

Androgens and Adult Neurogenesis in the Hippocampus

Samantha A. Blankers^{1,2} and Liisa A.M. Galea^{1-3,*}

Abstract

Adult neurogenesis in the hippocampus is modulated by steroid hormones, including androgens, in male rodents. In this review, we summarize research showing that chronic exposure to androgens, such as testosterone and dihydrotestosterone, enhances the survival of new neurons in the dentate gyrus of male, but not female, rodents, via the androgen receptor. However, the neurogenesis promoting the effect of androgens in the dentate gyrus may be limited to younger adulthood as it is not evident in middle-aged male rodents. Although direct exposure to androgens in adult or middle age does not significantly influence neurogenesis in female rodents, the aromatase inhibitor letrozole enhances neurogenesis in the hippocampus of middle-aged female mice. Unlike other androgens, androgenic anabolic steroids reduce neurogenesis in the hippocampus of male rodents. Collectively, the research indicates that the ability of androgens to enhance hippocampal neurogenesis in adult rodents is dependent on dose, androgen type, sex, duration, and age. We discuss these findings and how androgens may be influencing neuroprotection, via neurogenesis in the hippocampus, in the context of health and disease.

Keywords: dentate gyrus; sex differences; aging; dihydrotestosterone; testosterone; cognition

Introduction

Efforts to characterize the postnatal production of new neurons have been ongoing since the concept was first introduced through experiments by Joseph Altman in the 1960s.¹ Although Altman's findings were initially dismissed by the scientific community, the notion of structural plasticity of dendritic spines in the adult brain gained acceptance with time²⁻⁴ (reviewed in Bosch and Hayashi⁵) and studies in songbirds confirmed that new cells produced postnatally were morphologically similar to neurons.^{6,7} Great progress has been made since these initial findings, through extensive research in neurogenic brain regions, including the hippocampus and olfactory bulb.⁸⁻¹⁸ Even in the early studies by Altman, it was suspected that androgens may be involved in the regulation of neurogenesis in the hippocampus.¹ Indeed, decades later, it was

noted that seasonal fluctuations in hippocampal neurogenesis coincide with changes in gonadal hormone levels in meadow voles¹⁹ and in black-capped chickadees²⁰ of both sexes. Moreover, sex differences have been identified in the maturation and attrition of new neurons in the hippocampus of rats²¹ and in response to certain stimuli such as stress in rats^{22,23}; for review see Ref.²⁴ These findings warranted further investigation into the role of sex-steroid hormones in the regulation of hippocampal neurogenesis, including the sex-specific effects of estrogens and androgens. Thus, in this review, we discuss the stages and function of adult neurogenesis in the hippocampus, followed by the influence of androgens on different stages of hippocampal neurogenesis and in response to injury or disease. Within each section, we discuss sex differences in the influence of androgens, if known.

¹Graduate Program in Neuroscience, The University of British Columbia, Vancouver, Canada.

²Djavad Mowafaghian Centre for Brain Health, The University of British Columbia, Vancouver, Canada.

³Department of Psychology, The University of British Columbia, Vancouver, Canada.

*ORCID ID (<https://orcid.org/0000-0003-2874-9972>).

*Address correspondence to: Liisa A.M. Galea, PhD, Graduate Program in Neuroscience, The University of British Columbia, 2215 Wesbrook Mall, Vancouver V6T 1Z3, British Columbia, Canada, Email: liisa.galea@ubc.ca



Androgens

Androgens are sex steroid hormones produced in Leydig cells of the testes and thecal-interstitial cells of the ovaries, as well as in the zona reticularis of the adrenal glands in both males and females.^{25,26} In addition, androgens are produced in the brain itself, via either local production or *de novo* synthesis of steroids.^{27–30} It is also important to recognize that the systemic levels of androgens are associated with levels in the hippocampus³¹ and influence local and *de novo* production of androgens,³² although more studies are needed. Some of the widely studied androgens include testosterone, 5 α -dihydrotestosterone (DHT), androstenedione, and dehydroepiandrosterone (DHEA) among others.³³ Testosterone can be metabolized to other androgens or to estradiol, the most potent of the natural estrogens, and thus can exert its effects via androgen receptors (ARs) or estrogen receptors (ERs) depending on the availability of enzymes. Testosterone can be reduced to the potent androgen DHT via the enzyme 5 α -reductase. DHT can be further metabolized to 5 α -androstane-3 α ,17 β -diol (3 α -diol) and 5 α -androstane-3 β ,17 β -diol (3 β -diol).³⁴ Both 3 α -diol and 3 β -diol possess weak affinity for ARs,³⁵ although 3 β -diol displays preferential activity at ER β .³⁶ Testosterone can also be aromatized to estradiol,³⁴ which binds to ER α , ER β , and the G protein-coupled ER (GPER). The ARs and ERs are most often found in intracellular locations or on the nucleus, but they can also be located on the membrane.^{37–39} Ligand binding, when located in the intracellular compartment, causes the AR or ER to be transported to the nucleus, where they may influence transcription of target genes to elicit genomic effects that occur on a scale of hours to days.⁴⁰ In addition, estradiol can bind to membrane-associated ERs such as GPER to induce rapid non-genomic effects through second-messenger signaling.³⁹

The ERs are abundant in the hippocampus, with a high concentration of GPER in CA1, CA3, and the dentate gyrus⁴¹ and the highest density of ER α and ER β in CA3 region compared with the other subregions of the hippocampus in male and female rats.^{42,43} On the other hand, ARs are not abundant in the dentate gyrus relative to the CA1 and CA3 regions of the hippocampus in both sexes.^{44–48} Indeed, the highest expression of ARs in the hippocampus is in the CA1 region in both sexes, which will vary by hormonal status (intact vs. gonadectomized).^{44–48} Studies have found that the density of ARs in the hippocampus increases with age, as serum testosterone levels decline.^{47,49} Sex differences in AR expression have been detected, as the density of AR

in the CA1 region of males is greater than that of the CA1 region of females⁴⁸ but this is not seen in all studies,⁴⁷ likely due to differences in whether gonadectomized versus intact rats were used. Indeed, intact males had higher AR expression in the CA1 region compared with intact females⁴⁹ but there are no significant sex differences in AR expression when comparing gonadectomized rats.⁴⁷ In addition, the gene encoding the AR protein contains a polymorphic cytosine-adenine-guanine (CAG) microsatellite of variable lengths ranging from 6 to 39 repeats at the N-terminal transactivation domain.⁵⁰ The functionality of ARs depends on the number of CAG repeats, as transactivation of AR decreases with CAG repeat expansion.⁵¹ Therefore, androgens act through ERs or ARs that are expressed in the hippocampus and influenced by several factors such as age, sex, and genetics.

Adult Neurogenesis

Neurogenesis is defined as the creation and functional integration of new neurons produced from neural stem/progenitor cells. There are several stages of neurogenesis that include cell proliferation, migration, differentiation, and survival of newly generated neurons (Fig. 1). Various internal and external factors have been identified as regulators of neurogenesis in each one of these stages; therefore, it is critical to be aware of the stage of neurogenesis being evaluated within an individual experiment. There are many methods available for the detection of newly formed neurons, each with their own strengths and limitations.^{53,56,57} Endogenous proteins, such as Ki67, are expressed during all stages of the cell cycle except G₀ and are used to measure cell proliferation.⁵³ Doublecortin (DCX), another endogenous protein, is used to measure the presence of immature neurons, as it is expressed during proliferation until approximately day 21 in rats.⁵⁴ Exogenous DNA-markers, such as ³H-thymidine, or synthetic nucleosides, such as 5-bromo-2-deoxyuridine (BrdU), may be used to monitor the production and survival of new neurons depending on the time between injection and perfusion. These markers are incorporated during DNA synthesis and may be visualized postmortem to detect newly generated cells in a region of interest.^{58,59} The timing between administration of DNA synthesis markers and perfusion of the animal is significant, as different time spans will measure different stages of neurogenesis. If animals are perfused 24 h or less after BrdU administration, this will be a measure of cell proliferation, whereas a span of >24 h



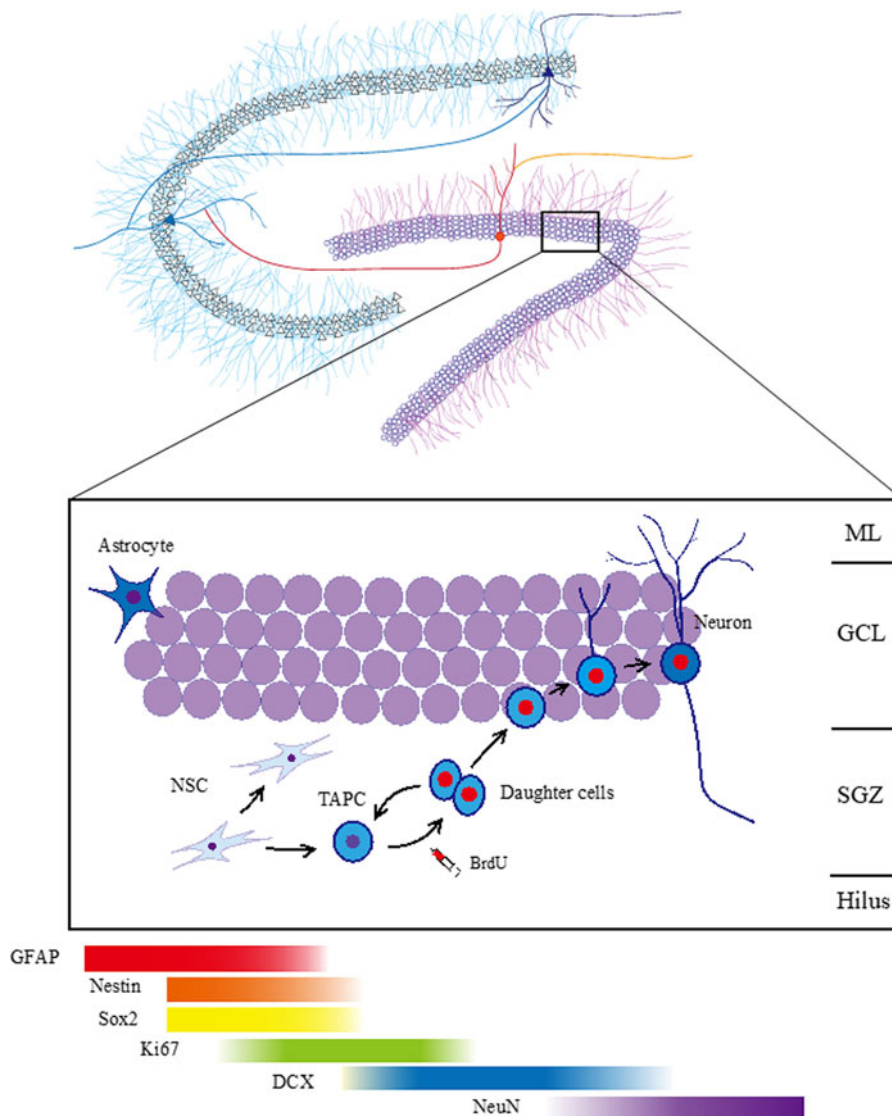


FIG. 1. Schematic image of a coronal section of the rodent hippocampus and stages of adult neurogenesis. An NSC can give rise to a TAPC, which is divided into two NPCs that migrate in the GCL of the dentate gyrus, differentiate into neurons, and send dendrites to the ML.⁵² The NSCs express the glia cell marker, GFAP, and NSC markers, Nestin, and Sox2.⁵² Ki67 is a nuclear protein that is expressed during all active phases of the cell cycle and thus is a marker of cell proliferation.⁵³ DCX is a microtubule-associated protein expressed in the cytoplasm of dividing NPC and immature neurons.⁵⁴ NeuN protein is expressed in new neurons beginning about 1 week after mitosis in rats and 2 weeks after mitosis in mice and is used as a nuclear marker for mature neurons.⁵⁵ BrdU is a DNA synthesis marker that is incorporated into cells that are actively synthesized DNA within the 2 h after injection (and in effect time stamps when the cell was produced). Depending on the length of time between injection and perfusion, BrdU-immunoreactive cells will label cell proliferation or the survival of new cells. Zif268 is an immediate early gene and will indicate whether a cell has been recently active (action potential). Reprinted with permission from Yagi et al.²¹ BrdU, 5-bromo-2-deoxyuridine; DCX, doublecortin; GCL, granular cell layer; GFAP, glial fibrillary acidic protein; ML, molecular layer; NeuN, neuronal nuclei; NPC, neural progenitor cells; NSC, neural stem cell; SGZ, subgranular zone; TAPC, transient amplifying progenitor cell.



will measure survival of the daughter cells.^{56,60} Further, the timing of experimental manipulation in relation to BrdU administration is important, as this will capture survival of new neurons either dependent or independent of the influence of cell proliferation; for review see Refs.^{56,61} It is noteworthy that endogenous markers of mature neurons must be used in combination with thymidine analogues to determine whether newly formed cells are new neurons; therefore, the co-expression of BrdU with endogenous markers such as neuronal nuclei is often used to detect new neurons versus the use of glial fibrillary acidic protein used for the detection of new glial cells.⁶²

As expected, neurogenesis occurs at a high level during development and diminishes in adulthood, and this has been observed across a wide variety of species, including primates^{8,9} and rodents.^{10,11} Under normal conditions, neurogenesis occurs in two distinct regions of the brain: the subventricular zone (SVZ), the lining of the lateral ventricles and the subgranular zone (SGZ) of the hippocampus. The SVZ contains mostly multipotent neural stem cells (NSCs), whereas the proliferative precursor cells of the SGZ were previously identified as mainly neural progenitor cells (NPC).^{63,64} However, studies have revealed the presence of multipotent NSCs in the SGZ,^{65–67} which ultimately give rise to NPC⁶⁸ that proliferate and migrate to the granular layer where most will differentiate into granule cells.^{1,69} Multipotent NSCs have the capacity for self-renewal and can give rise to multiple types of mature neural cells.⁷⁰ The NPC are distinct, as they have a limited capacity for self-renewal and usually differentiate into one specific type of neural cell.⁷⁰ The NPC originating in the SGZ proliferate and migrate to the granular layer, where most will differentiate into granule cells.^{1,69} Intriguingly, a few studies suggest that there is little decline in the number of new immature neurons throughout adulthood in humans.^{16–18} Evidence for neurogenesis in the hippocampus of humans has been demonstrated in a multitude of studies using a variety of techniques.^{12–18} Thus, these studies support the consensus that neurogenesis in the human hippocampus is evident throughout adult life, although this claim is not entirely undisputed.^{71,72} This review will focus on the influence of androgens in hippocampal neurogenesis, and the reader is directed to other reviews on hormones and the SVZ.^{73,74}

Function of Adult Hippocampal Neurogenesis

It is one thing to produce new neurons in the adult brain, but do these new neurons form meaningful con-

nections to alter the function of the hippocampus? It is important to recognize that a new neuron could make aberrant connections that might interfere with the normal function of the dentate gyrus.^{75,76} Ectopic new neurons can interfere with normal activity of the dentate gyrus, as seen with hippocampal neurogenesis after seizures in animal models of temporal lobe epilepsy.^{75,77} Populations of these new neurons form aberrant axon projections that are characteristic of those seen in postmortem humans with temporal lobe epilepsy,^{77,78} and this abnormal cytoarchitecture may result in long-term alterations of hippocampal circuitry.⁷⁵ Indeed, seizure-induced increases in neurogenesis is commensurate with reduced memory, and when neurogenesis is reduced in response to seizures, memory is improved.⁷⁵ On the other hand, voluntary wheel-running increases both hippocampal neurogenesis and performance on several new memory tasks in mice.^{79–81} Jakubs et al. used whole cell-patch clamp recordings to compare the properties of new neurons generated on exposure to either running or induced seizures in rats. Both running and seizures will increase neurogenesis in the hippocampus, but new neurons generated in runners demonstrated higher excitatory synaptic drive and lower inhibitory synaptic drive compared with new neurons generated with seizures.⁷⁶ Together, these findings demonstrate that new neurons may have distinct functional properties that are dependent on the circumstances of which neurogenesis was induced; thus, increased neurogenesis is not always beneficial to the individual if atypical integration occurs; for review see Refs.^{82–84}

New neurons appropriately integrated into the hippocampal circuit are believed to influence and support brain functioning in a variety of ways.^{24,52,61,68,85} The hippocampus is critical in forming contextually rich memories⁸⁶ due to the neuronal characteristics of the dentate gyrus. That is, discrete representations of memory are possible because neurons of the dentate gyrus are able to discriminate between small differences in cortical input patterns.⁸⁷ Briefly, adult hippocampal neurogenesis is critical in discrimination between highly similar situations,^{88–92} a phenomenon known as pattern separation.⁹³ Other studies have shown that hippocampal neurogenesis is critical for context encoding in spatial discrimination tasks and contextual/trace fear conditioning^{89,94–97} in both male and female mice; for review see Refs.^{98,99} Functional differences exist between the dorsal and ventral hippocampus due in part to their distinct connections to extra-hippocampal



structures, receptor density patterns, and gene expression patterns.^{100,101} The dorsal hippocampus is responsible for cognitive processing whereas the ventral hippocampus regulates emotional processing, including mood and stress.¹⁰² It has been proposed that hippocampal neurogenesis, particularly in the ventral hippocampus, is implicated in this emotional regulation.¹⁰³ Animal models of depression in male and female rodents decrease neurogenesis in the ventral dentate gyrus¹⁰⁴ and chronic, but not acute, treatment with pharmacological antidepressants restores neurogenesis in the dentate gyrus of male and female rodents.^{24,105,106} In primates, the anterior hippocampus is akin to the ventral hippocampus of rodents, and humans with major depressive disorder show decreased cell proliferation¹⁰⁷ which is increased with antidepressant exposure¹⁰⁷ in the anterior hippocampus, dependent on factors such as age and sex.¹⁰⁸ Further, Surget et al.¹⁰⁹ found that the antidepressant fluoxetine was not effective in restoring a dysregulated hippocampal-driven response to chronic stress unless an intact neurogenic system was present in the ventral hippocampus of male mice. Therefore, numerous lines of evidence suggest that neurogenesis in adulthood influences various aspects of brain function, including mood, stress, and cognition, dependent on the brain region in which new neurons are integrating.

Androgens and Adult Hippocampal Neurogenesis

In the following sections, we discuss the influence of androgen manipulations on hippocampal neurogenesis in males and females across the adult lifespan and in response to injury or disease.

Castration and Adult Hippocampal Neurogenesis

Reproductive status influences neurogenesis in the hippocampus of male rodents.^{110,111} Reproductively active male meadow voles had increased survival of new neurons compared with reproductively inactive male meadow voles.¹¹⁰ Castrated adult rats and mice show decreased survival of new neurons or fewer immature neurons in the dentate gyrus compared with intact males.^{111–114} On the other hand, castration before adolescence does not have the same effect on hippocampal neurogenesis as it increased neurogenesis in the hippocampus of male rhesus macaque monkeys¹¹⁵ and had no significant influence on hippocampal neurogenesis in male rats.¹¹⁶ These studies collectively indicate that

the timing of castration during the lifespan matters for the influence on hippocampal neurogenesis, with adult castration decreasing but adolescent castration either increasing or not affecting neurogenesis in the hippocampus.

Androgens and Adult Hippocampal Neurogenesis in Males

Chronic exposure to androgens generally upregulates neurogenesis in the hippocampus of male, but not female, rodents via enhancement of new neuron survival in a dose-dependent manner.^{47,111} Thirty days of testosterone or DHT exposure enhances neurogenesis by promoting the survival of new neurons in male rodents.¹¹¹ Interestingly, this effect is not seen with shorter testosterone replacement schedules of 15–21 days,^{106,117} indicating that 30 days of testosterone replacement is necessary to observe increased survival of new neurons in the hippocampus. The influence of testosterone to promote neurogenesis in the hippocampus may be independent of any effects on cell proliferation, as 21 days of testosterone did not influence cell proliferation.¹⁰⁶ However, long-term but not short-term castration in adulthood decreases cell proliferation in rats,^{111,112} suggesting that there may be an influence of longer-term androgen exposure on cell proliferation in the dentate gyrus.

Several lines of evidence support the notion that testosterone exerts its influence on hippocampal neurogenesis by interacting with the AR. First, treatment with estradiol has no significant effect on the survival of new neurons in adult males,¹¹¹ indicating that ER activation is not responsible for neurogenic effects of androgens in males. On the other hand, DHT, which directly stimulates the AR, increased the survival of new neurons, similar to testosterone.^{44,111} The neurogenesis-promoting effect of DHT was blocked in the presence of flutamide, a competitive AR antagonist.⁴⁴ Moreover, chronic testosterone had no effect on the survival of new neurons in rats with a testicular feminization mutation, which lack a functional AR,⁴⁴ indicating that androgens influence neurogenesis through an AR-mediated mechanism. However, considering that testosterone and DHT increase neurogenesis by promoting the survival of new neurons, it is a matter of curiosity that ARs are not located on immature neurons (DCX-expressing cells) in male rats or mice in the dentate gyrus.^{44,114} On the other hand, ARs are expressed in the CA3 region of the hippocampus,^{45–47} which is the axonal projection site of newly



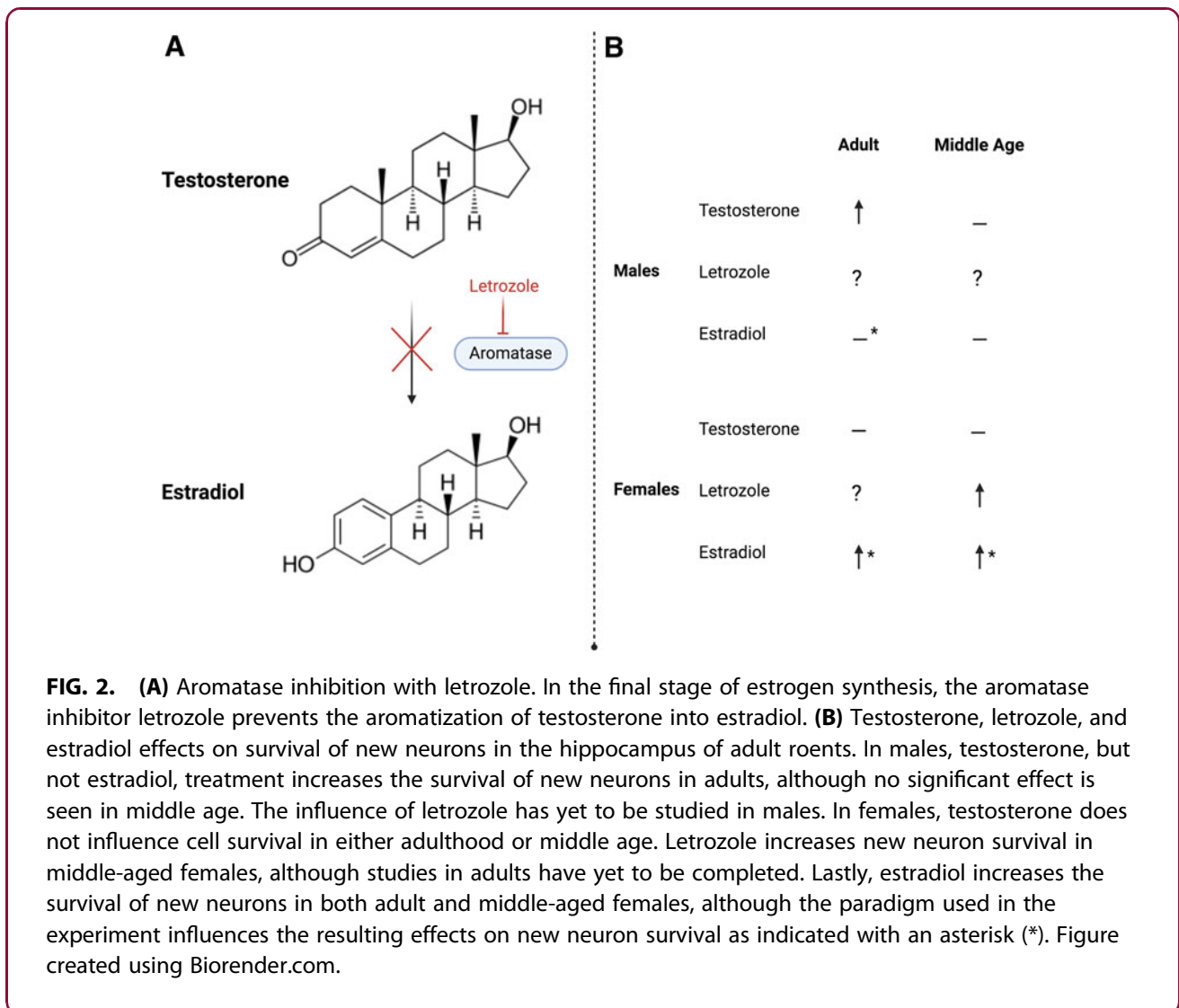
formed neurons.^{118,119} Androgens, via the AR, rapidly increase thorny excrescences in the CA3 region, which are the post-synaptic regions of the mossy fibers.¹²⁰ Therefore, it is possible that the small number of ARs located in the dentate gyrus and/or ARs located in the terminal region for granule cells in the CA3 region are responsible, at least in part, for the AR-dependent promotion of neurogenesis in the hippocampus.

Androgens and Adult Hippocampal Neurogenesis in Females

Although androgens promote neurogenesis in males, 30 days of testosterone or DHT treatment do not affect neurogenesis in young adult or middle-aged female rats.⁴⁷ Although these findings suggest that androgens do not support neurogenesis in females, Chaiton

et al.¹²¹ found that chronic treatment with letrozole, an aromatase inhibitor, increased neurogenesis (cell proliferation and density of immature neurons) in the hippocampus of middle-aged female mice. As letrozole inhibits the conversion of androgens into estradiol (Fig. 2), this result implies that the increase in neurogenesis may be related to the increase in androgens induced by letrozole, and/or the decrease in estradiol synthesis. Although letrozole increased the number of doublecortin cells (immature neurons) of middle-aged females, it suppresses the cell proliferation of cultured neurons from postnatal day 5 female rats,¹²² indicating age differences in the influence of letrozole on neurogenesis in the hippocampus.

In adult females, the influence of estradiol on adult hippocampal neurogenesis is complex, as it is dependent



on dose, sex, age, reproductive experience, and timing of estradiol exposure^{123–127}; for review see Ref.²⁴ As mentioned earlier, the survival of new neurons is enhanced by chronic exposure to testosterone and DHT, but not estradiol, in adult male rats.¹¹¹ However, other studies show that brief exposure to estradiol can increase neurogenesis in male meadow voles when administered during the time when new neurons are extending their axons.¹²⁵ Together, these results demonstrate that sex hormone-mediated regulation of hippocampal neurogenesis is distinct in males compared with females, with more neurogenic effects of androgens in males and more neurogenic effects of estrogens in females.

Age Effects of Androgens and Hippocampal Neurogenesis

Age is an additional factor that influences the effects of androgens on neurogenesis in males. Chronic DHT treatment increased new neuron survival in young, but not middle-aged, adult gonadectomized male rats.⁴⁷ Consistent with this, Moser et al.¹²⁸ found that 12 weeks of testosterone failed to increase neurogenesis (DCX-expressing cells) in both intact middle-aged and aged male rats. These findings are in contrast with the increased neurogenesis observed in young adult males with chronic testosterone or DHT,¹¹¹ suggesting that aging influences the ability of the dentate gyrus to respond to the pro-neurogenic effects of androgen signaling in male rats. This is a matter of curiosity, as ARs are evident in the dentate gyrus of middle-aged rodents, potentially at even higher levels than in young adults.^{47,49} Although AR levels may be higher in aged males compared with young males,^{47,49} it is possible that AR functionality decreases with age. Further, it is noteworthy that long-term castration resulted in loss of granule neurons in the dentate gyrus of adult male rats, which can be ameliorated by early but not late chronic DHT treatment,¹²⁹ indicating a critical window for the neuroplasticity-promoting effect of DHT. Whether or not this critical window exists with respect to DHT's ability to increase adult hippocampal neurogenesis has not yet been studied. However, it is evident that androgens are not promoting neurogenesis in intact or gonadectomized middle-aged animals, but it remains to be determined whether this has to do with dose, critical window, or the functionality of AR with aging.

Here, we concentrate our discussion on adult neurogenesis in the hippocampus, but it is important to consider that androgen exposure during gestation or the

early postpartum period influences neurogenesis of the developing offspring in both sexes.^{130–133} Briefly, mothers exposed to a hyperandrogenic environment during gestation produced offspring with decreased survival of new neurons in the dentate gyrus of both male and female rats.¹³⁰ On the other hand, there is evidence that androgens promote the survival of new neurons during the postpartum period in rats of both sexes.^{132,133} Thus, there are likely distinct mechanisms by which androgens influence neurogenesis during gestation and early development.

Effects of DHEA and Androstenedione on Hippocampal Neurogenesis

Although the majority of studies on androgens and neurogenesis have focused on testosterone and DHT, there is some evidence for the pro-neurogenic effects of DHEA.^{134–137} DHEA has distinct cellular effects from other androgens, such as DHT and testosterone, as DHEA preferentially activates ER β while also demonstrating affinity for ER α and AR^{138–140} along with other nuclear and membrane-bound receptors (reviewed in Prough et al.¹⁴¹). The NSCs derived from the human male fetal cortex demonstrated increased cell survival with DHEA treatment compared with controls.¹³⁷ In adult and middle-aged male rats, DHEA increases the survival of new neurons after repeated or chronic treatment.^{134,135} Short-term treatment (12 days) with DHEA increased the number of new neurons as well as cell proliferation in the dentate gyrus of adult male rats, and pretreatment with DHEA was protective against corticosterone injections, which inhibit neurogenesis.¹³⁵ The same study found that pretreatment with androstenedione had no influence on the inhibition of neurogenesis seen with corticosterone administration¹³⁵ but did not evaluate androstenedione alone. In adult male songbirds, although DHEA increased immature neurons (DCX-expressing cells) in the HVC, a brain region involved in song production, it did not do so in the hippocampus.¹³⁶ Thus, DHEA promotes the survival of new neurons in multiple species, increasing cell proliferation and protecting against stress in adult male rats specifically.

Androgenic Anabolic Steroids and Hippocampal Neurogenesis

Other compounds of interest include androgenic anabolic steroids (AAS) such as the testosterone analogue 19-nortestosterone, also known as nandrolone. Excessive use of AAS has been linked to mood



disturbances¹⁴² such as mania and major depression in humans. On the cellular level, nandrolone elicits its effects in the brain by binding to the AR and repeated administration causes AR upregulation.¹⁴³ A study utilizing rat NSCs in culture and in the dentate gyrus revealed that the administration of nandrolone for 5 days decreased the number of new neurons detected in the dentate gyrus through an AR-mediated mechanism.¹⁴⁴ Another study showed that chronic (28 days) treatment with supraphysiological doses of nandrolone eliminated the strength training-induced enhancement of cell proliferation in the dentate gyrus of adult male Wistar rats.¹⁴⁵ These studies indicate that AAS reduce hippocampal neurogenesis in a manner distinct from that of testosterone and DHT, which may be linked to the supraphysiological doses used or the molecular differences between these androgens.

Androgens and Adult Hippocampal Neurogenesis: Implications for Health and Disease

Androgen treatments in clinical disease may afford neuroprotection via their influence on neurogenesis in the hippocampus.¹⁴⁶ Low levels of androgens are associated with an increased risk for stroke, cerebrovascular disease, major depressive disorders, and dementia.^{147–150} Androgens have been used to treat these diseases with some success, which depends on a variety of factors.¹⁴⁶ In addition, androgens play a role in neuroprotective factors that boost neurogenesis, such as exercise. Exercise-induced neurogenesis is inhibited in the presence of the AR antagonist flutamide, and mild exercise stimulates DHT production in male rats, which, in turn, enhances neurogenesis in the hippocampus.¹⁵¹ Thus, androgens may be involved in some of the neurogenic responses to environmental factors and demonstrate neuroprotective effects against injury and disease.

Ischemic stroke is associated with impairments in memory along with increased neurogenesis,¹⁵² but as with seizure-induced neurogenesis in the hippocampus⁷⁶ these new neurons form aberrant connections.¹⁵² Indeed, the reduction of post-stroke neurogenesis aids in the retention of memory formation. Intriguingly, although castration and flutamide did not alter hippocampal neurogenesis 1 week after stroke, supraphysiological doses of testosterone and DHT reduced post-stroke hippocampal neurogenesis in the same study.¹⁵³ Thus, one way that androgens may afford protection in the face of stroke is to reduce ischemia-induced neurogenesis in the

hippocampus that may be related to aberrant connections tied to poorer memory outcomes.

Androgens play an important role in reducing depression in hypogonadal males, as meta-analyses demonstrate that testosterone treatment in hypogonadal men effectively reduces the symptoms of depression.^{146,154,155} Castration increased the susceptibility to depressive-like endophenotypes in the face of chronic unpredictable stress,¹¹² which was commensurate with reduced neurogenesis in the hippocampus. However, testosterone treatment did not rescue the decrease in cell proliferation observed under social isolation stress in castrated males.^{117,156} Interestingly, testosterone and the tricyclic antidepressant imipramine increased cell proliferation in the hippocampus more than antidepressant treatment alone in response to social isolation stress, an effect observed in males but not females.¹⁵⁶ Further, testosterone in conjunction with imipramine increased polysialylated neuronal cell adhesion molecule (PSA-NCAM)-ir cells in the dentate gyrus, but not neurogenesis *per se*, after exposure to chronic unpredictable stress.¹⁰⁶ In another model of depression using olfactory bulbectomy, the androgen DHEA prevented depressive-like behavior and increased the number of 1-week-old new neurons in response to bulbectomy in adult male mice.¹⁵⁷ DHEA has antidepressant effects¹⁵⁷ and works synergistically with fluoxetine, a selective serotonin reuptake inhibitor, causing an otherwise ineffective dose of fluoxetine to increase cell proliferation in the dentate gyrus of adult male rats.¹⁵⁸ Thus, collectively these data demonstrate that androgens can rescue depressive-like behavior and facilitate antidepressant efficacy, perhaps via its ability to stimulate neurogenesis.

In epilepsy, androgens can influence seizure susceptibility in males.^{159–163} In males with epilepsy, lower levels of free testosterone and sexual dysfunction can be observed¹⁶⁴ but the directionality of this effect and whether treatments for epilepsy compound this relationship are not clear.^{164,165} However, in animal studies, castrated rodents are more susceptible to pentylenetetrazol, picrotoxin, and perforant pathway stimulation-induced seizures compared with intact males, and testosterone administration decreases seizure activity in castrated males.^{159–161} There is also evidence to suggest that the testosterone metabolite 3 α -diol is protective against gamma-aminobutyric acid (GABA)_A receptor antagonist-induced seizures in male mice,^{162,163} which is interesting as 3 α -diol has a weak affinity for ARs.⁴¹ However, the relationship between sex steroid



hormones and epilepsy has proven difficult to discern as various factors such as experimental conditions and biological variability can greatly influence study outcomes (reviewed in Scharfman and MacLusky¹⁶⁶). Although the mechanisms of androgens' influence on seizure activity are not yet clear, it should be recalled that seizures induce aberrant hippocampal neurogenesis^{75,77}; thus, it is plausible that androgens may be protective via their effects on neurogenesis in the dentate gyrus to reduce the aberrant connectivity of new neurons.

Finally, androgens may play a protective role in neurodegenerative diseases such as Alzheimer's disease (AD); for review see Ref.¹⁶⁷ Low levels of free testosterone may be a risk factor for AD in males,^{168–170} and males with AD exhibit lower levels of testosterone in the periphery and in the brain compared with age-matched controls.^{149,171,172} Two copies of the *APOEε4* allele confer a greater risk to develop sporadic late-onset AD^{173,174} and female *APOEε4* carriers have disproportionately increased phosphorylated Tau (p-Tau), a neuropathological feature of AD, compared with male carriers.¹⁷⁵ Further, low serum testosterone in both males and females corresponded to increased p-Tau,¹⁷⁶ suggesting that the low levels of testosterone in both sexes may confer a greater risk of neuropathology related to AD. Testosterone treatment given to males with AD or mild cognitive impairment, a prodromal state to AD, increased scores on certain forms of memory, including spatial memory.¹⁷⁷ Thus, androgens may improve certain types of memory in both sexes with age and given that the hippocampus undergoes early degeneration with AD, it is possible that androgens may promote cognition during aging and in AD, via its effects on hippocampal neurogenesis. Intriguingly, neurogenesis in the hippocampus is decreased in people with AD when compared with healthy aging controls.^{12,13,178} In addition, low testosterone coupled with *APOEε4* genotype was related to lower hippocampal volume¹⁷⁹ and poorer verbal episodic memory.¹⁸⁰ Thus, several studies suggest that the androgens are related to hippocampal structure and function and more work is needed to explore the relationship between androgens and neurogenesis in AD and aging.

Conclusion

Chronic but not acute exposure to a variety of androgens generally increases neurogenesis in the hippocampus of adult males, an effect not seen in females or in middle-aged males. Despite the progress in this field,

it is currently not known how or where ARs work to promote neurogenesis in the dentate gyrus with chronic testosterone or DHT, and whether androgen-induced new neurons are contributing to the function of the dentate gyrus. The addition of new neurons in the dentate gyrus has functional implications, but improper integration of new neurons may be deleterious to the function of these circuits. In addition, environmental influences such as central nervous system injury, disease, stress, and exercise all play a role in the neurogenic response to androgens. Sex differences observed in the neurogenic response to sex steroid hormones indicate that males and females have distinct hormonal mechanisms for the control of neurogenesis, and further work should explore the benefits of androgens to influence neurogenesis in males and possibly females, particularly in relation to health and disease.

Authors' Contributions

S.A.B. and L.A.M.G. co-wrote the review.

Author Disclosure Statement

No competing financial interests exist.

Funding Information

This review was funded by the Natural Sciences and Engineering Research Council of Canada (NSERC) grant to L.A.M.G. (2018-04301) and a Canadian Institutes of Health Research grant to L.A.M.G. (MOP142308) and a CIHR PGS-M scholarship to S.A.B.

References

1. Altman J, Das GD. Autoradiographic and histological evidence of postnatal hippocampal neurogenesis in rats. *J Comp Neurol.* 1965;124(3):319–335.
2. Raisman G. Neuronal plasticity in the septal nuclei of the adult rat. *Brain Res.* 1969;14(1):25–48.
3. Van Harrevelde A, Fifkova E. Swelling of dendritic spines in the fascia dentata after stimulation of the perforant fibers as a mechanism of post-tetanic potentiation. *Exp Neurol.* 1975;49(3):736–749.
4. Greenough W, West R, DeVoogd T. Subsynaptic plate perforations: Changes with age and experience in the rat. *Science.* 1978;202(4372):1096–1098.
5. Bosch M, Hayashi Y. Structural plasticity of dendritic spines. *Curr Opin Neurobiol.* 2012;22(3):383–388.
6. Goldman SA, Nottebohm F. Neuronal production, migration, and differentiation in a vocal control nucleus of the adult female canary brain. *Proc Natl Acad Sci.* 1983;80(8):2390–2394.
7. Nottebohm F. Neuronal replacement in adulthood. *Ann N Y Acad Sci.* 1985;457(1):143–161.
8. Kornack DR, Rakic P. Continuation of neurogenesis in the hippocampus of the adult macaque monkey. *Proc Natl Acad Sci.* 1999;96(10):5768–5773.
9. Ngwenya LB, Peters A, Rosene DL. Maturation sequence of newly generated neurons in the dentate gyrus of the young adult rhesus monkey. *J Comp Neurol.* 2006;498(2):204–216.
10. Maslov AY, Barone TA, Plunkett RJ, Pruitt SC. Neural stem cell detection, characterization, and age-related changes in the subventricular zone of mice. *J Neurosci.* 2004;24(7):1726–1733.



11. Kempermann G, Gast D, Kronenberg G, Yamaguchi M, Gage FH. Early determination and long-term persistence of adult-generated new neurons in the hippocampus of mice. *Development*. 2003;130(2):391–399.
12. Moreno-Jiménez EP, Flor-García M, Terreros-Roncal J, et al. Adult hippocampal neurogenesis is abundant in neurologically healthy subjects and drops sharply in patients with Alzheimer's disease. *Nat Med*. 2019; 25:554–560.
13. Tobin MK, Musaraca K, Disouky A, et al. Human hippocampal neurogenesis persists in aged adults and Alzheimer's disease patients. *Cell Stem Cell*. 2019;24(6):974–982.
14. Eriksson PS, Perfilieva E, Björk-Eriksson T, et al. Neurogenesis in the adult human hippocampus. *Nat Med*. 1998;4:1313–1317.
15. Mathews KJ, Allen KM, Boerrigter D, Ball H, Shannon Weickert C, Double KL. Evidence for reduced neurogenesis in the aging human hippocampus despite stable stem cell markers. *Aging Cell*. 2017;16(5):1195–1199.
16. Knoth R, Singec I, Ditter M, et al. Murine features of neurogenesis in the human hippocampus across the lifespan from 0 to 100 years. *PLoS One*. 2010;5:e8809.
17. Boldrini M, Fulmore CA, Tartt AN, et al. Human hippocampal neurogenesis persists throughout aging. *Cell Stem Cell*. 2018;22(4):589–599.
18. Spalding KL, Bergmann O, Alkass K, et al. Dynamics of hippocampal neurogenesis in adult humans. *Cell*. 2013;153(6):1219–1227.
19. Galea LAM, McEwen BS. Sex and seasonal changes in the rate of cell proliferation in the dentate gyrus of adult wild meadow voles. *Neuroscience*. 1999;89(3):955–964.
20. Barnea A, Nottebohm F. Seasonal recruitment of hippocampal neurons in adult free-ranging black-capped chickadees. *Proc Natl Acad Sci U S A*. 1994;91(23):11217–11221.
21. Yagi S, Splinter JEJ, Tai D, Wong S, Wen Y, Galea LAM. Sex differences in maturation and attrition of adult neurogenesis in the hippocampus. *eNeuro*. 2020;7(4):1–14.
22. Falconer EM, Galea LAM. Sex differences in cell proliferation, cell death and defensive behavior following acute predator odor stress in adult rats. *Brain Res*. 2003;975(1–2):22–36.
23. Westenbroek C, Den Boer JA, Veenhuis M, Ter Horst GJ. Chronic stress and social housing differentially affect neurogenesis in male and female rats. *Brain Res Bull*. 2004;64(4):303–308.
24. Mahmoud R, Wainwright SR, Galea LAM. Sex hormones and adult hippocampal neurogenesis: Regulation, implications, and potential mechanisms. *Front Neuroendocrinol*. 2016;41:129–152.
25. Fortune JE, Armstrong DT. Androgen production by theca and granulosa isolated from proestrous rat follicles. *Endocrinology*. 1977;100(5):1341–1347.
26. Neville AM, O'Hare MJ. Functional activity of the adrenal cortex. In: *The Human Adrenal Cortex*. Berlin: Germany, Springer-Verlag. 1982; pp 68–98.
27. Corpechot C, Robel P, Axelson M, Sjoval J, Baulieu EE. Characterization and measurement of dehydroepiandrosterone sulfate in rat brain. *Proc Natl Acad Sci*. 1981;78:4704–4707.
28. Robel P, Synguelakis M, Halberg F, Baulieu EE. Persistence of the circadian rhythm of dehydroepiandrosterone in the brain, but not in the plasma, of castrated and adrenalectomized rats. *C R Acad Sci III*. 1986; 303(6):235–238.
29. Baulieu E-E, Robel P, Vatier O, Haug M, Le Goascogne C, Bourreau E. Neurosteroids: Pregnenolone and dehydroepiandrosterone in the brain. In: *Receptor-Receptor Interactions*. London: Palgrave Macmillan. 1987; pp 89–104.
30. Mukai H, Takata N, Ishii H-t, et al. Hippocampal synthesis of estrogens and androgens which are paracrine modulators of synaptic plasticity. *Synaptocrinology*. *Neuroscience*. 2006;138:757–764.
31. Caruso D, Pesaresi M, Abbiati F, et al. Comparison of plasma and cerebrospinal fluid levels of neuroactive steroids with their brain, spinal cord and peripheral nerve levels in male and female rats. *Psychoneuroendocrinology*. 2013;38:2278–2290.
32. Jalabert C, Ma C, Soma KK. Profiling of systemic and brain steroids in male songbirds: Seasonal changes in neurosteroids. *J Neuroendocrinol*. 2020;33(1):e12922.
33. Handelsman DJ. Androgen physiology, pharmacology, and abuse. *Endocrinology*. 2010:2469–2498.
34. McHenry J, Carrier N, Hull E, Kabbaj M. Sex differences in anxiety and depression: Role of testosterone. *Front Neuroendocrinol*. 2014;35(1): 42–57.
35. Handa RJ, Pak TR, Kudwa AE, Lund TD, Hinds L. An alternate pathway for androgen regulation of brain function: Activation of estrogen receptor beta by the metabolite of dihydrotestosterone, 5 α -androstane-3 β ,17 β -diol. *Horm Behav*. 2008;53(5):741–752.
36. Kuiper GG, Lemmen JG, Carlsson B, et al. Interaction of estrogenic chemicals and phytoestrogens with estrogen receptor β . *Endocrinology*. 1998;139(10):4252–4263.
37. Bennett NC, Gardiner RA, Hooper JD, Johnson DW, Gobe GC. Molecular cell biology of androgen receptor signalling. *Int J Biochem Cell Biol*. 2010;42(6):813–827.
38. Gatsson JW, Singh M. Activation of a membrane-associated androgen receptor promotes cell death in primary cortical astrocytes. *Endocrinology*. 2007;148(5):2458–2464.
39. Revankar CM, Cimino DF, Sklar LA, Arterburn JB, Prossnitz ER. A transmembrane intracellular estrogen receptor mediates rapid cell signaling. *Science*. 2005;307(5715):1625–1630.
40. Cutress ML, Whitaker HC, Mills IG, Stewart M, Neal DE. Structural basis for the nuclear import of the human androgen receptor. *J Cell Sci*. 2008;121: 957–968.
41. Matsuda KI, Sakamoto H, Mori H, et al. Expression and intracellular distribution of the G protein-coupled receptor 30 in rat hippocampal formation. *Neurosci Lett*. 2008;441(1):94–99.
42. Mehra RD, Sharma K, Nyakas C, Vij U. Estrogen receptor α and β immunoreactive neurons in normal adult and aged female rat hippocampus: A qualitative and quantitative study. *Brain Res*. 2005;1056(1):22–35.
43. Solum DT, Handa RJ. Localization of estrogen receptor alpha (ER α) in pyramidal neurons of the developing rat hippocampus. *Dev Brain Res*. 2001;128(2):165–175.
44. Hamson DK, Wainwright SR, Taylor JR, Jones BA, Watson NV, Galea LA. Androgens increase survival of adult-born neurons in the dentate gyrus by an androgen receptor-dependent mechanism in male rats. *Endocrinology*. 2013;154(9):3294–3304.
45. Simerly RB, Swanson LW, Chang C, Muramatsu M. Distribution of androgen and estrogen receptor mRNA-containing cells in the rat brain: An *in situ* hybridization study. *J Comp Neurol*. 1990;294(1): 76–95.
46. Tabori NE, Stewart LS, Znamensky V, et al. Ultrastructural evidence that androgen receptors are located at extranuclear sites in the rat hippocampal formation. *Neuroscience*. 2005;130(1):151–163.
47. Duarte-Guterman P, Lieblich SE, Wainwright SR, et al. Androgens enhance adult hippocampal neurogenesis in males but not females in an age-dependent manner. *Endocrinology*. 2019;160(9):2128–2136.
48. Xiao L, Jordan CL. Sex differences, laterality, and hormonal regulation of androgen receptor immunoreactivity in rat hippocampus. *Horm Behav*. 2002;42(3):327–336.
49. Wu D, Lin G, Gore AC. Age-related changes in hypothalamic androgen receptor and estrogen receptor α in male rats. *J Comp Neurol*. 2009; 512(5):688–701.
50. Edwards A, Hammond HA, Jin L, Caskey CT, Chakraborty R. Genetic variation at five trimeric and tetrameric tandem repeat loci in four human population groups. *Genomics*. 1992;12(2):241–253.
51. Chamberlain NL, Driver ED, Miesfeld RL. The length and location of CAG trinucleotide repeats in the androgen receptor N-terminal domain affect transactivation function. *Nucleic Acids Res*. 1994;22(15):3181–3186.
52. Braun SM, Jessberger S. Adult neurogenesis: Mechanisms and functional significance. *Development*. 2014;141(10):1983–1986.
53. Kee N, Sivalingam S, Boonstra R, Wojtowicz JM. The utility of Ki-67 and BrdU as proliferative markers of adult neurogenesis. *J Neurosci Methods*. 2002;115(1):97–105.
54. Brown JP, Couillard-Després S, Cooper-Kuhn CM, Winkler J, Aigner L, Kuhn HG. Transient expression of doublecortin during adult neurogenesis. *J Comp Neurol*. 2003;467(1):1–10.
55. Snyder JS, Choe JS, Clifford MA, et al. Adult-born hippocampal neurons are more numerous, faster maturing, and more involved in behavior in rats than in mice. *J Neurosci*. 2009;29(46):14484–14495.
56. Taupin P. BrdU immunohistochemistry for studying adult neurogenesis: Paradigms, pitfalls, limitations, and validation. *Brain Res Rev*. 2007;53(1): 198–214.
57. von Bohlen und Halbach O. Immunohistological markers for proliferative events, gliogenesis, and neurogenesis within the adult hippocampus. *Cell Tissue Res*. 2011;345:1–19.



58. Sidman RL, Miale IL, Feder N. Cell proliferation and migration in the primitive ependymal zone; An autoradiographic study of histogenesis in the nervous system. *Exp Neurol*. 1959;1(4):322–333.
59. Gratzner H. Monoclonal antibody to 5-bromo- and 5-iododeoxyuridine: A new reagent for detection of DNA replication. *Science*. 1982; 218(4571):474–475.
60. Miller MW, Nowakowski RS. Use of bromodeoxyuridine-immunohistochemistry to examine the proliferation, migration and time of origin of cells in the central nervous system. *Brain Res*. 1988;457(1):44–52.
61. Galea LA, Wainwright SR, Roes MM, Duarte-Guterman P, Chow C, Hamson DK. Sex, Hormones and neurogenesis in the hippocampus: Hormonal modulation of neurogenesis and potential functional implications. *J Neuroendocrinol*. 2013;25(11):1039–1061.
62. Wojtowicz JM, Kee N. BrdU assay for neurogenesis in rodents. *Nat Protoc*. 2006;1:1399–1405.
63. Seaberg RM, van der Kooy D. Adult rodent neurogenic regions: The ventricular subependyma contains neural stem cells, but the dentate gyrus contains restricted progenitors. *J Neurosci*. 2002;22(5):1784–1793.
64. Bull ND, Bartlett, PF. The adult mouse hippocampal progenitor is neurogenic but not a stem cell. *J Neurosci*. 2005;25(47):10815–10821.
65. Seri B, García-Verdugo Jose Manuel, McEwen BS, Alvarez-Buylla A. Astrocytes give rise to new neurons in the adult mammalian hippocampus. *J Neurosci*. 2001;21(18):7153–7160.
66. Steiner B, Klempin F, Wang L, Kott M, Kettenmann H, Kempermann G. Type-2 cells as link between glial and neuronal lineage in adult hippocampal neurogenesis. *Glia*. 2006;54(8):805–814.
67. Lagace DC, Whitman MC, Noonan MA, et al. Dynamic contribution of nestin-expressing stem cells to adult neurogenesis. *J Neurosci*. 2007; 27(46):12623–12629.
68. Kempermann G, Wiskott L, Gage FH. Functional significance of adult neurogenesis. *Curr Opin Neurobiol*. 2004;14(2):186–191.
69. Cameron HA, Woolley CS, McEwen BS, Gould E. Differentiation of newly born neurons and glia in the dentate gyrus of the adult rat. *Neuroscience*. 1993;56(2):337–344.
70. Seaberg RM, van der Kooy D. Stem and progenitor cells: The premature desertion of rigorous definitions. *Trends Neurosci*. 2003;26(3):125–131.
71. Sorrells SF, Paredes MF, Cebrian-Silla A, et al. Human hippocampal neurogenesis drops sharply in children to undetectable levels in adults. *Nature*. 2018;555:377–381.
72. Cipriani S, Ferrer I, Aronica E, et al. Hippocampal radial glial subtypes and their neurogenic potential in human fetuses and healthy and Alzheimer's disease adults. *Cereb Cortex*. 2018;28(7):2458–2478.
73. Ponti G, Farinetti A, Marraudino M, Panzica GC, Gotti S. Sex steroids and adult neurogenesis in the ventricular-subventricular zone. *Front Endocrinol*. 2018;9:156.
74. Peretto P, Paredes RG. Frontiers in neurosciencesocial cues, adult neurogenesis, and reproductive behavior. In: *Neurobiology of Chemical Communication* (Mucignat-Caretta C, ed). Boca Raton, FL: CRC Press, Taylor & Francis Group. 2014; pp 367–383.
75. Jessberger S, Zhao C, Toni N, Clemenson GD, Li Y, Gage FH. Seizure-associated, aberrant neurogenesis in adult rats characterized with retrovirus-mediated cell labeling. *J Neurosci*. 2007;27(35):9400–9407.
76. Jakubs K, Nanobashvili A, Bonde S, et al. Environment matters: Synaptic properties of neurons born in the epileptic adult brain develop to reduce excitability. *Neuron*. 2006;52(6):1047–1059.
77. Parent JM, Yu TW, Leibowitz RT, Geschwind DH, Sloviter RS, Lowenstein DH. Dentate granule cell neurogenesis is increased by seizures and contributes to aberrant network reorganization in the adult rat hippocampus. *J Neurosci*. 1997;17(10):3727–3738.
78. Houser CR. Granule cell dispersion in the dentate gyrus of humans with temporal lobe epilepsy. *Brain Res*. 1990;535(2):195–204.
79. van Praag H, Kempermann G, Gage FH. Running increases cell proliferation and neurogenesis in the adult mouse dentate gyrus. *Nat Neurosci*. 1999a;2:266–270.
80. van Praag H, Christie BR, Sejnowski TJ, Gage FH. Running enhances neurogenesis, learning, and long-term potentiation in mice. *Proc Natl Acad Sci*. 1999b;96(23):13427–13431.
81. Epp JR, Silva Mera R, Köhler S, Josselyn SA, Frankland PW. Neurogenesis-mediated forgetting minimizes proactive interference. *Nat Comm*. 2016; 7:10838.
82. Cameron HA, Christie BR. Do new neurons have a functional role in the adult hippocampus? *Debates Neurosci*. 2007;1:26–32.
83. Kempermann G. Why new neurons? Possible functions for adult hippocampal neurogenesis. *J Neurosci*. 2002;22(3):635–638.
84. Christian KM, Song H, Ming G-li. Functions and dysfunctions of adult hippocampal neurogenesis. *Annu Rev Neurosci*. 2014;37:243–262.
85. Zhao C, Deng W, Gage FH. Mechanisms and functional implications of adult neurogenesis. *Cell*. 2008;132(4):645–660.
86. Winocur G, Becker S, Luu P, Rosenzweig S, Wojtowicz JM. Adult hippocampal neurogenesis and memory interference. *Behav Brain Res*. 2012; 227(2):464–469.
87. Leutgeb JK, Leutgeb S, Moser M-B, Moser EI. Pattern separation in the dentate gyrus and CA3 of the hippocampus. *Science*. 2007;315(5814): 961–966.
88. Clelland CD, Choi M, Romberg C, Clemenson GD, Fragniere A, Tyers P, et al. A functional role for adult hippocampal neurogenesis in spatial pattern separation. *Science*. 2009;325(5937):210–213.
89. Tronel S, Belnoue L, Grosjean N, et al. Adult-born neurons are necessary for extended contextual discrimination. *Hippocampus*. 2010;22(2):292–298.
90. França TF, Bitencourt AM, Maximilla NR, Barros DM, Monserrat JM. Hippocampal neurogenesis and pattern separation: A meta-analysis of behavioral data. *Hippocampus*. 2017;27(9):937–950.
91. Kesner RP, Hui X, Sommer T, Wright C, Barrera VR, Fanselow MS. The role of postnatal neurogenesis in supporting remote memory and spatial metric processing. *Hippocampus*. 2014;24(12):1663–1671.
92. Danielson NB, Kaifosh P, Zaremba JD, et al. Distinct contribution of adult-born hippocampal granule cells to context encoding. *Neuron*. 2016;90(1):101–112.
93. Treves A, Tashiro A, Witter MP, Moser EI. What is the mammalian dentate gyrus good for? *Neuroscience*. 2008;154(4):1155–1172.
94. Saxe MD, Battaglia F, Wang J-W, et al. Ablation of hippocampal neurogenesis impairs contextual fear conditioning and synaptic plasticity in the dentate gyrus. *Proc Natl Acad Sci*. 2006;103(46):17501–17506.
95. Sahay A, Scobie KN, Hill AS, et al. Increasing adult hippocampal neurogenesis is sufficient to improve pattern separation. *Nature*. 2011;472: 466–470.
96. Shors TJ, Townsend DA, Zhao M, Kozorovitskiy Y, Gould E. Neurogenesis may relate to some but not all types of hippocampal-dependent learning. *Hippocampus*. 2002;12(5):578–584.
97. Winocur G, Wojtowicz JM, Sekeres M, Snyder JS, Wang S. Inhibition of neurogenesis interferes with hippocampus-dependent memory function. *Hippocampus*. 2006;16(3):296–304.
98. Miller SM, Sahay A. Functions of adult-born neurons in hippocampal memory interference and indexing. *Nat Neurosci*. 2019;22:1565–1575.
99. Anacker C, Hen R. Adult hippocampal neurogenesis and cognitive flexibility—Linking memory and mood. *Nat Rev Neurosci*. 2017;18:335–346.
100. Roberts GW, Woodhams PL, Polak JM, Crow TJ. Distribution of neuropeptides in the limbic system of the rat: The hippocampus. *Neuroscience*. 1984;11(1):35–77.
101. Fanselow MS, Dong H-W. Are the dorsal and ventral hippocampus functionally distinct structures? *Neuron*. 2010;65(1):7–19.
102. Bannerman DM, Sprengel R, Sanderson DJ, et al. Hippocampal synaptic plasticity, spatial memory and anxiety. *Nat Rev Neurosci*. 2014;15:181–192.
103. Snyder JS, Soumier A, Brewer M, Pickel J, Cameron HA. Adult hippocampal neurogenesis buffers stress responses and depressive behaviour. *Nature*. 2011;476:458–461.
104. Eid RS, Gobinath AR, Galea LAM. Sex differences in depression: Insights from clinical and preclinical studies. *Prog Neurobiol*. 2019; 176:86–102.
105. Tanti A, Westphal W-P, Girault V, et al. Region-dependent and stage-specific effects of stress, environmental enrichment, and antidepressant treatment on hippocampal neurogenesis. *Hippocampus*. 2013;23(9): 797–811.
106. Wainwright SR, Workman JL, Tehrani A, et al. Testosterone has antidepressant-like efficacy and facilitates imipramine-induced neuroplasticity in male rats exposed to chronic unpredictable stress. *Horm Behav*. 2016;79:58–69.
107. Boldrini M, Underwood MD, Hen R, et al. Antidepressants increase neural progenitor cells in the human hippocampus. *Neuropsychopharmacology*. 2009;34:2376–2389.
108. Epp JR, Beasley CL, Galea LAM. Increased hippocampal neurogenesis and p21 expression in depression: Dependent on antidepressants, sex,



- age, and antipsychotic exposure. *Neuropsychopharmacology*. 2013;38:2297–2306.
109. Surget A, Tanti A, Leonardo ED, et al. Antidepressants recruit new neurons to improve stress response regulation. *Mol Psychiatry*. 2011;16:1177–1188.
110. Ormerod BK, Galea LAM. Reproductive status influences the survival of new cells in the dentate gyrus of adult male meadow voles. *Neurosci Lett*. 2003;346(1–2):25–28.
111. Spritzer MD, Galea LAM. Testosterone and dihydrotestosterone, but not estradiol, enhance survival of new hippocampal neurons in adult male rats. *Dev Neurobiol*. 2007;67(10):1321–1333.
112. Wainwright SR, Lieblich SE, Galea LAM. Hypogonadism predisposes males to the development of behavioural and neuroplastic depressive phenotypes. *Psychoneuroendocrinology*. 2011;36(9):1327–1341.
113. Benice TS, Raber J. Castration and training in a spatial task alter the number of immature neurons in the hippocampus of male mice. *Brain Res*. 2010;1329:21–29.
114. Swift-Gallant A, Duarte-Guterman P, Hamson DK, Ibrahim M, Monks DA, Galea LA. Neural androgen receptors affect the number of surviving new neurons in the adult dentate gyrus of male mice. *J Neuroendocrinol*. 2018;30(4):e12578.
115. Allen KM, Fung SJ, Rothmond DA, Noble PL, Shannon Weickert C. Gonadectomy increases neurogenesis in the male adolescent rhesus macaque hippocampus. *Hippocampus*. 2013;24(2):225–238.
116. Allen KM, Purves-Tyson TD, Fung SJ, Shannon Weickert C. The effect of adolescent testosterone on hippocampal BDNF and TrkB mRNA expression: Relationship with cell proliferation. *BMC Neurosci*. 2015;16:4.
117. Spritzer MD, Ibler E, Inglis W, Curtis MG. Testosterone and social isolation influence adult neurogenesis in the dentate gyrus of male rats. *Neuroscience*. 2011;195:180–190.
118. Markakis EA, Gage FH. Adult-generated neurons in the dentate gyrus send axonal projections to field CA3 and are surrounded by synaptic vesicles. *J Comp Neurol*. 1999;406:449–460.
119. Hastings NB, Gould E. Rapid extension of axons into the CA3 region by adult-generated granule cells. *J Comp Neurol*. 1999;413:146–154.
120. Hatanaka Y, Mukai H, Mitsuhashi K, et al. Androgen rapidly Increases Dendritic thorns of ca3 neurons in male rat hippocampus. *Biochem Biophys Res Comm*. 2009;381:728–732.
121. Chaiton JA, Wong SJ, Galea LAM. Chronic aromatase inhibition increases ventral hippocampal neurogenesis in middle-aged female mice. *Psychoneuroendocrinology*. 2019;106:111–116.
122. Fester L, Ribeiro-Gouveia V, Prange-Kiel J, von Schassen C, Bottner M, Jarry H, et al. Proliferation and apoptosis of hippocampal granule cells require local oestrogen synthesis. *J Neurochem*. 2006;97(4):1136–1144.
123. Barker JM, Galea LAM. Repeated estradiol administration alters different aspects of neurogenesis and cell death in the hippocampus of female, but not male, rats. *Neuroscience*. 2008;152(4):888–902.
124. Tanapat P, Hastings NB, Gould E. Ovarian steroids influence cell proliferation in the dentate gyrus of the adult female rat in a dose- and time-dependent manner. *J Comp Neurol*. 2004;481(3):252–265.
125. Ormerod BK, Lee TT-Y, Galea LAM. Estradiol enhances neurogenesis in the dentate gyri of adult male meadow voles by increasing the survival of young granule neurons. *Neuroscience*. 2004;128(3):645–654.
126. Chiba S, Suzuki M, Yamanouchi K, Nishihara M. Involvement of granulin in estrogen-induced neurogenesis in the adult rat hippocampus. *J Reprod Dev*. 2007;53(2):297–307.
127. Barha CK, Galea LAM. Motherhood alters the cellular response to estrogens in the hippocampus later in life. *Neurobiol Aging*. 2011;32(11):2091–2095.
128. Moser VA, Christensen A, Liu J, et al. Effects of aging, high-fat diet, and testosterone treatment on neural and metabolic outcomes in male brown Norway rats. *Neurobiol Aging*. 2019;73:145–160.
129. Ramsden M, Nyborg AC, Murphy MP, et al. Androgens modulate β -amyloid levels in male rat brain. *J Neurochem*. 2003;87(4):1052–1055.
130. Cheng J, Wu H, Liu H, et al. Exposure of hyperandrogen during pregnancy causes depression- and anxiety-like behaviors, and reduced hippocampal neurogenesis in rat offspring. *Front Neurosci*. 2019;13:436.
131. Kight KE, McCarthy MM. Androgens and the developing hippocampus. *Biol Sex Differ*. 2020;11(30):1–14.
132. Waddell J, Bowers JM, Edwards NS, Jordan CL, McCarthy MM. Dysregulation of neonatal hippocampal cell genesis in the androgen insensitive Tfm rat. *Horm Behav*. 2013;64(1):144–152.
133. Zhang J-M, Konkle AT, Zup SL, McCarthy MM. Impact of sex and hormones on new cells in the developing rat hippocampus: A novel source of sex dimorphism? *Eur J Neurosci*. 2008;27(4):791–800.
134. Herrera-Pérez JJ, Martínez-Mota L, Jiménez-Rubio G, et al. Dehydroepiandrosterone increases the number and dendrite maturation of doublecortin cells in the dentate gyrus of middle age male Wistar rats exposed to chronic mild stress. *Behav Brain Res*. 2017;321:137–147.
135. Karishma KK, Herbert J. Dehydroepiandrosterone (DHEA) stimulates neurogenesis in the hippocampus of the rat, promotes survival of newly formed neurons and prevents corticosterone-induced suppression. *Eur J Neurosci*. 2002;16(3):445–453.
136. Wada H, Newman AEM, Hall ZJ, Soma KK, MacDougall-Shackleton SA. Effects of corticosterone and DHEA on doublecortin immunoreactivity in the song control system and hippocampus of adult song sparrows. *Dev Neurobiol*. 2013;74(1):52–62.
137. Suzuki M, Wright LS, Marwah P, Lardy HA, Svendsen CN. Mitotic and neurogenic effects of dehydroepiandrosterone (DHEA) on human neural stem cell cultures derived from the fetal cortex. *Proc Natl Acad Sci*. 2004;101(9):3202–3207.
138. Chen F, Knecht K, Birzin E, et al. Direct agonist/antagonist functions of dehydroepiandrosterone. *Endocrinology*. 2005;146:4568–4576.
139. Bruder JM, Sobek L, Oettel M. Dehydroepiandrosterone stimulates the estrogen response element. *J Steroid Biochem Mol Biol*. 1997;62:461–466.
140. Lu S-F, Mo Q, Hu S, Garippa C, Simon NG. Dehydroepiandrosterone upregulates neural androgen receptor level and transcriptional activity. *J Neurobiol*. 2003;57:163–171.
141. Prough RA, Clark BJ, Klinge CM. Novel mechanisms for dhea action. *J Mol Endocrinol*. 2016;56(3), R139–R155.
142. Pope HG, Katz DL. Psychiatric and medical effects of anabolic-androgenic steroid use. *Arch Gen Psychiatr*. 1994;51(5):375.
143. Menard CS, Harlan RE. Up-regulation of androgen receptor immunoreactivity in the rat brain by androgenic-anabolic steroids. *Brain Res*. 1993;622(1–2):226–236.
144. Brännvall K, Bogdanovic N, Korhonen L, Lindholm D. 19-Nortestosterone influences neural stem cell proliferation and neurogenesis in the rat brain. *Eur J Neurosci*. 2005;21(4):871–878.
145. Novaes Gomes FG, Fernandes J, Vannucci Campos D, et al. The beneficial effects of strength exercise on hippocampal cell proliferation and apoptotic signaling is impaired by anabolic androgenic steroids. *Psychoneuroendocrinology*. 2014;50:106–117.
146. Amanatou HR, Chibnall JT, Seo BW, Manepalli JN, Grossberg GT. Impact of exogenous testosterone on mood: A systematic review and meta-analysis of randomized placebo-controlled trials. *Ann Clin Psychiatry*. 2014;26(1):19–32.
147. Abi-Ghanem C, Robison LS, Zuloaga KL. Androgens' effects on cerebrovascular function in health and disease. *Biol Sex Differ*. 2020;11:35.
148. Zitzmann M. Testosterone, mood, behaviour and quality of life. *Andrology*. 2020;8(6):1598–1605.
149. Rosario ER. Age-related testosterone depletion and the development of Alzheimer disease. *JAMA*. 2004;292(12):1431–1432.
150. Jeppesen LL, Jørgensen HS, Nakayama H, Raaschou HO, Olsen TS, Winther K. Decreased serum testosterone in men with acute ischemic stroke. *Arterioscler Thromb Vasc Biol*. 1996;16(6):749–754.
151. Okamoto M, Hojo Y, Inoue K, et al. Mild exercise increases dihydrotestosterone in hippocampus providing evidence for androgenic mediation of neurogenesis. *Proc Natl Acad Sci*. 2012;109(32):13100–13105.
152. Cuartero MI, de la Parra J, Pérez-Ruiz A, et al. Abolition of aberrant neurogenesis ameliorates cognitive impairment after stroke in mice. *J Clin Investig*. 2019;129(4):1536–1550.
153. Zhang W, Cheng J, Vagnerova K, et al. Effects of androgens on early post-ischemic neurogenesis in mice. *Transl Stroke Res*. 2014;5:301–311.
154. Elliott J, Kelly SE, Millar AC. Testosterone therapy in hypogonadal men: A systematic review and network meta-analysis. *BMJ Open*. 2017;7(11):e015284.
155. Zarrouf FA, Artz S, Griffith J, Sirbu C, Kommor M. Testosterone and depression. *J Psychiatr Pract*. 2009;15(4):289–305.
156. Carrier N, Kabbaj M. Testosterone and imipramine have antidepressant effects in socially isolated male but not female rats. *Horm Behav*. 2012;61(5):678–685.
157. Moriguchi S, Shinoda Y, Yamamoto Y, et al. Stimulation of the sigma-1 receptor by DHEA enhances synaptic efficacy and neurogenesis in the



- hippocampal dentate gyrus of olfactory bulbectomized mice. *PLoS One*. 2013;8(4):e60863.
158. Pinnock SB, Lazic SE, Wong HT, Wong IHW, Herbert J. Synergistic effects of dehydroepiandrosterone and fluoxetine on proliferation of progenitor cells in the dentate gyrus of the adult male rat. *Neuroscience*. 2009; 158(4):1644–1651.
 159. Pesce ME, Acevedo X, Bustamante D, Miranda HF, Pinaridi G. Progesterone and testosterone modulate the convulsant actions of pentylenetetrazol and strychnine in mice. *Pharmacol Toxicol*. 2008;87:116–119.
 160. Schwartz-Giblin S, Korotzer A, Pfaff DW. Steroid hormone effects on picrotoxin-induced seizures in female and male rats. *Brain Res*. 1989;476: 240–247.
 161. Frye CA, Reed TAW. Androgenic neurosteroids: Anti-seizure effects in an animal model of epilepsy. *Psychoneuroendocrinology*. 1998;23:385–399.
 162. Frye CA, Rhodes ME, Walf AA, Harney JP. Testosterone reduces pentylenetetrazole-induced ictal activity of wildtype mice but not those deficient in type i 5 α -reductase. *Brain Res*. 2001;918:182–186.
 163. Reddy DS. Anticonvulsant activity of the testosterone-derived neurosteroid 3 α -androstane diol. *NeuroReport*. 2004;15:515–518.
 164. Hamed SA. The effect of epilepsy and antiepileptic drugs on sexual, reproductive and gonadal health of adults with epilepsy. *Exp Rev Clin Pharmacol*. 2016;9:807–819.
 165. Markoula S, Siarava E, Keramida A, et al. Reproductive health in patients with epilepsy. *Epilepsy Behav*. 2020;113:107563.
 166. Scharfman HE, MacLusky NJ. Sex differences in the neurobiology of epilepsy: A preclinical perspective. *Neurobiol Dis*. 2014;72:180–192.
 167. Cai Z, Li H. An updated review: Androgens and cognitive impairment in older men. *Front Endocrinol*. 2020;11.
 168. Hogervorst E, Combrinck M, Smith AD. Testosterone and gonadotropin levels in men with dementia. *Neuro Endocrinol Lett*. 2003;24(3–4):203–208.
 169. Hogervorst E, Bandelow S, Combrinck M, Smith AD. Low free testosterone is an independent risk factor for Alzheimer's disease. *Exp Gerontol*. 2004;39:1633–1639.
 170. Moffat SD, Zonderman AB, Metter EJ, et al. Free testosterone and risk for Alzheimer disease in older men. *Neurology*. 2004;62:188–193.
 171. Carcaillon L, Brailly-Tabard S, Ancelin M-L, et al. Low testosterone and the risk of dementia in elderly men: Impact of age and education. *Alzheimer's Dement*. 2013;10.
 172. Rosario ER, Chang L, Head EH, Stanczyk FZ, Pike CJ. Brain levels of sex steroid hormones in men and women during normal aging and in Alzheimer's disease. *Neurobiol Aging*. 2011;32:604–613.
 173. Coon KD, Myers AJ, Craig DW, et al. A high-density whole-genome association study reveals that APOE is the major susceptibility gene for sporadic late-onset Alzheimer's disease. *J Clin Psychiatry*. 2007;68:613–618.
 174. Reiman EM, Arboleda-Velasquez JF, Quiroz YT, et al. Exceptionally low likelihood of Alzheimer's dementia in Apoe2 homozygotes from a 5,000-person neuropathological study. *Nat Commun*. 2020;11.
 175. Duarte-Guterman P, Albert AY, Barha CK, Galea LAM. Sex influences the effects of APOE genotype and Alzheimer's diagnosis on Neuropathology and memory. *Psychoneuroendocrinology*. 2021;129:105248.
 176. Sundermann EE, Panizzon MS, Chen X, Andrews M, Galasko D, Banks SJ. Sex differences in Alzheimer's-related Tau biomarkers and a Mediating effect of testosterone. *Biol Sex Differ*. 2020;11.
 177. Cherrier MM, Matsumoto AM, Amory JK, et al. Testosterone improves spatial memory in men with Alzheimer disease and mild cognitive impairment. *Neurology*. 2005;64:2063–2068.
 178. Li B, Yamamori H, Tatebayashi Y, et al. Failure of neuronal maturation in Alzheimer disease dentate gyrus. *J Neuropathol Exp Neurol*. 2008;67: 78–84.
 179. Panizzon MS, Hauger R, Dale AM, et al. Testosterone modifies the effect of APOE genotype on hippocampal volume in middle-aged men. *Neurology*. 2010;75:874–880.
 180. Panizzon MS, Hauger R, Xian H, et al. Interaction of APOE genotype and testosterone on episodic memory in middle-aged men. *Neurobiol Aging*. 2014;35:1778.e1–1778.e8.

Cite this article as: Blankers SA, Galea LAM (2021) Androgens and adult neurogenesis in the hippocampus, *Androgens: Clinical Research and Therapeutics* 2.1, 203–215, DOI: 10.1089/andro.2021.0016.

Abbreviations Used

- 3 α -diol = 5 α -androstane-3 α ,17 β -diol
- 3 β -diol = 5 α -androstane-3 β ,17 β -diol
- AAS = androgenic anabolic steroids
- AD = Alzheimer's disease
- ARs = androgen receptors
- BrdU = 5-bromo-2-deoxyuridine
- CAG = cytosine-adenine-guanine
- DCX = doublecortin
- DHT = 5 α -dihydrotestosterone
- DHEA = dehydroepiandrosterone
- ERs = estrogen receptors
- GCL = granular cell layer
- GFAP = glial fibrillary acidic protein
- GPER = G protein-coupled ER
- ML = molecular layer
- NeuN = neuronal nuclei
- NPC = neural progenitor cells
- NSC = neural stem cell
- NSERC = Natural Sciences and Engineering Research Council of Canada
- p-Tau = phosphorylated Tau
- SGZ = subgranular zone
- SVZ = subventricular zone
- TAPC = transient amplifying progenitor cell

Publish in *Androgens*



- Immediate, unrestricted online access
- Rigorous peer review
- Compliance with open access mandates
- Authors retain copyright
- Highly indexed
- Targeted email marketing

liebertpub.com/ANDRO

