PROKARYOTES



Draft Genome Sequence of Acholeplasma laidlawii, a Common Contaminant of Cell Cultures

AMERICAN SOCIETY FOR MICROBIOLOGY

Franciele Maboni Siqueira,^a Samuel Paulo Cibulski,^{b*} Thais Fumaco Teixeira,^b Fabiana Quoos Mayer,^c Paulo Michel Roehe^b

Departamento de Patologia Clínica Veterinária, Faculdade de Veterinária, Universidade Federal do Rio Grande do Sul, Porto Alegro, Rio Grande do Sul, Brazil^a; Departamento de Microbiologia Imunologia e Parasitologia, Laboratório de Virologia, Universidade Federal do Rio Grande do Sul, Porto Alegro, Rio Grande do Sul, Brazil^b; Laboratório de Biologia Molecular, Instituto de Pesquisas Veterinárias Desidério Finamor (IPVDF), Fundação Estadual de Pesquisa Agropecuária, Porto Alegro, Rio Grande do Sul, Brazil^c

gen@meAnnouncements™

ABSTRACT *Mollicutes* are important cell culture contaminants which may eventually affect the results of biological assays or affect their interpretation. *Acholeplasma laid-lawii* is one of the most frequent contaminants of cell cultures. Here, we report the complete genome sequence of *A. laidlawii* strain MDBK/IPV, recovered from Madin-Darby bovine kidney (MDBK) cells.

choleplasma species comprise bacteria from the Mollicutes class that are described as saprophyte and commensal bacteria. It has been suggested that acholeplasmas and phytoplasmas come from an Acholeplasma-like last-common ancestor (1). Unlike other mycoplasmas, members of the family Acholeplasmataceae do not require sterols for cultivation and are able to synthesize fatty acids from precursors (2). Some mycoplasmal species, among them Acholeplasma laidlawii, could persist as culture contamination on cell lineages and consequently can affect the results of biological assays or affect their interpretation. Moreover, A. laidlawii infects plants, with phytopathogenic effects analogous to Phytoplasma infection; besides, these bacteria were detected as commensals in insects (3). To our knowledge, only one complete genome sequence of A. laidlawii (strain PG-8A) has been published (4). The analysis of this A. laidlawii genome revealed a richly equipped repertoire for metabolism, SOS response, repair systems, and extensive transcriptional regulation machinery, including the twocomponent systems, riboswitches and T-boxes (4, 5). Here, we report the complete genome sequence of A. laidlawii strain MDBK/IPV, recovered from a Madin-Darby bovine kidney (MDBK) cell lineage.

A. laidlawii MDBK/IPV DNA was extracted from preparations of MDBK cells as follows: the supernatant of MDBK monolayers was clarified by low-speed centrifugation, filtered (0.45- μ m pore size), and ultracentrifuged (6). DNA was extracted according to a phenol-chloroform protocol. Whole-genome paired-end sequencing was performed in the Illumina MiSeq platform (Illumina), with the 500-cycle kit (version 2). DNA libraries were prepared with a Nextera kit (Illumina). The original reads were imported into the Geneious software (version 8.1) and trimmed. The assembly of the *A. laidlawii* genome was accomplished by read mapping to the reference *A. laidlawii* genome (*A. laidlawii* PG-8A, GenBank accession no. CP000896) with the Geneious software and *de novo* assembly with the SPAdes assembler 3.6.0 (7). This yielded 12-fold average read depth. Gene prediction and annotation were performed automatically with the NCBI annotation tool (8). Statistics were generated by QUAST (9).

The genome of *A. laidlawii* MDBK/IPV consists of 29 contigs (largest contig, 212,101 bp; N_{50} , 86,549 bp), with a total size of 1,307,942 bp. The G+C content of

Received 28 November 2016 Accepted 30 November 2016 Published 2 February 2017

Citation Siqueira FM, Cibulski SP, Teixeira TF, Mayer FQ, Roehe PM. 2017. Draft genome sequence of *Acholeplasma laidlawii*, a common contaminant of cell cultures. Genome Announc 5:e01578-16. https://doi.org/10.1128/ genomeA.01578-16.

Copyright © 2017 Siqueira et al. This is an open-access article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Paulo Michel Roehe, proehe@gmail.com.

* Present address: Samuel Paulo Cibulski, Laboratório de Virologia, Faculdade de Veterinária, Universidade Federal do Rio Grande do Sul, Porto Alegro, Rio Grande do Sul, Brazil.

F.M.S. and S.P.C. contributed equally to this work.

genameAnnouncements[™]

A. laidlawii MDBK/IPV is 31.97%. The *A. laidlawii* MDBK/IPV genome contains 1,250 predicted genes, two rRNA gene operons (5S-16S-23S), 33 tRNA genes, three noncoding RNAs (ncRNAs), and 25 pseudogenes. Among 1,183 potential protein-coding genes, 56% encode proteins with assigned functional roles. A high genetic repertoire was observed in *A. laidlawii* MDBK/IPV, which is described as necessary for adaptation to changing environmental conditions (5).

A comparative analysis between *A. laidlawii* MDBK/IPV and *A. laidlawii* PG-8A genomes shows the absence of approximately 200 kb in the MDBK/IPV genome. Interestingly, this PG-8A region is composed mainly of insertion elements (IS) and phage sequences. This genomic difference can be related to genome condensation or duplication of genetic material and integration events, which are commonly described in acholeplasmas (5).

The genome sequence of *A. laidlawii* MDBK/IPV will enable further investigations to better understand the molecular biology process of this bacterium that is important in culture cell contamination and in insights of minimal cellular life functions.

Accession number(s). The draft genome of *A. laidlawii* strain MDBK/IPV has been deposited at DDBJ/ENA/GenBank under the accession no. MIPS00000000. The version described in this paper is version MIPS01000000.

ACKNOWLEDGMENTS

This work was supported by the National Council for Scientific and Technological Development (CNPq) and Financiadora de Estudos e Projetos (FINEP) (grant 01.10.0783.04).

P.M.R. is a CNPq 1A Research fellow.

REFERENCES

- Bai X, Zhang J, Ewing A, Miller SA, Jancso Radek A, Shevchenko DV, Tsukerman K, Walunas T, Lapidus A, Campbell JW, Hogenhout SA. 2006. Living with genome instability: the adaptation of phytoplasmas to diverse environments of their insect and plant hosts. J Bacteriol 188:3682–3696. https://doi.org/10.1128/JB.188.10.3682-3696.2006.
- Saito Y, Silvius JR, McElhaney N. 1977. Membrane lipid biosynthesis in *Acholeplasma laidlawii* B: *de novo* biosynthesis of saturated fatty acids by growing cells. J Bacteriol 132:497–504.
- McCoy RE, Caudwell A, Chang CJ, Chen TA, Chiykowski LN, Cousin MT, Dale JL, de Leeuw GTN, Golino DA, Hackett KJ, Kirkpatric BC, Marwitz R, Petzold H, Sinha RC, Sigiura M, Whitcomb RF, Yang IL, Zhu BM, Seemuller E. 1989. Plant diseases associated with mycoplasmalike organisms, p 545–560. *In* Whitcomb RF, Tully JG (ed), The mycoplasmas, vol 5. Academic Press, New York, NY.
- Lazarev VN, Levitskii SA, Basovskii YI, Chukin MM, Akopian TA, Vereshchagin VV, Kostrjukova ES, Kovaleva GY, Kazanov MD, Malko DB, Vitreschak AG, Sernova NV, Gelfand MS, Demina IA, Serebryakova MV, Galyamina MA, Vtyurin NN, Rogov SI, Alexeev DG, Ladygina VG, Govorun VM. 2011. Complete genome and proteome of *Acholeplasma laidlawii*. J Bacteriol 193:4943–4953. https://doi.org/10.1128/JB.05059-11.
- 5. Kube M, Siewert C, Migdoll AM, Duduk B, Holz S, Rabus R, Seemüller E,

Mitrovic J, Müller I, Büttner C, Reinhardt R. 2014. Analysis of the complete genomes of *Acholeplasma brassicae*, *A. palmae* and *A. laidlawii* and their comparison to the obligate parasites from *'Candidatus* Phytoplasma.' J Mol Microbiol Biotechnol 24:19–36. https://doi.org/10.1159/000354322.

- de Sales Lima FE, Cibulski SP, Witt AA, Franco AC, Roehe PM. 2015. Genomic characterization of two novel polyomaviruses in Brazilian insectivorous bats. Arch Virol 160:1831–1836. https://doi.org/10.1007/s00705 -015-2447-6.
- Bankevich A, Nurk S, Antipov D, Gurevich AA, Dvorkin M, Kulikov AS, Lesin VM, Nikolenko SI, Pham S, Prjibelski AD, Pyshkin AV, Sirotkin AV, Vyahhi N, Tesler G, Alekseyev MA, Pevzner PA. 2012. SPAdes: a new genome assembly algorithm and its applications to single-cell sequencing. J Comput Biol 19:455–477. https://doi.org/10.1089/cmb.2012.0021.
- Angiuoli SV, Gussman A, Klimke W, Cochrane G, Field D, Garrity G, Kodira CD, Kyrpides N, Madupu R, Markowitz V, Tatusova T, Thomson N, White O. 2008. Toward an online repository of Standard Operating Procedures (SOPs) for (meta)genomic annotation. Omics 12:137–141. https://doi.org/ 10.1089/omi.2008.0017.
- Gurevich A, Saveliev V, Vyahhi N, Tesler G. 2013. QUAST: quality assessment tool for genome assemblies. Bioinformatics 29:1072–1075. https:// doi.org/10.1093/bioinformatics/btt086.