MITOGENOME ANNOUNCEMENT

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The complete mitochondrial genome of a cryptic amphipod species from the *Gammarus fossarum* complex

Jan Niklas Macher^{a,b}, Florian Leese^{a,b}, Alexander Martin Weigand^{a,b} and Andrey Rozenberg^c

^aAquatic Ecosystem Research, Faculty of Biology, University of Duisburg-Essen, Universitätsstraße 5, Essen, Germany; ^bCentre for Water and Environmental Research, University of Duisburg-Essen, Universitätsstraße 2, Essen, Germany; ^cBeja's Lab, Faculty of Biology, Technion – Israel Institute of Technology, Haifa, Israel

ABSTRACT

The freshwater amphipod *Gammarus fossarum* is widely distributed throughout Europe and an important species for stream biomonitoring. It is known to consist of several cryptic species. We here report the complete mitochondrial genome of *G. fossarum* clade 11/type B with a length of 15,989 bp, encoding for 13 protein-coding genes, 22 tRNA genes, and 2 rRNA genes. Protein-coding and ribosomal genes have a similar arrangement as in other gammarid amphipods. A phylogenetic analysis clarifies the placement of *G. fossarum* within the Gammaridae.

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Amphipod crustaceans comprise a diverse taxon whose taxonomy is intensively studied (e.g. Myers & Lowry 2003; Hou & Sket 2016). The genus *Gammarus* is especially speciose and species are essential components of freshwater ecosystems, often forming important trophic links (MacNeil et al. 1997). Molecular studies revealed the common European *Gammarus fossarum* to be a species complex (Weiss et al. 2014), a widespread phenomenon in *Gammarus* (e.g. Mamos et al. 2016; Katouzian et al. 2016). Mitogenomes can help resolving amphipod phylogeny and diversity. Furthermore, mitogenomes are essential for future biomonitoring programmes using next-generation molecular tools (Crampton-Platt et al. 2016).

The sequenced specimen was collected in the Grundbächle (47.83N/7.93E) and belongs to G. fossarum clade 11/type B (Weiss & Leese 2016). It is stored in the UDE collection (accession number Gf_BF_S2). DNA was extracted as in Weiss and Leese (2016). A standard shotgun genomic library was sequenced on HiSeq 2000 (Otogenetics, USA), with 84,504,354 pairs of 101-nt reads obtained. Raw reads were trimmed with trimmomatic (v.0.22, Bolger et al. 2014) and mitochondrial reads were sampled with a blastn search against amphipod mitogenomes. The resulting data were assembled using MIRA (v.4.0, Chevreux 2005) and SOAPdenovo (v.2.04, Luo et al. 2012). Read re-mapping with bowtie2 (v.2.2.8, Langmead & Salzberg 2012) yielded 9095 read hits with a per-base coverage of 56.3. All three methods produced the same consensus sequence. Genes were annotated with MITOS WebServer (Bernt et al. 2013), and gene boundaries were refined manually. The complete circular mitogenome was 15,989 bp long (33.2% A, 22.0% C, 12.9% G, and 32.0% T) encompassing 13 protein-coding genes, 22 tRNA genes, 2 rRNA genes, and a \sim 1500 bp AT-rich control region. The gene order appeared identical to that of *G. duebeni* (Krebes & Bastrop 2012).

For phylogenetic reconstruction, in addition to G. fossarum nine closely related species and two outgroups were taken. The mitochondrial genes were obtained from annotated mitochondrial genomes (NCBI GenBank), unannotated genome or transcriptome assemblies (NCBI WGS and TSA) and from raw transcriptome data assembled anew (NCBI SRA) (Figure 1). The dataset included 13 protein-coding genes (fulllength except two cases), for each of which translation alignments were obtained with MAFFT (v.7.017, Katoh & Standley 2013). After manual trimming and concatenation, the resulting DNA alignment encompassed 10,863 nucleotides corresponding to 3621 amino acids. Four phylogenetic trees were produced using RAxML (v.8.2.9, Stamatakis 2014) and MrBayes (v.3.2.6, Ronquist & Huelsenbeck 2003) based on DNA and amino acid alignments with substitution models and partition schemes chosen with PartitionFinder (v.2.0.0, Lanfear et al. 2016).

Gammarus fossarum clade 11/B forms a monophylum with other *Gammarus* species, rendering the genus monophyletic, albeit without a strong branch support (Figure 1). This stands in contrast to the findings by Hou et al. (2014). The Gammaridae as a whole are nevertheless paraphyletic in relation to the other related families, supporting the broader concept of the family proposed by Hou & Sket (2016).

CONTACT Andrey Rozenberg 🖾 jaera@yandex.com 🖻 Beja's Lab, Faculty of Biology, Technion – Israel Institute of Technology, Haifa, Israel

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Figure 1. The Bayesian inference phylogenetic tree based on nucleotide alignment of 13 mitochondrial protein-coding genes for *Gammarus fossarum* (clade 11/type B) and 11 other amphipods. Both the current taxonomic status and the opinion of Hou and Sket (2016) are shown.

The mitochondrial genome of *G. fossarum* clade 11/B was deposited in GenBank under the accession number KY197961. Extended methods description and sequence data are available at figshare (DOI:10.6084/m9.figshare.4487912).

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Disclosure statement

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References

- Bernt M, Donath A, Jühling F, Externbrink F, Florentz C, Fritzsch G, Pütz J, Middendorf M, Stadler PF. 2013. MITOS: improved de novo metazoan mitochondrial genome annotation. Mol Phylogen Evol. 69:313–319.
- Bolger AM, Lohse M, Usadel B. 2014. Trimmomatic: a flexible trimmer for Illumina sequence data. Bioinformatics. 30:2114–2120.
- Chevreux B. 2005. MIRA: An automated genome and EST assembler [Ph.D. Thesis]. Heidelberg: German Cancer Research Center.
- Crampton-Platt A, Douglas WY, Zhou X, Vogler AP. 2016. Mitochondrial metagenomics: letting the genes out of the bottle. GigaScience. 5:1.
- Hou Z, Sket B, Li S. 2014. Phylogenetic analyses of Gammaridae crustacean reveal different diversification patterns among sister lineages in the Tethyan region. Cladistics. 30:352–365.
- Hou Z, Sket B. 2016. A review of Gammaridae (Crustacea: Amphipoda): the family extent, its evolutionary history, and taxonomic redefinition of genera. Zool J Linnean Soc. 176:323–348.
- Katoh K, Standley DM. 2013. MAFFT multiple sequence alignment software version 7: improvements in performance and usability. Mol Biol Evol. 30:772–780.

- Katouzian AR, Sari A, Macher JN, Weiss M, Saboori A, Leese F, Weigand AM. 2016. Drastic underestimation of amphipod biodiversity in the endangered Irano-Anatolian and Caucasus biodiversity hotspots. Sci Rep. 6:22507.
- Krebes L, Bastrop R. 2012. The mitogenome of *Gammarus duebeni* (Crustacea Amphipoda): a new gene order and non-neutral sequence evolution of tandem repeats in the control region. Comp Biochem Physiol Part D: Genomics Proteomics. 7:201–211.
- Lanfear R, Frandsen PB, Wright AM, Senfeld T, Calcott B. 2016. PartitionFinder 2: new methods for selecting partitioned models of evolution for molecular and morphological phylogenetic analyses. Mol Biol Evol. in press. DOI:10.1093/molbev/msw260.
- Langmead B, Salzberg SL. 2012. Fast gapped-read alignment with Bowtie 2. Nature Methods. 9:357–359.
- Luo R, Liu B, Xie Y, Li Z, Huang W, Yuan J, He G, Chen Y, Pan Q, Liu Y. 2012. SOAPdenovo2: an empirically improved memory-efficient short-read de novo assembler. GigaScience. 1:1.
- MacNeil C, Dick JT, Elwood RW. 1997. The trophic ecology of freshwater *Gammarus* spp. (Crustacea: Amphipoda): problems and perspectives concerning the functional feeding group concept. Biol Rev Cambridge Phil Soc. 72:349–364.
- Mamos T, Wattier R, Burzyński A, Grabowski M. 2016. The legacy of a vanished sea: a high level of diversification within a European freshwater amphipod species complex driven by 15 My of Paratethys regression. Mol Ecol. 25:795–810.
- Myers AA, Lowry JK. 2003. A phylogeny and a new classification of the Corophiidea Leach, 1814 (Amphipoda). J Crustacean Biol. 23:443–485.
- Ronquist F, Huelsenbeck JP. 2003. MRBAYES 3: Bayesian phylogenetic inference under mixed models. Bioinformatics. 19:1572–1574.
- Stamatakis A. 2014. RAxML version 8: a tool for phylogenetic analysis and post-analysis of large phylogenies. Bioinformatics. 30:1312–1313.
- Weiss M, Leese F. 2016. Widely distributed and regionally isolated! Drivers of genetic structure in *Gammarus fossarum* in a humanimpacted landscape. BMC Evol Biol. 16:153.
- Weiss M, Macher JN, Seefeldt MA, Leese F. 2014. Molecular evidence for further overlooked species within the *Gammarus fossarum* complex (Crustacea: Amphipoda). Hydrobiologia. 721:165–184.