

REVIEW

Type 1 interferons and myositis

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Abstract

Recent studies suggest a mechanistic role for molecules induced by type 1 interferons in the pathogenesis of some forms of myositis. For dermatomyositis, evidence that these molecules injure myofibers seems especially strong. In the group of disorders known as polymyositis, the study of blood samples suggests a potential role. It is unknown what drives the sustained presence of type 1 interferon-inducible molecules in these diseases, as the type 1 interferons themselves have not been specifically detected along with their downstream biomarkers. Therapeutic development for blockade of IFN α is in progress aided by the identification of blood genomic biomarkers.

Introduction

The inflammatory myopathies – including dermatomyositis (DM), inclusion body myositis, and polymyositis (PM) – are poorly understood autoimmune diseases affecting skeletal muscle. Evidence regarding the significance of type 1 interferons to these diseases, especially DM, is reviewed in the present article, with much of this material recently discussed elsewhere [1,2].

The type 1 interferons are a class of molecules that include IFN α and IFN β . After binding to the type 1 interferon receptor (IFNAR) on target cells, these cytokines can stimulate the transcription of a set of genes, the type 1 interferon-inducible genes. Proteins abundantly produced from these genes' transcripts – such as myxovirus resistance protein A, interferon-stimulated gene 15 (ISG15), and 2',5'-oligoadenylate synthetase 1 – remain inside cells. They normally function as defenses against viral infections through a variety of means, such as inhibiting viral transcription, translation, or assembly of viral nucleocapsids. It is possible that the chronic intracellular overproduction of these transcripts and proteins might

be directly harmful to cells, such as muscle fibers in myositis [3].

Myositis, pathology, and mechanisms

The varied forms of myositis have distinct clinical and pathological features (Figure 1), and probably involve distinct mechanisms of tissue injury. DM, in addition to clinical skin involvement, has two unique pathological features (perifascicular atrophy and endothelial cell tubuloreticular inclusions) that distinguish it from other muscle diseases. Inclusion body myositis has a unique clinical distribution of involvement, with substantial weakness of the quadriceps and wrist and finger flexors, as well as specific suggestive pathological features including rimmed vacuoles. The broad category of PM is mainly distinguished by a collection of otherwise individually nonspecific features. Although frequently lumped together, DM and PM probably involve entirely different mechanisms of tissue injury.

Because type 1 interferons may have many effects on cells of the immune system, they may have roles in the varied immune responses present across multiple myositis subtypes. Yet studies to date suggest that only in DM are these molecules strongly and directly influencing molecular events in muscle.

Why focus on type 1 interferons in dermatomyositis mechanism?

Recognition that cytokines are present in myositis muscle biopsy samples began with immunohistochemical studies for cytokine proteins [4]. This approach is confounded by a number of technical and biological limitations [5], including nonspecific immunoreactivity, transient expression of cytokines, and their low concentration. For these reasons, some investigators turned to examining cytokine mRNA transcripts in muscle homogenates [6]. Initial PCR-based studies of cytokine transcripts including IFN α and IFN γ generally found no myositis subtype-specific differences compared with nonmyositis muscle – except for granulocyte-macrophage colony-stimulating factor, which was detected in 12 out of 15 myositis samples but in none out of 10 controls [6]. Many subsequent studies of cytokine transcripts and proteins (discussed in [7-9]) have reported variable and often conflicting results.

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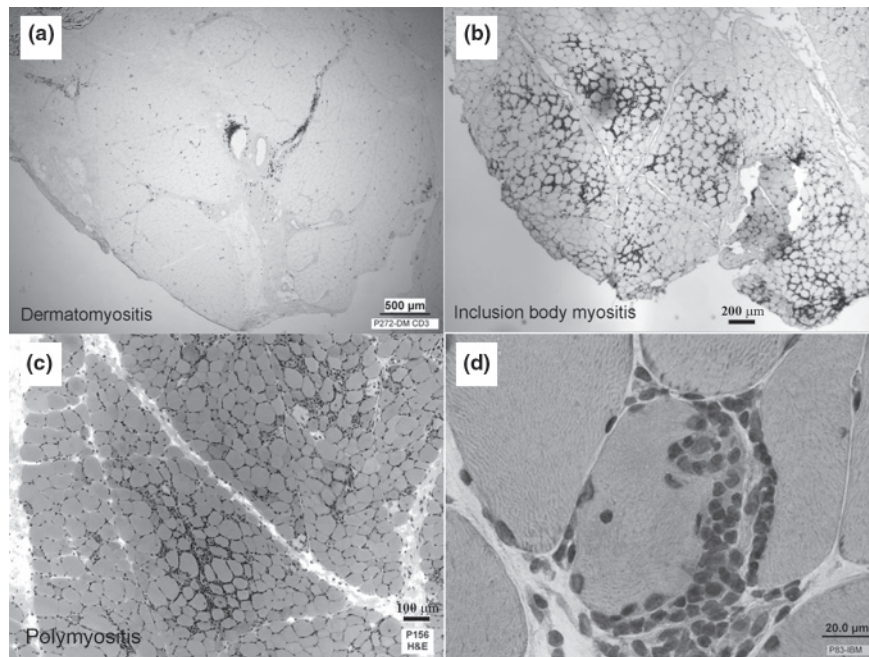


Figure 1. Differing pathologies in myositis subtypes. The distribution of immune system cells differs among myositis subtypes. (a) In dermatomyositis, immune system cells are predominantly in the regions of connective tissue that lie between muscle fascicles and include medium-sized and large blood vessels. In (b) inclusion body myositis and (c) polymyositis, immune cells surround myofibers. (d) Especially in inclusion body myositis, these may sometimes invade myofibers.

Because of the potential for transient expression and the low concentration of cytokines, downstream persisting effects of cytokines have been sought. Two types of biomarkers (macromolecular and molecular) provide strong evidence that DM muscle has experienced strong signaling of the type 1 interferon receptor. More than 25 years ago, tubuloreticular inclusions (also known as lupus inclusions) – macromolecular structures commonly visible with electron microscopy in DM muscle endothelial cells [10,11] and rarely seen in other forms of myositis – were recognized as downstream markers of type 1 interferon signaling. Tubuloreticular inclusions in circulating blood cells develop in patients treated with IFN α [12,13] and those in cultured endothelial cells and other cells develop directly in response to IFN α and IFN β [14-16], but not IFN γ [13]. For uncertain reasons, no PubMed indexed publication made a connection between this tubuloreticular inclusion literature and DM over 20 years [17].

Over the past 8 years the marked overproduction of type 1 interferon-inducible transcripts and proteins in muscle has been found to be remarkably unique to DM in comparison with all other muscle diseases studied [2,18,19]. Microarray gene expression studies of muscle biopsy specimens measuring approximately 18,000 transcripts in each of 113 muscle biopsy samples from patients with a wide range of myopathies showed that only DM samples with perifascicular atrophy have marked

elevation of type 1 interferon-inducible transcripts (Figure 2a) [2]. In analyses combining publicly available data, the remarkable specificity of these transcript abundances for DM is impressive. For example, the transcript for the type 1 interferon-inducible gene ISG15 was higher in muscle in all 28 biopsies from adults with DM and perifascicular atrophy and from children with juvenile DM than in every one of 199 non-DM biopsy samples from a wide range of neuromuscular diseases (Figure 2b).

Two type 1 interferon-inducible proteins have similarly been shown to be highly specific biomarkers of DM muscle. Myxovirus resistance protein A is impressively and uniquely (in comparison with other muscle diseases) abundant in DM myofibers with perifascicular atrophy and in DM capillaries (Figure 3) [18]. ISG15, a ubiquitin-like modifier, is furthermore attached to many other proteins in DM muscle, the identities of which have not been determined (Figure 2c). Exposure of human skeletal muscle cell cultures to IFN α or IFN β produces a similar picture of ISG15 conjugation present in human DM samples (Figure 2c) [2].

DM is a systemic disease, involving muscle, skin, and, variably, other tissues. Skin gene expression profiling, reported only in abstract format to date [20], has similarly shown marked abundance of type 1 interferon-inducible transcripts. The topology of keratinocyte injury in DM skin is similar to that of myofiber injury in DM muscle [21].

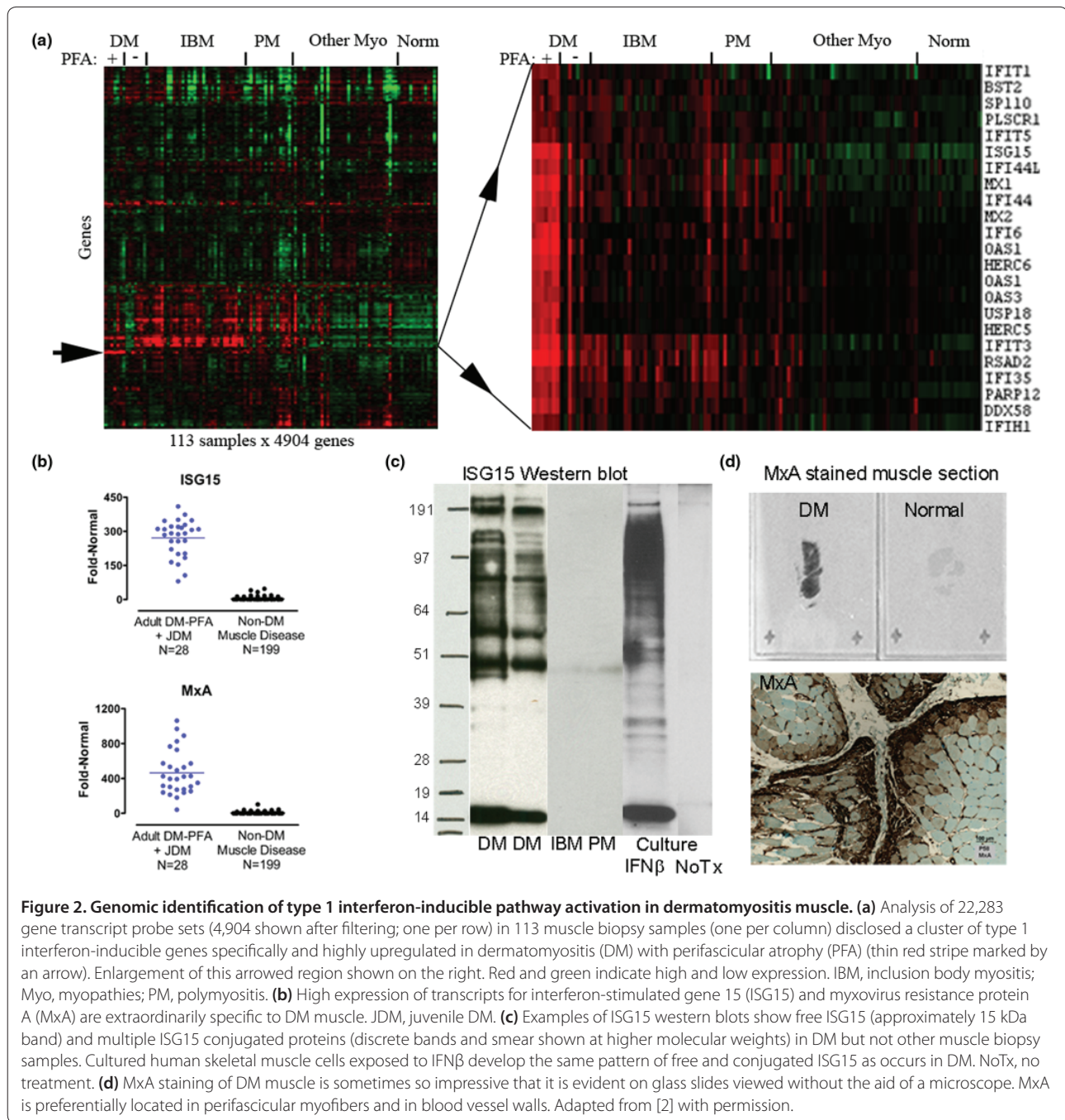


Figure 2. Genomic identification of type 1 interferon-inducible pathway activation in dermatomyositis muscle. (a) Analysis of 22,283 gene transcript probe sets (4,904 shown after filtering; one per row) in 113 muscle biopsy samples (one per column) disclosed a cluster of type 1 interferon-inducible genes specifically and highly upregulated in dermatomyositis (DM) with perifascicular atrophy (PFA) (thin red stripe marked by an arrow). Enlargement of this arrowed region shown on the right. Red and green indicate high and low expression. IBM, inclusion body myositis; Myo, myopathies; PM, polymyositis. (b) High expression of transcripts for interferon-stimulated gene 15 (ISG15) and myxovirus resistance protein A (MxA) are extraordinarily specific to DM muscle. JDM, juvenile DM. (c) Examples of ISG15 western blots show free ISG15 (approximately 15 kDa band) and multiple ISG15 conjugated proteins (discrete bands and smear shown at higher molecular weights) in DM but not other muscle biopsy samples. Cultured human skeletal muscle cells exposed to IFN β develop the same pattern of free and conjugated ISG15 as occurs in DM. NoTx, no treatment. (d) MxA staining of DM muscle is sometimes so impressive that it is evident on glass slides viewed without the aid of a microscope. MxA is preferentially located in perifascicular myofibers and in blood vessel walls. Adapted from [2] with permission.

Patients with dermatological features of DM who lack significant clinical evidence of muscle involvement have been classified as clinically amyopathic DM. Auto-antibodies to a classic type 1 interferon-inducible protein IFIH1 (interferon induced with helicase C domain; also called MDA-5) have been recently identified [22]. Using optimized cutoff values in an ELISA assay, the presence of anti-IFIH1 antibodies in clinically amyopathic DM among 262 patients with a range of connective tissue diseases was 69% sensitive and 99.6% specific. Significant

anti-IFIH1 autoantibody levels were present in 22 out of 32 patients with clinically amyopathic DM, but only in one of 35 patients with classic DM and in none of 53 patients with PM. These remarkably strong data both indicate a clinically valuable biomarker of clinically amyopathic DM and provide mechanistic evidence for some abnormality related to type 1 interferons in clinically amyopathic DM. The nature of this relationship is uncertain; one speculation is that IFIH1, a nuclear RNA helicase, is overproduced or altered in some way in



Figure 3. Myxovirus resistance protein A expression in dermatomyositis muscle. Image of whole muscle section stained for myxovirus resistance protein A (MxA) showing abundant myofiber protein expression (brown) preferentially in a perifascicular distribution. Adapted from [3] with permission.

clinically amyopathic DM, and evokes an autoantibody response.

A blood type 1 interferon-inducible signature

Blood gene expression profiling has also demonstrated marked abundance of these transcripts in patients with active DM, such as untreated patients, but also in PM (see below) [19]. One study that did not find marked type 1 interferon-inducible transcript abundance in DM blood samples had included almost only treated patients (11 out of 12 patients receiving prednisone; eight of these patients receiving an additional second immunosuppressant agent) [23]. Microarray experiments measure the abundance of 10,000s of transcripts simultaneously; any set of transcripts may be called a signature, but what is impressive about such experiments in DM and PM blood samples is the dominance of these gene expression patterns by type 1 interferon-inducible genes. In a study of 23 patients with DM and PM, at least 24 of the highest expressed 25 genes among approximately 38,000 genes studied are all known to be highly inducible by type 1 interferons [19].

The type 1 interferon-inducible transcript overproduction in DM and PM is highly correlated, within individual patients, with clinical measures of disease activity [19] (Figure 4 and unpublished data). Although the absolute magnitude of the upregulation of this signature does not highly correlate with disease severity across patients, within individual patients the signature does track disease activity – a situation similar to the commonly used clinical biomarker creatine kinase.

What drives the production of these downstream biomarkers of type 1 interferon signaling in DM muscle?

A fundamental question that has not been answered is what drives the production of type 1 interferon-inducible

molecules in DM muscle. Specific elevation of type 1 interferon transcripts or proteins has not been demonstrated in DM muscle. Both microarray and real-time quantitative PCR do not show impressive differences, compared with other myositis samples, in transcripts encoding a range of IFN α subtypes or IFN β in the same DM muscle samples that have marked upregulation (10-fold to 100-fold) of downstream type 1 interferon-inducible transcripts (unpublished data). Immunoblots from myositis samples similarly do not appear to show differential presence of IFN α or IFN β protein in DM muscle (preliminary unpublished data).

This situation parallels that seen in systemic lupus erythematosus. Although studies performed almost 30 years ago detected molecules believed to be IFN α in 60 to 76% of systemic lupus erythematosus blood samples using functional antiviral assays sometimes in combination with neutralizing antibodies [24-27], the literature has been notable for the absence of detection of IFN α in systemic lupus erythematosus blood or tissue samples by direct methods such as ELISA, immunoblot, or mass spectrometry. For example, one study found measurable levels of IFN α protein by ELISA in only two out of 38 patients, while most of these same 38 samples showed marked increases in type 1 interferon-inducible transcripts [28]. The lack of direct detection of IFN α protein has been attributed to potential technical limitations, although unexpected results in science have often been assumed to be erroneous. A more recent functional assay looking at type 1 interferon-inducible transcription has similarly detected activity in systemic lupus erythematosus plasma [29].

The interpretation of the results of functional assays is complicated by potential type 1 interferon autocrine mechanisms. In mouse cells, autostimulation of the IFNAR by early secreted type 1 interferons results in marked amplification of IFN α production [30-33]. Anti-IFN α antibodies used in functional assays to neutralize sample IFN α could potentially diminish type 1 interferon-inducible gene transcription through neutralizing early secreted reporter cell IFN α , although this possibility is speculative.

For juvenile DM, the functional assay for gene transcription has been used for detection of serum type 1 interferon-inducing activity and similarly interpreted as indicating the presence of IFN α in some blood samples [34]. What is particularly remarkable about this study was the marked range of interferon-inducing activity of serum from healthy children and adults, which varies by over 100-fold and includes many healthy people whose activity exceeded the mean value for the juvenile DM population.

These data suggest that the marked production of type 1 interferon-inducible transcripts and proteins in

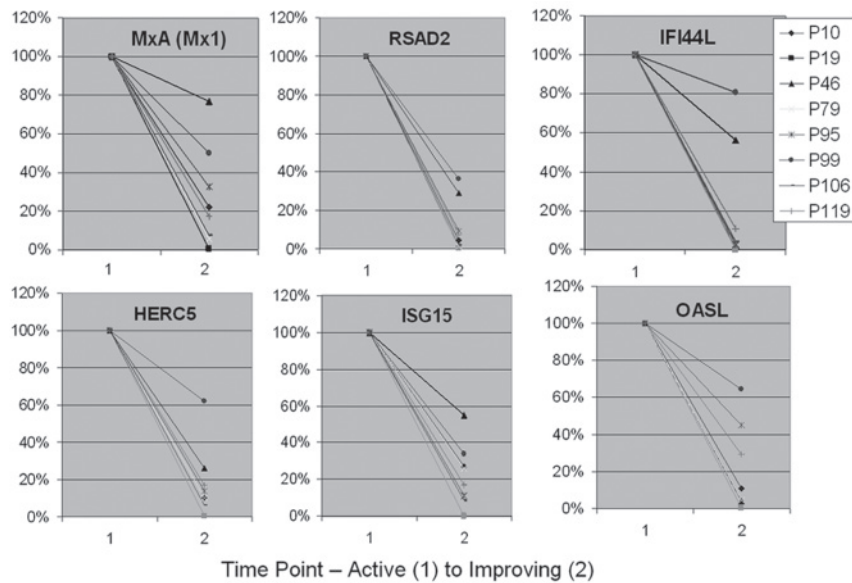


Figure 4. Blood type 1 interferon-inducible gene expression correlation with disease activity. Downregulation of six type 1 interferon-inducible genes in eight patients, correlating with improvement in clinical disease from time point 1 (active) to time point 2 (improving). Adapted from [19] with permission.

DM muscle, probably by myofibers, might result from sustained activation of the type 1 interferon receptor IFNAR in the absence of excessive (compared with the wide range of normal) type 1 interferons, or through mechanisms even further downstream that bypass IFNAR. The most natural interpretation of the data to date suggests that what may turn out to be most crucial with regard to DM myofiber injury is not the abundance of a type 1 interferon, but rather sustained abnormal function of the IFNAR or a further downstream process.

Potential role for type 1 interferons in polymyositis and inclusion body myositis

PM is an umbrella term for patients with various forms of myositis that are difficult to classify. DM and PM have substantial differences with regard to abundance of type 1 interferon-inducible molecules in muscle biopsy samples (Figure 5), yet marked overexpression of type 1 interferon-inducible genes has also been found in blood in PM [19]. Within PM muscle, inflammatory cells typically surround, displace, and sometimes invade muscle fibers. These cells include T cells, myeloid dendritic cells, macrophages, and plasma cells (reviewed in [3]). Type 1 interferons have multiple effects on these cell types, and it is possible that through these effects the type 1 interferon system is contributing to PM myofiber injury.

Patients with inclusion body myositis – a highly inflammatory disorder of muscle, as judged by abundance of immune system cells and transcripts in muscle – do not have high levels of muscle or blood type 1

interferon-inducible transcripts (Figure 5), although a small proportion of patients may have modest elevation of such transcripts in blood alone. As in PM, the mechanistic interpretation of blood expression of these transcripts is uncertain, and could reflect less-specific effects driving immune cell development.

Predicted exacerbations with TNF α inhibition

TNF α appears to have an antagonistic relationship with type 1 interferons [35]. It may directly inhibit the generation of plasmacytoid dendritic cells from progenitor cells and may inhibit plasmacytoid dendritic cell production of type 1 interferons. Studies of etanercept in Sjogren's syndrome indeed showed that this drug increased type 1 interferon activity [36]. Accordingly, models that propose a significant role for type 1 interferons in the pathogenesis of myositis predict that TNF α inhibition might exacerbate myositis. Published experience with TNF α inhibition in patients with myositis appears to support this model. Two open-label studies of infliximab have been terminated before completion or had substantial dropout rates for reasons that included disease progression [37,38]. Although this class of drugs may prove useful in the management of some patients, currently it appears unlikely to be of more general use for myositis.

Conclusion: diagnostics and therapeutic development

The presence of marked overproduction of type 1 interferon-inducible transcripts in blood specimens from

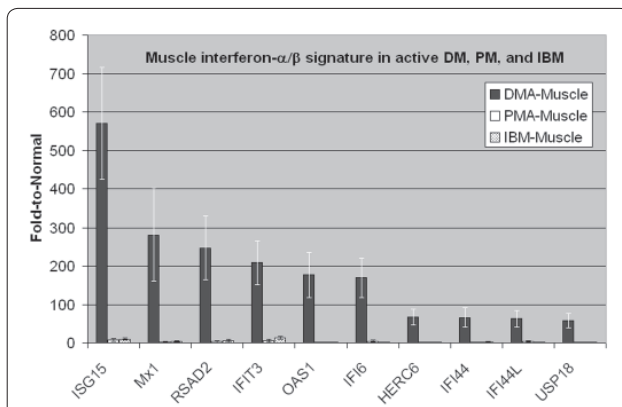


Figure 5. Distinct muscle expression of type 1 interferon-inducible genes in inflammatory myopathies. Distinct muscle expression of type 1 interferon-inducible genes in dermatomyositis (DM) compared with polymyositis (PM) and inclusion body myositis (IBM). Muscle microarray data shown for 20 patients (five each with active dermatomyositis (DMA), active polymyositis (PMA), untreated IBM, and normal) with plotted mean values and error bars for mean \pm standard error for each group. Highly expressed genes in DM muscle are orders of magnitude greater than in PM and IBM. Adapted from [19] with permission.

patients with active DM and PM has potential for diagnostic use [19]. These biomarkers may be able to distinguish these disorders from inclusion body myositis and other muscle diseases that sometimes present diagnostic uncertainty. Furthermore, they may be useful for therapeutic development. A phase 1b trial of anti-IFN α therapy has been initiated in DM and PM [39]. Entry criteria into this study include the presence of sufficiently high type 1 interferon-inducible gene expression in blood. Future studies targeting the IFNAR or further downstream events have strong rationale in DM, and perhaps in PM.

Abbreviations

DM = dermatomyositis; ELISA = enzyme-linked immunosorbent assay; IFN = interferon; IFNAR = type 1 interferon receptor; ISG15 = interferon-stimulated gene 15; PCR = polymerase chain reaction; PM = polymyositis; TNF = tumor necrosis factor.

Competing interests

SAG has worked as a consultant regarding clinical trial planning for MedImmune, LLC and has a Sponsored Research Agreement with MedImmune, LLC. SAG is an inventor of intellectual property pertaining to myositis diagnostics. SAG has National Institutes of Health funding and the publication of articles constitutes evidence of productivity that may be used to support future requests for National Institutes of Health funding.

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References

- Greenberg SA: Inflammatory myopathies: disease mechanisms. *Curr Opin Neurol* 2009, **22**:516-523.
- Salajegheh M, Kong SW, Pinkus JL, Walsh RJ, Liao A, Nazareno R, Amato AA, Krastins B, Morehouse C, Higgs BW, Jallal B, Yao Y, Sarracino DA, Parker KC, Greenberg SA: Interferon-stimulated gene 15 (ISG15) conjugates proteins in dermatomyositis muscle with perifascicular atrophy. *Ann Neurol* 2009, **67**:53-63.
- Greenberg SA: Proposed immunologic models of the inflammatory myopathies and potential therapeutic implications. *Neurology* 2007, **69**:2008-2019.
- Isenberg DA, Rowe D, Shearer M, Novick D, Beverley PC: Localization of interferons and interleukin 2 in polymyositis and muscular dystrophy. *Clin Exp Immunol* 1986, **63**:450-458.
- Emslie-Smith AM, Arahata K, Engel AG: Major histocompatibility complex class I antigen expression, immunolocalization of interferon subtypes, and T cell-mediated cytotoxicity in myopathies. *Hum Pathol* 1989, **20**:224-231.
- Lundberg I, Brengman JM, Engel AG: Analysis of cytokine expression in muscle in inflammatory myopathies, Duchenne dystrophy, and non-weak controls. *J Neuroimmunol* 1995, **63**:9-16.
- Figarella-Branger D, Civatte M, Bartoli C, Pellissier JF: Cytokines, chemokines, and cell adhesion molecules in inflammatory myopathies. *Muscle Nerve* 2003, **28**:659-682.
- Salomonsson S, Lundberg IE: Cytokines in idiopathic inflammatory myopathies. *Autoimmunity* 2006, **39**:177-190.
- Schmidt J, Barthel K, Wrede A, Salajegheh M, Bahr M, Dalakas MC: Interrelation of inflammation and APP in sIBM: IL-1 beta induces accumulation of beta-amyloid in skeletal muscle. *Brain* 2008, **131**(Pt 5):1228-1240.
- Banker BQ: Dermatomyositis of childhood, ultrastructural alterations of muscle and intramuscular blood vessels. *J Neuropathol Exp Neurol* 1975, **34**:46-75.
- Norton WL, Velayos E, Robison L: Endothelial inclusions in dermatomyositis. *Ann Rheum Dis* 1970, **29**:67-72.
- Grimley PM, Davis GL, Kang YH, Dooley JS, Strohmaier J, Hoofnagle JH: Tubuloreticular inclusions in peripheral blood mononuclear cells related to systemic therapy with alpha-interferon. *Lab Invest* 1985, **52**:638-649.
- Rich SA, Owens TR, Bartholomew LE, Gutterman JU: Immune interferon does not stimulate formation of alpha and beta interferon induced human lupus-type inclusions. *Lancet* 1983, **1**:127-128.
- Grimley PM, Rutherford MN, Kang YH, Williams T, Woody JN, Silverman RH: Formation of tubuloreticular inclusions in human lymphoma cells compared to the induction of 2'-5'-oligoadenylate synthetase by leucocyte interferon in dose-effect and kinetic studies. *Cancer Res* 1984, **44**:3480-3488.
- Kuyama J, Kanayama Y, Mizutani H, Katagiri S, Tamaki T, Yonezawa T, Tarui S, Morise H, Arimura H, Suyama T: Formation of tubuloreticular inclusions in mitogen-stimulated human lymphocyte cultures by endogenous or exogenous alpha-interferon. *Ultrastruct Pathol* 1986, **10**:77-85.
- Feldman D, Goldstein AL, Cox DC, Grimley PM: Cultured human endothelial cells treated with recombinant leukocyte A interferon. Tubuloreticular inclusion formation, antiproliferative effect, and 2',5' oligoadenylate synthetase induction. *Lab Invest* 1988, **58**:584-589.
- Greenberg SA, Amato AA: Uncertainties in the pathogenesis of adult dermatomyositis. *Curr Opin Neurol* 2004, **17**:359-364.
- Greenberg SA, Pinkus JL, Pinkus GS, Burleson T, Sanoudou D, Tawil R, Barohn RJ, Saperstein DS, Briemberg HR, Ericsson M, Park P, Amato AA: Interferon-alpha/beta-mediated innate immune mechanisms in dermatomyositis. *Ann Neurol* 2005, **57**:664-678.
- Walsh RJ, Kong SW, Yao Y, Jallal B, Kiener PA, Pinkus JL, Beggs AH, Amato AA, Greenberg SA: Type I interferon-inducible gene expression in blood is present and reflects disease activity in dermatomyositis and polymyositis. *Arthritis Rheum* 2007, **56**:3784-3792.
- Kea B, Pesich R, Chung LS, Brown PO, Fiorentino D: Genomic analyses identify lipid metabolism abnormalities in dermatomyositis patients [abstract]. *J Invest Dermatol* 2007, **127**(Suppl 1):S12.
- Greenberg SA, Fiorentino D: Similar topology of injury to keratinocytes and myofibers in dermatomyositis skin and muscle. *Br J Dermatol* 2009, **160**:464-465.

22. Sato S, Hoshino K, Satoh T, Fujita T, Kawakami Y, Fujita T, Kuwana M: **RNA helicase encoded by melanoma differentiation-associated gene 5 is a major autoantigen in patients with clinically amyopathic dermatomyositis: association with rapidly progressive interstitial lung disease.** *Arthritis Rheum* 2009, **60**:2193-2200.
23. Baechler EC, Bauer JW, Slattery CA, Ortmann WA, Espe KJ, Novitzke J, Ytterberg SR, Gregersen PK, Behrens TW, Reed AM: **An interferon signature in the peripheral blood of dermatomyositis patients is associated with disease activity.** *Mol Med* 2007, **13**:59-68.
24. Hooks JJ, Jordan GW, Cupps T, Moutsopoulos HM, Fauci AS, Notkins AL: **Multiple interferons in the circulation of patients with systemic lupus erythematosus and vasculitis.** *Arthritis Rheum* 1982, **25**:396-400.
25. Hooks JJ, Moutsopoulos HM, Geis SA, Stahl NI, Decker JL, Notkins AL: **Immune interferon in the circulation of patients with autoimmune disease.** *N Engl J Med* 1979, **301**:5-8.
26. Preble OT, Black RJ, Friedman RM, Klippel JH, Vilcek J: **Systemic lupus erythematosus: presence in human serum of an unusual acid-labile leukocyte interferon.** *Science* 1982, **216**:429-431.
27. Ytterberg SR, Schnitzer TJ: **Serum interferon levels in patients with systemic lupus erythematosus.** *Arthritis Rheum* 1982, **25**:401-406.
28. Baechler EC, Batliwalla FM, Karypis G, Gaffney PM, Ortmann WA, Espe KJ, Shark KB, Grande WJ, Hughes KM, Kapur V, Gregersen PK, Behrens TW: **Interferon-inducible gene expression signature in peripheral blood cells of patients with severe lupus.** *Proc Natl Acad Sci U S A* 2003, **100**:2610-2615.
29. Hua J, Kirou K, Lee C, Crow MK: **Functional assay of type I interferon in systemic lupus erythematosus plasma and association with anti-RNA binding protein autoantibodies.** *Arthritis Rheum* 2006, **54**:1906-1916.
30. Sato M, Suemori H, Hata N, Asagiri M, Ogasawara K, Nakao K, Nakaya T, Katsuki M, Noguchi S, Tanaka N, Taniguchi T: **Distinct and essential roles of transcription factors IRF-3 and IRF-7 in response to viruses for IFN-alpha/beta gene induction.** *Immunity* 2000, **13**:539-548.
31. Sato M, Tanaka N, Hata N, Oda E, Taniguchi T: **Involvement of the IRF family transcription factor IRF-3 in virus-induced activation of the IFN-beta gene.** *FEBS Lett* 1998, **425**:112-116.
32. Marie I, Durbin JE, Levy DE: **Differential viral induction of distinct interferon-alpha genes by positive feedback through interferon regulatory factor-7.** *Embo J* 1998, **17**:6660-6669.
33. Sato M, Hata N, Asagiri M, Nakaya T, Taniguchi T, Tanaka N: **Positive feedback regulation of type I IFN genes by the IFN-inducible transcription factor IRF-7.** *FEBS Lett* 1998, **441**:106-110.
34. Niewold TB, Kariuki SN, Morgan GA, Shrestha S, Pachman LM: **Elevated serum interferon-alpha activity in juvenile dermatomyositis: associations with disease activity at diagnosis and after thirty-six months of therapy.** *Arthritis Rheum* 2009, **60**:1815-1824.
35. Palucka AK, Blanck JP, Bennett L, Pascual V, Banchereau J: **Cross-regulation of TNF and IFN-alpha in autoimmune diseases.** *Proc Natl Acad Sci U S A* 2005, **102**:3372-3377.
36. Mavragani CP, Niewold TB, Moutsopoulos NM, Pillemer SR, Wahl SM, Crow MK: **Augmented interferon-alpha pathway activation in patients with Sjogren's syndrome treated with etanercept.** *Arthritis Rheum* 2007, **56**:3995-4004.
37. Hengstman GJ, De Bleecker JL, Feist E, Vissing J, Denton CP, Manoussakis MN, Slott Jensen H, van Engelen BG, van den Hoogen FH: **Open-label trial of anti-TNF-alpha in dermato- and polymyositis treated concomitantly with methotrexate.** *Eur Neurol* 2008, **59**:159-163.
38. Dastmalchi M, Grundtman C, Alexanderson H, Mavragani CP, Einarsson H, Helmers SB, Elvin K, Crow MK, Nennesmo I, Lundberg IE: **A high incidence of disease flares in an open pilot study of infliximab in patients with refractory inflammatory myopathies.** *Ann Rheum Dis* 2008, **67**:1670-1677.
39. **A study to evaluate safety of multi-dose MEDI-545 in adult patients with dermatomyositis or polymyositis** [<http://clinicaltrials.gov/ct2/show/NCT00533091>]

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