



# Set Up for Failure: Pre-Existing Autoantibodies in Lung Transplant

Alexander McQuiston<sup>1,2</sup>, Amir Emtiazjoo<sup>2</sup>, Peggi Angel<sup>3</sup>, Tiago Machuca<sup>4</sup>, Jason Christie<sup>5</sup> and Carl Atkinson<sup>2\*</sup>

<sup>1</sup> Department of Microbiology and Immunology, Medical University of South Carolina, Charleston, SC, United States, <sup>2</sup> Division of Pulmonary, Critical Care and Sleep Medicine, University of Florida, Gainesville, FL, United States, <sup>3</sup> Department of Cell and Molecular Pharmacology and Experimental Therapeutics, Medical University of South Carolina, Charleston, SC, United States, <sup>4</sup> Department of Surgery, University of Florida, Gainesville, FL, United States, <sup>5</sup> Department of Medicine, University of Pennsylvania, Philadelphia, PA, United States

Lung transplant patients have the lowest long-term survival rates compared to other solid organ transplants. The complications after lung transplantation such as primary graft dysfunction (PGD) and ultimately chronic lung allograft dysfunction (CLAD) are the main reasons for this limited survival. In recent years, lung-specific autoantibodies that recognize non-HLA antigens have been hypothesized to contribute to graft injury and have been correlated with PGD, CLAD, and survival. Mounting evidence suggests that autoantibodies can develop during pulmonary disease progression before lung transplant, termed pre-existing autoantibodies, and may participate in allograft injury after transplantation. In this review, we summarize what is known about pulmonary disease autoantibodies and lung transplantation, and potential mechanisms through which pre-existing autoantibodies contribute to graft injury and rejection.

Keywords: lung transplant, complement, Autoantibodies, primary graft dysfunction, chronic lung allograft dysfunction, glycans

# INTRODUCTION

Lung transplantation (LTx) is the only viable option for many chronic end-stage pulmonary diseases (CPD). The number of LTx performed annually in the US is rapidly increasing, and the demand for donor lungs far exceeds availability. Despite the improved surgical techniques and post-transplant management, the long-term survival has not significantly improved over the last decade and remains the lowest compared to other solid organ Tx (1). According to the International Society of Heart and Lung Transplantation (ISHLT) registry data, median survival after lung transplantation is 6.5 years, the worst amongst all solid organ transplantation, and is, in large part, the result of chronic allograft dysfunction (CLAD). Primary graft dysfunction (PGD) is a common early complication after LTx, and a major risk factor for development of CLAD. PGD occurs within the first 72 hours after transplantation and factors such as the recipient's underlying lung disease, donor medical history, recipient/donor interaction, severity of post-Tx complications all play an integral part in determining LTx success.

Arguably, organ transplantation was made possible through optimization of surgical techniques and subsequent manipulation of the immune system. Significant improvements have been made in

OPEN ACCESS

#### Edited by:

Seiamak Bahram, Université de Strasbourg, France

#### Reviewed by:

Henny Otten, University Medical Center Utrecht, Netherlands Rusan Ali Catar, Charité – Universitätsmedizin Berlin, Germany

> \*Correspondence: Carl Atkinson carl.atkinson@medicine.ufl.edu

#### Specialty section:

This article was submitted to Alloimmunity and Transplantation, a section of the journal Frontiers in Immunology

> Received: 17 May 2021 Accepted: 12 July 2021 Published: 11 August 2021

#### Citation:

McQuiston A, Emtiazjoo A, Angel P, Machuca T, Christie J and Atkinson C (2021) Set Up for Failure: Pre-Existing Autoantibodies in Lung Transplant. Front. Immunol. 12:711102. doi: 10.3389/fimmu.2021.711102

1

Pre-Existing Autoantibodies and Transplant Injury

controlling the recipient's immune system post-transplant, largely by modulating T cell immune responses. While modulation of adaptive immunity is essential for graft survival other factors occurring earlier in the transplant process also play a role in graft injury. Initial immune-mediated graft injury occurs upon reperfusion, referred to as ischemia reperfusion injury (IRI). Following IRI, activation of innate and adaptive immune response drives targeted graft damage. This damage manifests as post-Tx complications such as PGD, acute rejection, and CLAD, all of which predispose to increased risk of mortality. The generally accepted model of immune-mediated graft damage proposes a feed-forward mechanism starting with donor graft injury, IRI-mediated immune activation and graft damage, thereby promoting subsequent adaptive immune activation and T cell specific graft damage. However, this model does not take into account the recipients' lung specific pre-Tx immune system. Patients with end-stage lung diseases, such as chronic obstructive pulmonary disease (COPD) and interstitial lung disease (ILD) often have autoimmunity, which is increasingly being recognized as a potential driver of graft injury post transplantation (2, 3).

The presence of a lung-specific autoreactive immune system pre-transplant may well predispose targeted attack of the donor lung upon implantation, and further exacerbate the alloimmune effector mechanisms activated post-transplant. Immunemediated graft damage remains a major obstacle, and manipulation of the recipient immune system currently only occurs upon transplantation. Therefore, gaining a better understanding of the immune factors present within the LTx recipient prior to transplantation and how these factors contribute to shaping post-transplant graft outcomes can have a significant impact on the patient outcome. In this review, we will focus on the impact of pulmonary disease-associated autoantibodies and pulmonary disease-specific autoantibodies in LTx, and postulate on the clinical significance of autoantibodies identified in patients.

# PULMONARY DISEASES AND AUTOIMMUNITY: AUTOANTIBODIES

Over the last 20 years, accumulated data proports a role for autoimmunity in pulmonary disease pathogenesis and progression of certain lung pathologies (4). How pulmonary disease autoimmunity arises and its influence on disease progression is not fully understood; however, it is generally accepted that autoimmune factors, such as autoantibodies and autoreactive T cells, play critical roles in disease perpetuation.

Autoantibodies are antibodies produced by the immune system with reactivity to self-antigens. Multiple mechanisms can render host molecules antigenic, interestingly, those same mechanisms can occur during or as a result of pulmonary disease onset. For example, environmental exposure to pollutants, such as those found in cigarette smoke, a major risk factor for COPD development, can drive mutations and post-translational modifications such as oxidation, carbonylation, and citrullination to pulmonary peptides rendering them antigenic (5). Fibrotic-associated damage, commonly found in pulmonary diseases and directly correlated with ROS-mediated oxidative stress, can lead to the exposure of cryptic neoepitopes that act as pathogenic antigens that drive inflammation and injury (6). Autoantibodies against a wide spectrum of self-antigens have been identified in patients with lung disease and rodent models of disease (7–9). The clinical relevance of autoantibodies identified in lung disease patients is an area of intense debate. While it is clear that a majority of lung disease patients present with autoantibodies, whether autoantibodies contribute to pathogenesis, are useful biomarkers, or simply epiphenomenon of the underlying disease remains to be elucidated. Below, we highlight recent advances in the association between autoantibodies and COPD and ILD patients (**Table 1**).

# Autoantibodies in Chronic Obstructive Pulmonary Disease

Chronic obstructive pulmonary disease (COPD) is a group of progressive pulmonary disorders characterized by chronic pulmonary inflammation and damage to small airways and alveolar airspace that results in airflow limitation. The specific pathogenesis of COPD remains unknown, but it is widely accepted that cigarette smoke (CS) exposure is one of the leading causes. CS promotes innate and adaptive immune cell infiltration into the lung and airways of COPD patients (10, 11). Increased immune cell presence and activity in the lungs is hypothesized to promote progressive tissue destruction, a hallmark of COPD and critical in COPD pathogenesis.

Autoantibodies and autoantigens have been successfully identified in COPD patients and animal models of disease. A systematic review of all clinical studies investigating autoantibodies in the context of COPD was published in 2019 by Byrne et al. (12). The review focuses on 42 peer-reviewed articles that investigate one or multiple autoantibodies. The most well studied autoantibodies include anti-endothelial/epithelial cell autoantibodies (anti-AECA), Rheumatoid factor (RF), anticytokeratin (anti-CK), anti-nuclear autoantibodies (ANAs), anticollagen, anti-cyclic citrullinated peptide autoantibodies (anti-CCP), anti-elastin, anti-smooth muscle autoantibodies (ASMA), and antineutrophil cytoplasmic autoantibodies (ANCA) (12). All other autoantibodies mentioned (approximately 9) in the review were investigated in less than three studies. Overall, successful identification of multiple different autoantibodies supports the hypothesis that autoimmunity is prevalent in COPD patients, however, much more research is needed to determine the clinical significance of these autoantibodies.

Many of the most well-studied autoantibodies, such as the ones listed above, are common to other autoimmune diseases such as rheumatoid arthritis, systemic sclerosis, and lupus (13). While those autoantibodies may still be clinically relevant in COPD, identifying disease specific autoantibodies may prove to be more useful as biomarkers, and improve diagnoses and treatment. The complex heterogeneity among COPD patients makes it difficult to identify a single disease specific autoantibody/autoantigen common to all patients, therefore, establishing a panel or signature of autoantibodies may be

TABLE 1   Autoantibody titers correlate with disease	progression
--	-------------

Autoantibody	Ig Isotype	Disease	Association with Severity	Citation
HBEC	lgG/lgA	COPD	Most IgG/A Positive patients were GOLD Stage III and IV	(48)
Elastin	lgG	COPD	Decreased with severity (GOLD Stage)	(49)
Elastin	lgG	COPD	Increased with severity	(50)
Cytokeratin 18	lgG/lgA/lgM	COPD	Increased with severity (GOLD Stage); Correlated with FEV1 (L) and FEV1 (%) Predicted	(51, 52)
Cytokeratin 19	lgG/lgA/lgM	COPD	Increased with severity (GOLD Stage)	(51)
CD80	lgG	COPD	Increased with severity (GOLD Stage)	(53)
Carbonyl Modified Proteins	lgG	COPD	Increased with severity (GOLD Stage)	(54)
Serum Albumin	lgG	COPD	Increased with severity (GOLD Stage)	(54)
ANA	lgG	COPD	Increased with severity (GOLD Stage)	(55–57)
ASMA	lgG	COPD	Increased with severity (GOLD Stage)	(55)
Anti-Tissue	lgG	COPD	Increased with severity (GOLD Stage)	(57)
GRP78	lgG	COPD	Increased with severity (GOLD Stage)	(58)
B2-adrenergic receptor	lgG	COPD	Increased with severity (GOLD Stage)	(59)
Ro52	lgG	ILD	Increased with severity	(60)
MDA5	lgG	ILD	Increased with severity	(22)
Cyclic Citrullinated Peptides	lgG	ILD	Increased with severity	(61, 62)
CXCR3	lgG	ILD	Increased with severity	(63)
CXCR4	lgG	ILD	Increased with severity	(63)
Periplakin	IgG	ILD	Increased with severity	(64)

more useful. A recent study analyzed 19,000 human proteins by protein microarray and found that COPD patients had higher autoantibody titers towards extracellular proteins and neutrophil granule proteins compared to controls (14). Interestingly, IgM and IgG autoantibodies against intracellular antigens were lower in COPD patients, potentially because intracellular autoantibodies are associated with maintaining homeostasis (14–16). The fact that some autoantibody titers are increased while others are decreased in COPD patients compared to normal controls further supports the need for autoantibody profiling. Although the aforementioned data provides the start of a global understanding of the autoantibody landscape, multiple gaps in our knowledge remain.

The most glaring gap in the literature, as discussed in Duncan 2012, is the functional characterization of autoantibodies in COPD pathogenesis (17). Thus far, very little is known whether the autoantibodies identified in COPD patients are pathogenic, beneficial, biomarkers, or simply epiphenomena of the underlying disease. The accepted hypothesis is that the autoantibodies identified contribute to COPD pathogenesis by promoting immune activation, targeted tissue destruction, and cell death. How the autoantibodies promote such a response is unknown. There are multiple autoantibody characteristics and mechanisms that could promote different pathogenic processes, as we discuss in detail later in this review.

# Interstitial Lung Disease and Autoantibodies

Interstitial lung disease (ILD) is a group of multiple pulmonary diseases characterized by progressive scarring or fibrosis of the lungs. ILD is an extremely complex disease because it is commonly associated with connective tissue diseases (CTD-ILD) such as rheumatoid arthritis, lupus, and sclerosis, or nonconnective tissue disease origin referred to as interstitial pneumonia with autoimmune features (IPAF). In this context of CTD-ILD, ILD can be one of many manifestations of an established CTD or may be the only manifestation of the CTD. Interestingly, many CTDs can be categorized as autoimmune disorders suggesting that ILD may also comprise an autoimmune component. For example, ILD manifests and is the leading cause of mortality in Systemic-Sclerosis, an extensively studied autoimmune disease with anti-nuclear autoantibodies (ANA) (anti-centromere antibodies, anti-topoisomerase I antibodies, and anti-RNA polymerase antibodies) and anti-endothelial cell antibodies (AECA) in greater than 90% of patients with potential pathogenic roles (18). Additionally, ILD occurs in upwards of 80% of patients with myositis, a rare autoimmune disease that presents with myositis-specific autoantibodies (MSAs; antisynthetase and anti-CADM140/MDA5).

Whether or not the majority of autoantibodies identified in CTD-ILD patients are involved in disease pathogenesis remains unknown, but their clinical significance as diagnostic markers that shape patient is growing in significance. For example, a subset of clinically amyopathic dermatomyositis (CADM) patients with rapidly progressive ILD have been shown to present with circulating anti-CADM140/MDA5 autoantibody (19). Studies have demonstrated that anti-CADM140/MDA5 positive patients are at a higher risk for developing ILD, and are at a higher risk of ILD-related respiratory failure and death (20–23). While it remains unknown if anti-CADM140/MDA5 play a pathogenic role, its presence can dramatically change clinical intervention strategies.

In 2011, guidelines from the American Thoracic Society/ European Respiratory Society/Japanese Respiratory Society and Latin American Thoracic Association highlight ANA, anti-CCP, and RF as necessary autoantibodies to test for in all potential ILD patients (24). Not surprisingly, similar to COPD, numerous other autoantibodies have been discovered in ILD patients with a broad range of frequencies, associations, and techniques. An extensive review of autoantibodies in CTD-ILD and IPAF patients and their clinical importance was recently compiled by Jee et al. (25). The review article provides insight from over 100 articles evaluating the clinical relevance of autoantibodies in CTD-ILD and IPAF, demonstrating their utility, while also bringing light to hurdles that limit our understanding. The vast heterogeneity of CTD-ILD and IPAF disease phenotypes coupled with the lack of standardization in panels of autoantibodies tested for and techniques used makes it difficult to pool data (25). While there is great interest in the use of autoantibodies to predict outcomes and survival, lack of standardization and consensus hampers clinical utilization.

### PULMONARY DISEASE AUTOIMMUNITY AND LUNG TRANSPLANT

Pulmonary disease autoimmunity and lung transplant can be seen as a 'perfect storm' for graft injury and rejection. While several studies have investigated autoantibodies in context of lung transplantation few have attempted to determine the clinical significance or the mechanism of action of pre-existing autoantibodies, present prior to transplantation, on posttransplant outcomes. It is important to note that many of the autoantigens identified in pulmonary disease patients are not lung-specific, but rather have a broad biodistribution. While pulmonary disease-associated autoantigens that are not lungspecific may still be clinically relevant, it is possible that lungrestricted self-antigens may also be clinically relevant with regard to LTx outcomes because they could represent biomarkers of graft health/damage or contribute pathogenically to graft damage.

In this context, pre-existing autoantibodies that recognize lungrestricted self-antigens can be produced prior to LTx (Figure 1). One hypothesis for their production is that natural low-affinity IgM autoantibodies recognize damaged and abnormally exposed selfproteins as antigenic and act as templates for somatic hypermutation and class switching to high-affinity IgG/IgA autoantibodies (26). Lung-restricted proteins can become antigenic through tissue destruction and inherent pulmonary disease characteristics, as previously stated. Interestingly, similar processes that expose and render self-proteins antigenic in pulmonary disease can occur to the graft. This is interesting because it could explain a mechanism by which pre-existing autoantibodies bind to and target the donor graft. For example, donor brain death induces inflammation and immune cell infiltration into the lung that potentiate cell death and tissue damage (27). Other unavoidable injuries that impact the donor organ prior to transplantation such as organ procurement strategies, cold storage, transportation, time between organ procurement to transplantation, and reperfusion of the graft have all been demonstrated to reduce viability of the lung due to severe inflammatory responses and irreversible tissue damage (28, 29). The subsequent inflammation and tissue damage drives cell death, release of ROS, and degradation of proteins thereby promoting an environment for pre-existing recipient autoantibodies present prior to transplantation to bind to damaged lung-proteins early post-transplant and induce graft injury, however, definitive evidence for this mechanism is currently lacking.

Although there are not any studies directly comparing the tissue damage that occurs in pulmonary disease to graft damage, it is reasonable to hypothesize that similar enzymatic





4

degradation of extracellular matrix occurs early post-transplant. The innate immune cells associated with injury in both pulmonary disease and graft damage are essentially the same, and can induce post-translational modifications to ECM proteins through secretion of proteases and production of ROS. Along this line of thinking, the LTx recipient's pre-existing autoantibodies could recognize self-antigens that are present and exposed in the new graft. Therefore, the recipient's immune system is primed to target and attack the graft prior to transplant which could manifest as severe pre-existing autoantibody-mediated graft damage once the graft is implanted. Potential mechanisms by which pre-existing autoantibodies mediate graft damage could be increased complement activation and/or FcyR binding, which we discuss later in this review.

Although it is enticing to hypothesize about the potential pathogenicity and clinical relevance of pulmonary disease autoantibodies in LTx, definitive experimental and clinical data supporting potential mechanisms are lacking. The presence of pre-existing autoantibodies in transplant patients and their association with poor clinical outcomes has been well established in other solid organ transplants, specifically in renal and heart transplants; driving the hypothesis that pre-existing autoantibodies may also play a role in LTx outcomes (30, 31). As our understanding of autoimmune-mediated graft damage in LTx patients deepens, the association between pre-existing autoantibodies and post-LTx disorders has become an intense area of research. Identification and functional characterization of pre-existing autoantibodies in disorders such as primary graft dysfunction, chronic lung allograft dysfunction, and mortality has begun to accumulate (32-34). Below we will outline the research analyzing the association of pre-existing autoantibodies and lung transplant outcome.

# Pre-Existing Autoantibodies and Primary Graft Dysfunction

Primary graft dysfunction (PGD) is a progressive acute lung injury syndrome that manifests within the first 72 hours after lung transplantation and affects 25-30% of LTx patients (35). The ISHLT definition of PGD is based on radiographic infiltrates and P/F ratios, the ratio between arterial partial pressure and inspired oxygen, assessed at multiple time points post-LTx and is graded on a scale of Grade 0 to 3. PGD onset and grade are correlated with increased mechanical ventilation, hospital stay, cost, CLAD, and most importantly, early morbidity and mortality (36). The mechanisms responsible for PGD development are not completely understood, however, donor- and recipient-related clinical risk factors (age, gender, race, and comorbidities) and IRI have all been associated with PGD etiology [For detailed reviews see Diamond et al. (35), Suzuki et al. (37), and Altun et al. (38)].

Recently, the presence of pre-existing autoantibodies has been associated with increased risk of PGD in multiple retrospective clinical studies. The most well studied pre-existing autoantibodies correlated with PGD onset are Collagen V (Col V) and K-alpha1-tubulin (KAT). Col-V is a minor fibrillar collagen found in the lungs where it is normally sequestered from recognition by collagen I and collagen III overlays. Exposure to Col-V is hypothesized to occur upon matrix metalloprotease (MMP) degradation of the extracellular milieu during ischemia reperfusion injury, and immunogenic Col-V fragments can be found in BAL fluid (39-41). KAT is a gap junction protein that was observed to promote fibrogenic signaling after binding to airway epithelial cells (42). Clinically, anti-Col-V and anti-KAT autoantibodies have been identified in different CPD populations at different frequencies. Approximately 30% of ILD patients and 20% of COPD patients are positive for pre-existing anti-Col-V and anti-KAT autoantibodies (39). A second study also found that approximately 20% of LTx patients tested positive for preexisting anti-Col-V autoantibodies and about 60% of those patients remained positive for anti-Col-V pre-existing autoantibodies after LTx (43).

More literature focusing on the correlation between Col-V and KAT autoantibodies and PGD, and potential mechanisms were recently reviewed by Sureshbabu et al. (32). Briefly, the presence of pre-existing anti-Col-I, anti-Col-V, and anti-KAT autoantibodies correlated with increased risk of PGD development and high levels of proinflammatory cytokines (39, 44). In a syngeneic LTx mouse model, immunization with anti-Col-V or anti-KAT resulted in dose-dependent PGD-like phenotype development. Interestingly, immunization with KAT induced an immune response that led to de novo anti-Col-V antibody development post transplantation, and vice versa. Taken together these data suggest that autoantibodies to either self-antigen are capable of inducing lung injury that leads to epitope spreading post transplantation (45). These reductive models drive the hypothesis that pre-existing lung autoantibodies could impact post-transplant outcome in those patients with diverse high titers of autoantibodies associated with their relative CPD. The presence of pre-existing recipient autoantibodies pre-transplant may not only promote more intense graft injury but further promote diverse epitope spreading, de novo antibody production, and an increased antibody mediated graft injury (Figure 2). However, human studies demonstrating pre-existing autoantibody-mediated graft injury and epitope spreading by anti-Col-V or anti-KAT have not as yet been reported.

A potential mechanism for anti-Col-V autoantibodymediated damage can be found in elegant adoptive transfer studies performed by Iwata et al. (40). Transfer of total serum, purified serum IgG, and B cells from Col-V immunized rats into rat lung isograft recipients induced PGD-like phenotype, impaired graft function, and increased local expression of IFN-y, TNF-alpha, and IL-1ß. Interestingly, Col-V was shown to be apically expressed by epithelial cells, and when exposed to immunized rat serum, rat airway epithelial cells were sensitive to complement-mediated cytotoxicity (40). These data suggest that anti-Col-V autoantibodies promote graft damage *via* antibodydependent complement activity and by increasing injurious proinflammatory cytokine production. Similarly, novel studies by Patel et al. (33) explored the impact of CPD autoantibodies on lung transplant outcomes. Using a mouse model of emphysema, the



authors demonstrated that chronic exposure of mice to cigarette smoke led to the characteristic features of emphysema, such as airspace enlargement, immune cell infiltration and significantly elevated serum autoantibodies compared with non-smokeexposed age-matched controls. To determine the impact of a full pre-existing autoantibody repertoire on lung transplant IRI the authors transplanted BALB/c donor lungs into control or chronically cigarette smoke (CS) exposed recipients. CS recipients had significantly increased lung injury and immune cell infiltration after transplant. Immunofluorescence staining revealed increased IgM, IgG, and C3d deposition in CS recipients. To exclude confounding alloreactivity and confirm the role of pre-existing autoantibodies in IRI, syngeneic Rag1 knockout transplants were performed in which recipients were reconstituted with pooled serum from CS or control mice. Serum from CS-exposed mice significantly increased IRI compared with control mice characterized by increased antibody and complement deposition. To confirm the role of autoantibodies, serum from CS was immunoglobulin depleted and adoptively transferred. Immunoglobulin depleted serum had no significant impact on transplant outcomes as compared to controls. Taken together these data demonstrate that pre-transplant CS exposure was associated with increased IgM/IgG autoantibodies, which, upon transplant, bind to the donor lung, activate complement, and exacerbate post-transplant IRI. Further studies are needed to determine, if any, the long term sequalae that a broad spectrum of autoantibodies have on graft injury, epitope spreading, and rejection. Clinical studies are also needed to corroborate these findings.

Previous literature has largely focused on Collagen V and KAT autoantibodies. As discussed earlier, and in the studies by Patel et al. (33), a diverse spectrum of autoantibodies are likely present in the lung transplant population pre-transplantation, and therefore investigating if these antibodies pre and posttransplant are clinically relevant is urgently needed. To this end, a recent large proteomic/antigen microarray study using human patient sera has successfully identified multiple different preexisting autoantibodies that correlated with PGD. A microarray with 504 and 610 potential IgG and IgM targets, respectively, which covered 272 different proteins was created to identify new pre-existing autoantibodies. A total of 17 IgM/IgG novel autoantibodies were discovered that correlated with PGD onset (Table 2) (47). Another group performed a proteomic microarray analysis investigating 124 autoantigens identified 17 IgA and 3 IgG additional pre-existing autoantibodies that correlated with PGD development (See Table 2 for list of autoantibodies) (46). Interestingly, 6 of the 17 IgA autoantibodies correlated with worse survival and those 6 autoantibodies were able to correctly classify PGD positive from PGD negative patients. Survival was not statistically different for the 3 IgG autoantibodies (Periplakin, Muscarinic acetylcholine receptor type 3 (AChR3), and Angiotensin II

TABLE 2   Pre-existing autoantibodies and primary graft dysfunction develop	oment
---	-------

Autoantibody	lg Isotype	Associated with Survival	Citation
Collagen I	lgM/lgG	Yes	(44)
Collagen V	lgM/lgG	Yes	(44)
K-alpha Tubulin	lgM/lgG	Yes	(44)
Filaggrin	IgA	Yes	(46)
Factor P	IgA	Yes	(46)
Heparan sulfate	IgA	Yes	(46)
Laminin	IgA	Yes	(46)
RSV antigen	IgA	Yes	(46)
CRP antigen	IgA	Yes	(46)
Factor B	IgA	No	(46)
ERP29	IgA	No	(46)
Enolase	IgA	No	(46)
Endothelial cell extract	IgA	No	(46)
rhHSPG2	IgA	No	(46)
AGTR1	lgA/lgG	No	(46)
Proteopglycan	IgA	No	(46)
c-MYC	IgA	No	(46)
SDC1	IgA	No	(46)
Aggrecan	IgA	No	(46)
Complement C1q	IgA	No	(46)
Periplakin	IgG	No	(46)
Acetylcholine receptor (AchR3)	IgG	No	(46)
EGFR	IgM/IgG	No	(47)
MBP	IgM/IgG	No	(47)
MLANA	lgM/lgG	No	(47)
MUC1	lgM/lgG	No	(47)
MYCL1	lgM/lgG	No	(47)
PLCG1	lgM/lgG	No	(47)
PRKCA	lgM/lgG	No	(47)
HSP90AA1	lgM/lgG	No	(47)
IGF1R	lgM/lgG	No	(47)
RB1	lgM/lgG	No	(47)
CERK	laM/laG	No	(47)
HSPD1	lgM/lgG	No	(47)
TEP1	lgM/lgG	No	(47)
CYP3A4	lgM/lgG	No	(47)
SOCS3	laM/laG	No	(47)
TARP	laM/laG	No	(47)
TP53	laM/laG	No	(47)

Pre-existing autoantibodies found in COPD and ILD patients compared to healthy controls.

receptor type 1 (AT1R)) or the 11 remaining IgA autoantibodies (46). Anti-AT1R autoantibodies were the only antibodies discovered to have titers for both IgG and IgA subtypes. Although the 3 IgG and 11 remaining IgA autoantibodies do not correlate with reduced survival, they may play a role in PGD pathogenesis, but that is speculation. Anti-Periplakin IgG have been observed to decrease cell migration and induce bronchoalveolar lavage T lymphocyte proliferation in IPF, but their role in transplant is unknown (64). It is important to note that the mere presence of autoantibodies and their correlation with PGD development does not mean that they play a role in PGD pathogenesis. These autoantibodies could also represent novel predictive biomarkers for patients at risk of developing PGD.

Anti-AT1R autoantibodies have been previously identified in kidney, heart, and liver transplants with their presence associated with poor prognosis (65, 66). Specifically, the presence of pre-Tx anti-AT1R autoantibodies was associated with fibrosis and vessel occlusion in renal transplant, suggesting a potential mechanism through which anti-AT1R could promote PGD (67). Anti-AT1R acts as an agonist on AT1R, which leads to sustained AT1R activation which is thought to promote vasoconstriction, inflammation and fibrosis (68-70). In the context of kidney Tx, anti-AT1R-mediated prolonged signaling of AT1R which induced pro-inflammatory and pro-coagulant processes via activation of extracellular signaling-regulated kinase 1/2 and nuclear factor-kB in endothelial and vascular smooth muscle cells (69). Other studies have purported a pathogenic role of anti-AT1R through modulation of AT1R expressing immune cells (65). AT1R expression is observed on monocytes, T and B cells, and serum from anti-AT1R positive systemic sclerosis patients induced peripheral blood mononuclear cell production of proinflammatory chemokines, IL-8 and CCL18 (71). Interestingly, anti-AT1R antibodies were found to belong to the complement fixing IgG1 and IgG3 subclasses, however, its mechanism of action is reportedly complement independent based on the lack of complement deposition in anti-AT1R positive patients (The impact of IgG structure and subclass on function is discussed later in this review) (69, 72, 73). Although these elegant studies proposed pathogenic mechanisms for anti-AT1R in kidney transplant and systemic sclerosis patients, to the best of our knowledge no experimental evidence exists demonstrating pathogenic functions of anti-AT1R in the context of LTx and PGD onset.

One study identified anti-AT1R in conjunction with antiendothelin type A receptor (ETAR) in cystic fibrosis patients both pre- and post-Tx (74). A second study analyzing pre- and post-LTx sera of 162 patients from 3 centers also identified the presence of anti-AT1R and anti-ETAR. The presence of human leukocyte antigen (HLA)-specific antibodies and strong anti-AT1R and anti-ETAR binding significantly reduced freedom from *de novo* donor specific antigen (DSA) development post-LTx (75). *De novo* DSA development in LTx is well known to promote acute and chronic rejection, and reduce survival (76–78). Interestingly, the presence of anti-ETAR is highly associated with anti-AT1R in pediatric kidney transplant which may suggest another example of epitope spreading, however this has not been proven (79).

Overall, more research into pre-existing autoantibodies and PGD is needed. Large screenings, such as those used to discover autoantibodies in COPD and ILD patients, would help identify and measure pre-existing autoantibodies before and after LTx. Following identification, functional analyses of pre-existing autoantibodies are needed to determine if they are biomarkers, pathogenic, both, or neither. This information could hold the key to novel pre-Tx or perioperative treatments that reduce the risk of PGD development. Reducing the incidence of PGD is essential step to overcome poor survival. PGD is also a major risk factor for chronic lung allograft dysfunction (CLAD), the main cause of late-stage mortality (35, 80).

### Pre-Existing Autoantibodies and Chronic Lung Allograft Dysfunction

Chronic lung allograft dysfunction (CLAD) remains the largest obstacle to long term allograft survival. ISHLT defines CLAD as

the consistent decline in forced expiratory volume in 1 second (FEV<sub>1</sub>) from baseline FEV<sub>1</sub> following LTx. There are three CLAD phenotypes referred to as bronchiolitis obliterans syndrome (BOS), restrictive allograft syndrome (RAS, previously known as restrictive CLAD), and neutrophilic reversible allograft dysfunction (NRAD). However, NRAD was not included in the most recent ISHLT consensus report. BOS, the most common CLAD phenotype, accounts for approximately 60-75% of CLAD cases (81) [Extensive reviews of CLAD can be found in Verleden et al. (82) and Glanville et al. (83)].

To date, three studies have evaluated whether pre-existing autoantibodies are associated with CLAD development. In the first, patients with pre-existing anti-Col-V and anti-KAT autoantibodies were at increased risk for development of HLAantibodies and for BOS development (44). In the second, a single center study investigated the presence of pre-existing autoantibodies in 'stable BOS' and 'progressive BOS' patients. Using a proteomic microarray with 124 self-antigens, analysis revealed 16 IgG preexisting autoantibodies that were elevated in the progressive BOS patients as compared to stable BOS patients (See Table 3 for list of autoantibodies). A subset of 6 out of the 16 pre-existing autoantibodies correlated with worse BOS free survival. Those 6 IgG autoantibodies remained elevated three months and one-year post-Tx. Another microarray investigating 751 different antigens investigated autoantibodies in a subset of LTx patients with no-tomild BOS compared to moderate-to-severe BOS patients (84). Approximately 28 pre-existing autoantibodies were elevated in the moderate-to-severe BOS patient population compared to the no-tomild BOS patient population (Table 3). Interestingly, 6 of the IgM/ IgG autoantibodies identified were also elevated in a subset of patients that develop PGD, suggesting a connection between PGD and BOS development (47, 84). However, further studies are required to determine whether these pre-existing autoantibodies are pathogenic or epiphenomena of chronic rejection (85).

There are likely more pre-existing autoantibodies associated with CLAD that need to be identified. Interestingly, some of the same autoantibodies identified in the Kaza et al. (85) study have been described and associated with other solid organ transplants (30, 86-88). For example, anti-AGTR1, anti-LG3, and antivimentin pre-transplant autoantibodies have been correlated with high risk of graft failure in kidney and heart transplant (30, 86-88). The presence of anti-vimentin autoantibodies in patients awaiting solid organ transplantation (heart and lung transplantations) was previously observed to be low (approximately 6%) (89). Alternatively, a second study identified that circulating vimentin and anti-vimentin IgG autoantibodies were increased in IPF patients compared to healthy individuals (90). Interestingly, these data demonstrate that anti-vimentin autoantibodies have been identified in ILD patients and vimentin expression increases following LTx. However, any association between anti-vimentin autoantibodies and LTx outcome has not been studied (91). The list of potential pre-existing autoantibodies associated with CLAD is extensive based on the sheer number of different autoantibodies identified in COPD and ILD patients, and more research is needed to streamline and elucidate their clinical significance in post-LTx outcome (12, 25).

lg Isotype	CLAD Phenotype	Associated with Survival	Citation
lgM/lgG	BOS	Yes	(39–41)
lgM/lgG	BOS	Yes	(42, 136)
lgG	BOS	No	(85)
lgG	BOS	No	(85)
lgG	BOS	Yes	(85)
lgG	BOS	Yes	(85)
lgG	BOS	No	(85)
lgG	BOS	No	(85)
lgG	BOS	No	(85)
lgG	BOS	No	(85)
lgG	BOS	No	(85)
lgG	BOS	No	(85)
lgG	BOS	No	(85)
lgG	BOS	Yes	(85)
lgG	BOS	Yes	(85)
lgG	BOS	No	(85)
lgG	BOS	No	(85)
lgG	BOS	No	(85)
lgM/lgG	BOS	No	(84)
lgM/lgG	BOS	No	(84)
lgM/lgG	BOS	No	(84)
lgM/lgG	BOS	No	(84)
lgM/lgG	BOS	No	(84)
lgM/lgG	BOS	No	(84)
lgM/lgG	BOS	No	(84)
lgM/lgG	BOS	No	(84)
lgM/lgG	BOS	No	(84)
lgM/lgG	BOS	No	(84)
lgM/lgG	BOS	No	(84)
lgM/lgG	BOS	No	(84)
lgM/lgG	BOS	No	(84)
lgM/lgG	BOS	No	(84)
lgM/lgG	BOS	No	(84)
lgM/lgG	BOS	No	(84)
lgM/lgG	BOS	No	(84)
IgM/lgG	BOS	No	(84)
IgM/lgG	BOS	No	(84)
IgM/IgG	BOS	No	(84)
IgM/IgG	BOS	No	(84)
Igivi/igG	BOS	NO	(84)
IgIVI/IgG	BO2	INO	(84)
Igivi/igG	BOS	NO	(84)
IgIVI/IgG	BO2	INO	(84)
igivi/lgG	BO2	INO	(84)
igivi/lgG	BO2	INO N-	(84)
IgIVI/IgG	BO2	INO	(84)
igivi/lgG	BO2	INO	(84)
	Ig   Isotype   IgM/IgG   IgM/IgG   IgG   IgM/IgG   IgM/IgG	IgCLADIgotypePhenotypeIgM/IgGBOSIgM/IgGBOS <td>Ig IsotypeCLAD PhenotypeAssociated with SurvivalIgM/IgGBOSYesIgM/IgGBOSYesIgGBOSNoIgGBOSYesIgGBOSYesIgGBOSYesIgGBOSYesIgGBOSYesIgGBOSNoIgGBOSNoIgGBOSNoIgGBOSNoIgGBOSNoIgGBOSNoIgGBOSNoIgGBOSNoIgGBOSNoIgGBOSNoIgGBOSNoIgGBOSNoIgGBOSNoIgGBOSNoIgGBOSNoIgM/IgGBOSNoIgM/IgGBOSNoIgM/IgGBOSNoIgM/IgGBOSNoIgM/IgGBOSNoIgM/IgGBOSNoIgM/IgGBOSNoIgM/IgGBOSNoIgM/IgGBOSNoIgM/IgGBOSNoIgM/IgGBOSNoIgM/IgGBOSNoIgM/IgGBOSNoIgM/IgGBOSNoIgM/IgGBOSNoIgM/IgGBOSNoIgM/IgGBOSNoIgM/IgGBOSNoIgM/IgGBOSNo<tr< td=""></tr<></td>	Ig IsotypeCLAD PhenotypeAssociated with SurvivalIgM/IgGBOSYesIgM/IgGBOSYesIgGBOSNoIgGBOSYesIgGBOSYesIgGBOSYesIgGBOSYesIgGBOSYesIgGBOSNoIgGBOSNoIgGBOSNoIgGBOSNoIgGBOSNoIgGBOSNoIgGBOSNoIgGBOSNoIgGBOSNoIgGBOSNoIgGBOSNoIgGBOSNoIgGBOSNoIgGBOSNoIgGBOSNoIgM/IgGBOSNoIgM/IgGBOSNoIgM/IgGBOSNoIgM/IgGBOSNoIgM/IgGBOSNoIgM/IgGBOSNoIgM/IgGBOSNoIgM/IgGBOSNoIgM/IgGBOSNoIgM/IgGBOSNoIgM/IgGBOSNoIgM/IgGBOSNoIgM/IgGBOSNoIgM/IgGBOSNoIgM/IgGBOSNoIgM/IgGBOSNoIgM/IgGBOSNoIgM/IgGBOSNoIgM/IgGBOSNo <tr< td=""></tr<>

Pre-existing autoantibodies associated with CLAD compared to control (CoI-V and KAT) and pre-existing autoantibodies associated with progressive CLAD compared to stable CLAD (85).

# POTENTIAL MECHANISMS OF GRAFT INJURY BY PRE-EXISTING AUTOANTIBODIES

In addition to gaining a better understanding of the epitope targets, the functional characterization of pre-existing autoantibodies in transplantation is understudied. There is support and agreement that pre-existing autoantibodies are associated with graft injury and poor prognosis, however, conclusive evidence suggesting whether they are biomarkers of injury, pathogenic, beneficial, or simply epiphenomena of the diseases are lacking. Teasing out the biological function of preexisting autoantibodies may be difficult due to the interconnectedness of antibodies in multiple different immune responses (i.e., complement fixing and FcyR interactions) and the heterogeneity of antibody characteristics (i.e., Ig class and class subtypes). Nonetheless, functional characterization could lead to the development of patient risk stratification strategies if the autoantibodies prove to be biomarkers. Further, a better understanding of the type and function of autoantibodies could be leveraged to develop novel antibody-targeted therapeutics designed to reduce graft damage if autoantibodies are found to be pathogenic.

### Pre-Existing Autoantibody Immunoglobulin Classes, Subtypes, and Fab Specificity

The inherent heterogeneity among antibodies and their structure gives them the ability to mediate different immune responses. Antibodies, or immunoglobulins, are composed of two heavy and two light chains. The heavy chain, or constant domain (Fc), specifies effector function and the light chain, or variable domain (Fab), binds to antigen. There are five main Fc immunoglobulin (Ig) isotypes; IgM, IgG, IgA, IgE, and IgD. The IgG and IgA isotypes can be split into IgG1, IgG2, IgG3, and IgG4, and IgA1 and IgA2 subclasses, respectively.

The Fc immunoglobulin classes differ in various properties such as structure, complement fixation, and FcyR binding [For detailed review see: Schroeder and Cavacini (92)]. Historically, little attention was paid to IgA, IgD, and IgE in the context of autoantibodies because they function predominantly in mucosal response to pathogens, homeostasis, and allergens, respectively, and do not interact with FcyRs, and are incapable of complement fixation. Furthermore, while IgM makes up a group of antibodies termed natural antibodies that are capable of binding autoantigens, natural IgM antibodies have relatively low affinity and are not hypothesized to be drivers of autoimmune disease or pathogenesis (92). Natural IgM antibodies role in IRI and IgG (total IgG, not IgG subtypes) have received most of the attention from those studying autoantibodies (93-98). While focusing on total IgG autoantibodies in pulmonary disease has proven to be fruitful, we know little about the role if any of other Ig classes and IgG subtypes in autoreactive-mediated graft damage. Ig isotypes and subclasses have been studied in the context of alloantibodies (DSA and anti-HLA antibodies), however much less is known with regard to pre-existing autoantibodies (non-HLA autoantibodies). Comparatively fewer reports have investigated the mechanistic significance of Ig sub-classes of pre-existing autoantibodies in LTx. Elucidating whether Ig subclasses are clinically relevant in the context of LTx may well provide novel information that could be therapeutically leveraged to improve graft outcomes (33). Therefore, it is essential to study the multiple characteristics of pre-existing autoantibodies to comprehensively understand if they have any impact on graft injury. Below we will discuss different immunoglobulins and postulate on the potential impact they may have on pre-existing autoantibody-mediated graft damage.

IgA. IgA has been well studied in the context of mucosal immunity and renal function (IgA nephropathy), however, little is known about IgA and solid organ transplantation. A singular report has analyzed pre-existing IgA autoantibodies in LTx patients. Kaza et al. (46) performed a proteomic microarray of 124 potential antigens to measure serum profiles of IgG and IgA pre-existing autoantibodies. Although IgG is present at a higher concentration in the serum than IgA, the authors reported larger numbers of pre-existing IgA autoantibodies compared to preexisting IgG autoantibodies. Furthermore, only pre-existing IgA autoantibodies correlated with PGD development and survival (46). One explanation for higher IgA pre-existing autoantibodies could be the fact that IgA plays an important role in mucosal immunologic defense. Some of the IgA pre-existing autoantibodies identified were directed against proteoglycans that may be present in the respiratory mucosal system and extracellular matrix. IgA can promote pro-inflammatory responses when interacting with the FcaRI on neutrophils (99-101). Given the importance of the respiratory mucosal system, neutrophil infiltration and damage in LTx, it is reasonable to hypothesize that IgA-immune complexes may promote graft damage after binding mucosal and ECM autoantigens. Interestingly, recent evidence suggests that molecular phenotyping of mucosal biopsies can be used to detect and accurately diagnose rejection following LTx (102).

A group of studies examining pre-existing IgA autoantibodies, although in kidney Tx patients, identified a positive correlation with kidney graft survival. In contrast to most examples thus far, the authors demonstrated that the presence of pre-existing IgA autoantibodies directed against the Fab region of human IgG molecule increased the chance of kidney graft survival (103). Although the aforementioned studies were in a different solid organ Tx system, these studies suggest that whether an autoantibody is pathogenic or advantageous is dependent on the antigen. More support for IgA antibodies impacting transplant outcomes lies in the observation that low pre-transplant levels of systemic IgA antibodies have been associated with increased post-LTx mortality (104). Although no specific antigens were investigated in this study, increased mortality was hypothesized to result from increased risk of post-LTx infection, therefore, not necessarily autoantibody mediated. Nonetheless, these studies demonstrate that much more research is needed to better understand the relationship between IgA antibodies, their corresponding antigens, differences in IgA subclass, and their function/role in transplant outcome.

*IgG*. The connection between IgG and transplant success has been well studied for over forty years. IgG has been studied in the context of DSA and HLA- and non-HLA antibodies, antibody mediated rejection, and hypogammaglobulinemia, or immunoglobulin deficiency, in lung transplant. In general, studies examining total IgG indicate that expression levels and their antigen of target (HLA and non-HLA) play crucial roles in transplantation outcome. However, significantly less research has focused on the influence and importance of different IgG subclasses. The four IgG subclasses, IgG1, IgG2, IgG3, and IgG4, have different physical properties, abundance, and effector functions. For example, IgG1-IgG3 are complement fixing with IgG3 having highest binding efficiency to C1q followed by IgG1 and IgG2 (105). However, IgG4 does not appear to have any complement activating ability and is often associated with immunotolerance (106, 107). IgG subclasses also demonstrate different binding affinities for FcyRs: IgG1 and IgG3 bind FcyRI-III, while IgG4 binds FcyR I and II, and IgG2 only binds FcyRII (92). More details describing the differences in IgG subclasses can be found in elegant reviews by Schroeder and Cavacini (92) and in **Figure 3**. Overall, the differences in these properties between subclasses could greatly influence the effector functions of a pre-existing IgG autoantibody.

Unfortunately, very few papers have delved into characterizing the IgG subclasses of pre-existing autoantibodies in LTx. Our group recently performed a study measuring the level of anti-elastin preexisting autoantibody IgG subclasses in an emphysema mouse model. We discovered that all four IgG anti-elastin pre-existing autoantibodies are elevated in CS exposed mice compared to controls. These pre-existing autoantibodies were determined to be pathogenic, however, it remains unclear whether one subclass of IgG is more pathogenic than another (33). The hypothesized mechanism of action for pre-existing autoantibody-mediated damage includes complement fixation and activation, therefore, IgG3 and IgG1 would likely be the predominantly pathogenic IgG subclasses in this case, but further studies are needed.

Due to the lack of publications on pre-existing IgG subclass autoantibodies, support for this hypothesis can be found by investigating the role of IgG subclasses in donor specific antigen DSA HLA-antibodies. For example, a retrospective study analyzing individual IgG subclass DSAs following liver transplant found that patients positive for DSAs presented with IgG1 most often, followed by IgG3, and then IgG4 and IgG2. Patients that demonstrated chronic rejection often times had a combination of IgG subclasses, but patients with IgG3 DSAs were at increased risk of graft loss (108). A similar observation was made in patients following renal transplant. Antibody mediated rejection was more prevalent in patients with IgG1/IgG3 de novo DSA antibodies compared to a mixture of complement and non-complement fixing de novo DSA antibodies (109). In contrast, patients with pre-existing complement-fixing DSA IgG subclasses did not demonstrate significantly different incidence of antibody mediated rejection (AMR) as compared to patients with a combination of preexisting IgG1/IgG3 and IgG2/IgG4 DSA antibodies in kidney transplant (110). However, there was a trend that ABMR was higher 6 months post-Tx in patients with pre-existing complement fixing DSA antibodies. Taken together, these data suggest that IgG subclass DSA antibodies, whether pre-existing or developed de novo, impact outcomes. Therefore, it is reasonable to hypothesize that pre-existing non-HLA autoantibody Ig subtype analysis could prove beneficial in predicting graft outcomes.

*IgE/IgD*. To date, there are no studies investigating IgE or IgD autoantibodies in the context of transplantation.

### **Pre-Existing Autoantibody Glycan Profiles**

Antibody glycosylation is a fundamental biochemical reaction determined by various biological factors that significantly



FIGURE 3 | Pre-Existing Autoantibody Characteristics Impact Effector Functions. Autoantibodies bind to a variety of different antigens expressed on the cell surface of damaged cells or damaged/modified proteins that exist in the extracellular space. Upon binding, autoantibodies work to fine-tune the activation or suppression of the immune response. Inherent antibody characteristics such as antigen specificity, Ig class, IgG/A subclass, and glycosylation give autoantibodies the ability to regulate the immunological response to stimuli. Antigenic specificity is determined by the Fab' portion of the antibody while effector function is dependent on the Fc portion of the antibody. Immune responses can be pro-inflammatory or anti-inflammatory based on the target antigen. Ig classes (IgM/G/A/E/D) have different immune activation capabilities and activate different immune pathways. For example, IgG and IgM are able to fix complement while IgA, IgD, and IgE are not. IgG/A subclasses are also capable of modulating effector functions. IgG1-4 have different complement fixation capabilities and interact with different FcyR's, thereby, fine-tuning the immune response.

influence disease and immune states (111-121). All Ig families are glycosylated and previous review articles have elegantly highlighted the biological significance of Ig glycosylation in multiple disease settings (113, 114, 120, 121). A conserved site of N-glycosylation at N297 in the heavy chain (HC) region is common in the IgG class of antibodies which maintains the structure and stability of the Fc region (Figure 4) (92). IgG glycan alterations have significant impact on effector functions, such as immune cell FcvR interactions and altered complement activation, with some glycan signatures being predominantly pro-inflammatory, while others render IgG anti-inflammatory (120, 122–128). Given the importance of Ig in transplantation, it is clear that investigating Ig characteristics such as glycosylation state could well be essential in increasing our understanding of the role of antibodies in solid organ transplant. Unfortunately, to date little is known regarding Ig glycosylation status in solid organ transplant in general, and even less is known regarding autoantibodies in LTx.

Experimental evidence has demonstrated that serum and IgG N-glycan signatures change with disease and impact disease progression [Reviewed by: Goulabchand et al. (112); Maverakis et al. (121); and Zhou et al. (129)]. For example, COPD patients have more complex glycan structures and a decrease in monogalactosylated species compared to healthy individuals (130). In fact, the more complex glycan structures correlate with COPD GOLD stage progression. Similar serum N-glycan signature changes were observed in the IgG N-glycan signature (130). Therefore, it would be reasonable to hypothesize that LTx

patients present with pre-existing autoantibodies that have proinflammatory N-glycan signatures.

In support of this hypothesis, what little data that does exist on glycosylation in Tx has demonstrated that changes in the Nglycome are predictive of poorer outcomes in liver and kidney transplantation (131-134). Interestingly, one group studying IgG N-glycosylation in hematopoietic stem cell transplantation (HSCT) found that the recipients' IgG glycosylation status post-Tx does not mimic the donor profile, and instead is determined by environmental factors of the host (135). These data could suggest that the IgG N-glycan profile present pretransplant could be pro-inflammatory due to underlying pulmonary disease. A well-established pro-inflammatory environment resulting from pulmonary disease combined with pro-inflammatory processes inherent to transplantation may lead to environmental pressures that maintain proinflammatory IgG N-glycosylation states. Furthermore, unlike in HSCT, the LTx recipient's B cell population would remain of recipient origin and therefore already be primed to produce proinflammatory N-glycan signature IgG autoantibodies.

## **CONCLUDING REMARKS**

Current immunosuppressive strategies only start upon transplantation and have little impact on B cell functions and pre-existing antibodies. While de-sensitization strategies exist for





pre-sensitized patients no studies have investigated the impact of autoantibody depletion on lung transplant outcomes. Preexisting autoantibodies such as anti-Col-V, anti-KAT, and anti-elastin have been demonstrated to cause inflammatorymediated graft damage and promote epitope spreading in murine models. Furthermore, anti-Col-V and anti-KAT autoantibodies have been identified in patient samples and correlate with LTx rejection. Multiple other pre-existing autoantibodies have been inconsistently correlated with increased risk of PGD, and CLAD. While a greater understanding of the repertoires present is slowly emerging, little is still known regarding the antibody characteristics that may fine-tune the pathogenic mechanisms of autoantibodies in organ transplantation. Immunoglobulin class (IgM, IgG, IgA, IgE, IgD) and subclass (IgG1-4; IgA1-2) have all been shown to interact with different Fc receptors and have different complement fixation capabilities. Structural changes to the Fc portion of immunoglobulins such as glycosylation can also greatly impact the antibody-mediated inflammatory response. Unfortunately, little is known with regard to these antibody features and as such the pathogenic mechanisms and corresponding antibody characteristics of disease specific preexisting autoantibodies is largely still unknown. Gaining a better

### REFERENCES

- Thabut G, Mal H. Outcomes After Lung Transplantation. J Thorac Dis (2017) 9:2684–91. doi: 10.21037/jtd.2017.07.85
- Kheradmand F, Shan M, Xu C, Corry DB. Autoimmunity in Chronic Obstructive Pulmonary Disease: Clinical and Experimental Evidence. *Expert Rev Clin Immunol* (2012) 8:285–92. doi: 10.1586/eci.12.7
- Cojocaru M, Cojocaru IM, Silosi I, Vrabie CD. Pulmonary Manifestations of Systemic Autoimmune Diseases. In: *Maedica*, vol. 6. Springer International Publishing (2011). p. 224–9.
- Willemse BWM, Postma DS, Timens W, ten Hacken NHT. The Impact of Smoking Cessation on Respiratory Symptoms, Lung Function, Airway Hyperresponsiveness and Inflammation. *Eur Respir J* (2004) 23:464–76. doi: 10.1183/09031936.04.00012704
- Yang ML, Doyle HA, Clarke SG, Herold KC, Mamula MJ. Oxidative Modifications in Tissue Pathology and Autoimmune Disease. *Antioxidants Redox Signal* (2018) 29:1415–31. doi: 10.1089/ars.2017.7382
- Rasheed N, Rasheed Z. Oxidative Biomolecular Damage: A Possible Mechanism for Systemic Autoimmunity. Int J Health Sci (Qassim) (2019) 13:1–3.
- Wen L, Krauss-Etschmann S, Petersen F, Yu X. Autoantibodies in Chronic Obstructive Pulmonary Disease. *Front Immunol* (2018) 9:1–7. doi: 10.3389/ fimmu.2018.00066
- Feghali-Bostwick CA, Wilkes DS. Autoimmunity in Idiopathic Pulmonary Fibrosis. Am J Respir Crit Care Med (2011) 183:692–3. doi: 10.1164/ rccm.201010-1727ED
- Feghali-Bostwick CA, Gadgil AS, Otterbein LE, Pilewski JM, Stoner MW, Csizmadia E, et al. Autoantibodies in Patients With Chronic Obstructive Pulmonary Disease. *Am J Respir Crit Care Med* (2008) 177:156–63. doi: 10.1164/rccm.200701-014OC
- Majo J, Ghezzo H, Cosio MG. Lymphocyte Population and Apoptosis in the Lungs of Smokers and Their Relation to Emphysema. *Eur Respir J* (2001) 17:946–53. doi: 10.1183/09031936.01.17509460
- Strzelak A, Ratajczak A, Adamiec A, Feleszko W. Tobacco Smoke Induces and Alters Immune Responses in the Lung Triggering Inflammation, Allergy, Asthma and Other Lung Diseases: A Mechanistic Review. *Int J Environ Res Public Health* (2018) 15:1033. doi: 10.3390/ijerph15051033

understanding of the epitopes, Ig-sub class and glycosylation state may facilitate the development of novel personalized pharmacotherapeutics that can be leveraged to augment immunosuppression and improve the outcomes of lung transplant recipients.

## **AUTHOR CONTRIBUTIONS**

All authors contributed to the conceptualization, writing, and editing of the manuscript. All authors contributed to the article and approved the submitted version.

## FUNDING

These studies were supported by grants from the NIH (NHLBI R1R01HL 140470-01A1, NHLBI 3R01HL140470-03S1 **an**d NIAID U01 AI132894-01 to CA), TL1 TR001451 to AM, U01CA242096 to PMA (multi-PI), (NHLBI K24-HL115354, NHLBI R01HL087115, and NHLBI U01-HL145435 to JC).

- Byrne R, Todd I, Tighe PJ, Fairclough LC. Autoantibodies in Chronic Obstructive Pulmonary Disease: A Systematic Review. *Immunol Lett* (2019) 214:8–15. doi: 10.1016/j.imlet.2019.08.007
- Xiao ZX, Miller JS, Zheng SG. An Updated Advance of Autoantibodies in Autoimmune Diseases. *Autoimmun Rev* (2020) 20:102743. doi: 10.1016/ j.autrev.2020.102743
- Ma A, Wen L, Yin J, Hu Y, Yue X, Li J, et al. Serum Levels of Autoantibodies Against Extracellular Antigens and Neutrophil Granule Proteins Increase in Patients With COPD Compared to Non-COPD Smokers. *Int J COPD* (2020) 15:189–200. doi: 10.2147/COPD.S235903
- Avrameas S, Selmi C. Natural Autoantibodies in the Physiology and Pathophysiology of the Immune System. J Autoimmun (2013) 41:46–9. doi: 10.1016/j.jaut.2013.01.006
- Nagele EP, Han M, Acharya NK, DeMarshall C, Kosciuk MC, Nagele RG. Natural IgG Autoantibodies Are Abundant and Ubiquitous in Human Sera, and Their Number Is Influenced By Age, Gender, and Disease. *PloS One* (2013) 8:e60726. doi: 10.1371/journal.pone.0060726
- Duncan SR. Clues, Not Conclusions. Nature (2012) 489:S15. doi: 10.1038/ 489S15a
- Benfaremo D, Baroni SS, Manfredi L, Moroncini G, Gabrielli A. Putative Functional Pathogenic Autoantibodies in Systemic Sclerosis. *Eur J Rheumatol* (2020) 7:181–6. doi: 10.5152/eurjrheum.2020.19131
- 19. Sato S, Hoshino K, Satoh T, Fujita T, Kawakami Y, Fujita T, et al. RNA Helicase Encoded by Melanoma Differentiation-Associated Gene 5 Is a Major Autoantigen in Patients With Clinically Amyopathic Dermatomyositis: Association With Rapidly Progressive Interstitial Lung Disease. Arthritis Rheumatol (2009) 60:2193–200. doi: 10.1002/art.24621
- Allenbach Y, Leroux G, Suárez-Calvet X, Preusse C, Gallardo E, Hervier B, et al. Dermatomyositis With or Without Anti-Melanoma Differentiation-Associated Gene 5 Antibodies. *Am J Pathol* (2016) 186:691–700. doi: 10.1016/j.ajpath.2015.11.010
- Sontheimer RD. MDA5 Autoantibody-Another Indicator of Clinical Diversity in Dermatomyositis. Ann Transl Med (2017) 5:1–11. doi: 10.21037/atm.2017.03.94
- 22. Cao H, Pan M, Kang Y, Xia Q, Li X, Zhao X, et al. Clinical Manifestations of Dermatomyositis and Clinically Amyopathic Dermatomyositis Patients With Positive Expression of Anti-Melanoma Differentiation-Associated Gene 5 Antibody. Arthritis Care Res (2012) 64:1602–10. doi: 10.1002/acr.21728

- Chaisson NF, Paik J, Orbai AM, Casciola-Rosen L, Fiorentino D, Danoff S, et al. A Novel Dermato-Pulmonary Syndrome Associated With MDA-5 Antibodies: Report of 2 Cases and Review of the Literature. *Med (United States)* (2012) 91:220–8. doi: 10.1097/MD.0b013e3182606f0b
- 24. Fischer A, Antoniou KM, Brown KK, Cadranel J, Corte TJ, Du Bois RM, et al. An Official European Respiratory Society/American Thoracic Society Research Statement: Interstitial Pneumonia With Autoimmune Features. *Eur Respir J* (2015) 46:976–87. doi: 10.1183/13993003.00150-2015
- 25. Jee AS, Adelstein S, Bleasel J, Keir GJ, Nguyen M, Sahhar J, et al. Role of Autoantibodies in the Diagnosis of Connective-Tissue Disease ILD (CTD-ILD) and Interstitial Pneumonia With Autoimmune Features (IPAF). J Clin Med (2017) 6:51. doi: 10.3390/jcm6050051
- Elkon K, Casali P. Nature and Functions of Autoantibodies. Nat Clin Pract Rheumatol (2008) 4:491–8. doi: 10.1038/ncprheum0895
- Watts RP, Thom O, Fraser JF. Inflammatory Signalling Associated With Brain Dead Organ Donation: From Brain Injury to Brain Stem Death and Posttransplant Ischaemia Reperfusion Injury. J Transpl (2013) 2013:1–19. doi: 10.1155/2013/521369
- Petrenko A, Carnevale A, Somov A, Osorio A, Rodríguez A, Guibert A, et al. Organ Preservation Into the 2020s: The Era of Dynamic Intervention. *Transfus Med Hemotherapy* (2019) 46:151–72. doi: 10.1159/000499610
- Giwa S, Lewis JK, Alvarez L, Langer R, Roth AE, Church GM, et al. The Promise of Organ and Tissue Preservation to Transform Medicine HHS Public Access Author Manuscript. *Nat Biotechnol* (2017) 35:530–42. doi: 10.1038/nbt.3889
- Cardinal H, Dieudé M, Hébert MJ. The Emerging Importance of Non-HLA Autoantibodies in Kidney Transplant Complications. J Am Soc Nephrol (2017) 28:400–6. doi: 10.1681/ASN.2016070756
- Nunoda S. Impact of Pretransplant Antibodies on Outcomes After Heart Transplantation. *Curr Opin Organ Transpl* (2019) 24:220–6. doi: 10.1097/ MOT.00000000000629
- 32. Sureshbabu A, Fleming T, Mohanakumar T. Autoantibodies in Lung Transplantation. *Transpl Int* (2019) 8350:0–1. doi: 10.1111/tri.13487
- 33. Patel KJ, Cheng Q, Stephenson S, Allen D, Li P, Kilkenny C, et al. Emphysema-Associated Autoreactive Antibodies Exacerbate Post-Lung Transplant Ischemia-Reperfusion Injury. Am J Respir Cell Mol Biol (2019) 60:678–86. doi: 10.1165/rcmb.2018-0224OC
- 34. Vinson AD, Rampolla-Selles R, Cooper ES, Alquist CR. Non-Human Leukocyte Antigen Antibody-Mediated Lung Transplant Rejection: The Other Anti-a. Ochsner J (2018) 18:260–3. doi: 10.31486/toj.17.0068
- 35. Diamond JM, Lee JC, Kawut SM, Shah RJ, Localio AR, Bellamy SL, et al. Clinical Risk Factors for Primary Graft Dysfunction After Lung Transplantation. Am J Respir Crit Care Med (2013) 187:527–34. doi: 10.1164/rccm.201210-1865OC
- Burton CM, Iversen M, Milman M, Zemtsovski N, Carlsen M, Steinbrüchel J, et al. Outcome of Lung Transplanted Patients With Primary Graft Dysfunction. Eur J Cardio Thoracic Surg (2007) 31:75–82. doi: 10.1016/ j.ejcts.2006.10.024
- Suzuki Y, Cantu E, Christie J. Primary Graft Dysfunction. Semin Respir Crit Care Med (2013) 34:305–19. doi: 10.1055/s-0033-1348474
- Altun GT, Arslantaş MK, Cinel İ. Primary Graft Dysfunction After Lung Transplantation. *Turk Anesteziyoloji ve Reanimasyon Dern Derg* (2015) 43:418–23. doi: 10.5152/TJAR.2015.16443
- Tiriveedhi V, Gautam B, Sarma N, Askar J, Budev M, Aloush M, et al. Pre-Transplant Antibodies to Kα1 Tubulin and Collagen-V in Lung Transplantation: Clinical Correlations. J Hear Lung Transpl (2013) 32:807–14. doi: 10.1016/j.healun.2013.06.003
- Iwata T, Philipovskiy A, Fisher AJ, Presson RG, Chiyo M, Lee J, et al. Anti-Type V Collagen Humoral Immunity in Lung Transplant Primary Graft Dysfunction. J Immunol (2008) 181:5738–47. doi: 10.4049/jimmunol. 181.8.5738
- 41. Yoshida S, Haque A, Mizobuchi T, Iwata T, Chiyo M, Webb TJ, et al. Anti-Type V Collagen Lymphocytes That Express IL-17 and IL-23 Induce Rejection Pathology in Fresh and Well-Healed Lung Transplants. *Am J Transpl* (2006) 6:724–35. doi: 10.1111/j.1600-6143.2006.01236.x
- Goers TA, Ramachandran S, Aloush A, Trulock E, Patterson GA, Mohanakumar T, et al. De Novo Production of K-α1 Tubulin-Specific

Antibodies: Role in Chronic Lung Allograft Rejection. J Immunol (2008) 180:4487–94. doi: 10.4049/jimmunol.180.7.4487

- 43. Zaffiri L, Shah RJ, Stearman R, Rothhaar S, Emtiazjoo K, Yoshimoto AM, et al. Collagen Type-V Is a Danger Signal Associated With Primary Graft Dysfunction in Lung Transplantation. *Transpl Immunol* (2019) 56:101224. doi: 10.1016/j.trim.2019.101224
- 44. Bharat A, Saini D, Steward N, Hachem R, Trulock EP, Patterson GA, et al. Antibodies to Self-Antigens Predispose to Primary Lung Allograft Dysfunction and Chronic Rejection. Ann Thorac Surg (2010) 90:1094–101. doi: 10.1016/j.athoracsur.2010.06.009
- 45. Bharat A, Chiu S, Zheng Z, Sun H, Yeldandi A, DeCamp MM, et al. Lung-Restricted Antibodies Mediate Primary Graft Dysfunction and Prevent Allotolerance After Murine Lung Transplantation. Am J Respir Cell Mol Biol (2016) 55:532–41. doi: 10.1165/rcmb.2016-0077OC
- 46. Kaza V, Zhu C, Feng L, Torres F, Bollineni S, Mohanka M, et al. Pre-Existing Self-Reactive IgA Antibodies Associated With Primary Graft Dysfunction After Lung Transplantation. *Transpl Immunol* (2020) 59:101271. doi: 10.1016/j.trim.2020.101271
- Hagedorn PH, Burton CM, Sahar E, Domany E, Cohen I, Flyvbjerg R, et al. Integrative Analysis Correlates Donor Transcripts to Recipient Autoantibodies in Primary Graft Dysfunction After Lung Transplantation. *Immunology* (2011) 132:394–400. doi: 10.1111/j.1365-2567.2010.03373.x
- Cheng G, Zhang N, Wang Y, Rui J, Yin X, Cui T, et al. Antibodies of IgG, IgA and IgM Against Human Bronchial Epithelial Cell in Patients With Chronic Obstructive Pulmonary Disease. *Clin Lab* (2016) 62:1101–8. doi: 10.7754/ Clin.Lab.2015.151020
- Rinaldi M, Lehouck A, Heulens N, Lavend'homme R, Carlier V, Saint-Remy J-M, et al. Antielastin B-Cell and T-Cell Immunity in Patients With Chronic Obstructive Pulmonary Disease. *Thorax* (2012) 67:694–700. doi: 10.1136/ thoraxjnl-2011-200690
- Lee SH, Goswami S, Grudo A, Song L-Z, Bandi V, Goodnight-White S, et al. Antielastin Autoimmunity in Tobacco Smoking-Induced Emphysema. *Nat Med* (2007) 13:567–9. doi: 10.1038/nm1583
- 51. Xiong Y, Gao S, Luo G, Cheng G, Huang W, Jiang R, et al. Increased Circulating Autoantibodies Levels of IgG, IgA, IgM Against Cytokeratin 18 and Cytokeratin 19 in Chronic Obstructive Pulmonary Disease. Arch Med Res (2017) 48:79–87. doi: 10.1016/j.arcmed.2017.01.007
- Kuo Y, Chang CA, Wu YK, Hsieh MJ, Tsai CH, Chen KT, et al. Identification and Clinical Association of Anti-Cytokeratin 18 Autoantibody in COPD. *Immunol Lett* (2010) 128:131–6. doi: 10.1016/j.imlet.2009.12.017
- Luo XM, Liu XY, Tang JH, Yang W, Ni ZH, Chen QG, et al. Autoantibodies Against CD80 in Patients With COPD. *Clin Transl Immunol* (2016) 5:e102. doi: 10.1038/cti.2016.57
- 54. Kirkham PA, Caramori G, Casolari P, Papi AA, Edwards M, Shamji B, et al. Oxidative Stress-Induced Antibodies to Carbonyl-Modified Protein Correlate With Severity of Chronic Obstructive Pulmonary Disease. Am J Respir Crit Care Med (2011) 184:796–802. doi: 10.1164/rccm.201010-1605OC
- 55. Gouda TM, Yousof AA, Al Salahy MM, Al mehy GF, Essawy TS, El-Shaer OS, et al. Study of Anti Nuclear and Anti Smooth Muscle Antibodies in Patients With Chronic Obstructive Pulmonary Disease. *Egypt J Chest Dis Tuberc* (2014) 63:49–56. doi: 10.1016/j.ejcdt.2013.11.001
- Morissette MC, Jobse B, Thayaparan N, Nikota D, Shen JK, Labiris P, et al. Persistence of Pulmonary Tertiary Lymphoid Tissues and Anti-Nuclear Antibodies Following Cessation of Cigarette Smoke Exposure. *Respir Res* (2014) 15:1–10. doi: 10.1186/1465-9921-15-49
- Núñez B, Sauleda J, Antó JM, Julià MR, Orozco M, Monsó E, et al. Anti-Tissue Antibodies Are Related to Lung Function in Chronic Obstructive Pulmonary Disease. Am J Respir Crit Care Med (2011) 183:1025–31. doi: 10.1164/rccm.201001-0029OC
- Bon J, Kahloon R, Zhang Y, Xue J, Fuhrman CR, Tan J, et al. Autoreactivity to Glucose Regulated Protein 78 Links Emphysema and Osteoporosis in Smokers. *PloS One* (2014) 9:e105066. doi: 10.1371/journal.pone.0105066
- Hu JY, Liu BB, Du YP, Zhang Y, Zhang YW, Zhang YY, et al. Increased Circulating β2-Adrenergic Receptor Autoantibodies Are Associated With Smoking-Related Emphysema. *Sci Rep* (2017) 7:1–11. doi: 10.1038/ srep43962

- 60. Bauhammer J, Blank N, Max R, Lorenz HM, Wagner U, Krause D, et al. Rituximab in the Treatment of Jo1 Antibody-Associated Antisynthetase Syndrome: Anti-Ro52 Positivity as a Marker for Severity and Treatment Response. J Rheumatol (2016) 43:1566–74. doi: 10.3899/jrheum.150844
- Rocha-Muñoz AD, Ponce-Guarneros M, Gamez-Nava JI, Olivas-Flores EM, Mejía M, Juárez-Contreras P, et al. Anti-Cyclic Citrullinated Peptide Antibodies and Severity of Interstitial Lung Disease in Women With Rheumatoid Arthritis. J Immunol Res (2015) 2015.
- 62. Giles JT, Danoff SK, Sokolove J, Wagner CA, Winchester R, Pappas DA, et al. Association of Fine Specificity and Repertoire Expansion of Anticitrullinated Peptide Antibodies With Rheumatoid Arthritis Associated Interstitial Lung Disease. Ann Rheumatol Dis (2014) 73:1487– 94. doi: 10.1136/annrheumdis-2012-203160
- 63. Weigold F, Günther J, Pfeiffenberger M, Cabral-Marques O, Siegert E, Dragun D, et al. Antibodies Against Chemokine Receptors CXCR3 and CXCR4 Predict Progressive Deterioration of Lung Function in Patients With Systemic Sclerosis. *Arthritis Res Ther* (2018) 20:1–9. doi: 10.1186/s13075-018-1545-8
- 64. Taillé C, Grootenboer-Mignot S, Boursier C, Michel L, Debray MP, Fagart J, et al. Identification of Periplakin as a New Target for Autoreactivity in Idiopathic Pulmonary Fibrosis. Am J Respir Crit Care Med (2011) 183:759– 66. doi: 10.1164/rccm.201001-0076OC
- 65. Sorohan BM, Ismail G, Leca N, Tacu D, Obriscă B, Constantinescu I, et al. Angiotensin II Type 1 Receptor Antibodies in Kidney Transplantation: An Evidence-Based Comprehensive Review. *Transpl Rev* (2020) 34. doi: 10.1016/j.trre.2020.100573
- Reinsmoen NL. Role of Angiotensin II Type 1 Receptor-Activating Antibodies in Solid Organ Transplantation. *Hum Immunol* (2013) 74:1474-7. doi: 10.1016/j.humimm.2013.06.034
- 67. González-Almada A, Arreola-Guerra JM, López-Sánchez JA, Cuevas E, Vilatoba M, Contreras AG, et al. Pretransplant Angiotensin II Type 1-Receptor Antibodies Point to an Increase in Renal Graft Sub-Intimal Fibrosis in Living- Donor Kidney Transplant Recipients. *Transpl Immunol* (2019) 54:1–8. doi: 10.1016/j.trim.2018.10.003
- Lukitsch I, Kehr J, Chaykovska L, Wallukat G, Nieminen-Kelhä M, Batuman V, et al. Renal Ischemia and Transplantation Predispose to Vascular Constriction Mediated by Angiotensin II Type 1 Receptor-Activating Antibodies. *Transplantation* (2012) 94:8–13. doi: 10.1097/TP.0b013e3182529bb7
- Dragun D, Müller DN, Bräsen JH, Fritsche L, Nieminen-Kelhä M, Dechend R, et al. Angiotensin II Type 1–Receptor Activating Antibodies in Renal-Allograft Rejection. N Engl J Med (2005) 352:558–69. doi: 10.1056/NEJM0a035717
- Kawai T, Forrester SJ, O-Brien S, Baggett A, Rizzo V, Eguchi S, et al. AT1 Receptor Signaling Pathways in the Cardiovascular System. *Pharmacol Res* (2017) 125:4–13. doi: 10.1016/j.phrs.2017.05.008
- 71. Günther J, Kill A, Becker M, Heidecke O, Rademacher H, Siegert J, et al. Angiotensin Receptor Type 1 and Endothelin Receptor Type A on Immune Cells Mediate Migration and the Expression of IL-8 and CCL18 When Stimulated by Autoantibodies From Systemic Sclerosis Patients. *Arthritis Res Ther* (2014) 16:1–14. doi: 10.1186/ar4503
- 72. Lefaucheur C, Viglietti D, Bouatou Y, Philippe A, Pievani D, Aubert O, et al. Non-HLA Agonistic Anti-Angiotensin II Type 1 Receptor Antibodies Induce a Distinctive Phenotype of Antibody-Mediated Rejection in Kidney Transplant Recipients. *Kidney Int* (2019) 96:189–201. doi: 10.1016/ j.kint.2019.01.030
- Fuss A, Hope CM, Deayton S, Bennett GD, Holdsworth R, Carroll RP, et al. C4d-Negative Antibody-Mediated Rejection With High Anti-Angiotensin II Type I Receptor Antibodies in Absence of Donor-Specific Antibodies. Nephrology (2015) 20:467–73. doi: 10.1111/nep.12441
- 74. Budding K, van de Graaf E, Hoefnagel A, Kwakkel-van Erp T, van Kessel JM, Dragun D, et al. Anti-ETAR and Anti-AT1R Autoantibodies Are Elevated in Patients With Endstage Cystic Fibrosis. J Cyst Fibros (2015) 14:42–5. doi: 10.1016/j.jcf.2014.07.007
- 75. Reinsmoen NL, Lai CH, Heidecke H, Haas M, Cao K, Ong G, et al. Anti-Angiotensin Type 1 Receptor Antibodies Associated With Antibody Mediated Rejection in Donor HLA Antibody Negative Patients. *Transplantation* (2010) 90:1473–7. doi: 10.1097/TP.0b013e3181fd97f1
- 76. Hachem RR, Yusen RD, Meyers BF, Aloush AA, Mohanakumar T, Patterson GA, et al. Anti-Human Leukocyte Antigen Antibodies and Preemptive

Antibody-Directed Therapy After Lung Transplantation. J Hear Lung Transpl (2010) 29:973–80. doi: 10.1016/j.healun.2010.05.006

- 77. Morrell R, Pilewski JM, Gries CJ, Pipeling MR, Crespo MM, Ensor CR, et al. De Novo Donor-Specific HLA Antibodies Are Associated With Early and High-Grade Bronchiolitis Obliterans Syndrome and Death After Lung Transplantation. J Hear Lung Transpl (2014) 33:1288–94. doi: 10.1016/ j.healun.2014.07.018
- Safavi S, Robinson DR, Soresi S, Carby M, Smith JD. De Novo Donor HLA-Specific Antibodies Predict Development of Bronchiolitis Obliterans Syndrome After Lung Transplantation. J Hear Lung Transpl (2014) 33:1273–81. doi: 10.1016/j.healun.2014.07.012
- Pearl MH, Chen L, ElChaki R, Elashoff D, Gjertson D, Rossetti W, et al. Endothelin Type A Receptor Antibodies Are Associated With Angiotensin II Type 1 Receptor Antibodies, Vascular Inflammation, and Decline in Renal Function in Pediatric Kidney Transplantation. *Kidney Int Rep* (2020) 5:1925–36. doi: 10.1016/j.ekir.2020.09.004
- Chambers DC, Yusen R, Cherikh D, Goldfarb WS, Kucheryavaya SB, Khusch AY, et al. The Registry of the International Society for Heart and Lung Transplantation: Thirty-Fourth Adult Lung And Heart-Lung Transplantation Report—2017; Focus Theme: Allograft Ischemic Time. *J Hear Lung Transpl* (2017) 36:1047–59. doi: 10.1016/j.healun.2017.07.016
- Weigt S, DerHovanessian A, Wallace W, Lynch J, Belperio J. Bronchiolitis Obliterans Syndrome: The Achilles' Heel of Lung Transplantation. Semin Respir Crit Care Med (2013) 34:336–51. doi: 10.1055/s-0033-1348467
- Verleden GM, Glanville AR, Lease ED, Fisher AJ, Calabrese F, Corris PA, et al. Chronic Lung Allograft Dysfunction: Definition, Diagnostic Criteria, and Approaches to Treatment—A Consensus Report From the Pulmonary Council of the ISHLT. *J Hear Lung Transpl* (2019) 38:493–503. doi: 10.1016/ j.healun.2019.03.009
- Glanville AR, Verleden GM, Todd JL, Benden C, Calabrese F, Gottlieb J, et al. Chronic Lung Allograft Dysfunction: Definition and Update of Restrictive Allograft Syndrome—A Consensus Report From the Pulmonary Council of the ISHLT. J Hear Lung Transpl (2019) 38:483–92.
- Hagedorn PH, Burton C, Carlsen M, Steinbrüchel J, Andersen D, Sahar CB, et al. Chronic Rejection of a Lung Transplant Is Characterized by a Profile of Specific Autoantibodies. *Immunology* (2010) 130:427–35. doi: 10.1111/ j.1365-2567.2010.03246.x
- Kaza V, Zhu C, Terada L, Wang S, Torres L, Bollineni F, et al. Self-Reactive Antibodies Associated With Bronchiolitis Obliterans Syndrome Subtype of Chronic Lung Allograft Dysfunction. *Hum Immunol* (2020) 82:25–35. doi: 10.1016/j.humimm.2020.10.006
- Lopez-Soler RI, Borgia J, Kanangat A, Fhied S, Conti CL, Constantino DJ, et al. Anti-Vimentin Antibodies Present at the Time of Transplantation May Predict Early Development of Interstitial Fibrosis/Tubular Atrophy. *Transpl Proc* (2016) 48:2023–33. doi: 10.1016/j.transproceed.2016.04.009
- Zhang X, Reinsmoen NL. Impact of Non-Human Leukocyte Antigen-Specific Antibodies in Kidney and Heart Transplantation. *Front Immunol* (2017) 8:1–7. doi: 10.3389/fimmu.2017.00434
- Giral M, Foucher Y, Dufay A, Van Huyen JPD, Renaudin K, Moreau A, et al. Pretransplant Sensitization Against Angiotensin II Type 1 Receptor Is a Risk Factor for Acute Rejection and Graft Loss. *Am J Transpl* (2013) 13:2567–76. doi: 10.1111/ajt.12397
- Rose ML. Role of Anti-Vimentin Antibodies in Allograft Rejection. Hum Immunol (2013) 74:1459–62. doi: 10.1016/j.humimm.2013.06.006
- Li FJ, Surolia R, Li H, Wang Z, Kulkarni T, Liu G, et al. Autoimmunity to Vimentin Is Associated With Outcomes of Patients With Idiopathic Pulmonary Fibrosis. J Immunol (2017) 199:1596–605. doi: 10.4049/jimmunol.1700473
- 91. Yang Y, Fujita J, Bandoh S, Ohtsuki Y, Yamadori I, Yoshinouchi T, et al. Detection of Antivimentin Antibody in Sera of Patients With Idiopathic Pulmonary Fibrosis and Non-Specific Interstitial Pneumonia. *Clin Exp Immunol* (2002) 128:169–74. doi: 10.1046/j.1365-2249.2002.01811.x
- 92. Schroeder HW, Cavacini L. Structure and Function of Immunoglobulins. *J Allergy Clin Immunol* (2010) 125:S41–52. doi: 10.1016/j.jaci.2009.09.046
- Zhang M, Alicot EM, Chiu I, Li J, Verna N, Vorup-Jensen T, et al. Identification of the Target Self-Antigens in Reperfusion Injury. J Exp Med (2006) 203:141–52. doi: 10.1084/jem.20050390
- 94. Zhang M, Austen WG, Chiu I, Alicot EM, Hung R, Ma M, et al. Identification of a Specific Self-Reactive IgM Antibody That Initiates

Intestinal Ischemia/Reperfusion Injury. Proc Natl Acad Sci USA (2004) 101:3886–91.

- Elvington A, Atkinson C, Kulik L, Zhu H, Yu J, Kindy MS, et al. Pathogenic Natural Antibodies Propagate Cerebral Injury Following Ischemic Stroke in Mice. J Immunol (2012) 188:1460–8. doi: 10.4049/jimmunol.1102132
- 96. Kulik L, Fleming SD, Moratz C, Reuter JW, Novikov A, Chen K, et al. Pathogenic Natural Antibodies Recognizing Annexin IV Are Required to Develop Intestinal Ischemia-Reperfusion Injury. J Immunol (2009) 182:5363–73. doi: 10.4049/jimmunol.0803980. PMID: 1000000221
- Atkinson C, Qiao F, Yang X, Zhu P, Reaves N, Kulik L, et al. Targeting Pathogenic Postischemic Self-Recognition by Natural IgM to Protect Against Posttransplantation Cardiac Reperfusion Injury. *Circulation* (2015) 131:1171– 80. doi: 10.1161/CIRCULATIONAHA.114.010482
- Zhang M, Alicot EM, Carroll MC. Human Natural IgM can Induce Ischemia/Reperfusion Injury in a Murine Intestinal Model. *Mol Immunol* (2008) 45:4036–9. doi: 10.1016/j.molimm.2008.06.013
- Breedveld A, Van Egmond M. IgA and Fcαri: Pathological Roles and Therapeutic Opportunities. Front Immunol (2019) 10:553. doi: 10.3389/ fimmu.2019.00553
- 100. Hansen IS, Baeten DLP, den Dunnen J. The Inflammatory Function of Human IgA. Cell Mol Life Sci (2019) 76:1041–55. doi: 10.1007/s00018-018-2976-8
- 101. Aleyd E, Heineke MH, van Egmond M. The Era of the Immunoglobulin A Fc Receptor Fcαri; Its Function and Potential as Target in Disease. *Immunol Rev* (2015) 268:123–38. doi: 10.1111/imr.12337
- 102. Halloran K, Parkes MD, Timofte IL, Snell GI, Westall GP, Hachem R, et al. Molecular Phenotyping of Rejection-Related Changes in Mucosal Biopsies From Lung Transplants. Am J Transpl (2020) 20:954–66. doi: 10.1111/ ajt.15685
- 103. Susal C, Dohler B, Opelz G. Graft-Protective Role of High Pretransplantation IgA-Anti-Fab Autoantibodies: Confirmatory Evidence Obtained in More Than 4000 Kidney Transplants. The Collaborative Transplant Study. *Transplantation* (2000) 69:1337–40. doi: 10.1097/00007890-200004150-00021
- 104. Murthy SC, Avery RK, Budev M, Gupta S, Pettersson GB, Nowicki ER, et al. Low Pretransplant IgA Level Is Associated With Early Post–Lung Transplant Seromucous Infection. J Thorac Cardiovasc Surg (2018) 156:882–91.e8. doi: 10.1016/j.jtcvs.2018.03.165
- 105. Thomas KA, Valenzuela NM, Reed EF. The Perfect Storm: HLA Antibodies, Complement, Fcγrs, and Endothelium in Transplant Rejection. *Trends Mol Med* (2015) 21:319–29. doi: 10.1016/j.molmed.2015.02.004
- 106. Valenzuela NM, Hickey MJ, Reed EF. Antibody Subclass Repertoire and Graft Outcome Following Solid Organ Transplantation. *Front Immunol* (2016) 7:433. doi: 10.3389/fimmu.2016.00433
- 107. Koneczny I. A New Classification System for IgG4 Autoantibodies. Front Immunol (2018) 9:97. doi: 10.3389/fimmu.2018.00097
- 108. Kaneku H, O'Leary JG, Taniguchi M, Susskind BM, Terasaki PI, Klintmalm GB, et al. Donor-Specific Human Leukocyte Antigen Antibodies of the Immunoglobulin G3 Subclass Are Associated With Chronic Rejection and Graft Loss After Liver Transplantation. *Liver Transpl* (2012) 18:984–92. doi: 10.1002/lt.23451
- 109. Arnold ML, Ntokou ML, Doxiadis IS, Spriewald IIN, Boletis BM, Iniotaki JN, et al. Donor-Specific HLA Antibodies: Evaluating the Risk for Graft Loss in Renal Transplant Recipients With Isotype Switch From Complement Fixing IgG1/IgG3 to Noncomplement Fixing IgG2/IgG4 Anti-HLA Alloantibodies. *Transpl Int* (2014) 27:253–61. doi: 10.1111/tri.12206
- 110. Hönger G, Hopfer G, Arnold H, Spriewald ML, Schaub BM, Amico S, et al. Pretransplant IgG Subclasses of Donor-Specific Human Leukocyte Antigen Antibodies and Development of Antibody-Mediated Rejection. *Transplantation* (2011) 92:41–7. doi: 10.1097/TP.0b013e31821cdf0d
- Nose M, Wigzell H. Biological Significance of Carbohydrate Chains on Monoclonal Antibodies. *Proc Natl Acad Sci* (1983) 80:6632–6. doi: 10.1073/ pnas.80.21.6632
- 112. Goulabchand R, Vincent T, Batteux F, Eliaou J. François & Guilpain, P. Impact of Autoantibody Glycosylation in Autoimmune Diseases. *Autoimmun Rev* (2014) 13:742–50. doi: 10.1016/j.autrev.2014.02.005
- Reily C, Stewart TJ, Renfrow MB, Novak J. Glycosylation in Health and Disease. Nat Rev Nephrol (2019) 15:346–66. doi: 10.1038/s41581-019-0129-4
- 114. Cobb BA. The History of IgG Glycosylation and Where We Are Now. Glycobiology (2020) 30:202–13. doi: 10.1093/glycob/cwz065

- 115. Hipgrave Ederveen AL, de Haan N, Baerenfaenger M, Lefeber DJ, Wuhrer M. Dissecting Total Plasma and Protein-Specific Glycosylation Profiles in Congenital Disorders of Glycosylation. *Int J Mol Sci* (2020) 21:7635. doi: 10.3390/ijms21207635
- 116. Plomp R, Ruhaak LR, Uh HW, Reiding KR, Selman M, Houwing-Duistermaat JJ, et al. Subclass-Specific IgG Glycosylation Is Associated With Markers of Inflammation and Metabolic Health. Sci Rep (2017) 7:1– 10. doi: 10.1038/s41598-017-12495-0
- 117. Wuhrer M, Selman MH, McDonnell J, Kümpfel LA, Derfuss T, Khademi T, et al. Pro-Inflammatory Pattern of IgG1 Fc Glycosylation in Multiple Sclerosis Cerebrospinal Fluid. J Neuroinflamm (2015) 12:1–14. doi: 10.1186/s12974-015-0450-1
- Bondt A, Immunoglobulin G. (IgG) Fab Glycosylation Analysis Using a New Mass Spectrometric High-Throughput Profiling Method Reveals Pregnancy-Associated Changes. *Mol Cell Proteomics* (2014) 13:3029–39. doi: 10.1074/ mcp.M114.039537
- Gudelj I, Lauc G, Pezer M. Immunoglobulin G Glycosylation in Aging and Diseases. Cell Immunol (2018) 333:65–79. doi: 10.1016/j.cellimm.2018.07.009
- 120. Arnold JN, Wormald MR, Sim RB, Rudd PM, Dwek RA. The Impact of Glycosylation on the Biological Function and Structure of Human Immunoglobulins. Annu Rev Immunol (2007) 25:21–50. doi: 10.1146/ annurev.immunol.25.022106.141702
- 121. Maverakis E, Kim K, Shimoda M, Gershwin ME, Patel F, Wilken R, et al. Glycans in the Immune System and The Altered Glycan Theory of Autoimmunity: A Critical Review. J Autoimmun (2015) 57:1–13. doi: 10.1016/j.jaut.2014.12.002
- 122. Parekh RB, Dwek RA, Sutton BJ, Fernandes DL, Leung A, Stanworth D, et al. Association of Rheumatoid Arthritis and Primary Osteoarthritis With Changes in the Glycosylation Pattern of Total Serum IgG. *Nature* (1985) 316:452–7. doi: 10.1038/316452a0
- 123. Dekkers G, Rispens T, Vidarsson G. Novel Concepts of Altered Immunoglobulin G Galactosylation in Autoimmune Diseases. Front Immunol (2018) 9:553. doi: 10.3389/fimmu.2018.00553
- 124. Krapp S, Mimura Y, Jefferis R, Huber R, Sondermann P. Structural Analysis of Human IgG-Fc Glycoforms Reveals a Correlation Between Glycosylation and Structural Integrity. J Mol Biol (2003) 325:979–89. doi: 10.1016/S0022-2836(02)01250-0
- 125. Yamaguchi Y, Nishimura M, Nagano M, Yagi H, Sasakawa H, Uchida K, et al. Glycoform-Dependent Conformational Alteration of the Fc Region of Human Immunoglobulin G1 as Revealed by NMR Spectroscopy. *Biochim Biophys Acta -Gen Subj* (2006) 1760:693–700. doi: 10.1016/j.bbagen.2005.10.002
- 126. Dekkers G, Treffers L, Plomp R, Bentlage AEH, de Boer M, Koeleman CAM, et al. Decoding the Human Immunoglobulin G-Glycan Repertoire Reveals a Spectrum of Fc-Receptor- and Complement-Mediated-Effector Activities. *Front Immunol* (2017) 8:877. doi: 10.3389/fimmu.2017.00877
- 127. Wada R, Matsui M, Kawasaki N. Influence of N-Glycosylation on Effector Functions and Thermal Stability of Glycoengineered IgG1 Monoclonal Antibody With Homogeneous Glycoforms. *MAbs* (2019) 11:350–72. doi: 10.1080/19420862.2018.1551044
- Peschke B, Keller CW, Weber P, Quast I, Lünemann JD. Fc-Galactosylation of Human Immunoglobulin Gamma Isotypes Improves C1q Binding and Enhances Complement-Dependent Cytotoxicity. *Front Immunol* (2017) 8:646. doi: 10.3389/fimmu.2017.00646
- 129. Zhou X, Motta F, Selmi C, Ridgway WM, Gershwin ME, Zhang W, et al. Antibody Glycosylation in Autoimmune Diseases. Autoimmun Rev (2021) 20:102804. doi: 10.1016/j.autrev.2021.102804
- 130. Pavić T, Dilber D, Kifer D, Selak N, Keser T, Ljubičić CDS, et al. N-Glycosylation Patterns of Plasma Proteins and Immunoglobulin G in Chronic Obstructive Pulmonary Disease 11 Medical and Health Sciences 1102 Cardiorespiratory Medicine and Haematology. J Transl Med (2018) 16:1–15. doi: 10.1186/s12967-018-1695-0
- 131. Noro D, Yoneyama T, Hatakeyama S, Tobisawa Y, Mori K, Hashimoto Y, et al. Serum Aberrant N-Glycan Profile as a Marker Associated With Early Antibody-Mediated Rejection in Patients Receiving a Living Donor Kidney Transplant. *Int J Mol Sci* (2017) 18:1731. doi: 10.3390/ijms18081731
- 132. Verhelst X, Geerts A, Jochmans I, Vanderschaeghe D, Paradissis A, Vanlander A, et al. Glycome Patterns of Perfusate in Livers Before Transplantation Associate With Primary Nonfunction. *Gastroenterology* (2018) 154:1361–8. doi: 10.1053/j.gastro.2017.12.027

- 133. Verhelst X, Geerts A, Callewaert N, Van Vlierberghe H. The Potential of Glycomics as Prognostic Biomarkers in Liver Disease and Liver Transplantation. Acta Gastroenterol Belg (2019) 82:309–13.
- 134. Verhelst X, Dias AM, Colombel JF, Vermeire S, Van Vlierberghe H, Callewaert N, et al. Protein Glycosylation as a Diagnostic and Prognostic Marker of Chronic Inflammatory Gastrointestinal and Liver Diseases. *Gastroenterology* (2020) 158:95–110. doi: 10.1053/j.gastro.2019.08.060
- 135. de Haan N, van Tol MJD, Driessen GJ, Wuhrer M, Lankester AC. Immunoglobulin G Fragment Crystallizable Glycosylation After Hematopoietic Stem Cell Transplantation is Dissimilar to Donor Profiles. Front Immunol (2018) 9:1–10. doi: 10.3389/fimmu.2018.01238
- 136. Rao U, Sharma M, Mohanakumar T, Ahn C, Gao A, Kaza V, et al. Prevalence of Antibodies to Lung Self-Antigens (Kα1 Tubulin and Collagen V) and Donor Specific Antibodies to HLA in Lung Transplant Recipients and Implications for Lung Transplant Outcomes: Single Center Experience. *Transpl Immunol* (2019) 54:65–72. doi: 10.1016/j.trim. 2019.02.006

**Conflict of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

**Publisher's Note:** All claims expressed in this article are solely those of the authors and do not necessarily represent those of their affiliated organizations, or those of the publisher, the editors and the reviewers. Any product that may be evaluated in this article, or claim that may be made by its manufacturer, is not guaranteed or endorsed by the publisher.

Copyright © 2021 McQuiston, Emtiazjoo, Angel, Machuca, Christie and Atkinson. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.