
- [26] S. Kamamoto, et al., HPV vaccination and cervical cancer screening, *Lancet* 399 (10339) (2022) 1939–1940.
- [27] M. Vu, et al., Cervical cancer worldwide, *Current problems in cancer* 42 (5) (2018) 457–465.
- [28] K. Alfaro, et al., Removing global barriers to cervical cancer prevention and moving towards elimination, *Nat. Rev. Cancer* 21 (10) (2021) 607–608.
- [29] Z. Shaheen, et al., Clinical correlation between cervical cancer screening using Pap smear test, *Pakistan Journal of Medical & Health Sciences* 16 (7) (2022), 900–900.
- [30] R.A. Brooks, et al., Current recommendations and recent progress in endometrial cancer, *CA: a cancer journal for clinicians* 69 (4) (2019) 258–279.
- [31] E. Kalamokas, et al., Current approaches to the management of patients with endometrial cancer, *Cancers* 14 (18) (2022) 4500.
- [32] E.J. Crosbie, et al., Endometrial cancer, *Lancet* 399 (10333) (2022) 1412–1428.
- [33] T. Dörk, et al., Genetic susceptibility to endometrial cancer: risk factors and clinical management, *Cancers* 12 (9) (2020) 2407.
- [34] L.K. Nees, et al., Endometrial hyperplasia as a risk factor of endometrial cancer, *Arch. Gynecol. Obstet.* (2022) 1–15.
- [35] A.R. Bogdan, et al., Regulators of iron homeostasis: new players in metabolism, cell death, and disease, *Trends Biochem. Sci.* 41 (3) (2016) 274–286.
- [36] B. Galy, M. Conrad, M. Muckenthaler, Mechanisms controlling cellular and systemic iron homeostasis, *Nat. Rev. Mol. Cell Biol.* (2023) 1–23.
- [37] D.H. Manz, et al., Iron and cancer: recent insights, *Ann. N. Y. Acad. Sci.* 1368 (1) (2016) 149–161.
- [38] S.L. Pandrangi, et al., Role of dietary iron revisited: in metabolism, ferroptosis and pathophysiology of cancer, *Am. J. Cancer Res.* 12 (3) (2022) 974.
- [39] M. Salami, et al., Therapeutic potential of resveratrol in diabetic nephropathy according to molecular signaling, *Curr. Mol. Pharmacol.* 15 (5) (2022) 716–735.
- [40] H. Yu, et al., Ferroptosis, a new form of cell death, and its relationships with tumorous diseases, *J. Cell Mol. Med.* 21 (4) (2017) 648–657.
- [41] H. Sies, et al., Defining roles of specific reactive oxygen species (ROS) in cell biology and physiology, *Nat. Rev. Mol. Cell Biol.* 23 (7) (2022) 499–515.
- [42] J.Y. Cao, S.J. Dixon, Mechanisms of ferroptosis, *Cell. Mol. Life Sci.* 73 (2016) 2195–2209.
- [43] M. Liu, et al., The critical role and molecular mechanisms of ferroptosis in antioxidant systems: a narrative review, *Ann. Transl. Med.* 10 (6) (2022).
- [44] Y. Luo, et al., Ferroptosis and its potential role in glioma: from molecular mechanisms to therapeutic opportunities, *Antioxidants* 11 (11) (2022).
- [45] S. Ghavami, et al., Epigenetic regulation of autophagy in gastrointestinal cancers, *Biochimica et Biophysica Acta (BBA)-Molecular Basis of Disease* 1868 (11) (2022) 166512.
- [46] Z. Dashti, et al., Autophagy and the unfolded protein response shape the non-alcoholic fatty liver landscape: decoding the labyrinth, *Metabolism* (2024) 155811.
- [47] H. Behrouj, et al., Epigenetic regulation of autophagy in coronavirus disease 2019 (COVID-19), *Biochemistry and Biophysics Reports* 30 (2022) 101264.
- [48] X. Chen, et al., Cellular degradation systems in ferroptosis, *Cell Death Differ.* 28 (4) (2021) 1135–1148.
- [49] R. Singh, A. Letai, K. Sarosiek, Regulation of apoptosis in health and disease: the balancing act of BCL-2 family proteins, *Nat. Rev. Mol. Cell Biol.* 20 (3) (2019) 175–193.
- [50] X. Liu, et al., Induction of apoptotic program in cell-free extracts: requirement for dATP and cytochrome c, *Cell* 86 (1) (1996) 147–157.
- [51] A. Fraser, G. Evan, A license to kill, *Cell* 85 (6) (1996) 781–784.
- [52] S.J. Riedl, Y. Shi, Molecular mechanisms of caspase regulation during apoptosis, *Nat. Rev. Mol. Cell Biol.* 5 (11) (2004) 897–907.
- [53] T.V. Berghe, et al., Determination of apoptotic and necrotic cell death in vitro and in vivo, *Methods* 61 (2) (2013) 117–129.
- [54] A. Mafi, et al., Melatonin as a regulator of apoptosis in leukaemia: molecular mechanism and therapeutic perspectives, *Front. Pharmacol.* 14 (2023) 1224151.

- [55] W.J. Kaiser, et al., Toll-like receptor 3-mediated necrosis via TRIF, RIP3, and MLKL, *J. Biol. Chem.* 288 (43) (2013) 31268–31279.
- [56] S. He, et al., Toll-like receptors activate programmed necrosis in macrophages through a receptor-interacting kinase-3-mediated pathway, *Proc. Natl. Acad. Sci. USA* 108 (50) (2011) 20054–20059.
- [57] M. Brault, et al., Intracellular nucleic acid sensing triggers necroptosis through synergistic type I IFN and TNF signaling, *J. Immunol.* 200 (8) (2018) 2748–2756.
- [58] S.N. Schock, et al., Induction of necroptotic cell death by viral activation of the RIG-I or STING pathway, *Cell Death Differ.* 24 (4) (2017) 615–625.
- [59] Y.K. Dhuriya, D. Sharma, Necroptosis: a regulated inflammatory mode of cell death, *J. Neuroinflammation* 15 (2018) 1–9.
- [60] A. Zychlinsky, M.C. Prevost, P.J. Sansonetti, *Shigella flexneri* induces apoptosis in infected macrophages, *Nature* 358 (6382) (1992) 167–169.
- [61] B.T. Cookson, M.A. Brennan, Pro-inflammatory programmed cell death, *Trends Microbiol.* 9 (3) (2001) 113–114.
- [62] Y.-L. Gao, J.-H. Zhai, Y.-F. Chai, Recent advances in the molecular mechanisms underlying pyroptosis in sepsis, *Mediat. Inflamm.* (2018) 2018.
- [63] Z. Chen, et al., Ferroptosis as a potential target for cancer therapy, *Cell Death Dis.* 14 (7) (2023) 460.
- [64] Y.B. Zuo, et al., Ferroptosis in cancer progression: role of noncoding RNAs, *Int. J. Biol. Sci.* 18 (5) (2022) 1829–1843.
- [65] X. Jiang, B.R. Stockwell, M. Conrad, Ferroptosis: mechanisms, biology and role in disease, *Nat. Rev. Mol. Cell Biol.* 22 (4) (2021) 266–282.
- [66] D. Tang, et al., Ferroptosis: molecular mechanisms and health implications, *Cell Res.* 31 (2) (2021) 107–125.
- [67] W. Zeng, et al., The interplay of oncogenic signaling, oxidative stress and ferroptosis in cancer, *Int. J. Cancer* (2023).
- [68] S.J. Dixon, B.R. Stockwell, The hallmarks of ferroptosis, *Annual Review of Cancer Biology* 3 (2019) 35–54.
- [69] D.M. Kremer, et al., GOT1 inhibition promotes pancreatic cancer cell death by ferroptosis, *Nat. Commun.* 12 (1) (2021) 4860.
- [70] Y. Guo, et al., Tumor cell derived exosomal GOT1 suppresses tumor cell ferroptosis to accelerate pancreatic cancer progression by activating Nrf2/HO-1 axis via upregulating CCR2 expression, *Cells* 11 (23) (2022) 3893.
- [71] Y. Lu, et al., KLF2 inhibits cancer cell migration and invasion by regulating ferroptosis through GPX4 in clear cell renal cell carcinoma, *Cancer letters* 522 (2021) 1–13.
- [72] J. Hou, et al., ACSL4 as a potential target and biomarker for anticancer: from molecular mechanisms to clinical therapeutics, *Front. Pharmacol.* 13 (2022) 949863.
- [73] L. Feng, et al., SLC7A11 regulated by NRF2 modulates esophageal squamous cell carcinoma radiosensitivity by inhibiting ferroptosis, *J. Transl. Med.* 19 (2021) 1–16.
- [74] L. An, M. Li, Q. Jia, Mechanisms of radiotherapy resistance and radiosensitization strategies for esophageal squamous cell carcinoma, *Mol. Cancer* 22 (1) (2023) 140.
- [75] Q. He, et al., IL-1 β -induced elevation of solute carrier family 7 member 11 promotes hepatocellular carcinoma metastasis through up-regulating programmed death ligand 1 and colony-stimulating factor 1, *Hepatology* 74 (6) (2021) 3174–3193.
- [76] W. Liu, et al., Dysregulated cholesterol homeostasis results in resistance to ferroptosis increasing tumorigenicity and metastasis in cancer, *Nat. Commun.* 12 (1) (2021) 5103.
- [77] S.A.A. Al-Mahmood, Nanotechnology Approach for Targeted Treatment of Triple Negative Breast Cancer, Rutgers University-Graduate School, New Brunswick, 2016.
- [78] F. Xu, et al., The roles of ferroptosis regulatory gene SLC7A11 in renal cell carcinoma: a multi-omics study, *Cancer Med.* 10 (24) (2021) 9078–9096.
- [79] W. Huang, et al., ABCG5 facilitates the acquired resistance of sorafenib through the inhibition of SLC7A11-induced ferroptosis in hepatocellular carcinoma, *Neoplasia* 23 (12) (2021) 1227–1239.
- [80] L. Guo, et al., Mechanism of sorafenib resistance associated with ferroptosis in HCC, *Front. Pharmacol.* 14 (2023) 1207496.
- [81] Y. Wang, et al., Ferroptosis induction via targeting metabolic alterations in triple-negative breast cancer, *Biomed. Pharmacother.* 169 (2023) 115866.
- [82] E. Dai, et al., Ferroptotic damage promotes pancreatic tumorigenesis through a TMEM173/STING-dependent DNA sensor pathway, *Nat. Commun.* 11 (1) (2020) 6339.
- [83] J. Liu, et al., TMEM164 is a new determinant of autophagy-dependent ferroptosis, *Autophagy* 19 (3) (2023) 945–956.
- [84] X. Ma, et al., CD36-mediated ferroptosis dampens intratumoral CD8+ T cell effector function and impairs their antitumor ability, *Cell Metabol.* 33 (5) (2021) 1001–1012. e5.
- [85] Salehi, M., et al., Gastrointestinal cancer drug resistance: the role of exosomal miRNAs, *Mol. Biol. Rep.*: p. 1–12.
- [86] O. Vakili, et al., Circular RNAs in Alzheimer's disease: a new perspective of diagnostic and therapeutic targets, *CNS Neurol. Disord. - Drug Targets* (2023).
- [87] A. Movahedpour, et al., Exosomal noncoding RNAs: key players in glioblastoma drug resistance, *Mol. Cell. Biochem.* 476 (2021) 4081–4092.
- [88] A. Movahedpour, et al., Exosomal noncoding RNAs in prostate cancer, *Clin. Chim. Acta* (2022).
- [89] J.S. Mattick, et al., Long non-coding RNAs: definitions, functions, challenges and recommendations, *Nat. Rev. Mol. Cell Biol.* 24 (6) (2023) 430–447.
- [90] A. Mafi, et al., Recent insights into the microRNA-dependent modulation of gliomas from pathogenesis to diagnosis and treatment, *Cell. Mol. Biol. Lett.* 27 (1) (2022) 1–32.
- [91] S. Bahmyari, et al., microRNAs in female infertility: an overview, *Cell Biochem. Funct.* 39 (8) (2021) 955–969.
- [92] A. Mafi, et al., The significant role of microRNAs in gliomas angiogenesis: a particular focus on molecular mechanisms and opportunities for clinical application, *Cell. Mol. Neurobiol.* 43 (7) (2023) 3277–3299.
- [93] R. Mahmoudi-Lamouki, et al., Emerging role of miRNAs in the regulation of ferroptosis, *Front. Mol. Biosci.* 10 (2023) 1115996.
- [94] R. Qi, et al., The role of non-coding RNAs in ferroptosis regulation, *J. Trace Elem. Med. Biol.* 70 (2022) 126911.
- [95] S. Hu, et al., Emerging role of 12/15-Lipoxygenase (ALOX15) in human pathologies, *Prog. Lipid Res.* 73 (2019) 28–45.
- [96] H. Zhang, et al., CAF secreted miR-522 suppresses ferroptosis and promotes acquired chemo-resistance in gastric cancer, *Mol. Cancer* 19 (2020) 1–17.
- [97] X. Wang, et al., Effects of DNA, RNA, and protein methylation on the regulation of ferroptosis, *Int. J. Biol. Sci.* 19 (11) (2023) 3558.
- [98] A. Katscha, et al., Activation of EIF4E by Aurora kinase A depicts a novel druggable axis in everolimus-resistant cancer cells, *Clin. Cancer Res.* 23 (14) (2017) 3756–3768.
- [99] S. Hu, et al., Recent advances of ferroptosis in tumor: from biological function to clinical application, *Biomed. Pharmacother.* 166 (2023) 115419.
- [100] X.-D. Zhang, et al., Mechanisms and regulations of ferroptosis, *Front. Immunol.* 14 (2023).
- [101] M. Luo, et al., miR-137 regulates ferroptosis by targeting glutamine transporter SLC1A5 in melanoma, *Cell Death Differ.* 25 (8) (2018) 1457–1472.
- [102] Y. Niu, et al., RETRACTED: Physcion 8-O- β -Glucopyranoside Induced Ferroptosis via Regulating miR-103a-3p/GLS2 axis in Gastric Cancer, Elsevier, 2019.
- [103] Y. Wang, et al., Histone demethylase KDM3B protects against ferroptosis by upregulating SLC7A11, *FEBS Open Bio* 10 (4) (2020) 637–643.
- [104] Y. Chen, et al., Dihydroartemisinin-induced unfolded protein response feedback attenuates ferroptosis via PERK/ATF4/HSPA5 pathway in glioma cells, *J. Exp. Clin. Cancer Res.* 38 (1) (2019) 1–16.
- [105] T. Bai, et al., MicroRNA-214-3p enhances erastin-induced ferroptosis by targeting ATF4 in hepatoma cells, *J. Cell. Physiol.* 235 (7–8) (2020) 5637–5648.
- [106] K. Tomita, et al., MiR-7-5p is a key factor that controls radioresistance via intracellular Fe²⁺ content in clinically relevant radioresistant cells, *Biochemical and biophysical research communications* 518 (4) (2019) 712–718.
- [107] X. Li, et al., miR-335 promotes ferroptosis by targeting ferritin heavy chain 1 in vivo and in vitro models of Parkinson's disease, *Int. J. Mol. Med.* 47 (4) (2021) 1–12.
- [108] W.D. Bao, et al., Targeting miR-124/Ferroportin signaling ameliorated neuronal cell death through inhibiting apoptosis and ferroptosis in aged intracerebral hemorrhage murine model, *Aging Cell* 19 (11) (2020) e13235.
- [109] H. Zhang, et al., miR-30-5p-mediated ferroptosis of trophoblasts is implicated in the pathogenesis of preeclampsia, *Redox Biol.* 29 (2020) 101402.
- [110] K.R. Babu, M.U. Muckenthaler, miR-20a regulates expression of the iron exporter ferroportin in lung cancer, *Journal of molecular medicine* 94 (2016) 347–359.
- [111] R. Yan, et al., NRF2, a Superstar of ferroptosis, *Antioxidants* 12 (9) (2023) 1739.
- [112] D. Wang, et al., Regulatory pathways and drugs associated with ferroptosis in tumors, *Cell Death Dis.* 13 (6) (2022) 544.
- [113] S. Taghviimi, et al., Exosomal microRNAs and long noncoding RNAs: novel mediators of drug resistance in lung cancer, *J. Cell. Physiol.* 237 (4) (2022) 2095–2106.
- [114] M. Rezaee, et al., The landscape of exosomal non-coding RNAs in breast cancer drug resistance, focusing on underlying molecular mechanisms, *Front. Pharmacol.* 14 (2023) 1152672.
- [115] A. Chugunova, et al., Mining for small translated ORFs, *J. Proteome Res.* 17 (1) (2018) 1–11.
- [116] J.-W. Nam, S.-W. Choi, B.-H. You, Incredible RNA: dual functions of coding and noncoding, *Mol. Cell.* 39 (5) (2016) 367–374.
- [117] H. Sun, et al., Integration of mass spectrometry and RNA-Seq data to confirm human ab initio predicted genes and lncRNAs, *Proteomics* 14 (23–24) (2014) 2760–2768.
- [118] Z. Ji, et al., Many lncRNAs, 5'UTRs, and pseudogenes are translated and some are likely to express functional proteins, *Elife* 4 (2015) e08890.
- [119] J.-Z. Huang, et al., A peptide encoded by a putative lncRNA HOXB-AS3 suppresses colon cancer growth, *Molecular cell* 68 (1) (2017) 171–184. e6.
- [120] J. Xing, et al., LncRNA-encoded peptide: functions and predicting methods, *Frontiers in oncology* 10 (2021) 622294.
- [121] B. Kawahara, S. Sen, P.K. Mascharak, Reaction of carbon monoxide with cystathionine β -synthase: implications on drug efficacies in cancer chemotherapy, *Future Med. Chem.* 12 (4) (2020) 325–337.
- [122] M. Wang, et al., Long noncoding RNA LINC00336 inhibits ferroptosis in lung cancer by functioning as a competing endogenous RNA, *Cell Death Differ.* 26 (11) (2019) 2329–2343.
- [123] H. Bouchaoui, et al., ACSL4 and the lipoxygenases 15/15B are pivotal for ferroptosis induced by iron and PUFA dyshomeostasis in dopaminergic neurons, *Free Radic. Biol. Med.* 195 (2023) 145–157.
- [124] Y. Xie, et al., The tumor suppressor p53 limits ferroptosis by blocking DPP4 activity, *Cell Rep.* 20 (7) (2017) 1692–1704.
- [125] J. Lu, F. Xu, H. Lu, LncRNA PVT1 regulates ferroptosis through miR-214-mediated TFR1 and p53, *Life Sci.* 260 (2020) 118305.
- [126] Z. Wang, et al., A nuclear long non-coding RNA LINC00618 accelerates ferroptosis in a manner dependent upon apoptosis, *Mol. Ther.* 29 (1) (2021) 263–274.
- [127] A. He, et al., Zfas 1: a novel vital oncogenic lnc RNA in multiple human cancers, *Cell Prolif.* 52 (1) (2019) e12513.

- [128] Y. Yang, et al., lncRNA ZFAS1 promotes lung fibroblast-to-myofibroblast transition and ferroptosis via functioning as a ceRNA through miR-150-5p/SLC38A1 axis, *Aging (Albany NY)* 12 (10) (2020) 9085.
- [129] T. Qureshi, et al., The glutamine transporter Slc38a1 regulates GABAergic neurotransmission and synaptic plasticity, *Cerebr. Cortex* 29 (12) (2019) 5166–5179.
- [130] W. Luo, et al., lncRNA RP11-89 facilitates tumorigenesis and ferroptosis resistance through PROM2-activated iron export by sponging miR-129-5p in bladder cancer, *Cell Death Dis.* 12 (11) (2021) 1043.
- [131] S. Nazloomi, et al., Circular RNAs: emerging modulators in the pathophysiology of polycystic ovary syndrome and their clinical implications, *Curr. Mol. Med.* 24 (2) (2024) 153–166.
- [132] E. Heydarnia, et al., Circular RNAs and cervical cancer: friends or foes? A landscape on circRNA-mediated regulation of key signaling pathways involved in the onset and progression of HPV-related cervical neoplasms, *Cell Commun. Signal.* 22 (1) (2024) 107.
- [133] R. Salami, et al., Circular RNAs and glioblastoma multiforme: focus on molecular mechanisms, *Cell Commun. Signal.* 20 (1) (2022) 13.
- [134] S. Najafi, et al., Recent insights into the roles of circular RNAs in human brain development and neurologic diseases, *Int. J. Biol. Macromol.* 225 (2023) 1038–1048.
- [135] A. Mafi, et al., Circular RNAs; powerful microRNA sponges to overcome diabetic nephropathy, *Pathol. Res. Pract.* 227 (2021) 153618.
- [136] A. Rahmati, et al., Circular RNAs: pivotal role in the leukemogenesis and novel indicators for the diagnosis and prognosis of acute myeloid leukemia, *Front. Oncol.* 13 (2023) 1149187.
- [137] A. Rahmati, et al., Non-coding RNAs in leukemia drug resistance: new perspectives on molecular mechanisms and signaling pathways, *Ann. Hematol.* (2023) 1–28.
- [138] Z. Dorostgou, et al., Novel insights into the role of circular RNAs in Parkinson disease: an emerging renaissance in the management of neurodegenerative diseases, *J. Neurosci. Res.* 100 (9) (2022) 1775–1790.
- [139] J.-Y. Wei, et al., Circular RNA circTTBK2 facilitates non-small-cell lung cancer malignancy through the miR-873-5p/TEAD1/DERL1 axis, *Epigenomics* 14 (16) (2022) 931–949.
- [140] D.-H. Yuan, J. Zhao, G.-F. Shao, Circular RNA TTBK2 promotes the development of human glioma cells via miR-520b/EZH2 axis, *Eur. Rev. Med. Pharmacol. Sci.* 23 (24) (2019).
- [141] H.-Y. Zhang, et al., Circular RNA TTBK2 regulates cell proliferation, invasion and ferroptosis via miR-761/ITGB8 axis in glioma, *Eur. Rev. Med. Pharmacol. Sci.* 24 (5) (2020).
- [142] B. Zhou, et al., Ferroptosis is a type of autophagy-dependent cell death, in: *Seminars in Cancer Biology*, Elsevier, 2020.
- [143] F. Chen, et al., Autophagy-dependent ferroptosis in cancer, *Antioxidants Redox Signal.* 39 (1–3) (2023) 79–101.
- [144] H. Zhu, et al., ALKBH5 inhibited autophagy of epithelial ovarian cancer through miR-7 and BCL-2, *J. Exp. Clin. Cancer Res.* 38 (2019) 1–15.
- [145] Z. Liu, et al., Circular RNA ciARS regulates ferroptosis in HCC cells through interacting with RNA binding protein ALKBH5, *Cell death discovery* 6 (1) (2020) 72.
- [146] C. Li, et al., Retracted ARTICLE: circ_0008035 contributes to cell proliferation and inhibits apoptosis and ferroptosis in gastric cancer via miR-599/EIF4A1 axis, *Cancer Cell Int.* 20 (1) (2020) 1–15.
- [147] Z. Jin, et al., RNA modifications in hematological malignancies, *Int. J. Hematol.* 117 (6) (2023) 807–820.
- [148] P. Wu, et al., Circular RNA circEPST11 accelerates cervical cancer progression via miR-375/409-3P/515-5p-SLC7A11 axis, *Aging (Albany NY)* 13 (3) (2021) 4663.
- [149] R. Liu, Y. Zhou, Y. Cao, CircRNA and ferroptosis in human disease: insights for new treatments, *Animal Models and Experimental Medicine* (2023).
- [150] S. Cai, et al., SIRT6 silencing overcomes resistance to sorafenib by promoting ferroptosis in gastric cancer, *Biochemical and biophysical research communications* 577 (2021) 158–164.
- [151] X.-s. Ding, et al., Ferroptosis in Parkinson's disease: molecular mechanisms and therapeutic potential, *Ageing Res. Rev.* (2023) 102077.
- [152] Y. Wang, et al., Frizzled-7 identifies platinum-tolerant ovarian cancer cells susceptible to ferroptosis, *Cancer Res.* 81 (2) (2021) 384–399.
- [153] M. Alimohammadi, et al., Circular RNAs: novel actors of Wnt signaling pathway in lung cancer progression, *EXCLI journal* 22 (2023) 645.
- [154] M. Do, et al., A FZD7-specific Antibody-drug Conjugate induces ovarian tumor regression in preclinical models, *Mol Cancer Ther* 21 (1) (2022) 113–124.
- [155] V.H.L. Nguyen, et al., Wnt/ β -catenin signalling in ovarian cancer: insights into its hyperactivation and function in tumorigenesis, *J. Ovarian Res.* 12 (1) (2019) 122.
- [156] D. Zhang, et al., MiR-1-3p enhances the sensitivity of ovarian cancer cells to ferroptosis by targeting FZD7, *Zhong Nan Da Xue Xue Bao Yi Xue Ban* 47 (11) (2022) 1512–1521.
- [157] H. Du, et al., MicroRNA-424-5p acts as a potential biomarker and inhibits proliferation and invasion in hepatocellular carcinoma by targeting TRIM29, *Life Sci.* 224 (2019) 1–11.
- [158] Y. Zhou, et al., miR424-5p functions as an anti-oncogene in cervical cancer cell growth by targeting KDM5B via the Notch signaling pathway, *Life Sci.* 171 (2017) 9–15.
- [159] L.L. Ma, et al., Tumor suppressor miR-424-5p abrogates ferroptosis in ovarian cancer through targeting ACSL4, *Neoplasma* 68 (1) (2021) 165–173.
- [160] Y. Chen, et al., Targeting tumor-associated macrophages: a potential treatment for solid tumors, *J. Cell. Physiol.* 236 (5) (2021) 3445–3465.
- [161] Y. Luo, et al., The suppression of cervical cancer ferroptosis by macrophages: the attenuation of ALOX15 in cancer cells by macrophages-derived exosomes, *Acta Pharm. Sin. B* 13 (6) (2023) 2645–2662.
- [162] Y. Shen, et al., Inhibition of miR-660-5p expression suppresses tumor development and metastasis in human breast cancer, *Genet. Mol. Res.* 16 (1) (2017).
- [163] Y. Wu, et al., MiR-660-5p promotes the progression of hepatocellular carcinoma by interaction with YWHAH via PI3K/Akt signaling pathway, *Biochem. Biophys. Res. Commun.* 531 (4) (2020) 480–489.
- [164] Z. Li, et al., Targeting ferroptosis in breast cancer, *Biomark. Res.* 8 (1) (2020) 58.
- [165] K.L. Britt, J. Cuzick, K.A. Phillips, Key steps for effective breast cancer prevention, *Nat. Rev. Cancer* 20 (8) (2020) 417–436.
- [166] C. Liu, et al., Lidocaine inhibits the metastatic potential of ovarian cancer by blocking Na(V) 1.5-mediated EMT and FAK/Paxillin signaling pathway, *Cancer Med.* 10 (1) (2021) 337–349.
- [167] P. Yadav, et al., Lidocaine Inhibits and Intraoperative Lidocaine Infusion to Reduce Persistent Neuropathic Pain after Breast Cancer Surgery: a Multicenter, Factorial, Randomized, Controlled Pilot Trial, *J. Pain* 20 (8) (2019) 980–993.
- [168] D. Sun, Y.C. Li, X.Y. Zhang, Lidocaine promoted ferroptosis by targeting miR-382-5p/SLC7A11 Axis in ovarian and breast cancer, *Front. Pharmacol.* 12 (2021) 681223.
- [169] P. Yadav, et al., SLC7A11/xCT is a target of miR-5096 and its restoration partially rescues miR-5096-mediated ferroptosis and anti-tumor effects in human breast cancer cells, *Cancer Lett.* 522 (2021) 211–224.
- [170] N. Saini, X. Yang, Metformin as an anti-cancer agent: actions and mechanisms targeting cancer stem cells, *Acta Biochim. Biophys. Sin.* 50 (2) (2018) 133–143.
- [171] C. Deng, et al., Metformin induces ferroptosis through the Nrf2/HO-1 signaling in lung cancer, *BMC Pulm. Med.* 23 (1) (2023) 360.
- [172] J. Yang, et al., Metformin induces Ferroptosis by inhibiting UFMylation of SLC7A11 in breast cancer, *J. Exp. Clin. Cancer Res.* 40 (1) (2021) 206.
- [173] Y. Hou, et al., Metformin induces ferroptosis by targeting miR-324-3p/GPX4 axis in breast cancer, *Acta Biochim. Biophys. Sin.* 53 (3) (2021) 333–341.
- [174] L. Jing, et al., Exosomal miR-499a-5p inhibits endometrial cancer growth and metastasis via targeting VAV3, *Cancer Manag. Res.* 12 (2020) 13541–13552.
- [175] L. Zhao, et al., Downregulation of miR-499a-5p predicts a poor prognosis of patients with non-small cell lung cancer and Restrains the tumorigenesis by targeting fibroblast growth factor 9, *Technol. Cancer Res. Treat.* 19 (2020) 1533033820957001.
- [176] X. Gu, et al., MiR-499a-5p inhibits proliferation, invasion, migration, and epithelial-Mesenchymal transition, and enhances radiosensitivity of cervical cancer cells via targeting eIF4E, *OncoTargets Ther.* 13 (2020) 2913–2924.
- [177] J. Liu, et al., TMEM189 promotes breast cancer through inhibition of autophagy-regulated ferroptosis, *Biochem. Biophys. Res. Commun.* 622 (2022) 37–44.
- [178] D. Fan, et al., TMEM189 as a target gene of MiR-499a-5p regulates breast cancer progression through the ferroptosis pathway, *J. Clin. Biochem. Nutr.* 73 (2) (2023) 154–160.
- [179] W. Cui, et al., Peroxisome-driven ether-linked phospholipids biosynthesis is essential for ferroptosis, *Cell Death Differ.* 28 (8) (2021) 2536–2551.
- [180] E.R. Werner, et al., The TMEM189 gene encodes plasmalethanolamine desaturase which introduces the characteristic vinyl ether double bond into plasmalogens, *Proc Natl Acad Sci U S A* 117 (14) (2020) 7792–7798.
- [181] Q. Li, et al., A risk score model Incorporating three m6A RNA methylation regulators and a related network of miRNAs-m6A regulators-m6A target genes to predict the prognosis of patients with ovarian cancer, *Front. Cell Dev. Biol.* 9 (2021) 703969.
- [182] N. Bley, et al., IGF2BP1 is a targetable SRC/MAPK-dependent driver of invasive growth in ovarian cancer, *RNA Biol.* 18 (3) (2021) 391–403.
- [183] Y. Jin, et al., lncRNA CACNA1G-AS1 up-regulates FTH1 to inhibit ferroptosis and promote malignant phenotypes in ovarian cancer cells, *Oncol. Res.* 31 (2) (2023) 169–179.
- [184] L. Cai, et al., Long non-coding RNA ADAMTS9-AS1 attenuates ferroptosis by Targeting microRNA-587/solute carrier family 7 member 11 axis in epithelial ovarian cancer, *Bioengineered* 13 (4) (2022) 8226–8239.
- [185] P. Wang, et al., lncRNA ADAMTS9-AS1 inhibits the stemness of lung adenocarcinoma cells by regulating miR-5009-3p/NPNT axis, *Genomics* 115 (3) (2023) 110596.
- [186] A.R. Javanmard, et al., ADAMTS9-AS1 long non-coding RNA sponges miR-128 and miR-150 to regulate Ras/MAPK signaling pathway in glioma, *Cell. Mol. Neurobiol.* 43 (5) (2023) 2309–2322.
- [187] S. Fang, Y. Zhao, X. Hu, lncRNA ADAMTS9-AS1 Restrains the aggressive traits of breast carcinoma cells via sponging miR-513a-5p, *Cancer Manag. Res.* 12 (2020) 10693–10703.
- [188] M. Wang, et al., Long noncoding RNA LINC00336 inhibits ferroptosis in lung cancer by functioning as a competing endogenous RNA, *Cell Death Differ.* 26 (11) (2019) 2329–2343.
- [189] Z. Ai, et al., Overcoming cisplatin resistance of ovarian cancer cells by targeting HIF-1-regulated cancer metabolism, *Cancer Lett.* 373 (1) (2016) 36–44.
- [190] P.S. Macklin, et al., Hypoxia and HIF pathway in cancer and the placenta, *Placenta* 56 (2017) 8–13.
- [191] Q. Xu, et al., lncRNA TLR8-AS1 promotes metastasis and chemoresistance of ovarian cancer through enhancing TLR8 mRNA stability, *Biochem. Biophys. Res. Commun.* 526 (4) (2020) 857–864.
- [192] H. Gu, et al., ELK1 activated-long noncoding RNA LBX2-AS1 aggravates the progression of ovarian cancer through targeting miR-4784/KDM5C axis, *J. Mol. Histol.* 52 (1) (2021) 31–44.

- [193] J. Cao, et al., LBX2-AS1 promotes ovarian cancer progression by facilitating E2F2 gene expression via miR-455-5p and miR-491-5p sponging, *J. Cell Mol. Med.* 25 (2) (2021) 1178–1189.
- [194] S. Yang, et al., Construction of ovarian cancer prognostic model based on the investigation of ferroptosis-related lncRNA, *Biomolecules* 13 (2) (2023).
- [195] S. Feng, et al., Integrated clinical characteristics and omics analysis identifies a ferroptosis and iron-metabolism-related lncRNA signature for predicting prognosis and therapeutic responses in ovarian cancer, *J. Ovarian Res.* 15 (1) (2022) 10.
- [196] M.M. Gao, et al., Multi-label fusion collaborative matrix factorization for predicting lncRNA-disease associations, *IEEE J Biomed Health Inform* 25 (3) (2021) 881–890.
- [197] Y. Liu, et al., lncRNA TONSL-AS1 regulates miR-490-3p/CDK1 to affect ovarian epithelial carcinoma cell proliferation, *J. Ovarian Res.* 13 (1) (2020) 60.
- [198] Q. Yang, Y.J. Dong, lncRNA SNHG20 promotes migration and invasion of ovarian cancer via modulating the microRNA-148a/ROCK1 axis, *J. Ovarian Res.* 14 (1) (2021) 168.
- [199] Y. Zhao, et al., lncRNA-MSC-AS1 inhibits the ovarian cancer progression by targeting miR-425-5p, *J. Ovarian Res.* 14 (1) (2021) 109.
- [200] P.A. Cohen, et al., Cervical cancer, *Lancet* 393 (10167) (2019) 169–182.
- [201] Z. Jiang, et al., A ferroptosis-related lncRNA model to enhance the predicted value of cervical cancer, *J Oncol* 2022 (2022) 6080049.
- [202] W. Ma, et al., Immune-related lncRNAs as predictors of survival in breast cancer: a prognostic signature, *J. Transl. Med.* 18 (1) (2020) 442.
- [203] Z. Li, D. Wang, H. Yin, A seven immune-related lncRNA signature predicts the survival of patients with colon adenocarcinoma, *Am J Transl Res* 12 (11) (2020) 7060–7078.
- [204] R. Cristescu, et al., Pan-tumor genomic biomarkers for PD-1 checkpoint blockade-based immunotherapy, *Science* 362 (6411) (2018).
- [205] P. Li, et al., The role of ferroptosis-related molecules and significance of ferroptosis score in cervical cancer, *J Oncol* 2022 (2022) 7835698.
- [206] W. Qi, et al., lncRNA GABPB1-AS1 and GABPB1 regulate oxidative stress during erastin-induced ferroptosis in HepG2 hepatocellular carcinoma cells, *Sci. Rep.* 9 (1) (2019) 16185.
- [207] C. Mao, et al., A G3BP1-interacting lncRNA promotes ferroptosis and apoptosis in cancer via nuclear sequestration of p53, *Cancer Res.* 78 (13) (2018) 3484–3496.
- [208] Y. Yang, et al., lncRNA ZFAS1 promotes lung fibroblast-to-myofibroblast transition and ferroptosis via functioning as a ceRNA through miR-150-5p/SLC38A1 axis, *Aging (Albany NY)* 12 (10) (2020) 9085–9102.
- [209] L. Cui, et al., LINC01116 promotes proliferation and migration of endometrial stromal cells by targeting FOXP1 via sponging miR-9-5p in endometriosis, *J. Cell Mol. Med.* 25 (4) (2021) 2000–2012.
- [210] X. Wang, Q. Yu, Endometriosis-related ceRNA network to identify predictive biomarkers of endometrial receptivity, *Epigenomics* 11 (2) (2019) 147–167.
- [211] Y. Wan, et al., Long noncoding RNA ADAMTS9-AS1 represses ferroptosis of endometrial stromal cells by regulating the miR-6516-5p/GPX4 axis in endometriosis, *Sci. Rep.* 12 (1) (2022) 2618.
- [212] A. Qin, et al., Ferroptosis-related lncRNA model based on CFAP58-DT for predicting prognosis and immunocytes infiltration in endometrial cancer, *Ann. Transl. Med.* 11 (3) (2023) 151.
- [213] S. Wang, et al., RUNX1-IT1 favors breast cancer carcinogenesis through regulation of IGF2BP1/GPX4 axis, *Discov Oncol* 14 (1) (2023) 42.
- [214] X. Tong, et al., lncRNA HCP5-encoded protein regulates ferroptosis to promote the progression of triple-negative breast cancer, *Cancers* 15 (6) (2023).
- [215] J.K. Kulski, Long noncoding RNA HCP5, a Hybrid HLA class I endogenous Retroviral gene: structure, expression, and disease associations, *Cells* 8 (5) (2019).
- [216] Y. Zhu, et al., Long noncoding RNA linc00460 promotes breast cancer progression by regulating the miR-489-5p/FGF7/AKT axis, *Cancer Manag. Res.* 11 (2019) 5983–6001.
- [217] C. Zhang, et al., LINC00460 facilitates cell proliferation and inhibits ferroptosis in breast cancer through the miR-320a/MAL2 axis, *Technol. Cancer Res. Treat.* 22 (2023) 15330338231164359.
- [218] Z.Y. Yao, et al., Development and validation of ferroptosis-related lncRNAs as prognosis and diagnosis biomarkers for breast cancer, *BioMed Res. Int.* 2022 (2022) 2390764.
- [219] J. Lai, et al., Molecular characterization of breast cancer: a potential novel immune-related lncRNAs signature, *J. Transl. Med.* 18 (1) (2020) 416.
- [220] F. Wang, et al., Predictors of breast cancer cell types and their prognostic power in breast cancer patients, *BMC Genom.* 19 (1) (2018) 137.
- [221] P.O. Angrand, et al., The role of long non-coding RNAs in genome formatting and expression, *Front. Genet.* 6 (2015) 165.
- [222] L. Yan, et al., Regulation of tumor cell migration and invasion by the H19/let-7 axis is antagonized by metformin-induced DNA methylation, *Oncogene* 34 (23) (2015) 3076–3084.
- [223] J. Chen, et al., Metformin may induce ferroptosis by inhibiting autophagy via lncRNA H19 in breast cancer, *FEBS Open Bio* 12 (1) (2022) 146–153.
- [224] X. Fan, et al., lncFASA promotes cancer ferroptosis via modulating PRDX1 phase separation, *Sci. China Life Sci.* (2023) 1–16.
- [225] Y. Wang, Y. Xu, Y. Zhang, A novel ferroptosis-related long noncoding RNA signature for relapse free survival prediction in patients with breast cancer, *Medicine (Baltim.)* 101 (31) (2022) e29573.
- [226] N. Zhou, J. Bao, FerrDb: a manually curated resource for regulators and markers of ferroptosis and ferroptosis-disease associations, *Database* (2020) 2020.
- [227] X. Wang, et al., The long non-coding RNA CYTOR drives colorectal cancer progression by interacting with NCL and Sam68, *Mol. Cancer* 17 (1) (2018) 110.
- [228] M. Chen, et al., Long non-coding RNA USP30-AS1 aggravates the malignant progression of cervical cancer by sequestering microRNA-299-3p and thereby overexpressing PTP4A1, *Oncol. Lett.* 22 (1) (2021) 505.
- [229] R. Vishnubalaji, et al., Long non-coding RNA (lncRNA) transcriptional landscape in breast cancer identifies LINC01614 as non-favorable prognostic biomarker regulated by TGFβ and focal adhesion kinase (FAK) signaling, *Cell Death Discov* 5 (2019) 109.
- [230] X. Li, F. Jin, Y. Li, A novel autophagy-related lncRNA prognostic risk model for breast cancer, *J. Cell Mol. Med.* 25 (1) (2021) 4–14.
- [231] S. Shen, et al., A novel prognostic ferroptosis-related lncRNA signature associated with immune landscape in invasive breast cancer, *Dis. Markers* 2022 (2022) 9168556.
- [232] X. Fan, et al., lncFASA promotes cancer ferroptosis via modulating PRDX1 phase separation, *Sci. China Life Sci.* 67 (3) (2024) 488–503.
- [233] C. Xin, et al., Roles of circRNAs in cancer chemoresistance, *Oncol. Rep.* 46 (4) (2021) (Review).
- [234] K. Qin, et al., circRNA circSnx12 confers Cisplatin chemoresistance to ovarian cancer by inhibiting ferroptosis through a miR-194-5p/SLC7A11 axis, *BMB Rep* 56 (2) (2023) 184–189.
- [235] J. An, W. Lv, Y. Zhang, lncRNA NEAT1 contributes to paclitaxel resistance of ovarian cancer cells by regulating ZEB1 expression via miR-194, *OncoTargets Ther.* 10 (2017) 5377–5390.
- [236] K. Nakamura, et al., Downregulation of miR-194-5p induces paclitaxel resistance in ovarian cancer cells by altering MDM2 expression, *Oncotarget* 10 (6) (2019) 673–683.
- [237] W. Wei, N. Wang, L. Lin, Prognostic value of hsa_circ_0007615 in epithelial ovarian cancer and its regulatory effect on tumor progression, *Horm. Metab. Res.* (2023).
- [238] J. Zhu, et al., CircRNA-ACAP2 contributes to the invasion, migration, and anti-apoptosis of neuroblastoma cells through targeting the miRNA-143-3p-hexokinase 2 axis, *Transl. Pediatr.* 10 (12) (2021) 3237–3247.
- [239] Y. Liu, et al., Circular RNA circACAP2 suppresses ferroptosis of cervical cancer during malignant progression by miR-193a-5p/GPX4, *J Oncol* 2022 (2022) 5228874.
- [240] H.L. Nielsen, et al., Identification of EPSTI1, a novel gene induced by epithelial-stromal interaction in human breast cancer, *Genomics* 79 (5) (2002) 703–710.
- [241] B. Chen, et al., circEPSTI1 as a prognostic marker and mediator of triple-negative breast cancer progression, *Theranostics* 8 (14) (2018) 4003–4015.
- [242] P. Wu, et al., Circular RNA circEPSTI1 accelerates cervical cancer progression via miR-375/409-3p/515-5p/SLC7A11 axis, *Aging (Albany NY)* 13 (3) (2021) 4663–4673.
- [243] R. Ou, et al., Circular RNA circLMO1 suppresses cervical cancer growth and metastasis by triggering miR-4291/ACSL4-mediated ferroptosis, *Front. Oncol.* 12 (2022) 858598.
- [244] J. Zhang, et al., CircRAPGEF5 interacts with RBFOX2 to confer ferroptosis resistance by modulating alternative splicing of TFRC in endometrial cancer, *Redox Biol.* 57 (2022) 102493.
- [245] J. Yi, et al., Oncogenic activation of PI3K-AKT-mTOR signaling suppresses ferroptosis via SREBP-mediated lipogenesis, *Proc Natl Acad Sci U S A* 117 (49) (2020) 31189–31197.
- [246] J.L. Hecht, G.L. Mutter, Molecular and pathologic aspects of endometrial carcinogenesis, *J. Clin. Oncol.* 24 (29) (2006) 4783–4791.
- [247] F. Xing, et al., CMTM6 overexpression confers trastuzumab resistance in HER2-positive breast cancer, *Mol. Cancer* 22 (1) (2023) 6.
- [248] S. Wang, et al., A novel circular RNA confers trastuzumab resistance in human epidermal growth factor receptor 2-positive breast cancer through regulating ferroptosis, *Environ. Toxicol.* 37 (7) (2022) 1597–1607.
- [249] F.A. Elian, et al., FOXQ1 is differentially expressed across breast cancer subtypes with low expression associated with poor overall survival, *Breast Cancer* 13 (2021) 171–188.
- [250] X. Huang, et al., FOXQ1 inhibits breast cancer ferroptosis and progression via the circ_0000643/miR-153/SLC7A11 axis, *Exp. Cell Res.* 431 (1) (2023) 113737.
- [251] H.D. Zhang, et al., Circular RNA hsa_circ_0052112 promotes cell migration and invasion by acting as sponge for miR-125a-5p in breast cancer, *Biomed. Pharmacother.* 107 (2018) 1342–1353.
- [252] Y.Y. Tang, et al., Circular RNA hsa_circ_0001982 promotes breast cancer cell carcinogenesis through decreasing miR-143, *DNA Cell Biol.* 36 (11) (2017) 901–908.
- [253] H. Zhang, et al., Circular RNA RHOT1 promotes progression and inhibits ferroptosis via miR-106a-5p/STAT3 axis in breast cancer, *Aging (Albany NY)* 13 (6) (2021) 8115–8126.
- [254] M. Bazhabayi, et al., CircGFRA1 facilitates the malignant progression of HER-2-positive breast cancer via acting as a sponge of miR-1228 and enhancing AIFM2 expression, *J. Cell Mol. Med.* 25 (21) (2021) 10248–10256.
- [255] K. Shimada, et al., Global survey of cell death mechanisms reveals metabolic regulation of ferroptosis, *Nat. Chem. Biol.* 12 (7) (2016) 497–503.
- [256] S.J. Dixon, et al., Ferroptosis: an iron-dependent form of nonapoptotic cell death, *Cell* 149 (5) (2012) 1060–1072.
- [257] H.-Y. Lin, et al., The evolving role of ferroptosis in breast cancer: translational implications present and future, *Cancers* 13 (18) (2021) 4576.
- [258] J. Li, et al., Ferroptosis: past, present and future, *Cell Death Dis.* 11 (2) (2020) 88.
- [259] M.J. Hangauer, et al., Drug-tolerant persister cancer cells are vulnerable to GPX4 inhibition, *Nature* 551 (7679) (2017) 247–250.
- [260] V.S. Viswanathan, et al., Dependency of a therapy-resistant state of cancer cells on a lipid peroxidase pathway, *Nature* 547 (7664) (2017) 453–457.

- [261] M. Sato, et al., Loss of the cystine/glutamate antiporter in melanoma abrogates tumor metastasis and markedly increases survival rates of mice, *Int. J. Cancer* 147 (11) (2020) 3224–3235.
- [262] Y. Zhang, et al., Imidazole ketone erastin induces ferroptosis and slows tumor growth in a mouse lymphoma model, *Cell Chem. Biol.* 26 (5) (2019) 623–633. e9.
- [263] S. Doll, et al., FSP1 is a glutathione-independent ferroptosis suppressor, *Nature* 575 (7784) (2019) 693–698.
- [264] R. Gao, et al., FSP1-mediated ferroptosis in cancer: from mechanisms to therapeutic applications, *Apoptosis* (2024) 1–19.
- [265] S.-E. Yoo, et al., Gpx4 ablation in adult mice results in a lethal phenotype accompanied by neuronal loss in brain, *Free Radic. Biol. Med.* 52 (9) (2012) 1820–1827.
- [266] M. Conrad, D.A. Pratt, The chemical basis of ferroptosis, *Nat. Chem. Biol.* 15 (12) (2019) 1137–1147.
- [267] W.S. Yang, et al., Regulation of ferroptotic cancer cell death by GPX4, *Cell* 156 (1) (2014) 317–331.
- [268] M.-H. Larrauffie, et al., Incorporation of metabolically stable ketones into a small molecule probe to increase potency and water solubility, *Bioorg. Med. Chem. Lett* 25 (21) (2015) 4787–4792.
- [269] G.A. Calin, et al., MiR-15a and miR-16-1 cluster functions in human leukemia, *Proc. Natl. Acad. Sci. USA* 105 (13) (2008) 5166–5171.