GENOME SEQUENCES





Draft Genome Sequence of Serratia sp. 1D1416

Diaa Alabed,^a Naxin Huo,^{a,b} Yong Gu,^a Kent F. McCue,^a James G. Thomson^a

^aUSDA-ARS Crop Improvement and Genetics, Western Regional Research Center, Albany, California, USA ^bDepartment of Plant Sciences, University of California, Davis, California, USA

ABSTRACT This work reports the draft genome of *Serratia* sp. 1D1416. The assembled genome contains a 5,552,016-bp circular chromosome. The strain was discovered in a mixed culture from a gall isolated from *Euonymus japonicas*.

A lthough strains of *Serratia* are often associated with human infections, such as *Serratia marcescens* and the *Serratia liquefaciens* complex, other *Serratia* spp. ("unusual" *serratiae*) are isolated from the environment (1, 2). Published unusual *Serratia* spp. include *Serratia ficaria, Serratia fonticola, Serratia odorifera, Serratia plymuthica*, and *Serratia rubidaea* (3). *Serratia ficaria* was the first nonpathogen microbe associated with plant growth and was characterized in 1979 as an important part of the fig tree ecosystem (4). *Serratia plymuthica, S. liquefaciens*, and *S. rubidaea* have been observed as part of the rhizosphere microbiome (5, 6) and may have antibacterial or antifungal properties or both (7). The recently isolated strain *Serratia* sp. 1D1416 appears to have low-level resistance to tetracycline but is susceptible to chloramphenicol, kanamycin, and gentamicin. This is a typical feature of *Serratia* spp. and may include the entire genus *Serratia* (3). Here we present the novel genome from *Serratia* sp. 1D1416, isolated from a mixed culture along with an *Agrobacterium tumefaciens* isolated from a *Euonymus japonicus* (Japanese spindle) gall within the UC Davis microbe collection of Dr. Kobe. The strain was grown in Luria broth at 30°C with shaking at 200 rpm.

Genomic DNA was isolated from our strains (8) using the Qiagen blood and cell culture DNA Maxi kit (number 13362) and genomic DNA buffer set (number 19060) (9). DNA samples were evaluated with gel electrophoresis and quantified with a 2100 Nanodrop spectrophotometer (Thermo Fisher Scientific) and a Qubit fluorimeter (Invitrogen) with the Qubit dsDNA HS assay kit (Invitrogen). The genomic DNA was sheared with g-TUBE (Covaris). A 20-kb DNA library was constructed according to the manufacturer's instructions and the BluePippin size selection system, and it was sequenced using singlemolecule real-time (SMRT) sequencing technology on a PacBio RS system. SMRT sequencing data were generated at an average coverage of $88.53 \times$, with a mean read length of 16,608 bp. De novo genome assembly was conducted with the sequence reads using the Hierarchical Genome Assembly Process (HGAP) workflow (SMRT Portal; Pacific Biosciences), protocol RS_HGAP_Assembly.3 (10), and SMRTAnalysis_2.3.0 software (https://www.pacb.com/wp-content/uploads/2015/09/SMRT-Analysis-Software-Installation-v2.3.0.pdf). This allowed the generation of a single contig with an N_{50} contig length of 5,577,755 bp. The linear DNA was manually circularized with a 25,739-bp chimeric overlap. The circular chromosome is composed of 5,552,016 bp, with a GC content of 59.6%. Assembled and raw read sequences were entered into the National Center for Biotechnology Information (NCBI), and BLAST was used for identification (http://blast.ncbi.nlm.nih.gov/). During genome comparison with other published Serratia, a unique cluster of opine genes were detected. We used this gene cluster to develop primers as a marker to identify specifically the Serratia sp. 1D1416 genome. Primers STNop 5304 F61 5' ggtgctgaaaagataatgaatccgcacagg and STNop 5304 R61 5' cctggttggaaatgaagaaattcaggtaagcactg were used to produce a unique 1,560-bp am-

Citation Alabed D, Huo N, Gu Y, McCue KF, Thomson JG. 2019. Draft genome sequence of *Serratia* sp. 1D1416. Microbiol Resour Announc 8:e01354-18. https://doi.org/10.1128/MRA .01354-18.

Editor Julie C. Dunning Hotopp, University of Maryland School of Medicine

This is a work of the U.S. Government and is not subject to copyright protection in the United States. Foreign copyrights may apply.

Address correspondence to James G. Thomson, James.Thomson@ars.usda.gov.

Received 22 October 2018 Accepted 14 December 2018 Published 17 January 2019 plicon from a unique ABC transporter substrate-binding protein gene. Automated annotation was performed with the Rapid Annotation Using Subsystem Technology (RAST) Pipeline for annotation of the genome (11). *Serratia* sp. 1D1416 contains 5,234 predicted coding sequences, 570 subsystems, and 118 predicted RNA-coding genes.

Data availability. The whole-genome assembly for *Serratia* sp. 1D1416 has been deposited in DDBJ/ENA/GenBank under the accession number CP032738, BioProject accession number PRJNA493633, and SRA accession number SRX5036357.

ACKNOWLEDGMENTS

This work was supported by the USDA Agricultural Research Service CRIS project 2030-21000-020. Mention of trade names or commercial products is solely for the purpose of providing specific information and does not imply recommendation or endorsement by the U.S. Department of Agriculture. The USDA is an equal opportunity provider and employer.

REFERENCES

- Singh BR, Singh Y, Tiwari AK. 1997. Characterization of virulence factors of *Serratia* strains isolated from foods. Intl J Food Microbiol 34:259–266. https://doi.org/10.1016/S0168-1605(96)01196-8.
- Carrero P, Garrote JA, Pacheco S, Garcia AI, Gil R, Carbajosa SG. 1995. Report of six cases of human infection by *Serratia plymuthica*. J Clin Microbiol 33:275–276.
- Stock I, Burak S, Sherwood KJ, Grüger T, Wiedemann B. 2003. Natural antimicrobial susceptibilities of strains of "unusual" Serratia species: S. ficaria, S. fonticola, S. odorifera, S. plymuthica and S. rubidaea. J Antimicrob Chemother 51:865–885. https://doi.org/10.1093/jac/dkg156.
- Grimont PAD, Grimont F, Starr MP. 1979. Serratia ficaria sp. nov., a bacterial species associated with Smyrna figs and the fig wasp Blastophaga psenes. Curr Microbiol 2:277–282. https://doi.org/10.1007/ BF02602859.
- Alström S, Gerhardson B. 1987. Characteristics of a Serratia plymuthica isolated from plant rhizospheres. Plant Soil 103:185–189. https://doi.org/ 10.1007/BF02370387.
- Berg G. 2000. Diversity of antifungal and plant-associated Serratia plymuthica strains. J Appl Microbiol 88:952–960. https://doi.org/10.1046/j .1365-2672.2000.01064.x.

- Kalbe C, Marten P, Berg G. 1996. Strains of the genus Serratia as beneficial rhizobacteria of oilseed rape with antifungal properties. Microbiol Res 151: 433–439. https://doi.org/10.1016/S0944-5013(96)80014-0.
- Wise AA, Liu Z, Binns AN. 2006. Nucleic acid extraction from Agrobacterium strains, p 67–76. In Wang K (ed), Agrobacterium protocols, 2nd ed, vol1. Humana Press, Totowa, NJ. https://doi.org/10.1385/1-59745-130 -4:67.
- 9. Qiagen. CLC genomics workbench 8.5. Qiagen, Redwood City, CA. https://www.qiagenbioinformatics.com/.
- Chin C-S, Alexander DH, Marks P, Klammer AA, Drake J, Heiner C, Clum A, Copeland A, Huddleston J, Eichler EE, Turner SW, Korlach J. 2013. Nonhybrid, finished microbial genome assemblies from long-read SMRT sequencing data. Nat Methods 10:563–569. https://doi.org/10.1038/ nmeth.2474.
- Brettin T, Davis JJ, Disz T, Edwards RA, Gerdes S, Olsen GJ, Olson R, Overbeek R, Parrello B, Pusch GD, Shukla M, Thomason JA, III, Stevens R, Vonstein V, Wattam AR, Xia F. 2015. RASTtk: a modular and extensible implementation of the RAST algorithm for building custom annotation pipelines and annotating batches of genomes. Sci Rep 5:8365. https:// doi.org/10.1038/srep08365.