# **Review** Article

# Perturbed Biochemical Pathways and Associated Oxidative Stress Lead to Vascular Dysfunctions in Diabetic Retinopathy

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Diabetic retinopathy (DR) is a vascular insult that accompanies the hyperglycemic state. Retinal vasculature holds a pivotal role in maintaining the integrity of the retina, and any alteration to retinal vasculature affects retinal functions. The blood retinal barrier, a prerequisite to vision acuity, is most susceptible to damage during the progression of DR. This is a consequence of impaired biochemical pathways such as the polyol, advanced end glycation products (AGE), hexosamine, protein kinase C (PKC), and tissue renin-angiotensin system (RAS) pathways. Moreover, the role of histone modification and altered miRNA expression is also emerging as a major contributor. Epigenetic changes create a link between altered protein function and redox status of retinal cells, creating a state of metabolic memory. Although various biochemical pathways underlie the etiology of DR, the major insult to the retina is due to oxidative stress, a unifying factor of altered biochemical pathways. This review primarily focuses on the critical biochemical pathways altered in DR leading to vascular dysfunctions and discusses antioxidants as plausible treatment strategies.

# 1. Introduction

The retina, a transparent tissue of the eye, has an intricate arrangement of neurons and requires highly dedicated circulation to meet its metabolic requirements and functioning of neurotransmission, phototransduction, and complex interaction of metabolites, growth factors, and vasoactive agents. Retinal circulation is a regular geometrically arranged network of vessels with a complex threedimensional architecture. It mainly is supplied by two vasculatures: the choroid and retinal vessels, where the endothelial cells lining the vessels integrate the normal physiology of the retina [1]. The central retinal artery enters via the optic nerve ensuring blood flow and exchange of gases and nutrients, while the central retinal vein is involved in the removal of waste products that move away from the retina. An important normal physiological function of retinal vasculature is maintenance of the inner blood-retinal barrier (iBRB), which prevents nonspecific permeation of retinal neuropile by macromolecules yet facilitates exchange of respiratory gases, amino acids, salts, sugars, and some peptides [2].

The most sensitive part of the retina is the outer region which constitutes one-third of the retina and is devoid of blood vessels. The absence of blood vessels serves as a special adaptation for visual functioning, but poses a great challenge in the maintenance of continuous energy requirements [1]. The outer blood-retinal barrier that is formed between the tight junctions of retinal pigment cells maintains ionic concentrations in the avascular region of the retina and the interstitial space for neurotransmission. Alternatively, the metabolic need of photoreceptor cells is maintained by choroid vessels. Thus, these efficient blood retinal barriers serve as major anatomical adaptation, with attainment of demanding metabolic requirements of the retina and without compromising its conductive extracellular microenvironment [1, 2]. This intricate retinal vasculature is sensitive to various systemic disorders with diabetes being the most common and perhaps well-studied metabolic insult that has a profound influence on retinal vessels. Retinal vascular dysfunction commences soon after the onset of diabetes and is characterized by impaired microvasculature and transport through the blood retinal barrier which may be an important factor in the initiation and progression of the vascular lesions

# 2. Diabetic Retinopathy

tion are key features of diabetic retinopathy [5].

Diabetic retinopathy is one of the leading causes of visual impairment and morbidity across the globe [6]. Type 1 and type 2 diabetes damages the blood vessels in the retina, which may lead to microvasculature complications; however, the incidence of DR is higher in type 1 patients than in those with type 2 diabetes [7]. Amongst 468 million people estimated with diabetes mellitus worldwide [8], approximately 90 million suffer with some form of diabetic retinopathy, [7].

DR is classified into nonproliferative DR (NPDR) and proliferative DR (PDR) stages on the basis of the presence of visible ophthalmologic changes and manifestation of retinal neovascularization [9]. In the NPDR stage, sex, onset, and duration of type 1 diabetes and HbA1c levels are suggested to be the key pointers implicated in NPDR development [10]. Diabetic maculopathy accompanies the NPDR stage and has been considered as the main reason for the loss of vision. The NPDR stage is primarily consequent to hyperglycemic conditions which weaken the capillary walls resulting in microaneurysms. This is followed by the rupture of vessels which leads to accumulation of fatty deposits and lipid by-products [11]. Ensuing this, an obstruction in the nerve fibre layer is observed that results in white fluffy spots known as cotton wool spots. The NPDR stage ranges from mid, moderate, and severe, where the microaneurysms are followed by venous beading and cotton wool spots along with severe microvascular complications [12].

NPDR is then followed by a proliferative state of the retinal tissue. The PDR stage is a consequence of ischemic conditions that arise due to obstruction in the NPDR stage. The higher metabolic requirement of retinal tissue poses the need for neovascularization which is due to the release of angiogenic signals. Retinal detachment and this neovascularization with the proliferation of the fibrovascular tissue is a characteristic of the PDR stage [13]. These newly formed vessels are leaky, fragile, and misdirected, and with age the shrinkage of the vitreous humour causes them to tear and result in sudden vision loss. If a greater force is created, it may lead to tractional retinal detachment. Despite severe complications in the PDR stage, macular oedema is the main reason behind the loss of visual acuity. Regardless of these varied stages characterising DR, the major progressive change leading to retinopathy in diabetic patients remains speculative in terms of microvascular and biochemical complications leading to oxidative/nitrative stress.

#### 3. Microvascular Complication

Microcirculation involves the transfer of nutrients and removal of waste products and regulates the fluctuating hydrostatic pressure of the eye. Normally, fundamental metabolic and myogenic autoregulatory mechanisms guarantee satisfactory advancement of these microcirculatory functions [14]. Microvascular endothelial cells are believed to be targets of hyperglycemic damage since they are not capable of lowering the glucose transport rate when glucose concentrations are high, stimulating intracellular hyperglycemia. This has been thought to be the key event in microvascular endothelial damage, diminished accessibility of nitric oxide, increased permeability, increased leukocyte attachment, and procoagulant action [14].

In one of the recent studies on high sucrose- (HSu-) treated rats, a decrease in the thickness of the inner retinal layers was observed. Nevertheless, neither apoptotic cells nor retinal neural markers were identified in the retinas of HSu-treated animals. Also, no progression was recognized in the permeability of the blood-retinal barrier as well as tight junction proteins. Likewise, these parameters stayed unaltered in the retina regardless of the increase in the number of retinal microglial cells. Thus, a prediabetic rodent demonstrates that the retinal structure is influenced by the diminishing internal layers, without vascular and inflammatory changes [15]. Therefore, inconspicuous auxiliary changes may be seen as an early unsettling influence in the development of DR which could be reversed by preventive strategies at this stage, before it results in irreversible damage to the retina. Progression from diabetes to diabetic retinopathy underlies changes in the haemodynamics or vascular geometry [16]. Haemodynamic factors like perfusion pressure, vascular resistance, blood viscosity, and vascular geometry influence the circulation of blood flow to the retina. Regardless of this fact, the components involved in haemodynamic modifications have not been fully elucidated [14, 16].

In an earlier study, an analysis was performed to assess the relationship between the assessed haemodynamic highlights and the progression of DR. Vessel bifurcations on fundus images and factors like nodal pressure, volumetric blood flow, wall shear stress, blood flow velocity, and Reynolds number were investigated over a period of three years [17]. The analysis revealed critical changes in haemodynamic parameters related with perceptible changes to vessel geometry, especially in the venular network, and these changes were articulated to be three years prior to DR onset [17]. Therefore, these findings altogether suggest a role of the microvasculature and its geometry in the progression of DR, but further studies in this area are yet to be undertaken to fully elucidate the role of altered vascular biology in the progression of DR.

However, vascular modifications are considered to be the major insult for the onset and progression of DR, including altered blood flow, dyslipidemia, basement membrane thickening, loss of pericytes, and platelet aggregation along with neuroglial damage [18]. These changes have been suggested to be the result of alteration/disruption of biochemical mechanisms required for the normal functioning of a cell. A unifying mechanism of hyperglycemia and induced oxidative stress in retinal cells has been regarded as one of the crucial players in causing alteration(s) in various biochemical pathways that have been shown to be interconnected with vascular insult [19, 20]. Furthermore, an increase in the stressful conditions eventually leading to apoptosis of the cells associated with retinal vasculature is the major effect of these pathways. Cardinal biochemical pathways proposed to be involved in the DR includes increased flux of the polyol pathway [21, 22], advanced glycation end product/receptors of advanced glycation end product (AGE/RAGE) pathway [10, 23], hexosamine pathway [24], PKC activation [25], tissue (renin-angiotensin system) RAS [26], and histone modifications which are emerging as key events in the development of DR. The overall effects of these metabolic abnormalities are hypothesized to result in augmentation of (reactive oxygen species) ROS and (reactive nitrative species) RNS production and associated oxidative and nitrosative damage [20, 27] which are key mediators in inducing vascular dysfunctions and related insult to retinal circulation (Figure 1).

# 4. Increased Flux in the Polyol Pathway

Although the polyol pathway is a minor glucose metabolism pathway, it is considered to play a pivotal role in retinopathy [21, 28]. The first and rate-limiting step of this pathway is the conversion of excess glucose to sorbitol, using NADPH as a cofactor, a reaction catalyzed by an enzyme aldose reductase. The sorbitol formed is then converted to fructose by a slow reaction involving sorbitol dehydrogenase [22]. Also, the existence of polymorphism in the aldose reductase gene (C106T) has been shown to be associated with the increased susceptibility to retinopathy in individuals with type 1 (diabetes mellitus) DM [29]. Numerous studies have been carried out to establish the role of increased polyol production in DR. In one of the studies, both rat and human retinal endothelial cells showed increased aldose reductase immunoreactivity. In addition, rat and human retinas exposed to high glucose in organ culture increased the production of sorbitol corroborating excess aldose reductase activity to be one of the mechanisms in the development of DR [30]. Dagher et al. [30] have suggested that the polyol pathway mediates the increase in apoptosis of neurons and attenuation of GFAP- (glial fibrillary acidic protein-) immunostained astrocytes along with the increase in the levels of sorbitol and fructose in the retina. Indeed, this is due to the nonpermeability of sorbitol which results in osmotic damage [21, 31]. Moreover, fructose produced by the polyol pathway gets phosphorylated to fructose-3-phosphate [32], which in turn is broken down to 3-deoxyglucosone; both these molecules are strong glycosylating agents that result in the formation of AGEs. Additionally, the increased flux of the polyol pathway results in depletion of cellular NADPH, affecting the production of reduced glutathione and nitric oxide thereby resulting in antioxidant imbalance [33]. Moreover, it has further been suggested that the polyol pathway is the only mechanism of glucose toxicity responsible for the spectrum of neural and vascular abnormalities [34]. Aldose reductase has been extensively studied as a molecular target for DR, and inhibitors targeting aldose reductase are expected to be beneficial against DR. Inhibitors such as sorbinil and beta-glucogallin (BGG) have been shown to deplete sorbitol accumulation and reduce oxidative stress [33]. Various in silico studies have identified 2-benzoxazolinone derivatives effective against ALR2 (aldose reductase 2) and can reduce AGE and oxidative stress [35]. Additionally, studies on human

aldolase reductase have revealed huperzine A, rosmarinic acid, and luteolin 78 to possess aldose reductase inhibition potential [36] and hence could be potential molecules in targeted therapeutics.

# 5. AGE/RAGE Pathway

Elevated glucose and coupled perturbations in the pathways regulating glucose levels result in the formation of advanced glycation end products (AGEs) which are made from nonenzymatic glycoxidation and glycation of various biomolecules and sugar metabolites [37]. AGEs bind to their receptors termed as RAGE (receptor for advanced glycation end product) and trigger the cascade of inflammatory signals [38]. The AGE-modified plasma proteins have been also found to bind to AGE receptors on cells like macrophages, vascular endothelial cells, and vascular smooth cells affecting their functionality [37, 38]. AGEs accumulate in the circulation due to their inefficient renal clearance. Moreover, exogenous AGEs or dietary AGEs have also shown to be the reason for their accumulation in diabetic patients [39] and are considered to be the pivotal participants in inducing the ROS formation by altering the proteins, enzymes, and the genetic material of the mitochondria by glycation [40]. Their accumulation increases the vessel thickening and platelet aggregation leading to ischemic situation, a condition also responsible for the induction of growth factors and neovascularization [41]. Both intracellular and extracellular formations of AGEs in the retina are involved in destructive roles, as the alteration in protein chemistry distorts their structure [39]. Additionally, oxidative stress has also shown to accelerate the formation of the AGEs. Although the human body is self-sufficient in degrading the AGEs by ubiquitination and autophagy, the excess formation or intake results in their accumulation [37]. Additionally, AGEs are also accountable for permanent dysfunction of the mitochondrial enzymes due to glycation of the mitochondrial genetic material leading to "metabolic memory," a severe condition that is observed in DR [42]. It is an unresponsive state where even glucose control cannot prevent complications of DR. Besides these, the components of the extracellular matrix are subject to modification by AGE precursors.

AGEs have been corroborated to upregulate the expression of RAGE in retinal pericytes via ROS production [43] which is the earliest known alteration in the diabetic retinal vasculature [44]. AGE-RAGE interactions are responsible for NADPH-mediated ROS generation via activation of mitogen-activated protein kinase [38]. These interactions are also responsible for the translocation of NF- $\kappa$ B, decrease in the ratio of Bcl-2/Bax, and increased expression of vascular endothelial growth factor (VEGF), inflammatory cytokines, and adhesion molecules [45], which correlate with the development of DR. During diabetes, AGEs have been shown to accumulate in retinal pericytes, decreasing their survival, breakdown of the blood retinal barrier, and progression towards diabetic retinopathy [9]. Even the major AGE precursor methylglyoxal has been shown to mediate oxidative stress and impair nitric oxide- (NO-) mediated vasorelaxation and upregulate inflammatory markers in an animal



FIGURE 1: Effect of glucotoxicity on plausible biochemical pathways involved in pathogenesis of DR.

model of type 2 diabetes where the rats were fed with methylglyoxal [46]. Also, methylglyoxal in a similar study was shown to reduce retinal pigment epithelial cell viability via ER stress-dependent ROS production, mitochondrial membrane potential loss, and intracellular calcium increase [47]. In a recent study, retinal pigment epithelial cells were treated with chrysin, a naturally occurring flavonoid, to exploit its retinoprotective effects against diabetes-associated visual cycle impairment by targeting the AGE-RAGE pathway. It was demonstrated that chrysin treatment restored the retinoid visual cycle through blocking ER stress via AGE-RAGE activation in glucose-stimulated retinal pigment epithelial cells and diabetic eyes [48], highlighting the importance of the AGE-RAGE pathway in impaired visual cycle associated with diabetic retinopathy.

#### 6. Hexosamine Pathway

Another crucial pathway involved in the pathogenesis of DR is the hexosamine pathway which itself is relatively a minor branch of glycolysis where fructose-6-phosphate is converted to glucosamine-6-phosphate, a reaction catalyzed by the first and rate-limiting enzyme, glutamine:fructose-6-phosphate amidotransferase (GFAT) [49]. Chronic hyperglycemia leads to an enhanced influx through the hexosamine pathway which results in perturbation of retinal cells [50, 51]. In this pathway, glucose is metabolized into UDP-N-acetylglucosamine (UDPGlcNAc) [19]. Specific O-GlcNAc transferases (OGT) then modify various cytoplasmic and nuclear proteins by addition of the amino sugar N-acetylglucosamine (O-GlcNAc) from

UDP-GlcNAc similar to phosphorylation modification at the ser/thr residues. Pancreatic  $\beta$ -cells express large amounts of both O-linked  $\beta$ -N-acetylglucosamine transferase (OGT) and O-GlcNAcase (OGA), suggesting the importance of O-GlcNAc in pancreatic  $\beta$ -cell functioning and survival under normal glucose conditions [52]. In addition, hyperglycemia-mediated enhanced O-GlcNAc modifications contribute towards increased  $\beta$ -cell death [53]. This imbalanced O-GlcNAc modification has been implicated in the etiology of microvascular complications related to DR as it regulates the fate of retinal vascular cells [54, 55]. Moreover, it has been observed that in retinal neuronal cells, this modification alters the neuroprotective effect of the insulin/Akt pathway [56].

Nucleoside diphosphate kinases (NDPK) are the enzymes which provide nucleoside triphosphates to the cells and hence play a pivotal role in mediating fundamental cellular processes [57-59]. NDPKs are histidine protein kinases, which transfer the phosphoryl group from the phosphohistidine active site to the histidine residue of the target protein. These histidine kinases maintain the metabolic status of the cell by regulating the levels of NTP in the cell [57]. The B isoform of this enzyme, NDPK-B, has been shown to phosphorylate the  $\beta$  subunit of the G protein, potassium channels  $(K_{Ca}3.1)$ , and calcium channels (TRPV5), thus modulating their functions [60]. Moreover, NDPK-B has been documented in modulating vascular integrity [61]. NDPK deficiency has been shown to mimic vascular regression similar to DR. O-GlcNAcylation of FoxO1, a transcription factor, upregulates Ang 2 (angiopoietin 2) which is an initiator of vascular regression suggesting that the hexosamine pathway is responsible for O-GlcNAcylation of various proteins to

be the prime culprit underlying the changes in the molecular signals associated with microvasculature [62]. Corroborating the similar fact, another study has stated the role of NDPK-B deficiency in causing a diabetes-like vascular pathology by upregulating endothelial angiopoietin 2 in the retina of mice [49]. Hyperglycemia increases O-GlcNAcylation of retinal proteins in DR. O-GlcNAcylation of the p65 subunit of NF- $\kappa$ B in streptozotocin-induced DR mice has been shown to be responsible for hyperglycemia-induced activation of NF- $\kappa$ B and retinal ganglion cell death [51], further linking the involvement of imbalanced O-GlcNAc modification in the etiology of the microvascular complications in DR.

#### 7. PKC Pathway

Diacylglycerol and PKC are also amongst the key players altered by hyperglycemia in DR [63, 64]. Mainly, three isoforms of PKC are reported in biological systems; viz., the conventional PKC isoforms (PKC- $\alpha$ ,  $\beta$ 1,  $\beta$ 2, and  $\gamma$ ) are activated by phosphatidylserine, calcium, and DAG or phorbol esters. The novel PKCs (PKC- $\delta$ , - $\theta$ , - $\eta$ , and - $\varepsilon$ ) are activated by phosphatidylserine, DAG, or PMA (phorbol 12myristate 13-acetate), and the atypical PKCs (PKC- $\zeta$  and  $-\iota/\lambda$ ) are not activated by either calcium, DAG, or PMA [65]. In the retina, hyperglycemia persistently elevates diacylglycerol (DAG) and activates downstream protein kinase C, evidently interconnecting PKC with associated microvascular changes [21, 66, 67]. The beta and delta isoforms of PKC are mainly activated, but increases in other isoforms have also been found in the retina [68]. Hyperglycemia activates PKC isoforms indirectly through the AGE-RAGE pathway [69] and polyol pathway [70] by increasing ROS. The DAG-PKC signaling pathway plays roles in vascular cells by regulation of permeability, contractility, extracellular matrix (ECM), cell growth, angiogenesis, cytokine actions, and leukocyte adhesions, processes seen to be altered in diabetes [71, 72]. Numerous studies have demonstrated the contribution of PKC activation in decreasing retinal blood flow. Studies focussed on PKC agonists and antagonists have revealed decreased or increased retinal blood flow, respectively. Introduction of phorbol esters, an agonist of PKC into the retina, declines retinal blood flow while this decrease in blood flow has been shown to be resolved by PKC inhibitors [67].

Amid many targets, a plausible mechanism employed by PKC to cause vasoconstriction and decreased retinal blood flow is the increase in the expression of a potent vasoconstrictor, endothelin A (ET-A) [73]. Its expression has been shown to increase in the retina of diabetic rats while intravitreous injection of the endothelin-A (ET-A) receptor antagonist prevented the decrease in retinal blood flow [73]. This declined retinal blood flow causes hypoxic conditions, which in turn is a robust inducer of VEGF, causing increases in permeability and microaneurysms [74, 75].  $\delta$  isoform of PKC and p38 $\alpha$  mitogen-activated protein kinase (MAPK) activation increases the expression of Src homology 2 domain-containing phosphatase-1 (SHP-1), a protein tyrosine phosphatase, which dephosphorylates the PDGF $\beta$  receptor and induces pericyte apoptosis [38, 76]. Previously, it was shown

that antiamyotrophic lateral sclerosis (ALS) drug riluzole attenuates pathological changes in oxygen-induced retinopathy, a surrogate model of DR [77]. More recently, a similar study on cultured retinal pericytes as well as in diabetic rats targeting PKC $\beta$  showed that anti-ALS drug riluzole attenuates monocyte chemotactic protein (MCP1), a cytokine elevated in vitreous humour and serum during DR [78] most probably by preventing the abnormal activation of PKC.

## 8. Renin-Angiotensin System

One of the hallmarks and early events of DR is the breakdown of the blood-retinal barrier (BRB), and one of the plausible reasons of this breakdown is renin-angiotensin system-(RAS-) mediated altered vascular permeability [79]. Tissue RAS, a paracrine system, is a property of numerous organs such as eye, brain, vessels, adrenal gland, testis, and kidney, which locally produces angiotensin (Ang) [80]. This system involves prorenin which binds to its receptor, termed as prorenin receptor (P)RR, which has been implicated in the pathogenesis of DR [81] and induces the production of VEGF through ERK1/2, and this signal cascade is known as the receptor-associated prorenin system (RAPS) [82, 83] which is considered to be responsible for the dysfunctional blood retinal barrier. A study on PDR patients has shown the levels of (P)RR to be high in their vitreous fluid samples than in nondiabetic control eyes, strengthening the implication of (P)RR in DR [84, 85]. Also, (P)RR and other RAS system components have also been detected in human PDR fibrovascular tissues, normal ocular tissues, and various human retinal cell lines, including retinal pigment epithelial cells [84, 86], while vitreous prorenin and Ang 2 levels have been reported to increase in PDR eyes [84, 87].

Ang 2, a vasoactive and angiogenic agent, along with VEGF which is a proangiogenic stimulus has been observed to be elevated in the vitreous fluid of PDR patients [88]. Additionally, an increase in VEGF and VEGFR-2 gene expression and an elevation of ocular active renin are indicative of the interaction of tissue RAS and VEGF wherein endothelial cell proliferation is observed as an outcome of the activated tissue RAS system [89], thus correlating the tissue RAS system and VEGF in the progression of DR. In addition, ATP6AP2 or prorenin receptor (P)RR is shown to interact and colocalize with the PDHB subunit of the PDH (pyruvate dehydrogenase) complex. It was observed that PDH activity is downregulated due to ATP6AP2 knockdown, and it leads to suppression of glucose-induced ROS generation in retinal pigment epithelial cells. Thus, ATP6AP2 is considered pathogenic due to its role in RAPS activation and mitochondrial ROS generation [86]. Therefore, blockade of (P)RR and other players of the RAS system might inhibit a series of events vital for vascular abnormalities represented in DR.

## 9. Metabolic Memory and Epigenetic Modifications

Numerous studies have suggested epigenetic modifications to be a significant contributor in DR development [90–92]. The duration of hyperglycemia decides whether improved glycaemic control would be effective in DR [85], implicating that hyperglycemia exposure results in a phenomenon of metabolic memory and could be attributed to epigenetics [91]. Earlier, an anatomic observation of gradual reduction of capillary cells in DR suggested that "retinopathy neither appears promptly after the onset of hyperglycaemia nor arrests promptly on correction of the hyperglycaemia" [90].

The most primitive epigenetic modification is DNA methylation which has a correlation with DR progression as indicated by various studies. DNA methylation is a phenomenon where the methyl group is transferred from S-adenosylmethionine (SAM) to DNA molecules, a reaction catalyzed by DNA methyltransferases. DR patients have shown a significantly higher level of DNA methylation as compared to those without DR [92], indicating that higher DNA methylation is a key component in DR development. Moreover, the study also showed that the levels of DNA methylation remain constant in these DR patients, suggesting that this epigenetic modification occurred only during the early stage of the disease. Another study on DR patients found altered methylated CpG sites, further highlighting the role of epigenetics in DR [93]. Studies using animal models have also strengthened this data where modified methylation patterns have been seen under hyperglycemic conditions [94]. Another study revealed methylation and activation of the matrix metalloproteinase 9 (MMP-9) gene which is known to be associated with DR [95] that plays a role in accelerating the apoptosis of retinal vascular endothelia. In addition, transcription of MMP-9 is regulated by nuclear factor kappa B (NF-kB) whose activation is modulated by the acetylation of its p65 subunit. Histone deacetylase plays an important role in the acetylation-deacetylation of p65. In diabetic mice, histone deacetylase activity was found to be decreased and p65 acetylation was elevated leading to an increase in MMP-9 expression [96].

Another epigenetic alteration, histone modification, has also been a key contributor in DR pathophysiology [97]. The transcription activity of HDAC1/2/8 (histone deacetylase) was elevated in retinal endovascular cells, while the activity of HAT (histone acetyltransferase) and the expression of acetylated histone H3 were both decreased in streptozotocin- (STZ-) induced diabetic models. Moreover, these changes were found to be irreversible after the blood glucose of the rats was restored to normal level, indicating that DR development is to be associated with histone modifications and might be participants in the formation of the "metabolic memory" phenomenon.

Several studies implicated a major role of mitochondrial alteration due to epigenetic modifications as the key process in the induction of metabolic memory in DR. During DR, the mitochondrial homeostasis and dynamics are altered, creating a vicious cycle where the alteration of mitochondrial enzymes induces superoxide formation which in turn alters the organelle physiology. The sensitivity of the mitochondria is exclaimed due to the close proximity of the mitochondrial DNA (mtDNA) to the electron transport chain (ETC) and lack of histones. An increase in 8-OHdG in the diabetic retina confirms the mitochondrial susceptibility [94]. Additionally, the dysfunction of the repair pathways further complicates the mitochondrial damage [98]. The mtDNA replication also plays an important role in mtDNA damage experienced by the retina in diabetes, and these are under the control of superoxide, which is well known to be altered under hyperglycemic conditions. Thus, the regulation of mtDNA replication/repair machinery has the potential to prevent mitochondrial dysfunction and the development of diabetic retinopathy [99].

Histone modifications of the molecules regulating the redox status of the cell has been extensively studied, wherein the mitochondrial superoxide dismutase SOD2 depletion and inhibition of Nrf2 (nuclear factor- (erythroid-derived 2-) like 2), a transcription factor affecting antioxidants, has been observed. During the state of oxidative stress, Nrf2 translocates to the nucleus where it binds to the antioxidant response element (ARE). Keap1, an inhibitor of Nrf2, tethers it in the cytosol and leads to proteasomal degradation through cullin-3-dependent degradation [100]. Mishra et al. [101] have observed that hyperglycemia increased the binding of Sp1, a transcription factor at the Keap1 promoter, and enriched H3K4me1 and activated SetD7 (methyl transferase). This leads to inhibition of Nrf2 binding on antioxidant response element (ARE) leading to oxidative stress in the cell. In earlier studies, deletion in MnSOD (manganese superoxide dismutase) and Sod2 activity in vivo has been shown to increase oxidative damage in mitochondria and alters the mitochondrial function [102]. In another study, streptozotocin- (STZ-) induced diabetic rats showed an increase in H4K20me3, acetyl H3K9, and NF-κB p65 at the promoter and enhancer of retinal Sod2. Even the reversal of hyperglycemia failed to prevent increases in H4K20me3, acetyl H3K9, and NF-kB p65 at Sod2. Thus, increased H4K20me3 at Sod2 contributes to its downregulation and is responsible for the development of DR and metabolic memory phenomenon [103].

Dysregulated mitochondrial biogenesis also contributes to the phenomenon of metabolic memory. The nuclearmitochondrial transcriptional factors and translocation of transcription factor A (TFAM) to the mitochondria are essential for transcription and replication of mitochondria and thus tightly control the biogenesis of the organelle [104]. An earlier study which investigated the effects of diabetes on nuclear-mitochondrial communication in the retina has uncovered that retinal mitochondrial biogenesis is under the control of superoxide radicals and is debilitated in diabetes, perhaps by diminished transport of TFAM to the mitochondria. Hence, regulation of biogenesis by pharmaceutical or molecular means might provide potential means to impede the development/progression of diabetic retinopathy [105]. Additionally, a good glucose control along with lipoic acid supplementation has shown to retard the progression of DR indicative of a major role of mitochondrial function in the progression of disease [105, 106].

#### 10. Role of miRNA

MicroRNAs (miRNAs) are a class of 19 to 25 nucleotide bases, noncoding RNAs that regulate gene expression at the posttranscriptional level by annealing to their partially

complementary sequences in the target mRNAs resulting in translational repression or degradation of mRNAs, thereby depleting the protein expression [107]. miRNAs have been implicated in the regulation of genes involved in DR development, thus playing a role in epigenetic alterations of DR [108]. A total of 11 miRNAs (miR-182, miR-96, miR-183, miR-211, miR-204, miR-124, miR-135b, miR-592, miR-190b, miR-363, and miR-29c) displayed increased expression in the retinas of DM rats, whereas the expression of 6 miR-NAs (miR-10b, miR-10a, miR-219-2-3p, miR-144, miR-338, and miR-199a-3p) was found to be decreased [109]. Moreover, it was observed in cultured human endothelium cells that miR-23b-3p regulates high-glucose-induced cellular metabolic memory through an SIRT1-dependent signaling pathway [110], while in a diabetic rat model, miR-126 was found to play a potential role in the pathogenesis of DR [111, 112]. Additionally, miRNA has also been implicated in regulating retinal neovascularization [112].

# 11. Interplay of Nitrosative Stress/Oxidative Stress and Inflammation

The biochemical mechanisms which are altered in DR probably via hyperglycemic conditions eventually culminate into cellular stress affecting retinal homeostasis. This stressful condition is induced either by the increased flux of these biochemical pathways along with alterations in the proteins involved in maintaining the metabolic energy homeostasis [96]. Also, the mitochondrial complexes are the prime victims due to an increase in reactive oxidative species [96] and nitrative species [113].

11.1. Nitrosative Stress. Nitrosative stress is prompted by a response of superoxide with nitric oxide (NO), which creates peroxynitrite and is ensnared in diabetic conditions. Increased nitrative stress could be prompted by protein nitration and damage membrane proteins and fatty acids, prompting changes in cell signaling transduction and upregulation of inflammatory reaction and initiating the apoptotic pathway. Hyperglycemic episodes are connected with increased nitrative stress, which can trigger the advancement in diabetic complications [113].

Mitochondria are additionally equipped for creating both RNS and ROS. NOS exist in three isoforms (endothelial (eNOS), neuronal (nNOS), and inducible (iNOS)) which catalyze the change of L-arginine into citrulline and NO. This response additionally requires flavin adenine dinucleotide, flavin mononucleotide, tetrahydrobiopterin (BH4), heme, and calmodulin. These cofactors are essential (e.g., when BH4 levels are restricted) in light of the fact that NOS may move toward becoming uncoupled and produce superoxide rather than NO. Another plausibility is that, in states of high oxidative stress, NO and superoxide collaborate to produce ONOO-, an exceptionally receptive species equipped for nitrating tyrosine residues, thereby enhancing oxidative damage [113]. The accumulation of RNS shifts the homeostasis of the cell, resulting in nitrosative stress, which is a major phenomenal consequence of the stress conditions in the diabetic retina and is now emerging as a new insight for DR research [113]. NO, a multifunctional molecule that can change proteins by means of nitrosylation, is majorly found in reactive form during DR progression. In addition, nicotinamide adenine dinucleotide phosphate diaphorase (NADPH-d), a specific marker for NO-producing neurons, was revealed to be positive in immunoreactive experiments on diabetic retina suggesting a role of NO in development of DR [114].

Augmentation in nitrosative stress has been revealed as the key player in worsening the pathogenesis of PDR [115]. Additionally, LPO (lipid peroxide), NO, and GSH (glutathione) levels were also shown to be significantly associated with the severity of diabetic retinopathy [116]. An increase of oxidative/nitrative pressure markers, for example, 4hydroxynonenal and nitrotyrosine, was more elevated in retinal vasculature of diabetic rodents when contrasted with normoglycemic mature and adult rodent retinas [117].

On similar lines, a study showed that the onset of diabetes results in an increase in the nitrated proteins in the retina [114]. Furthermore, nitrotyrosine immunolabelling in the photoreceptor layer, ganglion cell layer, inner nuclear layer, and some Muller cell processes in the retina of diabetic rats points towards the impinging role of nitrosative stress in DR [114]. Conjointly, recent preclinical and clinical studies have indicated a plethora of factors contributing towards the role of NO in the pathogenesis of DR and have pointed towards a further investigation in the plight of therapeutics [118]. Another finding suggests that aminoguanidine treatment hinders capillary cell death and development of diabetic retinopathy [119, 120]. Moreover, it has also been found to decrease the hyperglycemia-induced increase in NO and its sequelae raise a plausibility that (RNS) assumes a major role in the pathogenesis of retinopathy. In any case, it is not conceivable at present to infer that aminoguanidine represses retinopathy exclusively through hindrance of NO generation. Therefore, approaches that specifically repress NO-intervened procedures will be important to absolutely assess the role of NO in the development of DR [121].

Calcium dobesilate (calcium,2,5-dihydroxybenzenesulfonic corrosive (CaD)), which is considered an angioprotective agent, is recommended as an elective treatment for diabetic retinopathy and other vascular diseases, in spite of the debate with respect to its clinical viability. Some clinical preliminaries did not report any helpful impact of CaD treatment, particularly in patients at the later phases of diabetic retinopathy. Notwithstanding, a few different clinical trials demonstrated enhancement in visual acuity after oral treatment. Diabetes increases tyrosine nitration in the retina, fundamentally in the ganglion cell layer, and treatment with CaD attenuates this increase in tyrosine nitration initiated by diabetes [122].

11.2. Oxidative Stress. Besides nitrative stress, a shift in the normal physiology due to oxidative stress is also a chief suspect in DR pathophysiology [123]. The balance of oxidants and antioxidants is a mediator of the fundamental cellular process including maintenance of the vascular system [124], which if disrupted can lead to threatening situations depending upon the severity of stress imposed and availability of their clearance system [125]. Unfortunately, imbalance

Oxidants	Involvement in DR
Superoxide radicals	Accumulation in retinal cell mitochondria leads to mutations in the mtDNA and induces a phenomenon of "metabolic memory" [132, 133].
Hydroxy radicals	Generated by Fenton reaction and responsible for damage to retinal cells membrane and mtDNA as well as reduction of thinning of outer and inner nuclear layers of the retina [133–135].
Peroxyl radical/lipid peroxides	Lipid peroxidation chain reaction damages retinal cell membrane and creates a redox milieu, as evident by an increase in the vitreous of PDR patients [136, 137].
Hydrogen peroxide	Toxic radical species are increased in retinal cells, and treatment with obestatin prevents the $H_2O_2$ -induced (retinal ganglion cells) RGC damage [133, 138].
Singlet oxygen	Excitation of oxygen through sunlight and radiation and higher oxygen consumption by retinal cells adds to the impaired redox status of the cell in DR [131].
Peroxynitrite	The reactive nitrogen species shifts the redox status of the cell towards destruction causing apoptosis of retinal endothelial cells [139].

TABLE 1: Oxidants elevated in diabetic retinopathy.

TABLE 2: Antioxidants depleted in diabetic retinopathy.

Antioxidants	Mechanism of action	Effect on DR
Thioredoxin (Trx)	The negative regulator of thioredoxin, thioredoxin-interacting protein (TXNIP), and the dissociation of apoptosis signal regulating kinase-1 (ASK-1) from the oxidised thioredoxin are key players inducing apoptosis during DR [140, 141].	ASK-1 induces apoptosis of Neuro2a cells during DR. TXNIP is upregulated in Muller cells and leads to apoptosis of pericytes [140, 142].
Superoxide dismutase (SOD)	First line of defence against hyperglycemic induced superoxide anion radicals in the mitochondria, and the highest SOD activity is present in the retina to help scavenge the superoxide radicals generated via metabolism [143–145].	SOD downregulation in retinal endothelial cells induces apoptosis [146] and upon overexpression ameliorates and protects the mtDNA from oxidative damage in DR [144].
NADPH oxidase (Nox)	Nox4, an isoform of Nox enzyme, promotes retinal neovascularization through ROS-dependent regulation of the VEGF/VEGFR2 signalling pathway [147] and via various inflammatory signalling pathways [148].	Nox4 isoform gene ( <i>NOX4</i> ) is involved in DR [149] and is the predominant isoform expressed in the human retinal endothelial cells [150].
Vitamin E (α-tocopherol)	A nonenzymatic antioxidant which donates hydrogen atom to peroxy radicals and other radicals to maintain the redox status of the cell [151].	Its supplementation reduces oxidative stress in NPDR and PDR patients [152] interlinking an antioxidant role in the prevention of DR.
Vitamin C (ascorbic acid)	An antioxidant or a reducing agent that donates electrons to various radical species [153].	Prevents high-glucose and RAGE-induced apoptosis in pericytes and endothelial cells. Also preserves NO generated by endothelial cells and tightens the leaky endothelial permeability barrier [154].

in ROS formation and the scavenging system plays a role in the pathogenesis of diseases including DR as increased oxidative stress has been seen in the retinas of animal models of diabetes [126] as well as in DR patients. In addition, ROS buildup in the eye is considered to be a trigger for degeneration of retinal cells, neural cells, and vascular lesions in the progression of DR. Accumulation of ROS in the eye gradually activates the NF- $\kappa$ B and MAPK cascades that result in inflammation in the retinal tissue. Hence, the interplay of ROS and inflammatory cytokines has been a major area of target and a platform to prevent prognosis of the condition [9]. Along with inflammation, neurodegeneration is another potential target of oxidative stress. Degeneration of retinal ganglion cells (RGC) has been reported to be in close association with oxidative stress and inflammation, wherein the activation of microglial cells results in the neurotoxicity and apoptosis of the RGC. It has been observed that highglucose free fatty acid cotreatment results in the upregulation of CD11b and ionized calcium-binding adapter 1 (Iba-1), markers of microglial cells, corroborating oxidative species as key players in neurodegeneration, mediated via inflammatory response [127]. Moreover, crocin, a bioactive component of saffron, has been proved to be a good therapeutic agent, due to observed reduction in ROS and NO levels and IL-1 $\beta$  and TNF- $\alpha$  and additionally with its neuroprotective effect by activating the PI-3/Akt pathway [127]. Another insight into the cause of vascular lesions is neural photoreceptors inducing oxidative stress and inflammatory changes. These photoreceptors are a major source of superoxide radicals (correct this other place also) in the diabetic retina which

Antioxidant	Effect on DR
Resveratrol (RSV)	Reduces ROS levels and cleavage of caspase 3 in BREC (bovine retinal endothelial cells). Additionally, RSV shows antiapoptotic effects <i>in vitro</i> and <i>in vivo</i> on Muller cells with addition of miR-29b [155, 156].
Citrus flavones (hesperidin)	Modulation of mitochondrial function and inhibition of caspase activation via a ROS-dependent p38 and JNK signalling pathway. Also protects retinal pigment cells from hyperglycemic effects [157, 158].
Citrus flavones (hesperetin)	Prevents early- or late-stage microvasculopathy by its antiangiogenic properties. Protects Muller cell processes and photoreceptors with an increase in basement membrane thickness in diabetic retina [159, 160].
Lipoic acid	Reduces VEGF levels and preserves retinal layer thickness and protects ganglion cells. Also, safeguards injured the retinas of diabetic rats by decreasing oxidative stress, partially via AMPK activation [161, 162].
Telmisartan	Increases neurotrophic factors such as BDNF, CNTF, and TH by decreasing caspase-3 activity and increasing GSH levels in the serum and diabetic retina [163].
Astaxanthin	Reduces hyperglycemia-induced abnormal proliferation and oxidative stress in retinal pigmented epithelial cells. Also downregulates retinal ganglion cell apoptosis by inhibiting oxidative stress [164, 165].
Hydrogen sulphide	Suppresses oxidative stress and exhibits neuroprotective effects on the retina and ablates oxidative stress and inflammation in STZ-induced diabetic rats. However, during PDR stage, increased H <sub>2</sub> S levels are detected in the vitreous cavity and require further studies to understand H <sub>2</sub> S's role in therapeutics [166, 167].
Tauroursodeoxycholic acid (TUDCA)	Suppresses inflammatory cytokines and molecules such as NF-kappa B, ICAM-1, and NOS (nitric oxide synthase). Also decreases the levels of VEGF and exerts neuroprotective effects in an experimental retinal detachment model [168, 169].
Curcumin	Exhibits hypoglycemic, antioxidant, and anti-inflammatory properties in diabetic rats. Additionally downregulates VEGF and has neuroprotective properties [170, 171].

TABLE 3: Antioxidants as treatment strategies in diabetic retinopathy.

is due to the contributory effect of both mitochondria and NADPH oxidase. This is further supplemented by the fact that diabetes-induced induction of the proinflammatory molecules iNOS and ICAM-1 was not observed by the removal of photoreceptor cells [128].

Epigenetic changes in the mitochondrial enzymes induced by ROS form a metabolic memory such that even controlled glycemic levels do not respite the symptoms of DR [98]. In retinal endothelial cells, hyperglycemia leads to ROS production, decreasing the levels of a class III histone deacetylase and increasing inflammatory responses from NF- $\kappa$ B [129]. Moreover, this high glucose-mediated ROS production and SIRT1 have been considered important in mediating memory phenomena of retinal endothelial cells [129].

The various species that are involved in oxidative stress includes free radical molecules like superoxide radicals, hydroxyl radicals, nonradicals like hydrogen peroxides and ozone, and reactive lipids like ketosamine and ketoaldehyde groups. These reactive species can be generated by endogenous factors including the electron transport chain of the mitochondria or from the polymorphonuclear cells [130]. Exogenous factors like UV and infrared radiations also contribute to the radical formation. These superoxide species can also lead to formation of peroxynitrate and other reactive nitrogen species, leading to nitrative stress. The eye which allows the light to penetrate every layer makes it susceptible to damage by oxidative or nitrative stress via exogenous factors [131]. Some of the culprit oxidants of DR are mentioned in Table 1. Moreover, oxidative stress also leads to a decline in certain antioxidants thus making it easier for the DR to manifest and progress. These include thioredoxin, superoxide dismutase, NADPH oxidase, Nrf2, vitamin C, and vitamin E and are listed in Table 2.

Although prevention of the progression of retinopathy in diabetic patients lies in the sole strategy of a better diet regimen and maintenance of normal glycemic conditions, new avenues for treatment are increasing with the evolution of better pharmacological targets to treat the severity of retinopathy. Since standard therapies pose a drawback in clinic such as resistance to anti-VEGF intravitreal injection and inflammatory conditions like macular oedema, an hour of need is to find better treatment strategies with lower side effects. Moreover, the balance between prooxidants and antioxidants is disrupted in DR; thus, clinical research involving the use of antioxidant therapies serves as a new direction in relieving the severity of this condition. Some of the antioxidant molecules studied as treatment strategies against DR are mentioned in Table 3. These molecules have shown to be involved in reducing the severity of the disease by exhibiting their effects on the pathways that lead to cellular damage. However, none of the treatments is efficient enough to revert the symptoms completely; further studies are needed to evaluate more potential antioxidants with specific bioactive properties in combating retinal pathophysiology associated with DR.



FIGURE 2: Perturbations in diabetic retinopathy.

# 12. Conclusions

Diabetic retinopathy, an acquired blindness, is amongst one of the leading conditions around the globe. The major insult in the progression of DR is the vascular dysfunction in the retinal blood barrier. This is a consequence of the impaired biochemical pathways such as the polyol, AGE/RAGE, hexosamine, PKC, tissue RAS, histone modifications, and altered miRNAs which are considered as major contributors in DR. These perturbations are summarized in Figure 2. The state of metabolic memory has also been implicated along with oxidative stress-induced damage to the mtDNA. This alteration in the mitochondrial complexes is the major inducer of radical species eventually resulting in depletion of antioxidants, leading to inflammation and apoptosis of retinal and endothelial cells during DR. Several treatment strategies aforementioned as the emerging area cardinally target the prooxidant/antioxidant status of the cells to protect from damage in DR. However, more studies are warranted to understand DR development and progression for the benefit of those individuals with retinal damage from diabetes.

# **Conflicts of Interest**

The authors declare no conflicts of interest.

#### References

- P. A. Campochiaro, "Molecular pathogenesis of retinal and choroidal vascular diseases," *Progress in Retinal and Eye Research*, vol. 49, pp. 67–81, 2015.
- [2] J. Cunha-Vaz, R. Bernardes, and C. Lobo, "Blood-retinal barrier," *European Journal of Ophthalmology*, vol. 21, Supplement 6, pp. 3–S9, 2018.
- [3] E. M. Kohner, V. Patel, and S. M. B. Rassam, "Role of blood flow and impaired autoregulation in the pathogenesis of diabetic retinopathy," *Diabetes*, vol. 44, no. 6, pp. 603–607, 1995.
- [4] V. A. Alder, E. N. Su, D. Y. Yu, S. Cringle, and P. Yu, "Overview of studies on metabolic and vascular regulatory changes in early diabetic retinopathy," *Australian and New Zealand Journal of Ophthalmology*, vol. 26, no. 2, pp. 141–148, 1998.

- [5] L. Schmetterer and M. Wolzt, "Ocular blood flow and associated functional deviations in diabetic retinopathy," *Diabetologia*, vol. 42, no. 4, pp. 387–405, 1999.
- [6] S. R. Flaxman, R. R. A. Bourne, S. Resnikoff et al., "Global causes of blindness and distance vision impairment 1990-2020: a systematic review and meta-analysis," *The Lancet Global Health*, vol. 5, no. 12, pp. e1221–e1234, 2017.
- [7] J. W. Y. Yau, S. L. Rogers, R. Kawasaki et al., "Global prevalence and major risk factors of diabetic retinopathy," *Diabetes Care*, vol. 35, no. 3, pp. 556–564, 2012.
- [8] K. Ogurtsova, J. D. da Rocha Fernandes, Y. Huang et al., "IDF Diabetes Atlas: global estimates for the prevalence of diabetes for 2015 and 2040," *Diabetes Research and Clinical Practice*, vol. 128, pp. 40–50, 2017.
- [9] A. R. Santiago, R. Boia, I. D. Aires, A. F. Ambrósio, and R. Fernandes, "Sweet stress: coping with vascular dysfunction in diabetic retinopathy," *Frontiers in Physiology*, vol. 9, no. 9, 2018.
- [10] M. S. Ola, M. I. Nawaz, M. M. Siddiquei, S. Al-Amro, and A. M. Abu El-Asrar, "Recent advances in understanding the biochemical and molecular mechanism of diabetic retinopathy," *Journal of Diabetes and its Complications*, vol. 26, no. 1, pp. 56–64, 2012.
- [11] J. Vislisel and T. A. Oetting, *Diabetic Retinopathy: From One Medical Student to Another*, Eye Rounds, 2010.
- [12] A. M. Hendrick, M. V. Gibson, and A. Kulshreshtha, "Diabetic retinopathy," *Primary Care: Clinics in Office Practice*, vol. 42, no. 3, pp. 451–464, 2015.
- [13] Y. Zheng, M. He, and N. Congdon, "The worldwide epidemic of diabetic retinopathy," *Indian Journal of Ophthalmology*, vol. 60, no. 5, pp. 428–431, 2012.
- [14] C. D. A. Stehouwer, "Microvascular dysfunction and hyperglycemia: a vicious cycle with widespread consequences," *Diabetes*, vol. 67, no. 9, pp. 1729–1741, 2018.
- [15] M. Alves, R. Boia, E. J. Campos et al., "Subtle thinning of retinal layers without overt vascular and inflammatory alterations in a rat model of prediabetes," *Molecular Vision*, vol. 24, pp. 353–366, 2018.
- [16] R. Donnelly and E. Horton, Vascular Complications of Diabetes: Current Issues in Pathogenesis and Treatment, John Wiley & Sons, 2008.

- [17] F. Caliva, G. Leontidis, P. Chudzik, A. Hunter, L. Antiga, and B. Al-Diri, "Hemodynamics in the retinal vasculature during the progression of diabetic retinopathy," *Journal for Modeling in Ophthalmology*, vol. 1, no. 4, pp. 6–15, 2017.
- [18] E. Fletcher, J. Phipps, M. Ward, T. Puthussery, and J. Wilkinson-Berka, "Neuronal and glial cell abnormality as predictors of progression of diabetic retinopathy," *Current Pharmaceutical Design*, vol. 13, no. 26, pp. 2699–2712, 2007.
- [19] M. Brownlee, "The pathobiology of diabetic complications: a unifying mechanism," *Diabetes*, vol. 54, no. 6, pp. 1615–1625, 2005.
- [20] A. Maugeri, M. G. Mazzone, F. Giuliano et al., "Curcumin modulates DNA methyltransferase functions in a cellular model of diabetic retinopathy," *Oxidative Medicine and Cellular Longevity*, vol. 2018, Article ID 5407482, 12 pages, 2018.
- [21] K. H. Gabbay, "The sorbitol pathway and the complications of diabetes," *The New England Journal of Medicine*, vol. 288, no. 16, pp. 831–836, 1973.
- [22] J. M. Tarr, K. Kaul, M. Chopra, E. M. Kohner, and R. Chibber, "Pathophysiology of diabetic retinopathy," *ISRN Ophthalmology*, vol. 2013, Article ID 343560, 13 pages, 2013.
- [23] S. Roy, T. S. Kern, B. Song, and C. Stuebe, "Mechanistic insights into pathological changes in the diabetic retina: implications for targeting diabetic retinopathy," *American Journal of Pathology*, vol. 187, no. 1, pp. 9–19, 2017.
- [24] M. G. Buse, "Hexosamines, insulin resistance and the complications of diabetes: current status," *American Journal of Physiology-Endocrinology and Metabolism*, vol. 290, no. 1, pp. E1–E8, 2006.
- [25] P. Geraldes and G. L. King, "Activation of protein kinase C isoforms and its impact on diabetic complications," *Circulation Research*, vol. 106, no. 8, pp. 1319–1331, 2010.
- [26] J. A. Groves, A. Lee, G. Yildirir, and N. E. Zachara, "Dynamic O-GlcNAcylation and its roles in the cellular stress response and homeostasis," *Cell Stress Chaperones*, vol. 18, no. 5, pp. 535–558, 2013.
- [27] Y. Wu, L. Tang, and B. Chen, "Oxidative stress: implications for the development of diabetic retinopathy and antioxidant therapeutic perspectives," *Oxidative Medicine and Cellular Longevity*, vol. 2014, Article ID 752387, 12 pages, 2014.
- [28] M. Lorenzi, "The polyol pathway as a mechanism for diabetic retinopathy: attractive, elusive, and resilient," *Journal* of *Diabetes Research*, vol. 2007, Article ID 61038, 10 pages, 2007.
- [29] M. Zhou, P. Zhang, X. Xu, and X. Sun, "The relationship between aldose reductase C106T polymorphism and diabetic retinopathy: an updated meta-analysis," *Investigative Ophthalmology & Visual Science*, vol. 56, no. 4, pp. 2279–2289, 2015.
- [30] Z. Dagher, Y. S. Park, V. Asnaghi, T. Hoehn, C. Gerhardinger, and M. Lorenzi, "Studies of rat and human retinas predict a role for the polyol pathway in human diabetic retinopathy," *Diabetes*, vol. 53, no. 9, pp. 2404–2411, 2004.
- [31] F. Willermain, L. Scifo, C. Weber, L. Caspers, J. Perret, and C. Delporte, "Potential interplay between hyperosmolarity and inflammation on retinal pigmented epithelium in pathogenesis of diabetic retinopathy," *International Journal of Molecular Sciences*, vol. 19, no. 4, 2018.
- [32] B. Szwergold, F. Kappler, and T. Brown, "Identification of fructose 3-phosphate in the lens of diabetic rats," *Science*, vol. 247, no. 4941, pp. 451–454, 1990.

- [33] K.-C. Chang, A. Snow, D. V. LaBarbera, and J. M. Petrash, "Aldose reductase inhibition alleviates hyperglycemic effects on human retinal pigment epithelial cells," *Chemico-Biological Interactions*, vol. 234, pp. 254–260, 2015.
- [34] B. Giri, S. Dey, T. Das, M. Sarkar, J. Banerjee, and S. K. Dash, "Chronic hyperglycemia mediated physiological alteration and metabolic distortion leads to organ dysfunction, infection, cancer progression and other pathophysiological consequences: an update on glucose toxicity," *Biomedicine & Pharmacotherapy*, vol. 107, pp. 306–328, 2018.
- [35] B. Vyas, S. Choudhary, P. K. Singh et al., "Identification of 2-benzoxazolinone derivatives as lead against molecular targets of diabetic complications," *Chemical Biology & Drug Design*, vol. 92, no. 6, pp. 1981–1987, 2018.
- [36] S. Khan, T. Bhardwaj, P. Somvanshi et al., "Inhibition of C298S mutant of human aldose reductase for antidiabetic applications: evidence from in silico elementary mode analysis of biological network model," *Journal of Cellular Biochemistry*, vol. 119, no. 8, pp. 6961–6973, 2018.
- [37] E. Bejarano and A. Taylor, "Too sweet: problems of protein glycation in the eye," *Experimental Eye Research*, vol. 178, pp. 255–262, 2019.
- [38] M. Brownlee, "Biochemistry and molecular cell biology of diabetic complications," *Nature*, vol. 414, no. 6865, pp. 813– 820, 2001.
- [39] N. C. Chilelli, S. Burlina, and A. Lapolla, "AGEs, rather than hyperglycemia, are responsible for microvascular complications in diabetes: a "glycoxidation-centric" point of view," *Nutrition, Metabolism & Cardiovascular Diseases*, vol. 23, no. 10, pp. 913–919, 2013.
- [40] M. B. Manigrasso, J. Juranek, R. Ramasamy, and A. M. Schmidt, "Unlocking the biology of RAGE in diabetic microvascular complications," *Trends in Endocrinology & Metabolism*, vol. 25, no. 1, pp. 15–22, 2014.
- [41] L. Sun, T. Huang, W. Xu, J. Sun, Y. Lv, and Y. Wang, "Advanced glycation end products promote VEGF expression and thus choroidal neovascularization via Cyr61-PI3-K/AKT signaling pathway," *Scientific Reports*, vol. 7, no. 1, article 14925, 2017.
- [42] R. A. Kowluru, "Diabetic retinopathy, metabolic memory and epigenetic modifications," *Vision Research*, vol. 139, pp. 30– 38, 2017.
- [43] S. Yamagishi, K. Nakamura, and T. Imaizumi, "Advanced glycation end products (AGEs) and diabetic vascular complications," *Current Diabetes Reviews*, vol. 1, no. 1, pp. 93–106, 2005.
- [44] H. P. Hammes, Y. Feng, F. Pfister, and M. Brownlee, "Diabetic retinopathy: targeting vasoregression," *Diabetes*, vol. 60, no. 1, pp. 9–16, 2010.
- [45] J. Xu, L. J. Chen, J. Yu et al., "Involvement of advanced glycation end products in the pathogenesis of diabetic retinopathy," *Cellular Physiology and Biochemistry*, vol. 48, no. 2, pp. 705–717, 2018.
- [46] C. M. Sena, P. Matafome, J. Crisóstomo et al., "Methylglyoxal promotes oxidative stress and endothelial dysfunction," *Pharmacological Research*, vol. 65, no. 5, pp. 497–506, 2012.
- [47] C. M. Chan, D. Y. Huang, Y. P. Huang et al., "Methylglyoxal induces cell death through endoplasmic reticulum stress-associated ROS production and mitochondrial dysfunction," *Journal of Cellular and Molecular Medicine*, vol. 20, no. 9, pp. 1749–1760, 2016.

- [48] M.-K. Kang, E.-J. Lee, Y.-H. Kim et al., "Chrysin ameliorates malfunction of retinoid visual cycle through blocking activation of AGE-RAGE-ER stress in glucose-stimulated retinal pigment epithelial cells and diabetic eyes," *Nutrients*, vol. 10, no. 8, 2018.
- [49] Y. Qiu, D. Zhao, V. M. Butenschön et al., "Nucleoside diphosphate kinase B deficiency causes a diabetes-like vascular pathology via up-regulation of endothelial angiopoietin-2 in the retina," *Acta Diabetologica*, vol. 53, no. 1, pp. 81–89, 2016.
- [50] G. W. Hart, C. Slawson, G. Ramirez-Correa, and O. Lagerlof, "Cross talk between O-GlcNAcylation and phosphorylation: roles in signaling, transcription, and chronic disease," *Annual Review of Biochemistry*, vol. 80, no. 1, pp. 825–858, 2011.
- [51] S. J. Kim, W. S. Yoo, M. Choi, I. Chung, J. M. Yoo, and W. S. Choi, "Increased O-GlcNAcylation of NF-κB enhances retinal ganglion cell death in streptozotocin-induced diabetic retinopathy," *Current Eye Research*, vol. 41, no. 2, pp. 249–257, 2016.
- [52] S. Zraika, M. Dunlop, J. Proietto, and S. Andrikopoulos, "The hexosamine biosynthesis pathway regulates insulin secretion via protein glycosylation in mouse islets," *Archives of Biochemistry and Biophysics*, vol. 405, no. 2, pp. 275–279, 2002.
- [53] C. D'Alessandris, F. Andreozzi, M. Federici et al., "Increased O-glycosylation of insulin signaling proteins results in their impaired activation and enhanced susceptibility to apoptosis in pancreatic  $\beta$ -cells," *FASEB Journal*, vol. 18, no. 9, pp. 959–961, 2004.
- [54] Z. Gurel, B. W. Zaro, M. R. Pratt, and N. Sheibani, "Identification of O-GlcNAc modification targets in mouse retinal pericytes: implication of p53 in pathogenesis of diabetic retinopathy," *PLoS One*, vol. 9, no. 5, article e95561, 2014.
- [55] Z. Gurel, K. M. Sieg, K. D. Shallow, C. M. Sorenson, and N. Sheibani, "Retinal O-linked N-acetylglucosamine protein modifications: implications for postnatal retinal vascularization and the pathogenesis of diabetic retinopathy," *Molecular Vision*, vol. 19, pp. 1047–1059, 2013.
- [56] Z. Gurel and N. Sheibani, "O-linked β-N-acetylglucosamine (O-GlcNAc) modification: a new pathway to decode pathogenesis of diabetic retinopathy," *Clinical Science*, vol. 132, no. 2, pp. 185–198, 2018.
- [57] Y. Kang, D. C. Lee, J. Han et al., "NM23-H2 involves in negative regulation of diva and Bcl2L10 in apoptosis signaling," *Biochemical and Biophysical Research Communications*, vol. 359, no. 1, pp. 76–82, 2007.
- [58] K. S. Krishnan, R. Rikhy, S. Rao et al., "Nucleoside diphosphate kinase, a source of GTP, is required for dynamin dependent synaptic vesicle recycling," *Neuron*, vol. 30, no. 1, pp. 197–210, 2001.
- [59] F. Palacios, J. K. Schweitzer, R. L. Boshans, and C. D'Souza-Schorey, "ARF6-GTP recruits Nm23-H1 to facilitate dynamin-mediated endocytosis during adherens junctions disassembly," *Nature Cell Biology*, vol. 4, no. 12, pp. 929– 936, 2002.
- [60] P. V. Attwood and R. Muimo, "The actions of NME1/ NDPK-A and NME2/NDPK-B as protein kinases," *Laboratory Investigation*, vol. 98, no. 3, pp. 283–290, 2018.
- [61] Y. Feng, S. Gross, N. M. Wolf et al., "Nucleoside diphosphate kinase B regulates angiogenesis through modulation of vascular endothelial growth factor receptor type 2 and endothelial adherens junction proteins," *Arteriosclerosis, Thrombosis, and Vascular Biology*, vol. 34, no. 10, pp. 2292–2300, 2014.

- [62] S. Shan, A. Chatterjee, Y. Qiu, H. P. Hammes, T. Wieland, and Y. Feng, "O-GlcNAcylation of FoxO1 mediates nucleoside diphosphate kinase B deficiency induced endothelial damage," *Scientific Reports*, vol. 8, no. 1, article 10581, 2018.
- [63] G. L. King, H. Ishii, and D. Koya, "Diabetic vascular dysfunctions: a model of excessive activation of protein kinase C," *Kidney International*, vol. 60, pp. S77–S85, 1997.
- [64] R. A. Kowluru and P. S. Chan, "Oxidative stress and diabetic retinopathy," *Experimental Diabetes Research*, vol. 2007, Article ID 43603, 12 pages, 2007.
- [65] S. F. Steinberg, "Structural basis of protein kinase C isoform function," *Physiological Reviews*, vol. 88, pp. 1341–1378, 2008.
- [66] B. A. Wolf, J. R. Williamson, R. A. Easom, K. Chang, W. R. Sherman, and J. Turk, "Diacylglycerol accumulation and microvascular abnormalities induced by elevated glucose levels," *Journal of Clinical Investigation*, vol. 87, no. 1, pp. 31–38, 1991.
- [67] T. Shiba, T. Inoguchi, J. R. Sportsman, W. F. Heath, S. Bursell, and G. L. King, "Correlation of diacylglycerol level and protein kinase C activity in rat retina to retinal circulation," *American Journal of Physiology-Endocrinology and Metabolism*, vol. 265, no. 5, pp. E783–E793, 1993.
- [68] D. Koya and G. L. King, "Protein kinase C activation and the development of diabetic complications," *Diabetes*, vol. 47, no. 6, pp. 859–866, 1998.
- [69] D. Portilla, G. Dai, J. M. Peters, F. J. Gonzalez, M. D. Crew, and A. D. Proia, "Etomoxir-induced PPARα-modulated enzymes protect during acute renal failure," *American Journal* of *Physiology-Renal Physiology*, vol. 278, no. 4, pp. F667–F675, 2000.
- [70] R. J. Keogh, M. E. Dunlop, and R. G. Larkins, "Effect of inhibition of aldose reductase on glucose flux, diacylglycerol formation, protein kinase C, and phospholipase A2 activation," *Metabolism*, vol. 46, no. 1, pp. 41–47, 1997.
- [71] Y. Nishizuka, "Intracellular signaling by hydrolysis of phospholipids and activation of protein kinase C," *Science*, vol. 258, no. 5082, pp. 607–614, 1992.
- [72] Y. Nishizuka, "Protein kinase C and lipid signaling for sustained cellular responses," *The FASEB Journal*, vol. 9, no. 7, pp. 484–496, 1995.
- [73] C. Takagi, S. E. Bursell, Y. W. Lin et al., "Regulation of retinal hemodynamics in diabetic rats by increased expression and action of endothelin-1," *Investigative Ophthalmology & Visual Science*, vol. 37, no. 12, pp. 2504–2518, 1996.
- [74] L. P. Aiello, R. L. Avery, P. G. Arrigg et al., "Vascular endothelial growth factor in ocular fluid of patients with diabetic retinopathy and other retinal disorders," *New England Journal of Medicine*, vol. 331, no. 22, pp. 1480–1487, 1994.
- [75] B. Williams, B. Gallacher, H. Patel, and C. Orme, "Glucose-induced protein kinase C activation regulates vascular permeability factor mRNA expression and peptide production by human vascular smooth muscle cells in vitro," *Diabetes*, vol. 46, no. 9, pp. 1497–1503, 1997.
- [76] P. Geraldes, J. Hiraoka-Yamamoto, M. Matsumoto et al., "Activation of PKC-δ and SHP-1 by hyperglycemia causes vascular cell apoptosis and diabetic retinopathy," *Nature Medicine*, vol. 15, no. 11, pp. 1298–1306, 2009.
- [77] D. E. Sims, "The pericyte—a review," *Tissue & Cell*, vol. 18, no. 2, pp. 153–174, 1986.
- [78] J. A. Choi, Y. R. Chung, H. R. Byun, H. Park, J. Y. Koh, and Y. H. Yoon, "The anti-ALS drug riluzole attenuates pericyte

loss in the diabetic retinopathy of streptozotocin-treated mice," *Toxicology and Applied Pharmacology*, vol. 315, pp. 80–89, 2017.

- [79] J. H. Kim, J. H. Kim, Y. S. Yu, C. S. Cho, and K. W. Kim, "Blockade of angiotensin II attenuates VEGF-mediated blood—retinal barrier breakdown in diabetic retinopathy," *Journal of Cerebral Blood Flow & Metabolism*, vol. 29, no. 3, pp. 621–628, 2008.
- [80] M. Bader and D. Ganten, "Update on tissue renin-angiotensin systems," *Journal of Molecular Medicine*, vol. 86, no. 6, pp. 615–621, 2008.
- [81] A. Kanda, K. Noda, and S. Ishida, "ATP6AP2/(pro)renin receptor contributes to glucose metabolism via stabilizing the pyruvate dehydrogenase E1 β subunit," *Journal of Biological Chemistry*, vol. 290, no. 15, pp. 9690–9700, 2015.
- [82] G. Nguyen, F. Delarue, C. Burcklé, L. Bouzhir, T. Giller, and J. D. Sraer, "Pivotal role of the renin/prorenin receptor in angiotensin II production and cellular responses to renin," *Journal of Clinical Investigation*, vol. 109, no. 11, pp. 1417– 1427, 2002.
- [83] A. Ichihara, M. Hayashi, Y. Kaneshiro et al., "Inhibition of diabetic nephropathy by a decoy peptide corresponding to the "handle" region for nonproteolytic activation of prorenin," *Journal of Clinical Investigation*, vol. 114, no. 8, pp. 1128–1135, 2004.
- [84] A. Kanda, K. Noda, W. Saito, and S. Ishida, "(Pro)renin receptor is associated with angiogenic activity in proliferative diabetic retinopathy," *Diabetologia*, vol. 55, no. 11, pp. 3104– 3113, 2012.
- [85] A. Kanda, K. Noda, W. Saito, and S. Ishida, "Vitreous renin activity correlates with vascular endothelial growth factor in proliferative diabetic retinopathy," *British Journal of Ophthalmology*, vol. 97, no. 5, pp. 666–668, 2013.
- [86] S. Satofuka, A. Ichihara, N. Nagai et al., "(Pro)renin receptor-mediated signal transduction and tissue renin-angiotensin system contribute to diabetes-induced retinal inflammation," *Diabetes*, vol. 58, no. 7, pp. 1625–1633, 2009.
- [87] A. H. J. Danser, M. A. van den Dorpel, J. Deinum et al., "Renin, prorenin, and immunoreactive renin in vitreous fluid from eyes with and without diabetic retinopathy," *The Journal of Clinical Endocrinology & Metabolism*, vol. 68, no. 1, pp. 160–167, 1989.
- [88] H. Funatsu, H. Yamashita, Y. Nakanishi, and S. Hori, "Angiotensin II and vascular endothelial growth factor in the vitreous fluid of patients with proliferative diabetic retinopathy," *British Journal of Ophthalmology*, vol. 86, no. 3, pp. 311– 315, 2002.
- [89] C. J. Moravski, S. L. Skinner, A. J. Stubbs et al., "The renin-angiotensin system influences ocular endothelial cell proliferation in diabetes," *The American Journal of Pathology*, vol. 162, no. 1, pp. 151–160, 2003.
- [90] R. L. Engerman and T. S. Kern, "Progression of incipient diabetic retinopathy during good glycemic control," *Diabetes*, vol. 36, no. 7, pp. 808–812, 1987.
- [91] R. N. Frank, "Diabetic retinopathy," New England Journal of Medicine, vol. 350, no. 1, pp. 48–58, 2004.
- [92] Z. Maghbooli, A. Hossein-nezhad, B. Larijani, M. Amini, and A. Keshtkar, "Global DNA methylation as a possible biomarker for diabetic retinopathy," *Diabetes/Metabolism Research and Reviews*, vol. 31, no. 2, pp. 183–189, 2015.

- [93] E. Agardh, A. Lundstig, A. Perfilyev et al., "Genome-wide analysis of DNA methylation in subjects with type 1 diabetes identifies epigenetic modifications associated with proliferative diabetic retinopathy," *BMC Medicine*, vol. 13, no. 1, p. 182, 2015.
- [94] S. Tewari, Q. Zhong, J. M. Santos, and R. A. Kowluru, "Mitochondria DNA replication and DNA methylation in the metabolic memory associated with continued progression of diabetic retinopathy," *Investigative Ophthalmology & Visual Science*, vol. 53, no. 8, pp. 4881–4888, 2012.
- [95] Q. Zhong and R. A. Kowluru, "Regulation of matrix metalloproteinase-9 by epigenetic modifications and the development of diabetic retinopathy," *Diabetes*, vol. 62, no. 7, pp. 2559–2568, 2013.
- [96] R. A. Kowluru, J. M. Santos, and Q. Zhong, "Sirt1, a negative regulator of matrix metalloproteinase-9 in diabetic retinopathy," *Investigative Ophthalmology & Visual Science*, vol. 55, no. 9, pp. 5653–5660, 2014.
- [97] X. Zhang, S. Bao, D. Lai, R. W. Rapkins, and M. C. Gillies, "Intravitreal triamcinolone acetonide inhibits breakdown of the blood-retinal barrier through differential regulation of VEGF-A and its receptors in early diabetic rat retinas," *Diabetes*, vol. 57, no. 4, pp. 1026–1033, 2008.
- [98] S. Jarrett, H. Lin, B. Godley, and M. Boulton, "Mitochondrial DNA damage and its potential role in retinal degeneration," *Progress in Retinal and Eye Research*, vol. 27, no. 6, pp. 596–607, 2008.
- [99] X. Zhang, L. Zhao, B. Hambly, S. Bao, and K. Wang, "Diabetic retinopathy: reversibility of epigenetic modifications and new therapeutic targets," *Cell & Bioscience*, vol. 7, no. 1, p. 42, 2017.
- [100] J. W. Kaspar, S. K. Niture, and A. K. Jaiswal, "Nrf2:INrf2 (Keap1) signaling in oxidative stress," *Free Radical Biology* and Medicine, vol. 47, no. 9, pp. 1304–1309, 2009.
- [101] M. Mishra, Q. Zhong, and R. A. Kowluru, "Epigenetic modifications of Keap1 regulate its interaction with the protective factor Nrf2 in the development of diabetic retinopathy," *Investigative Ophthalmology & Visual Science*, vol. 55, no. 11, pp. 7256–7265, 2014.
- [102] M. D. Williams, H. van Remmen, C. C. Conrad, T. T. Huang, C. J. Epstein, and A. Richardson, "Increased oxidative damage is correlated to altered mitochondrial function in heterozygous manganese superoxide dismutase knockout mice," *Journal of Biological Chemistry*, vol. 273, no. 43, pp. 28510– 28515, 1998.
- [103] Q. Zhong and R. A. Kowluru, "Epigenetic changes in mitochondrial superoxide dismutase in the retina and the development of diabetic retinopathy," *Diabetes*, vol. 60, no. 4, pp. 1304–1313, 2011.
- [104] R. C. Scarpulla, "Nuclear control of respiratory chain expression by nuclear respiratory factors and PGC-1-related coactivator," Annals of the New York Academy of Sciences, vol. 1147, no. 1, pp. 321–334, 2008.
- [105] J. M. Santos, S. Tewari, A. F. X. Goldberg, and R. A. Kowluru, "Mitochondrial biogenesis and the development of diabetic retinopathy," *Free Radical Biology and Medicine*, vol. 51, no. 10, pp. 1849–1860, 2011.
- [106] J. M. Santos and R. A. Kowluru, "Role of mitochondria biogenesis in the metabolic memory associated with the continued progression of diabetic retinopathy and its regulation by lipoic acid," *Investigative Ophthalmology & Visual Science*, vol. 52, no. 12, pp. 8791–8798, 2011.

- [107] R. W. Carthew and E. J. Sontheimer, "Origins and mechanisms of miRNAs and siRNAs," *Cell*, vol. 136, no. 4, pp. 642–655, 2009.
- [108] Q. Gong, J. Xie, Y. Liu, Y. Li, and G. Su, "Differentially expressed MicroRNAs in the development of early diabetic retinopathy," *Journal of Diabetes Research*, vol. 2017, Article ID 4727942, 10 pages, 2017.
- [109] J. H. Wu, Y. Gao, A. J. Ren et al., "Altered microRNA expression profiles in retinas with diabetic retinopathy," *Ophthalmic Research*, vol. 47, no. 4, pp. 195–201, 2012.
- [110] S. Zhao, T. Li, J. Li et al., "MiR-23b-3p induces the cellular metabolic memory of high glucose in diabetic retinopathy through a SIRT1-dependent signalling pathway," *Diabetologia*, vol. 59, no. 3, pp. 644–654, 2016.
- [111] R. Mortuza, B. Feng, and S. Chakrabarti, "miR-195 regulates SIRT1-mediated changes in diabetic retinopathy," *Diabetologia*, vol. 57, no. 5, pp. 1037–1046, 2014.
- [112] M. Lagos-Quintana, R. Rauhut, J. Meyer, A. Borkhardt, and T. Tuschl, "New microRNAs from mouse and human," *RNA*, vol. 9, no. 2, pp. 175–179, 2003.
- [113] N. Kaludercic and V. Giorgio, "The dual function of reactive oxygen/nitrogen species in bioenergetics and cell death: the role of ATP synthase," *Oxidative Medicine and Cellular Longevity*, vol. 2016, Article ID 3869610, 17 pages, 2016.
- [114] E. Hernández-Ramírez, G. Sánchez-Chávez, L. A. Estrella-Salazar, and R. Salceda, "Nitrosative stress in the rat retina at the onset of streptozotocin-induced diabetes," *Cellular Physiology and Biochemistry*, vol. 42, no. 6, pp. 2353–2363, 2017.
- [115] L. K. Mandal, S. Choudhuri, D. Dutta et al., "Oxidative stress-associated neuroretinal dysfunction and nitrosative stress in diabetic retinopathy," *Canadian Journal of Diabetes*, vol. 37, no. 6, pp. 401–407, 2013.
- [116] S. Sharma, S. Saxena, K. Srivastav et al., "Nitric oxide and oxidative stress is associated with severity of diabetic retinopathy and retinal structural alterations," *Clinical & Experimental Ophthalmology*, vol. 43, no. 5, pp. 429–436, 2015.
- [117] F. Lamoke, S. Shaw, J. Yuan et al., "Increased oxidative and nitrative stress accelerates aging of the retinal vasculature in the diabetic retina," *PLoS One*, vol. 10, no. 10, article e0139664, 2015.
- [118] R. Opatrilova, P. Kubatka, M. Caprnda et al., "Nitric oxide in the pathophysiology of retinopathy: evidences from preclinical and clinical researches," *Acta Ophthalmologica*, vol. 96, no. 3, pp. 222–231, 2018.
- [119] T. S. Kern, J. Tang, M. Mizutani et al., "Response of capillary cell death to aminoguanidine predicts the development of retinopathy: comparison of diabetes and galactosemia," *Investigative Ophthalmology & Visual Science*, vol. 41, no. 12, pp. 3972–3978, 2000.
- [120] T. S. Kern and R. L. Engerman, "Pharmacological inhibition of diabetic retinopathy: aminoguanidine and aspirin," *Diabetes*, vol. 50, no. 7, pp. 1636–1642, 2001.
- [121] Y. Du, M. A. Smith, C. M. Miller, and T. S. Kern, "Diabetes-induced nitrative stress in the retina, and correction by aminoguanidine," *Journal of Neurochemistry*, vol. 80, no. 5, pp. 771–779, 2002.
- [122] P. Voabil, J. Liberal, E. C. Leal et al., "Calcium dobesilate is protective against inflammation and oxidative/nitrosative stress in the retina of a type 1 diabetic rat model," *Ophthalmic Research*, vol. 58, no. 3, pp. 150–161, 2017.

- [123] R. E. Anderson, L. M. Rapp, and R. D. Wiegand, "Lipid peroxidation and retinal degeneration," *Current Eye Research*, vol. 3, no. 1, pp. 223–227, 1984.
- [124] J. L. Wilkinson-Berka, I. Rana, R. Armani, and A. Agrotis, "Reactive oxygen species, Nox and angiotensin II in angiogenesis: implications for retinopathy," *Clinical Science*, vol. 124, no. 10, pp. 597–615, 2013.
- [125] V. Calabrese, E. Guagliano, M. Sapienza et al., "Redox regulation of cellular stress response in aging and neurodegenerative disorders: role of vitagenes," *Neurochemical Research*, vol. 32, no. 4-5, pp. 757–773, 2007.
- [126] M. Barot, M. R. Gokulgandhi, and A. K. Mitra, "Mitochondrial dysfunction in retinal diseases," *Current Eye Research*, vol. 36, no. 12, pp. 1069–1077, 2011.
- [127] X. Yang, F. Huo, B. Liu et al., "Crocin inhibits oxidative stress and pro-inflammatory response of microglial cells associated with diabetic retinopathy through the activation of PI3K/Akt signaling pathway," *Journal of Molecular Neuroscience*, vol. 61, no. 4, pp. 581–589, 2017.
- [128] Y. Du, A. Veenstra, K. Palczewski, and T. S. Kern, "Photoreceptor cells are major contributors to diabetes-induced oxidative stress and local inflammation in the retina," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 110, no. 41, pp. 16586–16591, 2013.
- [129] H. Karbasforooshan and G. Karimi, "The role of SIRT1 in diabetic retinopathy," *Biomedicine & Pharmacotherapy*, vol. 97, pp. 190–194, 2018.
- [130] G. D. Calderon, O. H. Juarez, G. E. Hernandez, S. M. Punzo, and Z. D. de la Cruz, "Oxidative stress and diabetic retinopathy: development and treatment," *Eye*, vol. 31, no. 8, pp. 1122–1130, 2017.
- [131] P. N. Youssef, N. Sheibani, and D. M. Albert, "Retinal light toxicity," *Eye*, vol. 25, no. 1, pp. 1–14, 2011.
- [132] S. A. Madsen-Bouterse, Q. Zhong, G. Mohammad, Y. S. Ho, and R. A. Kowluru, "Oxidative damage of mitochondrial DNA in diabetes and its protection by manganese superoxide dismutase," *Free Radical Research*, vol. 44, no. 3, pp. 313–321, 2010.
- [133] D. C. Guzman, H. J. Olguín, E. H. García, A. V. Peraza, D. Z. de la Cruz, and M. P. Soto, "Mechanisms involved in the development of diabetic retinopathy induced by oxidative stress," *Redox Report*, vol. 22, no. 1, pp. 10–16, 2017.
- [134] C. C. Winterbourn, "Toxicity of iron and hydrogen peroxide: the Fenton reaction," *Toxicology Letters*, vol. 82-83, pp. 969– 974, 1995.
- [135] Y. Shirai, A. Mori, T. Nakahara, K. Sakamoto, and K. Ishii, "Deferiprone protects against photoreceptor degeneration induced by tunicamycin in the rat retina," *Biological and Pharmaceutical Bulletin*, vol. 38, no. 7, pp. 1076–1080, 2015.
- [136] A. J. Augustin, W. Breipohl, T. Böker, J. Lutz, and M. Spitznas, "Increased lipid peroxide levels and myeloperoxidase activity in the vitreous of patients suffering from proliferative diabetic retinopathy," *Graefe's Archive for Clinical and Experimental Ophthalmology*, vol. 231, no. 11, pp. 647–650, 1993.
- [137] X. Chen, H. Hall, J. P. Simpson, W. D. Leon-Salas, D. F. Ready, and V. M. Weake, "Cytochrome b5 protects photoreceptors from light stress-induced lipid peroxidation and retinal degeneration," *npj Aging and Mechanisms of Disease*, vol. 3, no. 1, p. 18, 2017.
- [138] Q. Liu, F. Zhang, X. Zhang et al., "Fenofibrate ameliorates diabetic retinopathy by modulating Nrf2 signaling and

NLRP3 inflammasome activation," *Molecular and Cellular Biochemistry*, vol. 445, no. 1-2, pp. 105–115, 2018.

- [139] R. A. Kowluru, "Diabetic retinopathy: mitochondrial dysfunction and retinal capillary cell death," *Antioxidants & Redox Signaling*, vol. 7, no. 11-12, pp. 1581–1587, 2005.
- [140] X. Ren, H. Ma, Y. Qiu et al., "The downregulation of thioredoxin accelerated Neuro2a cell apoptosis induced by advanced glycation end product via activating several pathways," *Neurochemistry International*, vol. 87, pp. 128–135, 2015.
- [141] L. Perrone, T. S. Devi, K. I. Hosoya, T. Terasaki, and L. P. Singh, "Inhibition of TXNIP expression in vivo blocks early pathologies of diabetic retinopathy," *Cell Death & Disease*, vol. 1, no. 8, article e65, 2010.
- [142] L. P. Singh, "Thioredoxin interacting protein (TXNIP) and pathogenesis of diabetic retinopathy," *Journal of Clinical & Experimental Ophthalmology*, vol. 4, no. 4, 2013.
- [143] T. Fukai and M. Ushio-Fukai, "Superoxide dismutases: role in redox signaling, vascular function, and diseases," *Antioxidants & Redox Signaling*, vol. 15, no. 6, pp. 1583– 1606, 2011.
- [144] A. Behndig, B. Svensson, S. L. Marklund, and K. Karlsson, "Superoxide dismutase isoenzymes in the human eye," *Investigative Ophthalmology & Visual Science*, vol. 39, no. 3, pp. 471–475, 1998.
- [145] R. A. Kowluru, L. Atasi, and Y. S. Ho, "Role of mitochondrial superoxide dismutase in the development of diabetic retinopathy," *Investigative Opthalmology & Visual Science*, vol. 47, no. 4, p. 1594, 2006.
- [146] J. Li, J. J. Wang, and S. X. Zhang, "NADPH oxidase 4-derived H<sub>2</sub>O<sub>2</sub> promotes aberrant retinal neovascularization via activation of VEGF receptor 2 pathway in oxygen-induced retinopathy," *Journal of Diabetes Research*, vol. 2015, Article ID 963289, 13 pages, 2015.
- [147] J. C. Rivera, R. Dabouz, B. Noueihed, S. Omri, H. Tahiri, and S. Chemtob, "Ischemic retinopathies: oxidative stress and inflammation," *Oxidative Medicine and Cellular Longevity*, vol. 2017, Article ID 3940241, 16 pages, 2017.
- [148] S. Z. Safi, R. Qvist, S. Kumar, K. Batumalaie, and I. S. B. Ismail, "Molecular mechanisms of diabetic retinopathy, general preventive strategies, and novel therapeutic targets," *BioMed Research International*, vol. 2014, Article ID 801269, 18 pages, 2014.
- [149] A. S. Ibrahim, S. Elshafey, H. Sellak et al., "A lipidomic screen of hyperglycemia-treated HRECs links 12/15-lipoxygenase to microvascular dysfunction during diabetic retinopathy via NADPH oxidase," *Journal of Lipid Research*, vol. 56, no. 3, pp. 599–611, 2015.
- [150] W. Meng, K. P. Shah, S. Pollack et al., "A genome-wide association study suggests new evidence for an association of the NADPH oxidase 4 (*NOX4*) gene with severe diabetic retinopathy in type 2 diabetes," *Acta Ophthalmologica*, vol. 96, no. 7, pp. e811–e819, 2018.
- [151] A. Konior, A. Schramm, M. Czesnikiewicz-Guzik, and T. J. Guzik, "NADPH oxidases in vascular pathology," *Antioxidants & Redox Signaling*, vol. 20, no. 17, pp. 2794–2814, 2014.
- [152] M. G. Traber and J. F. Stevens, "Vitamins C and E: beneficial effects from a mechanistic perspective," *Free Radical Biology and Medicine*, vol. 51, no. 5, pp. 1000–1013, 2011.

- [153] I. P. Chatziralli, G. Theodossiadis, P. Dimitriadis et al., "The effect of vitamin E on oxidative stress indicated by serum malondialdehyde in insulin-dependent *type 2* diabetes mellitus patients with retinopathy," *The Open Ophthalmology Journal*, vol. 11, no. 1, pp. 51–58, 2017.
- [154] J. M. May, "Ascorbic acid repletion: a possible therapy for diabetic macular edema?," *Free Radical Biology and Medicine*, vol. 94, pp. 47–54, 2016.
- [155] J. Li, S. Yu, J. Ying, T. Shi, and P. Wang, "Resveratrol prevents ROS-induced apoptosis in high glucose-treated retinal capillary endothelial cells via the activation of AMPK/Sirt1/ PGC-1α pathway," Oxidative Medicine and Cellular Longevity, vol. 2017, Article ID 7584691, 10 pages, 2017.
- [156] K. Zeng, Y. Wang, N. Yang et al., "Resveratrol inhibits diabetic-induced Müller cells apoptosis through micro-RNA-29b/specificity protein 1 pathway," *Molecular Neurobiology*, vol. 54, no. 6, pp. 4000–4014, 2017.
- [157] W. Liu, S. S. Liou, T. Y. Hong, and I. M. Liu, "Protective effects of hesperidin (citrus flavonone) on high glucose induced oxidative stress and apoptosis in a cellular model for diabetic retinopathy," *Nutrients*, vol. 9, no. 12, article 1312, 2017.
- [158] W. Liu, S.-S. Liou, T.-Y. Hong, and I.-M. Liu, "Hesperidin prevents high glucose-induced damage of retinal pigment epithelial cells," *Planta Medica*, vol. 84, no. 14, pp. 1030–1037, 2018.
- [159] B. Kumar, S. K. Gupta, B. P. Srinivasan, T. C. Nag, S. Srivastava, and R. Saxena, "Hesperetin ameliorates hyperglycemia induced retinal vasculopathy via anti-angiogenic effects in experimental diabetic rats," *Vascular Pharmacology*, vol. 57, no. 5-6, pp. 201–207, 2012.
- [160] B. Kumar, S. K. Gupta, B. P. Srinivasan et al., "Hesperetin rescues retinal oxidative stress, neuroinflammation and apoptosis in diabetic rats," *Microvascular Research*, vol. 87, pp. 65– 74, 2013.
- [161] Y. S. Kim, M. Kim, M. Y. Choi et al., "Alpha-lipoic acid reduces retinal cell death in diabetic mice," *Biochemical and Biophysical Research Communications*, vol. 503, no. 3, pp. 1307–1314, 2018.
- [162] E. Kan, Ö. Alici, E. K. Kan, and A. Ayar, "Effects of alphalipoic acid on retinal ganglion cells, retinal thicknesses, and VEGF production in an experimental model of diabetes," *International Ophthalmology*, vol. 37, no. 6, pp. 1269–1278, 2017.
- [163] M. S. Ola, M. M. Ahmed, H. M. Abuohashish, S. S. al-Rejaie, and A. S. Alhomida, "Telmisartan ameliorates neurotrophic support and oxidative stress in the retina of streptozotocininduced diabetic rats," *Neurochemical Research*, vol. 38, no. 8, pp. 1572–1579, 2013.
- [164] L. Y. Dong, J. Jin, G. Lu, and X. L. Kang, "Astaxanthin attenuates the apoptosis of retinal ganglion cells in *db/db* mice by inhibition of oxidative stress," *Marine Drugs*, vol. 11, no. 12, pp. 960–974, 2013.
- [165] F. Desheng and T. Xiaofeng, "Effect of astaxanthin on retinal pigment epithelial cells in high glucose: an *in-vitro* study," *Biomedical Research*, vol. 28, no. 15, pp. 6839–6843, 2017.
- [166] Y. F. Si, J. Wang, J. Guan, L. Zhou, Y. Sheng, and J. Zhao, "Treatment with hydrogen sulfide alleviates streptozotocin-induced diabetic retinopathy in rats," *British Journal of Pharmacology*, vol. 169, no. 3, pp. 619–631, 2013.
- [167] J. Du, H. Jin, and L. Yang, "Role of hydrogen sulfide in retinal diseases," *Frontiers in Pharmacology*, vol. 8, p. 588, 2017.

- [168] C. F. Wang, J. R. Yuan, D. Qin et al., "Protection of tauroursodeoxycholic acid on high glucose-induced human retinal microvascular endothelial cells dysfunction and streptozotocin-induced diabetic retinopathy rats," *Journal* of *Ethnopharmacology*, vol. 185, pp. 162–170, 2016.
- [169] D. Mantopoulos, Y. Murakami, J. Comander et al., "Tauroursodeoxycholic acid (TUDCA) protects photoreceptors from cell death after experimental retinal detachment," *PLoS One*, vol. 6, no. 9, article e24245, 2011.
- [170] S. K. Gupta, B. Kumar, T. C. Nag et al., "Curcumin prevents experimental diabetic retinopathy in rats through its hypoglycemic, antioxidant, and anti-inflammatory mechanisms," *Journal of Ocular Pharmacology and Therapeutics*, vol. 27, no. 2, pp. 123–130, 2011.
- [171] F. Yang, J. Yu, F. Ke et al., "Curcumin alleviates diabetic retinopathy in experimental diabetic rats," *Ophthalmic Research*, vol. 60, no. 1, pp. 43–54, 2018.