



Structural elucidation of a hydroxy–cineole product obtained from cytochrome P450 monooxygenase CYP101J2 catalysed transformation of 1,8-cineole

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1,8-Cineole is an abundant natural product that has the potential to be transformed into other building blocks that could be suitable alternatives to petroleum-based chemicals. Monohydroxylation of 1,8-cineole can potentially occur at eight different carbon sites around the bicyclic ring system. Using cytochrome P450 monooxygenase CYP101J2 from *Sphingobium yanoikuyae* B2, the hydroxylation can be regioselectively directed at the C atom adjacent to the methyl-substituted quaternary bridgehead atom of 1,8-cineole. The unambiguous location of the hydroxyl functionality and the stereochemistry at this position was determined by X-ray crystal analysis. The monohydroxylated compound derived from this microorganism was determined to be (1*S*)-2*a*-hydroxy-1,8-cineole (trivial name) or (1*S*,4*R*,6*S*)-1,3,3-trimethyl-2-oxabicyclo[2.2.2]octan-6-ol (V) (systematic), C₁₀H₁₈O₂. In the solid state this compound exhibits an interesting O–H···O hydrogen-bonding motif.

1. Chemical context

The terpenoid compound commonly known as 1,8-cineole, or less easily identified using systematic nomenclature as 1,3,3-trimethyl-2-oxabicyclo[2.2.2]octane (I) (Fig. 1), is a key component of the leaf oil from eucalypts and is also found in a variety of plant types, such as sage, thyme and fruit extracts, albeit in lower quantities (Fig. 1). Its natural abundance makes it a suitable bio-derived feedstock from which other useful chemical building blocks could be accessed and used as an alternative to petrochemical based-materials. Although continued research into the chemical and biochemical trans-

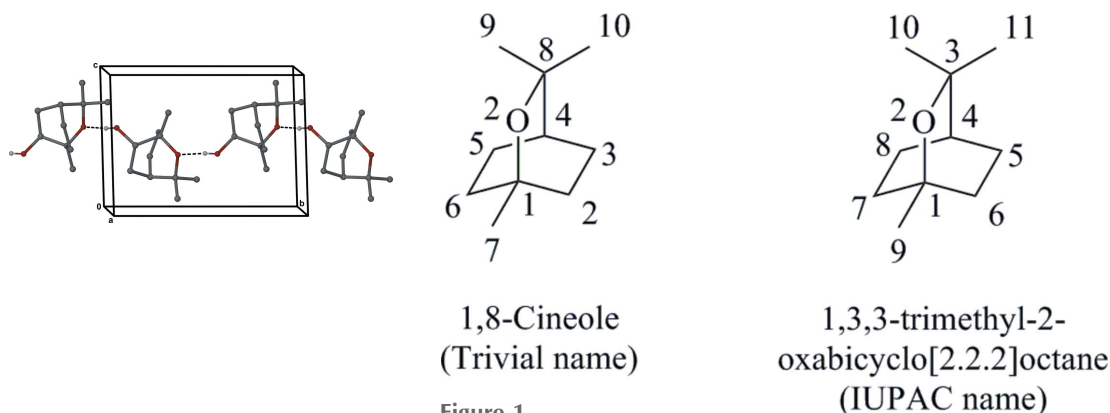


Figure 1
Trivial and systematic naming and atom numbering used for compound (I).

formation of 1,8-cineole (I) is being directed towards accessing high quality and commercial quantities of these derivatives, the naming of these products by using non-systematic nomenclature, coupled with the chiral nature of these products has created inconsistencies and made it challenging to compare data of these derivatives in the literature. To address this Azerad (2014) recently published an extremely useful review article capturing all the oxidation products of 1,8-cineole (I) by providing trivial and systematic names along with characterization data (i.e. melting point, optical rotation and proton and carbon NMR spectroscopic information).

In continuing our research activities on the biocatalytic mono-hydroxylation of 1,8-cineole (I) at the C atom adjacent to the quaternary C1 bridgehead atom (i.e. labelled 6 or 7 following IUPAC rules) four possible stereoisomers [Fig. 2, compounds (II), (III), (IV) and (V)] could be formed. However, there is no current crystallographic information of these pure materials to support these assignments. Knowing the inconsistencies with the nomenclature of these compounds and to gain a better understanding of how to control the regio-

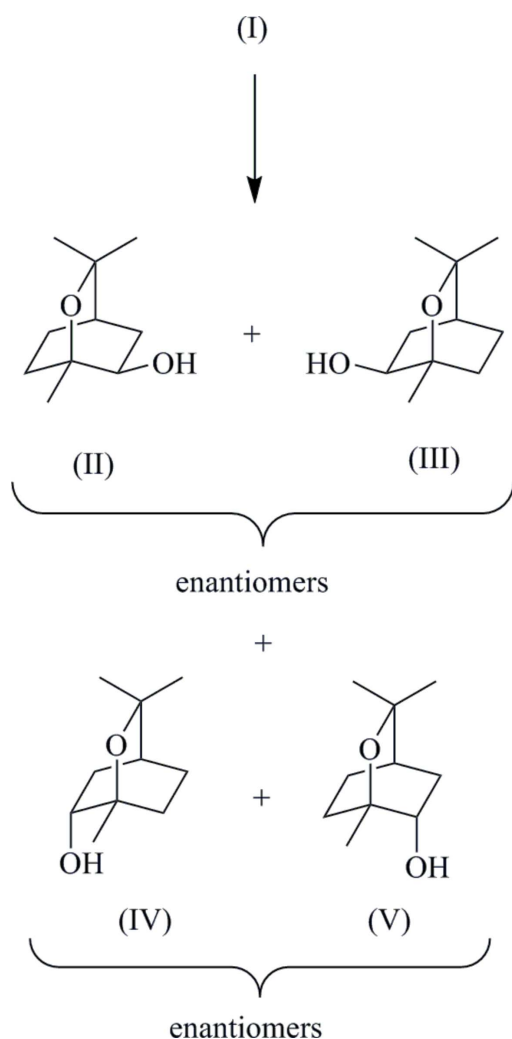
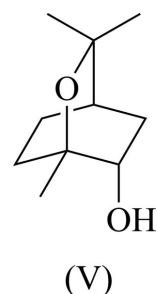


Figure 2
Biotransformation of 1,8-cineole (I) by *S. yanoikuyae* B2 to produce four possible isomeric mono-hydroxylated products (Unterweger *et al.*, 2016).

and stereo-chemistry at the different sites around the 1,8-cineole bicyclic ring system, we sought confirmation of the absolute configuration by undertaking X-ray crystallographic studies.

2. Structural commentary

Suitable crystals for X-ray diffraction were prepared by the slow diffusion of petroleum ether into a solution of the compound dissolved in ethyl acetate. The X-ray crystal structure of the purified mono-hydroxylated 1,8-cineole (V) (Fig. 3) was solved in the $P2_1$ space group and revealed the location of the hydroxyl group to be in the 6 position (IUPAC) (Fig. 1). The absolute configuration was determined by the method of Parsons *et al.* (2013) and confirmed the proposed stereochemistry (i.e. structure (V) see above, Fig. 2).



The presence of the axial hydroxyl substituent in (V) breaks the crystallographic symmetry of the parent 1,8-cineole (I), with $P2_1/m$ space group (Bond & Davies, 2001), resulting in a slight twisting of the molecular framework as shown by the torsion angle C1–O2–C7–C4 of $-12.8(2)^\circ$ and is presumably steric in origin. For the related 1,8-cineole-5,6-diol, three

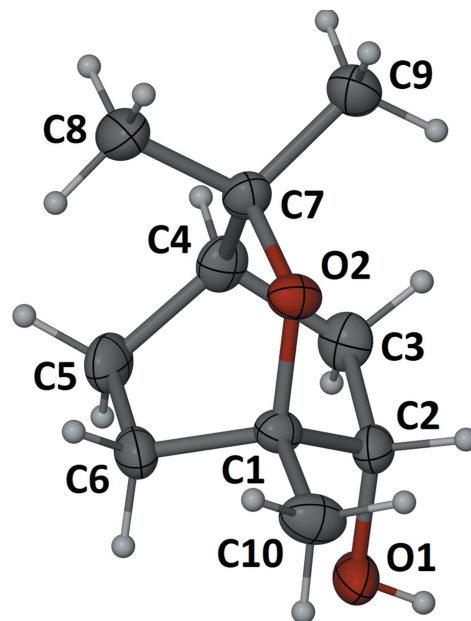


Figure 3
Molecular structure of (1*S*,4*R*,6*S*)-1,3,3-trimethyl-2-oxabicyclo[2.2.2]-octan-6-ol (V) with non-H atoms represented by 50% displacement ellipsoids and H atoms as spheres of arbitrary size.

Table 1
Hydrogen-bond geometry (Å, °).

$D-H\cdots A$	$D-H$	$H\cdots A$	$D\cdots A$	$D-H\cdots A$
$O1-H1\cdots O2^i$	0.80 (3)	1.97 (3)	2.7530 (19)	170 (3)

Symmetry code: (i) $-x + 1, y - \frac{1}{2}, -z + 1$.

of the four possible diastereoisomers have been structurally characterized and only the one with the 6α hydroxyl group showed a similar distortion (Farlow *et al.*, 2013).

3. Supramolecular features

Individual molecules of (V) are connected by $O-H\cdots O$ hydrogen bonds between the hydroxyl and ether moieties (Table 1) and form spiral chains parallel to the b axis (Fig. 4).

4. Database survey

A search of the Cambridge Structural Database (V5.38; Groom *et al.*, 2016) for the 1,3,3-trimethyl-2-oxabicyclo[2.2.2]octane (cineole) skeleton gave the parent structure (I) (ref code MOFPAY; Bond & Davies, 2001) and the oxidation products, 5,6-dihydroxycineole (three stereoisomers: ref codes DIFJAF, DIFJEJ and DIFJIN; Farlow *et al.*, 2013), 6-(1,3-dioxolan-2-yl)-5-ketocineole and 5-(1,3-dioxolan-2-yl)-6-ketocineole (ref codes DIFHOR and DIFHUX; Farlow *et al.*, 2013).

5. Synthesis and crystallization

1,8-Cineole (I) was mono-hydroxylated using a recombinant *Escherichia coli* whole-cell fed-batch process using CYP101J2 in combination with suitable redox partner proteins from *S. yanoikuyae* B2 to provide a major product (Unterweger, 2016). The isolated material was further purified by recrystallization from diethyl ether/petroleum ether to afford white needles. The melting point (this work m.p. 371.2–371.8 K, lit. m.p. 371–372 K (Carman *et al.*, 1986), 370, 370, 369, 368, 371–372, 371–372, 371–372 369–372, 372 and 370 K as cited in Azerad (2014)) and 1H NMR spectrum are in agreement with

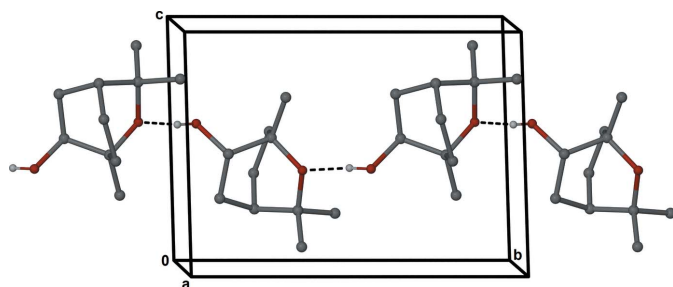


Figure 4
Ball-and-stick representation of a hydrogen-bonded chain of molecules of (V). Only selected H atoms are shown and $O-H\cdots O$ contacts are indicated as dashed bonds.

Table 2
Experimental details.

Crystal data	
Chemical formula	$C_{10}H_{18}O_2$
M_r	170.24
Crystal system, space group	Monoclinic, $P2_1$
Temperature (K)	123
a, b, c (Å)	6.3121 (1), 10.5611 (2), 7.9925 (2)
β (°)	112.126 (3)
V (Å ³)	493.57 (2)
Z	2
Radiation type	Cu $K\alpha$
μ (mm ⁻¹)	0.62
Crystal size (mm)	0.25 × 0.10 × 0.02
Data collection	
Diffractometer	Oxford Gemini Ultra CCD
Absorption correction	Multi-scan (CrysAlis PRO; Rigaku OD, 2015)
T_{min}, T_{max}	0.650, 1.000
No. of measured, independent and observed [$I > 2\sigma(I)$] reflections	6839, 1746, 1728
R_{int}	0.027
$(\sin \theta/\lambda)_{max}$ (Å ⁻¹)	0.596
Refinement	
$R[F^2 > 2\sigma(F^2)], wR(F^2), S$	0.029, 0.076, 1.05
No. of reflections	1746
No. of parameters	113
No. of restraints	1
H-atom treatment	H atoms treated by a mixture of independent and constrained refinement
$\Delta\rho_{max}, \Delta\rho_{min}$ (e Å ⁻³)	0.13, -0.11
Absolute structure	Flack x determined using 804 quotients $[(I^+) - (I^-)] / [(I^+) + (I^-)]$ (Parsons <i>et al.</i> , 2013)
Absolute structure parameter	0.07 (9)

Computer programs: CrysAlis PRO (Rigaku OD, 2015), SHELXS97 (Sheldrick, 2015), SHELXL2016 (Sheldrick, 2015), X-SEED (Barbour, 2001) and publCIF (Westrip, 2010).

cited literature values (Azerad, 2014) for either compound (IV) and/or (V). Optical rotation {this work $[a]_D +32.0$ (c 1.3, EtOH), lit $[a]_D +31.9$ (c 1.3, EtOH)}. The experimental data for the current material produced from the biotransformation of cineole is well aligned with one set of literature data (Carman *et al.*, 1986).

6. Refinement

Crystal data, data collection and structure refinement details are summarized in Table 2. H atoms potentially involved in hydrogen-bonding interactions were located by difference methods and were freely refined. Other H atoms were included in the refinement at calculated positions with $C-H = 0.95-0.98$ Å and treated as riding with $U_{iso}(H) = 1.2U_{eq}(C)$ or $1.52U_{eq}(O$ or methyl C).

Acknowledgements

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References

- Azerad, R. (2014). *ChemPlusChem* **79**, 634–655.
- Barbour, L. J. (2001). *J. Supramol. Chem.* **1**, 189–191.
- Bond, A. D. & Davies, J. E. (2001). *Aust. J. Chem.* **54**, 683–684.
- Carman, R. M., Macrae, I. C. & Perkins, M. V. (1986). *Aust. J. Chem.* **39**, 1739–1746.
- Farlow, A. J., Bernhardt, P. V. & De Voss, J. J. (2013). *Tetrahedron Asymmetry*, **24**, 324–333.
- Groom, C. R., Bruno, I. J., Lightfoot, M. P. & Ward, S. C. (2016). *Acta Cryst.* **B72**, 171–179.
- Parsons, S., Flack, H. D. & Wagner, T. (2013). *Acta Cryst.* **B69**, 249–259.
- Rigaku OD (2015). *CrysAlis PRO*. Rigaku Oxford Diffraction, Yarnton, England.
- Sheldrick, G. M. (2015). *Acta Cryst.* **C71**, 3–8.
- Unterweger, B. (2016). PhD thesis, Monash University, Victoria, Australia.
- Unterweger, B., Bulach, D. M., Scoble, J., Midgley, D. J., Greenfield, P., Lyras, D., Johanesen, P. & Dumsday, G. J. (2016). *Appl. Environ. Microbiol.* **82**, 6507–6517.
- Westrip, S. P. (2010). *J. Appl. Cryst.* **43**, 920–925.

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Computing details

Data collection: *CrysAlis PRO* (Rigaku OD, 2015); cell refinement: *CrysAlis PRO* (Rigaku OD, 2015); data reduction: *CrysAlis PRO* (Rigaku OD, 2015); program(s) used to solve structure: *SHELXS97* (Sheldrick, 2015); program(s) used to refine structure: *SHELXL2016* (Sheldrick, 2015); molecular graphics: *X-SEED* (Barbour, 2001); software used to prepare material for publication: *publCIF* (Westrip, 2010).

(1S,4R,6S)-1,3,3-Trimethyl-2-oxabicyclo[2.2.2]octan-6-ol

Crystal data

$C_{10}H_{18}O_2$	$F(000) = 188$
$M_r = 170.24$	$D_x = 1.146 \text{ Mg m}^{-3}$
Monoclinic, $P2_1$	Cu $K\alpha$ radiation, $\lambda = 1.54184 \text{ \AA}$
$a = 6.3121 (1) \text{ \AA}$	Cell parameters from 4695 reflections
$b = 10.5611 (2) \text{ \AA}$	$\theta = 7.6\text{--}66.8^\circ$
$c = 7.9925 (2) \text{ \AA}$	$\mu = 0.62 \text{ mm}^{-1}$
$\beta = 112.126 (3)^\circ$	$T = 123 \text{ K}$
$V = 493.57 (2) \text{ \AA}^3$	Plate, colourless
$Z = 2$	$0.25 \times 0.10 \times 0.02 \text{ mm}$

Data collection

Oxford Gemini Ultra CCD diffractometer	6839 measured reflections
Radiation source: fine focus sealed tube	1746 independent reflections
Mirror monochromator	1728 reflections with $I > 2\sigma(I)$
Detector resolution: $10.3389 \text{ pixels mm}^{-1}$	$R_{\text{int}} = 0.027$
ω scans	$\theta_{\text{max}} = 66.7^\circ$, $\theta_{\text{min}} = 7.6^\circ$
Absorption correction: multi-scan (CrysAlis PRO; Rigaku OD, 2015)	$h = -7 \rightarrow 7$
$T_{\text{min}} = 0.650$, $T_{\text{max}} = 1.000$	$k = -12 \rightarrow 12$
	$l = -9 \rightarrow 9$

Refinement

Refinement on F^2	H atoms treated by a mixture of independent and constrained refinement
Least-squares matrix: full	$w = 1/[\sigma^2(F_o^2) + (0.0408P)^2 + 0.077P]$
$R[F^2 > 2\sigma(F^2)] = 0.029$	where $P = (F_o^2 + 2F_c^2)/3$
$wR(F^2) = 0.076$	$(\Delta/\sigma)_{\text{max}} < 0.001$
$S = 1.05$	$\Delta\rho_{\text{max}} = 0.13 \text{ e \AA}^{-3}$
1746 reflections	$\Delta\rho_{\text{min}} = -0.11 \text{ e \AA}^{-3}$
113 parameters	
1 restraint	
Hydrogen site location: mixed	

Absolute structure: Flack x determined using
804 quotients $[(F^-)-(F)]/[(F^+)+(F)]$ (Parsons *et al.*,
2013)
Absolute structure parameter: 0.07 (9)

Special details

Geometry. All esds (except the esd in the dihedral angle between two l.s. planes) are estimated using the full covariance matrix. The cell esds are taken into account individually in the estimation of esds in distances, angles and torsion angles; correlations between esds in cell parameters are only used when they are defined by crystal symmetry. An approximate (isotropic) treatment of cell esds is used for estimating esds involving l.s. planes.

Fractional atomic coordinates and isotropic or equivalent isotropic displacement parameters (\AA^2)

	x	y	z	$U_{\text{iso}}^*/U_{\text{eq}}$
O1	0.3440 (2)	0.06028 (14)	0.5824 (2)	0.0344 (4)
H1	0.421 (5)	0.000 (3)	0.584 (4)	0.049 (8)*
C1	0.3290 (3)	0.28323 (16)	0.5241 (2)	0.0245 (4)
O2	0.3425 (2)	0.37248 (12)	0.39090 (15)	0.0273 (3)
C2	0.3671 (3)	0.15220 (17)	0.4598 (2)	0.0268 (4)
H2	0.525368	0.147597	0.459797	0.032*
C3	0.1930 (3)	0.13171 (17)	0.2663 (3)	0.0327 (5)
H3A	0.273802	0.126032	0.181704	0.039*
H3B	0.109504	0.051339	0.259698	0.039*
C4	0.0233 (3)	0.24262 (19)	0.2122 (3)	0.0311 (4)
H4	-0.095250	0.227897	0.088837	0.037*
C5	-0.0898 (3)	0.2491 (2)	0.3515 (3)	0.0357 (5)
H5A	-0.152629	0.165173	0.363326	0.043*
H5B	-0.217014	0.310951	0.312108	0.043*
C6	0.0917 (3)	0.28989 (18)	0.5342 (3)	0.0299 (4)
H6A	0.060505	0.377427	0.562909	0.036*
H6B	0.085833	0.233283	0.631090	0.036*
C7	0.1531 (3)	0.36548 (19)	0.2146 (2)	0.0277 (4)
C8	0.0064 (4)	0.4834 (2)	0.1933 (3)	0.0394 (5)
H8A	-0.121668	0.480975	0.076148	0.059*
H8B	-0.052847	0.486220	0.290291	0.059*
H8C	0.099284	0.558972	0.199665	0.059*
C9	0.2605 (3)	0.3681 (2)	0.0727 (2)	0.0360 (4)
H9A	0.139808	0.363569	-0.048097	0.054*
H9B	0.347117	0.446788	0.084261	0.054*
H9C	0.363568	0.295535	0.090656	0.054*
C10	0.5167 (4)	0.3200 (2)	0.7013 (3)	0.0377 (5)
H10A	0.515033	0.262157	0.796667	0.056*
H10B	0.665254	0.314862	0.689156	0.056*
H10C	0.491166	0.406855	0.732737	0.056*

Atomic displacement parameters (\AA^2)

	U^{11}	U^{22}	U^{33}	U^{12}	U^{13}	U^{23}
O1	0.0373 (8)	0.0291 (7)	0.0455 (8)	0.0094 (6)	0.0253 (7)	0.0102 (6)

C1	0.0236 (9)	0.0271 (10)	0.0230 (8)	-0.0020 (7)	0.0089 (7)	0.0028 (7)
O2	0.0264 (6)	0.0293 (7)	0.0244 (6)	-0.0054 (5)	0.0076 (5)	0.0019 (6)
C2	0.0226 (8)	0.0290 (9)	0.0315 (9)	0.0034 (7)	0.0133 (7)	0.0045 (8)
C3	0.0341 (10)	0.0280 (11)	0.0349 (10)	-0.0016 (8)	0.0116 (8)	-0.0065 (8)
C4	0.0226 (8)	0.0352 (11)	0.0303 (9)	-0.0013 (7)	0.0041 (7)	-0.0033 (8)
C5	0.0217 (9)	0.0414 (11)	0.0444 (11)	0.0002 (8)	0.0128 (8)	0.0045 (9)
C6	0.0300 (9)	0.0296 (10)	0.0357 (9)	0.0051 (7)	0.0187 (8)	0.0028 (8)
C7	0.0253 (8)	0.0329 (9)	0.0223 (8)	0.0028 (8)	0.0061 (6)	-0.0001 (8)
C8	0.0438 (12)	0.0399 (12)	0.0381 (11)	0.0136 (9)	0.0194 (10)	0.0101 (9)
C9	0.0347 (9)	0.0462 (11)	0.0279 (9)	0.0039 (10)	0.0127 (7)	0.0026 (9)
C10	0.0380 (11)	0.0445 (12)	0.0266 (9)	-0.0091 (9)	0.0078 (8)	0.0018 (8)

Geometric parameters (Å, °)

O1—C2	1.425 (2)	C5—H5A	0.9900
O1—H1	0.80 (3)	C5—H5B	0.9900
C1—O2	1.448 (2)	C6—H6A	0.9900
C1—C10	1.515 (2)	C6—H6B	0.9900
C1—C2	1.526 (2)	C7—C8	1.523 (3)
C1—C6	1.532 (2)	C7—C9	1.525 (3)
O2—C7	1.4665 (18)	C8—H8A	0.9800
C2—C3	1.539 (3)	C8—H8B	0.9800
C2—H2	1.0000	C8—H8C	0.9800
C3—C4	1.535 (3)	C9—H9A	0.9800
C3—H3A	0.9900	C9—H9B	0.9800
C3—H3B	0.9900	C9—H9C	0.9800
C4—C7	1.531 (3)	C10—H10A	0.9800
C4—C5	1.534 (3)	C10—H10B	0.9800
C4—H4	1.0000	C10—H10C	0.9800
C5—C6	1.540 (3)		
C2—O1—H1	109 (2)	H5A—C5—H5B	108.4
O2—C1—C10	106.18 (14)	C1—C6—C5	109.29 (14)
O2—C1—C2	106.40 (13)	C1—C6—H6A	109.8
C10—C1—C2	112.35 (16)	C5—C6—H6A	109.8
O2—C1—C6	109.76 (13)	C1—C6—H6B	109.8
C10—C1—C6	112.00 (15)	C5—C6—H6B	109.8
C2—C1—C6	109.91 (14)	H6A—C6—H6B	108.3
C1—O2—C7	114.93 (12)	O2—C7—C8	107.90 (15)
O1—C2—C1	108.43 (14)	O2—C7—C9	106.51 (13)
O1—C2—C3	112.08 (15)	C8—C7—C9	108.91 (16)
C1—C2—C3	108.81 (14)	O2—C7—C4	107.07 (14)
O1—C2—H2	109.2	C8—C7—C4	113.06 (15)
C1—C2—H2	109.2	C9—C7—C4	113.05 (16)
C3—C2—H2	109.2	C7—C8—H8A	109.5
C4—C3—C2	109.53 (15)	C7—C8—H8B	109.5
C4—C3—H3A	109.8	H8A—C8—H8B	109.5
C2—C3—H3A	109.8	C7—C8—H8C	109.5

C4—C3—H3B	109.8	H8A—C8—H8C	109.5
C2—C3—H3B	109.8	H8B—C8—H8C	109.5
H3A—C3—H3B	108.2	C7—C9—H9A	109.5
C7—C4—C5	110.26 (16)	C7—C9—H9B	109.5
C7—C4—C3	109.25 (15)	H9A—C9—H9B	109.5
C5—C4—C3	107.20 (16)	C7—C9—H9C	109.5
C7—C4—H4	110.0	H9A—C9—H9C	109.5
C5—C4—H4	110.0	H9B—C9—H9C	109.5
C3—C4—H4	110.0	C1—C10—H10A	109.5
C4—C5—C6	108.51 (14)	C1—C10—H10B	109.5
C4—C5—H5A	110.0	H10A—C10—H10B	109.5
C6—C5—H5A	110.0	C1—C10—H10C	109.5
C4—C5—H5B	110.0	H10A—C10—H10C	109.5
C6—C5—H5B	110.0	H10B—C10—H10C	109.5
C10—C1—O2—C7	-171.24 (15)	C3—C4—C5—C6	68.1 (2)
C2—C1—O2—C7	68.89 (16)	O2—C1—C6—C5	63.37 (18)
C6—C1—O2—C7	-50.00 (18)	C10—C1—C6—C5	-178.95 (17)
O2—C1—C2—O1	-177.12 (13)	C2—C1—C6—C5	-53.32 (19)
C10—C1—C2—O1	67.09 (18)	C4—C5—C6—C1	-11.6 (2)
C6—C1—C2—O1	-58.34 (18)	C1—O2—C7—C8	109.18 (16)
O2—C1—C2—C3	-54.97 (17)	C1—O2—C7—C9	-134.02 (16)
C10—C1—C2—C3	-170.76 (15)	C1—O2—C7—C4	-12.81 (18)
C6—C1—C2—C3	63.81 (17)	C5—C4—C7—O2	65.78 (18)
O1—C2—C3—C4	113.35 (17)	C3—C4—C7—O2	-51.77 (18)
C1—C2—C3—C4	-6.56 (19)	C5—C4—C7—C8	-52.9 (2)
C2—C3—C4—C7	61.95 (19)	C3—C4—C7—C8	-170.47 (16)
C2—C3—C4—C5	-57.52 (19)	C5—C4—C7—C9	-177.24 (15)
C7—C4—C5—C6	-50.8 (2)	C3—C4—C7—C9	65.2 (2)

Hydrogen-bond geometry (\AA , $^\circ$)

$D-H\cdots A$	$D-H$	$H\cdots A$	$D\cdots A$	$D-H\cdots A$
O1—H1 \cdots O2 ⁱ	0.80 (3)	1.97 (3)	2.7530 (19)	170 (3)

Symmetry code: (i) $-x+1, y-1/2, -z+1$.