



RESEARCH

Open Access

# The conformation change of Bcl-2 is involved in arsenic trioxide-induced apoptosis and inhibition of proliferation in SGC7901 human gastric cancer cells

Yihu Zheng<sup>1,2</sup>, Mengtao Zhou<sup>1</sup>, Aifang Ye<sup>3</sup>, Qiu Li<sup>4</sup>, Yongheng Bai<sup>2</sup> and Qiyu Zhang<sup>\*1,2</sup>

## Abstract

**Background:** Arsenic trioxide has been established as a first-line agent for treating acute promyelocytic leukemia. Experimental data suggest that arsenic trioxide also can have a potential use as chemotherapeutic agent for other malignancies. The precise mechanisms of action of arsenic trioxide have though not been elucidated. As the role of Bcl-2 in arsenic trioxide-mediated cell apoptosis and conformation change of Bcl-2 in response to arsenic trioxide treatment has not been studied. The aim of the present study was to determine whether conformation change of Bcl-2 is involved in the action of arsenic trioxide.

**Methods:** Human gastric cancer SGC7901 cells were exposed to different concentrations of arsenic trioxide. Proliferation was measured by using the Kit-8 cell counting assay. Analysis of nuclear morphology was observed by DAPI staining. The apoptosis rates of cells treated with arsenic trioxide were analyzed by flow cytometry using Annexin V-FITC staining. The conformation change of Bcl-2 and Bax activation were detected by immunostaining and Western blot analysis. Total expression of Bcl-2 and Bax were examined by Western blot analysis.

**Results:** Arsenic trioxide inhibited the growth of human gastric cancer SGC7901 cells and induced apoptosis. There were two Bcl-2 phenotypes coexisting in SGC7901 cells and the Bcl-2 cytoprotective phenotype could change into a cytotoxic phenotype following conformational change of Bcl-2, triggered by arsenic trioxide exposure. Bax activation might also be involved in arsenic trioxide-induced Bcl-2 conformational change. Arsenic trioxide did not change levels of total Bcl-2 expression, but up-regulated total Bax expression for the treatment time ranging from 3 to 24 hours.

**Conclusion:** Arsenic trioxide induces apoptosis through induction of Bcl-2 conformational change, Bax activation and up-regulation of total Bax expression rather than affecting total Bcl-2 expression in human gastric cancer SGC7901 cells. The conformational change of Bcl-2 may be a novel described mechanism of arsenic trioxide-induced apoptosis in cancer cells.

## Background

Arsenic trioxide, one member of the three inorganic forms of arsenic, is formed by heating realgar, which is formed as an arsenic complex with sulfur. Although arsenic trioxide is highly toxic, it has been shown to have a therapeutic potential. It has for long been used as a drug in traditional Chinese medicine to treat a variety of dis-

eases, including malaria, psoriasis, syphilis, rheumatosis and cancer [1-3]. Contemporary studies show that arsenic trioxide is an effective therapeutic agent for the treatment of various hematological malignancies and especially acute promyelocytic leukemia [4-7]. More recent experimental data have demonstrated that arsenic trioxide may have effects in the treatment of several other malignancies in the experimental setting, including gastric cancer, lung cancer, breast cancer, hepatocellular carcinoma, gallbladder carcinoma, and neuroblastoma [8-

\* Correspondence: qiyuz@126.com

<sup>1</sup> Department of Surgery, The First Affiliated Hospital of Wenzhou Medical College, Wenzhou 325000, China

Full list of author information is available at the end of the article



13]. However, arsenic trioxide exerts its effect through different cellular and physiological pathways. The mechanisms of action of arsenic trioxide related to the induction of apoptosis in cancer cells remain controversial. Arsenic trioxide affects the activities of Akt, JNK kinases, NF- $\kappa$ B, glutathione, calcium signaling, ROS, Caspases, as well as pro- and anti-apoptotic proteins [14-17]. Down-regulation of Bcl-2, an "anti-apoptotic" protein, has also been considered as one of its significant mechanism of action [12,18-20].

Bcl-2 is considered as an important anti-apoptotic member of the Bcl-2 family, its expression manifests either cytoprotective or cytotoxic phenotypes, depending on the cellular context [21]. The anti-apoptotic Bcl-2 family members Bcl-2 and Bcl-XL have hydrophobic properties on their surfaces, essential for their anti-apoptotic effect, whereas their BH3 domains are buried. In contrast, pro-apoptotic Bcl-2 family members have an exposed BH3 domain, which binds to the hydrophobic pockets of anti-apoptotic Bcl-2 members to inhibit their survival effect [22]. Subsequent research showed that the dual phenotypes of Bcl-2 are controlled by its protein conformation [23]. When the loop of Bcl-2 interacts with an external factor, the hydrophobic binding groove of Bcl-2 undergoes a large-scale realignment, resulting in exposure of its BH3 domain [23,24]. This conformational change is responsible for the conversion of Bcl-2 from a cytoprotective to a cytotoxic molecule.

The present study aimed at determining whether arsenic trioxide inhibits proliferation and induces apoptosis in SGC7901 human gastric cancer cells, accompanied by conformational changes of Bcl-2 and changes in total Bcl-2 levels.

## Methods

### Materials

Arsenic trioxide was purchased from Sigma. A 5 mM stock solution of arsenic trioxide was obtained by dissolving arsenic trioxide in 1.65 M NaOH and by diluting in PBS, followed by adjustment of the pH to 7.0. RPMI Medium1640 and FBS were purchased from Invitrogen. Anti-Bcl-2 antibody (sc-492), anti-Bcl-2 antibody (sc-7382), anti-Bax antibody (sc-7480), anti-Bax (6A7) antibody (sc-23959) and anti- $\beta$ -actin antibody (sc-47778) were from Santa Cruz Biotechnology. Anti-Bcl-2 BH3 (AP1303a) was from Abgent. Goat anti-mouse and rabbit secondary antibody conjugated to horseradish peroxidase (A0216, A0208), Cy3-labeled anti-rabbit IgG (A0516), FITC-labeled anti-mouse IgG (A0568) and DAPI were purchased from Beyotime Institute of Biotechnology.

### Cell culture and treatment

SGC7901 human gastric cancer cells were purchased from Shanghai Institutes for Biological Sciences and cul-

tured in RPMI Medium1640 containing 10% FBS in a humidified atmosphere containing 5% CO<sub>2</sub> at 37°C. Cells were split every 2-3 days by trypsinization and centrifugation, followed by aspiration of the culture medium. Before arsenic trioxide exposure, cell density was adjusted to  $1.5 \times 10^4$  cells per square centimeter.

### Proliferation Analysis

Proliferation was measured by using the Cell Counting Assay Kit-8 (Dojindo Molecular Technologies) according to the manufacturer's protocol. One hundred microliters of SGC7901 human gastric cancer cells were plated on 96-well plates at a density of  $1.5 \times 10^4$  cells per square centimeter and cultured for 24 hours. Cells were starved for 24 hours by replacing the media with serum-free media containing 0.1% BSA, followed by exposure to different concentrations of arsenic trioxide (0  $\mu$ mol/L, 5  $\mu$ mol/L, 10  $\mu$ mol/L, 15  $\mu$ mol/L and 20  $\mu$ mol/L) for 24 and 48 hours. Ten microliters of Cell Counting Assay Kit-8 solution was added to each well, the cells were incubated for another 2 hours, and the absorbance at 450 nm was measured by using a microplate reader (BioTek Instruments). The amount of the formazan dye, generated by the activities of dehydrogenases in cells, is directly proportional to the number of living cells. Inhibitory rate of cellular growth was calculated as the following formula: Inhibitory rate (%) =  $(1 - A \text{ value in experimental group} / A \text{ value in control group}) \times 100\%$ . The 0  $\mu$ mol/L group was used as black control group. The IC<sub>50</sub> value (the concentration of the drug which is capable of bringing about 50% inhibition of cell survival) of the drug used for treatment was determined by plotting a graph with inhibitory rate of cell growth (Y-axis) against the concentrations of the arsenic trioxide (X-axis).

### Analysis of nuclear morphology by DAPI staining

Cells grew in 6 well plates at a seeding density of  $1.5 \times 10^4$  cells per square centimeter and were then treated with 10  $\mu$ mol/L arsenic trioxide in complete media for 24 hours. Cells were fixed with 4% paraformaldehyde prior to washing with PBS. Washed cells were then stained with 1  $\mu$ g/ml DAPI for 15 min in the dark. Slides were viewed with a fluorescent microscope at 340-380 nm and  $\times 1000$  magnification (Carl Zeiss). Cells were evaluated as normal or apoptotic depending on morphological characteristics. Normal nuclei (smooth nuclear) and apoptotic nuclei (condensed or fragmented chromatin) were easily distinguished. Thus, analysis of nuclear morphology was observed in three independent experiments.

### Apoptosis Analysis

Cells treated with different concentrations of arsenic trioxide (0  $\mu$ mol/L, 5  $\mu$ mol/L, 10  $\mu$ mol/L, 15  $\mu$ mol/L, 20  $\mu$ mol/L, 25  $\mu$ mol/L and 30  $\mu$ mol/L) in serum-free

medium for 24 hours were collected and stained with Annexin V/propidium iodide (PI) using Vybrant apoptosis assay kit No. 2 (Molecular Probes) and analyzed by flow cytometry. The 0  $\mu\text{mol/L}$  group was used as black control group.

#### Immunofluorescence microscopy

Cells treated with 15  $\mu\text{mol/L}$  arsenic trioxide for 12 hours were cultured in serum-free medium overnight on glass coverslips. The cells were fixed in 4% paraformaldehyde in PBS for 15 min and washed twice with PBS. The cells were then permeabilized with 1% Triton X-100 in PBS for 5 min. Fixed cells were preincubated for 45 min in PBS containing 5% bovine serum albumin at room temperature, followed by incubation with various primary antibodies at 4°C overnight and detected by Cy3-labeled anti-rabbit IgG (1:300) or FITC-labeled anti-mouse IgG (1:300) at room temperature for 2 hours. Cells were stained with 1  $\mu\text{g/ml}$  DAPI to visualize the nuclei. The images were taken under a fluorescent microscope. The primary antibodies included anti-Bcl-2 antibody (sc-492) (Santa Cruz, 1:200), anti-Bcl-2 BH3 antibody (Abgent, 1:200) and anti-Bax(6A7) antibody (sc-23959) (Santa Cruz, 1:200).

#### Protein extraction and western blot analysis

Cells were treated with 15  $\mu\text{mol/L}$  arsenic trioxide for different time. Both adherent and floating cells were harvested and lysed with Mammalian Protein Extraction Reagent (Pierce) according to the manufacturer's protocol. Equal amounts of protein were separated by SDS-PAGE or Native-PAGE, and then transferred onto a PVDF membrane (Millipore). The membrane was blocked for 1 hour in a non-fat dried milk solution containing 1% Tween-20. The membrane was then incubated with various primary antibodies overnight at 4°C, followed by incubation with anti-mouse (1:5,000) secondary antibodies for 1 hour. Finally, protein bands were detected by using the Chemiluminescent Substrate (HRP) Kit from Pierce. The dilutions of the primary antibodies were anti-Bcl-2 antibody (sc-7382) in 1:800, anti-Bax antibody (sc-7480) in 1:800, anti-Bcl-2 BH3 antibody in 1:500, anti-Bax (6A7) antibody in 1:500. The blots were reprobed with anti- $\beta$ -actin antibody for loading control.

#### Statistical analysis

The results of each series of experiments (performed in triplicates) were expressed as the mean values  $\pm$  standard deviation of the mean (SD). Statistical significance of differences between groups was analyzed by using ANOVA analysis.  $P < 0.05$  was considered statistically significant.

## Results

#### Proliferation Analysis

SGC7901 cells were treated with different concentrations of arsenic trioxide (5  $\mu\text{mol/L}$ , 10  $\mu\text{mol/L}$ , 15  $\mu\text{mol/L}$  and 20  $\mu\text{mol/L}$ ) at 24 and 48 hours. The inhibitory rates of cell growth were  $16.50 \pm 0.55\%$ ,  $50.83 \pm 0.75\%$ ,  $65.50 \pm 1.05\%$ ,  $73.50 \pm 1.05\%$ ;  $41.83 \pm 0.75\%$ ,  $61.67 \pm 0.82\%$ ,  $71.17 \pm 0.75\%$ ,  $76.67 \pm 0.82\%$ , respectively. By using curve fitting, the IC50 was about 10  $\mu\text{mol/L}$  for 24 hours. Arsenic trioxide obviously could inhibit the proliferation of SGC7901 cells in concentration and time-dependent manner (Fig. 1A).

#### Morphologic characteristic of apoptosis

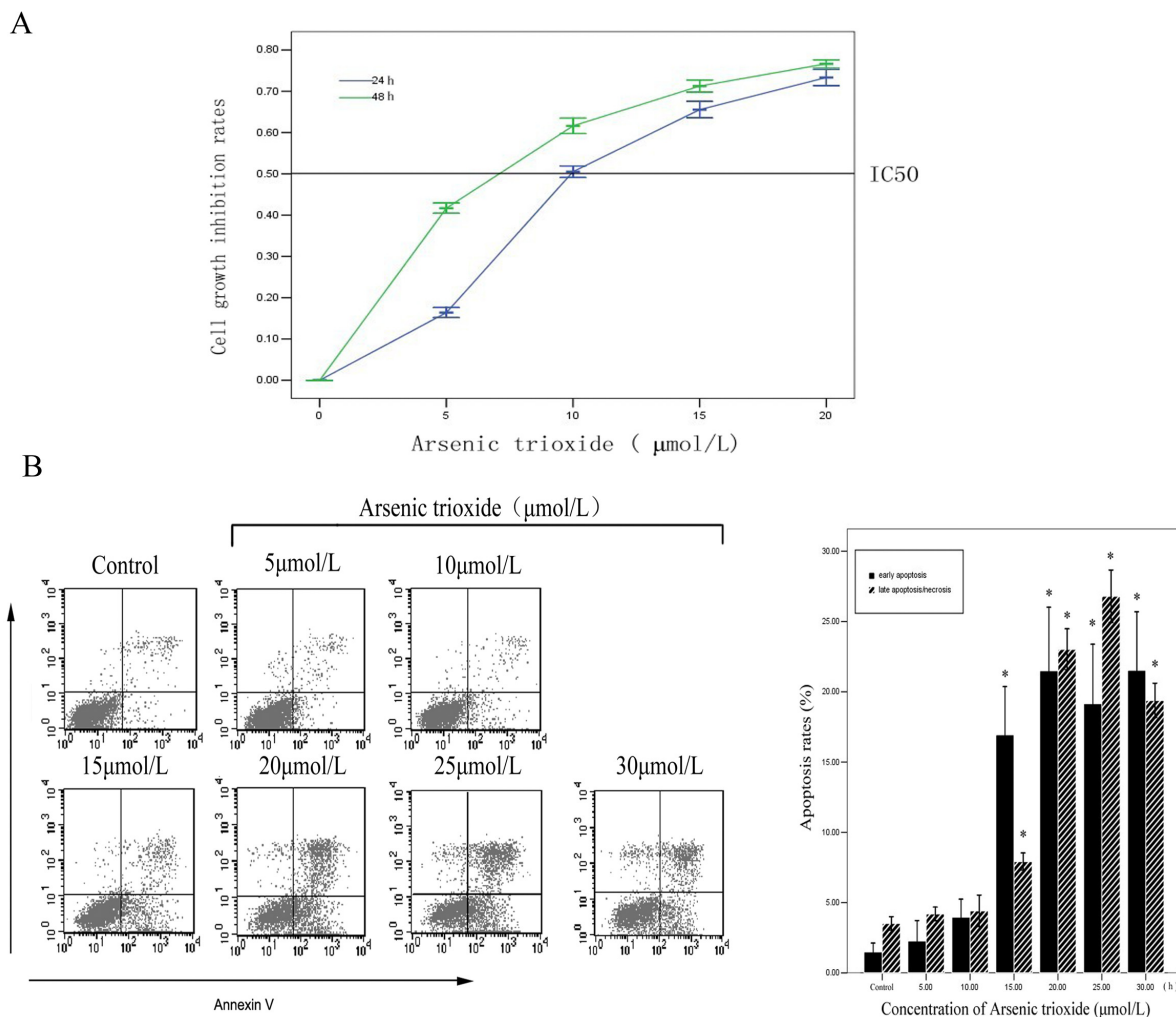
Nuclear morphology analysis showed characteristic apoptotic changes, such as convoluted nuclei with cavitations, clumps of chromatin abutting to inner regions of the nuclear envelope between the nuclear pores, breakdown of nuclear envelope, chromatin condensation and dissociation of DNA fragments in SGC7901 cells after treatment with arsenic trioxide for 24 hours (Fig. 2A).

#### Apoptosis Analysis

SGC7901 cells were treated with different concentrations of arsenic trioxide (0  $\mu\text{mol/L}$ , 5  $\mu\text{mol/L}$ , 10  $\mu\text{mol/L}$ , 15  $\mu\text{mol/L}$ , 20  $\mu\text{mol/L}$ , 25  $\mu\text{mol/L}$  and 30  $\mu\text{mol/L}$ ) for 24 hours. The early and late apoptosis/necrosis rates were  $11.49 \pm 0.63\%$ ,  $2.28 \pm 1.46\%$ ,  $3.97 \pm 1.28\%$ ,  $16.94 \pm 3.42\%$ ,  $21.50 \pm 4.51\%$ ,  $19.16 \pm 4.21\%$ ,  $21.53 \pm 4.16\%$ ;  $3.52 \pm 0.49\%$ ,  $4.21 \pm 0.48\%$ ,  $4.42 \pm 1.12\%$ ,  $7.92 \pm 0.61\%$ ,  $23.02 \pm 1.46\%$ ,  $26.80 \pm 1.86\%$ ,  $19.39 \pm 1.23\%$ , respectively, which suggested that arsenic trioxide induce apoptosis (Fig. 1B).

#### Arsenic trioxide induced Bcl-2 conformational change and Bax activation

SGC7901 cells were strongly stained by anti-Bcl-2 N terminus antibody. It showed that SGC7901 cells highly expressed total Bcl-2 protein (Fig. 2B). Moreover, an enhanced immunostaining by anti-Bcl-2 BH3 antibody, as compared to the "black control group" (0 hour), was observed in SGC7901 cells treated with 15  $\mu\text{mol/L}$  arsenic trioxide for 12 hours using immunofluorescence. The black control group (0 hour) did not immunostain by the anti-Bax (6A7) antibody, suggesting that Bax was inactive in the cells. However, SGC7901 cells treated with 15  $\mu\text{mol/L}$  arsenic trioxide displayed strong immunostaining with the anti-Bax (6A7) antibody, demonstrating that arsenic trioxide could activate Bax (Fig. 2C). After treatment of 15  $\mu\text{mol/L}$  arsenic trioxide for the indicated times (0 hour, 3 hours, 6 hours, 12 hours and 24 hours), Western blot showed that the expression of BH3 domain exposed Bcl-2 had an upward trend and reached a peak at 12 hours and the difference as compared with 0 hour was statistically significant ( $P < 0.05$ ). By time, the activated Bax also presented an upward trend and reached a peak



**Figure 1 A. Inhibitory effects of arsenic trioxide on SGC7901 cell. B** The effects of arsenic trioxide on early and late apoptosis/necrosis of SGC7901 cell. (\* represents  $p < 0.05$  compared to the black control group and arsenic trioxide treated groups).

at 24 hours and the difference compared to 0 hour was statistically significant ( $P < 0.05$ ) (Fig. 3A). Arsenic trioxide-treated SGC7901 cells, detected by western blot and stained by both anti-Bcl-2 (BH3) and anti-Bax (6A7) antibodies, express conformational change of Bcl-2, which may play a role in arsenic trioxide-induced apoptosis and Bax activation.

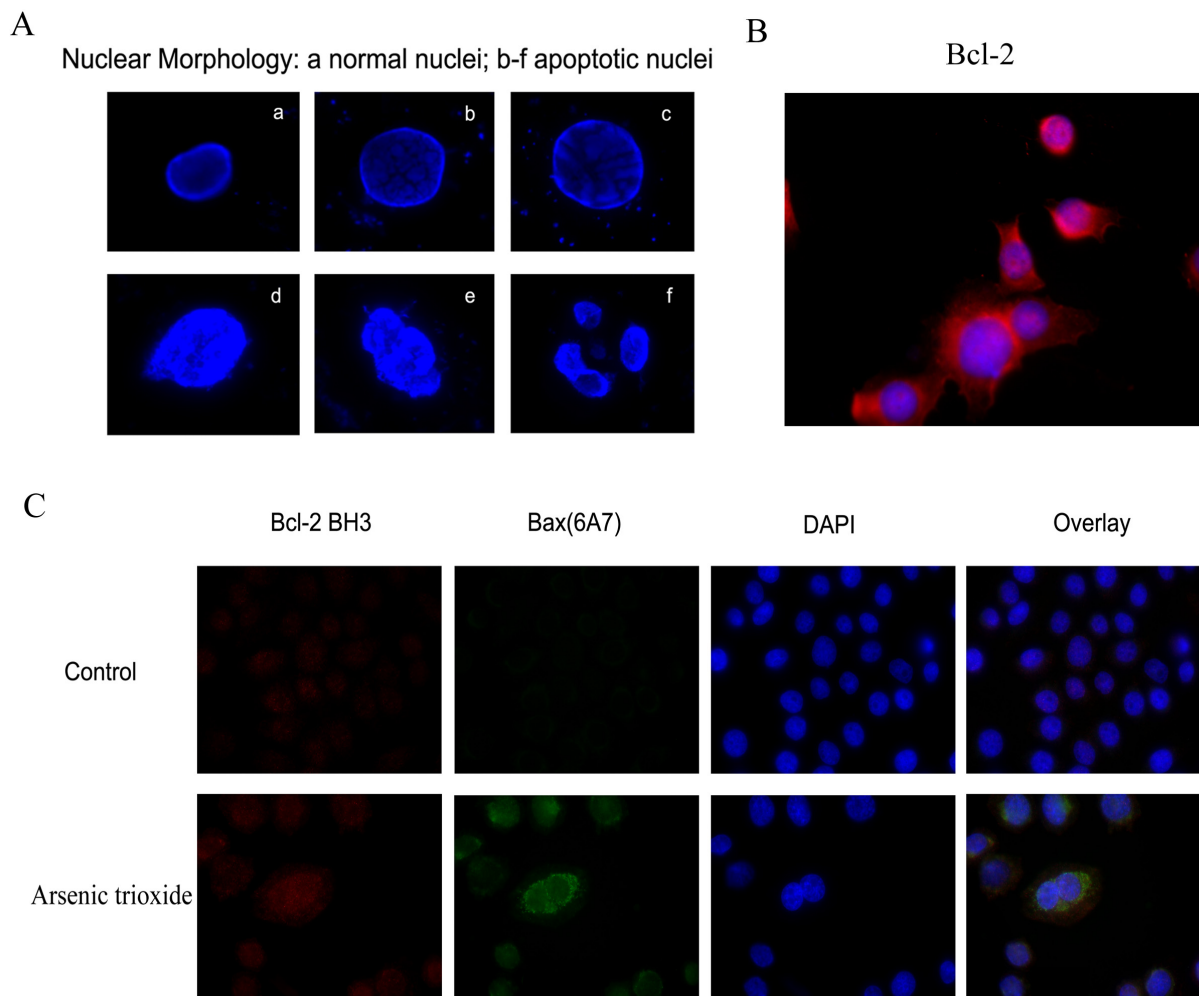
**Arsenic trioxide did not affect total Bcl-2 expression, but up-regulated total Bax expression**

After 15 μmol/L arsenic trioxide exposure for various times (0 hour, 3 hours, 6 hours, 12 hours and 24 hours), the change in total Bcl-2 expression was unobvious and the differences compared to the different groups did not statistically differ ( $P > 0.05$ ). Total Bax had a higher expression and reached a peak at 3 hours and the differences compared to 0 hour was statistically significant ( $P <$

0.05), though the levels descended at 3 hours (Fig. 3B). The results showed that arsenic trioxide did not cause any apparent change in levels of Bcl-2, but Bax expression was up-regulated for treatment times ranging from 3 to 24 hours.

**Discussion**

Arsenic is a well-known environmental toxic and carcinogenic substance, and an effective chemotherapeutic drug. Due to the dual capability of arsenic, the agent carries significant risks for medical applications. The underlying mechanisms are, however, not fully understood. Arsenic exerts its effect by inhibiting the activities of several enzymes, especially those involved in cellular signaling pathways and DNA synthesis and repair. During the past centuries, a number of arsenic compounds have

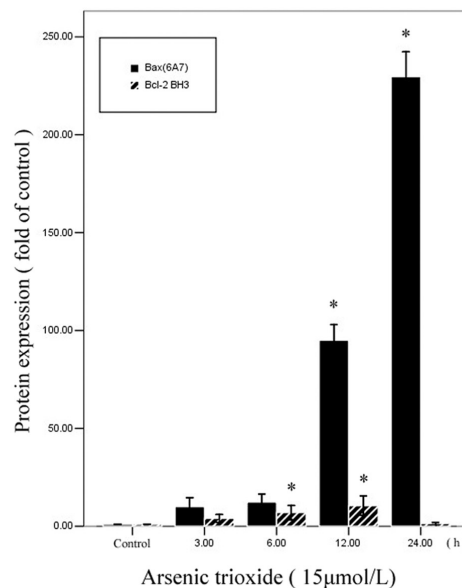
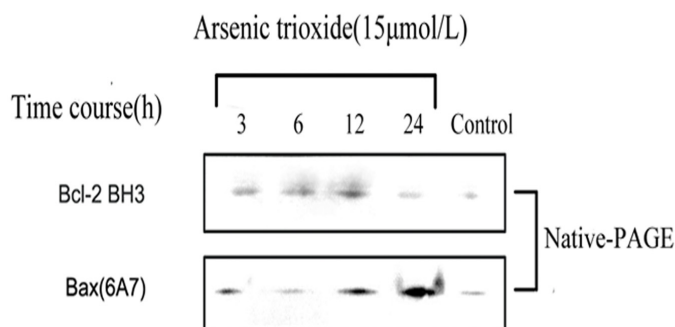


**Figure 2 A. Nuclear morphologic changes showing features of apoptosis in SGC7901 cells after treatment with arsenic trioxide for 24 hours.** (a) Untreated SGC7901 cells; (b-f) 15  $\mu\text{mol/L}$  arsenic trioxide-treated SGC7901 cells. The cells were stained using DAPI staining. **B.** Untreated SGC7901 cells stained by anti-Bcl-2 N terminus antibody. **C** SGC7901 cell stained by anti-Bcl-2 BH3 and anti-Bax(6A7) antibody before and after exposure to 15  $\mu\text{mol/L}$  arsenic trioxide for 12 hours.

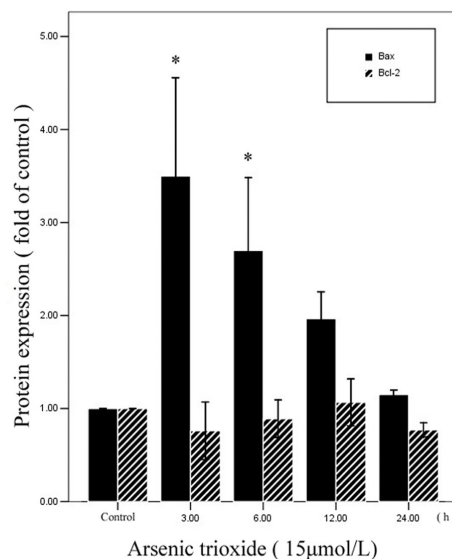
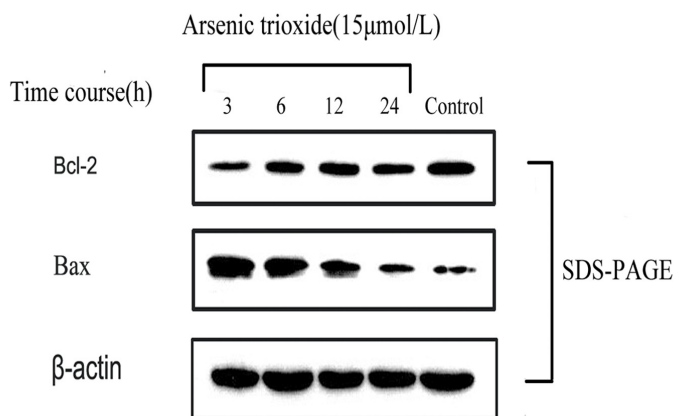
been used as medicines. Arsenic trioxide, one form of arsenicals, has been used in a variety of ways over the past hundred years, but most commonly in the treatment of malignancies. Owing to the impressive effects of arsenic trioxide in hematological cancers and solid tumor cells in vitro, the mechanisms of arsenic trioxide-mediated cell death have recently come under increasing scrutiny. Arsenic trioxide may be a promising candidate for the treatment of other malignancies. The combination therapy of arsenic trioxide and other chemotherapeutic agents have been applied experimentally for treatment of refractory malignant tumors. In the current study, we observed that arsenic trioxide had a strong anti-proliferative effect, most likely by

induction of apoptosis, on human gastric cancer SGC7901 cells in a dose and time dependent manner. As has previously been reported, the cellular and biochemical effects of arsenic were performed using concentrations greater than 5  $\mu\text{mol/L}$ , often 50  $\mu\text{mol/L}$ , and the 50% inhibitory concentration (IC<sub>50</sub>) of arsenic trioxide on proliferation of SGC7901 cells was about 10  $\mu\text{mol/L}$  for 24 hours. Maybe it was much too high than relevant to therapeutic levels (1 to 2  $\mu\text{mol/L}$ ) [25,26]. However, from the 24 and 48 hours curve fitting, we could suppose that the 50% inhibitory concentration (IC<sub>50</sub>) for 72 hours may be similar to clinically therapeutic levels, which also has been described by others [8,9,27]. This suggests that

A



B



**Figure 3 A.** The expression of BH3 exposed Bcl-2 and activated Bax after exposed to 15 μmol/L arsenic trioxide in SGC7901 cell. **B** The expression of total Bcl-2 and total Bax after exposure to 15 μmol/L arsenic trioxide in SGC7901 cell. (\* represents  $p < 0.05$  between black control group and arsenic trioxide treated groups).

SGC7901 human gastric cancer cells are sensitive to arsenic trioxide.

The mechanisms of arsenic trioxide-induced anti-proliferation have been extensively investigated. Apoptosis appears to be the main phenomenon resulting in significant cell death and cell growth inhibition. Arsenic trioxide is known to modulate multiple signal transduction pathways, including inhibition of telomerase activity, induction of reactive oxygen species release, and inhibition of survival pathways involving extracellular signal

regulated kinase, Akt, calcium signaling and NF-κB activities [14-17,28]. Interestingly, the apoptotic effect of arsenic trioxide largely depends on a Bcl-2-controlled pathway [12,18-20].

Bcl-2, an anti-apoptotic Bcl-2 family member, for which an increased expression has been associated with a more aggressive malignant phenotype and drug resistance to various categories of chemotherapeutic drugs in malignancies. Small molecule inhibitors of the Bcl-2 family proteins, designed to bind the hydrophobic groove of

anti-apoptotic Bcl-2 proteins in place of BH3-only proteins, are potential agents to treat cancers. They can oligomerize Bax or Bak, which subsequently depolarize in the mitochondrial membrane potential to release cytochrome c and induce apoptosis [29]. Agents targeting anti-apoptotic Bcl-2 family members have preclinical activity as single agents and also affect combination with other anti-neoplastic agents. Recent researches have demonstrated that Bcl-2 could manifest opposing phenotypes, induced by interactions with proteins, such as Nur77, suggesting novel strategies for regulating apoptosis in cancers and other diseases [30]. This phenotype change of Bcl-2 is controlled by its protein conformational change. When the loop of Bcl-2 interacts with an external factor, the hydrophobic binding groove of Bcl-2 undergoes a large-scale realignment, resulting in exposure of its BH3 domain [21,22]. It was also reported that paclitaxel could directly target Bcl-2 in the loop domain, mimics activity of Nur77, thereby facilitating the initiation of apoptosis [31].

Whether Bcl-2 phenotype changes phenomenon occur in arsenic trioxide-induced cell apoptosis is still unknown. In the present study, we used anti-Bcl-2 BH3 antibody to detect the conformational change of Bcl-2. When Bcl-2 undergoes conformational change, the hydrophobic binding groove of Bcl-2 gives rise to a large-scale realignment, resulting in exposure of its cryptic BH3 domain and can be recognized by Bcl-2 BH3 antibody. We used Bax (6A7) antibody to detect the activated Bax, Bax undergoes a conformational change and oligomerization during early apoptosis, which can be followed by exposure of cryptic antibody epitopes (the N-terminal residues 1-21). This type of Bax can be recognized by anti-Bax (6A7) antibody. The novel finding from this work was that SGC7901 cells highly expressed Bcl-2, but they were weakly stained by the anti-Bcl-2 BH3 antibody, suggesting that there were two Bcl-2 phenotypes coexisting in SGC7901 cells and mostly Bcl-2 was anti-apoptotic. The results showed that Bcl-2 anti-apoptotic phenotype could change into a pro-apoptotic phenotype following exposure to arsenic trioxide. Also Bax activation was involved in arsenic trioxide-induced conformational change of Bcl-2 by immunostaining SGC7901 cells with anti-Bax (6A7) antibody that recognizes activated Bax. Arsenic trioxide caused no apparent change in the levels of Bcl-2, but up-regulated Bax for treatment times ranging from 3 to 24 hours. Thus, Bcl-2 conformational change, Bax activation and up-regulation of total Bax expression involved arsenic trioxide-induced apoptosis rather than affecting total Bcl-2 expression in human gastric cancer SGC7901 cells.

Although the anti-apoptotic effect of Bcl-2 is well established, the role of Bcl-2 in cancer response to therapy and drug resistance has not been completely explored. The

mechanism how it promotes cell death has recently gained increasing interest. In general, over-expression and up-regulation of Bcl-2 has been associated with resistance to chemotherapy in various human cancers [29,32], and many studies have shown that over-expression of Bcl-2 is a poor prognostic factor in various cancers. It was found that Bcl-2 expression tended to be associated with a worsened survival in olfactory neuroblastoma (ONB) [33]. Also the expression of Bcl-2 and Bax proteins, evaluated by immunohistochemical staining in specimens from 110 patients with oral squamous cell carcinoma (OSCC) showed that the 5-year survival rate was significantly higher in patients with a ratio of Bcl-2/Bax  $\leq 1$  as compared to those with Bcl-2/Bax  $> 1$  [34]. On the opposite side, high Bcl-2 expression also correlated with favorable parameters and a better prognosis in other cancers. A recent systematic review of the literature showed that over-expression of Bcl-2 was a good prognostic factor for survival in patients with non-small cell lung cancer [35]. Bcl-2 expression also correlated with a favorable prognosis in colorectal cancer [36], and with improved overall survival rate in oral squamous cell carcinoma [37]. Our finding of conformational change of Bcl-2 in SGC7901 cells following exposure to arsenic trioxide is important for founding an explanation accounting for the opposing biological activities of Bcl-2. This may also represent that arsenic trioxide may be a promising candidate for the future treatment of malignancies that over-express endogenous Bcl-2, though substantial experimental and clinical research remains to validate its potential value.

## Conclusion

Our results show that arsenic trioxide is an effective anti-cancer agent with potential for human gastric cancer. Arsenic trioxide can reduce proliferation and induce apoptosis in SGC7901 human gastric cancer cells. There are two Bcl-2 phenotypes coexisting in SGC7901 cells and the Bcl-2 cytoprotective phenotype can change into a cytotoxic phenotype following arsenic trioxide exposure. Also Bax activation is involved in arsenic trioxide-induced conformational change of Bcl-2 in SGC7901 cells. The conformational change of Bcl-2 may be the new mechanism explaining arsenic trioxide-induced apoptosis, other than the ones affecting the total Bcl-2 expression in some cancer cells.

## Abbreviations

As<sub>2</sub>O<sub>3</sub>: arsenic trioxide; APL: acute promyelocytic leukemia; JNK: c-jun terminal kinase; NF- $\kappa$ B: nuclear factor  $\kappa$ B; ROS: reactive oxygen species; PBS: phosphate buffered saline; FBS: fetal bovine serum; HSP: heat shock proteins; PVDF: polyvinylidene fluoride; SDS-PAGE: sodium dodecyl sulfate-polyacrylamide gel electrophoresis.

## Competing interests

The authors declare that they have no competing interests.

#### Authors' contributions

YZ and AY are researchers working in cancer biology and carried the study. QL and YB undertook the Statistical analysis. QZ along with MZ designed the work and interpreted the results. QZ and MZ contributed to the writing of the manuscript. All the authors read and approved the final manuscript.

#### Acknowledgements

This study was sponsored by Zhejiang Provincial Top Key Discipline in Surgery and Wenzhou Key Laboratory Project in Surgery.

#### Author Details

<sup>1</sup>Department of Surgery, The First Affiliated Hospital of Wenzhou Medical College, Wenzhou 325000, China, <sup>2</sup>Key Laboratory of Surgery, The First Affiliated Hospital of Wenzhou Medical College, Wenzhou 325000, China, <sup>3</sup>Department of Laboratory, The First Affiliated Hospital of Wenzhou Medical College, Wenzhou 325000, China and <sup>4</sup>Department of Internal Medicine, The First Affiliated Hospital of Wenzhou Medical College, Wenzhou 325000, China

Received: 5 December 2009 Accepted: 20 April 2010

Published: 20 April 2010

#### References

1. Miller WH Jr, Schipper HM, Lee JS, Singer J, Waxman S: **Mechanisms of action of arsenic trioxide.** *Cancer Res* 2002, **62**(14):3893-3903.
2. Evens AM, Tallman MS, Gartenhaus RB: **The potential of arsenic trioxide in the treatment of malignant disease: past, present, and future.** *Leuk Res* 2004, **28**(9):891-900.
3. Gazitt Y, Akay C: **Arsenic trioxide: an anti cancer missile with multiple warheads.** *Hematology* 2005, **10**(3):205-213.
4. Soignet SL, Frankel SR, Douer D, Tallman MS, Kantarjian H, Calleja E, Stone RM, Kalaycio M, Scheinberg DA, Steinherz P, Sievers EL, Coutre S, Dahlberg S, Ellison R, Warrell RP Jr: **United States multicenter study of arsenic trioxide in relapsed acute promyelocytic leukemia.** *J Clin Oncol* 2001, **19**(18):3852-3860.
5. Murgu AJ, McBee WL, Cheson BD: **Clinical trials referral resource. Clinical trials with arsenic trioxide.** *Oncology (Williston Park)* 2000, **14**(2): 206, 211, 215-206 passim.
6. Hayakawa F, Privalsky ML: **Phosphorylation of PML by mitogen-activated protein kinases plays a key role in arsenic trioxide-mediated apoptosis.** *Cancer Cell* 2004, **5**(4):389-401.
7. Coe E, Schimmer AD: **Catalase activity and arsenic sensitivity in acute leukemia.** *Leuk Lymphoma* 2008, **49**(10):1976-1981.
8. Shao QS, Ye ZY, Ling ZQ, Ke JJ: **Cell cycle arrest and apoptotic cell death in cultured human gastric carcinoma cells mediated by arsenic trioxide.** *World J Gastroenterol* 2005, **11**(22):3451-3456.
9. Han B, Zhou G, Zhang Q, Zhang J, Wang X, Tang W, Kakudo K: **Effect of arsenic trioxide (ATO) on human lung carcinoma PG cell line: ATO induced apoptosis of PG cells and decreased expression of Bcl-2, Pgp.** *J Exp Ther Oncol* 2004, **4**(4):335-342.
10. Cheng Y, Chang LW, Tsou TC: **Mitogen-activated protein kinases mediate arsenic-induced down-regulation of survivin in human lung adenocarcinoma cells.** *Arch Toxicol* 2006, **80**(6):310-318.
11. Luo L, Qiao H, Meng F, Dong X, Zhou B, Jiang H, Kanwar JR, Krissansen GW, Sun X: **Arsenic trioxide synergizes with B7H3-mediated immunotherapy to eradicate hepatocellular carcinomas.** *Int J Cancer* 2006, **118**(7):1823-1830.
12. Ai Z, Lu W, Qin X: **Arsenic trioxide induces gallbladder carcinoma cell apoptosis via downregulation of Bcl-2.** *Biochem Biophys Res Commun* 2006, **348**(3):1075-1081.
13. Cheung WM, Chu PW, Kwong YL: **Effects of arsenic trioxide on the cellular proliferation, apoptosis and differentiation of human neuroblastoma cells.** *Cancer Lett* 2007, **246**(1-2):122-128.
14. Izdebska M, Grzanka A, Szczepanski MA, Litwiniec A: **Selected mechanisms of the therapeutic effect of arsenic trioxide in cancer treatment.** *Postepy Hig Med Dosw (Online)* 2008, **62**:463-467.
15. Gao F, Yi J, Shi GY, Li H, Shi XG, Tang XM: **The sensitivity of digestive tract tumor cells to As<sub>2</sub>O<sub>3</sub> is associated with the inherent cellular level of reactive oxygen species.** *World J Gastroenterol* 2002, **8**(1):36-39.
16. Bowling BD, Doudican N, Manga P, Orlow SJ: **Inhibition of mitochondrial protein translation sensitizes melanoma cells to arsenic trioxide cytotoxicity via a reactive oxygen species dependent mechanism.** *Cancer Chemother Pharmacol* 2008, **63**(1):37-43.
17. Florea AM, Busselberg D: **Anti-cancer drugs interfere with intracellular calcium signaling.** *Neurotoxicology* 2009, **30**(5):803-810.
18. Deng Y, Lin C, Zheng J, Liang X, Chen J, Fu M, Xiao P, Wu M: **Mechanisms of arsenic trioxide induced apoptosis of human cervical cancer HeLa cells and protection by Bcl-2.** *Sci China C Life Sci* 1999, **42**(6):635-643.
19. Scholz C, Richter A, Lehmann M, Schulze-Osthoff K, Dorken B, Daniel PT: **Arsenic trioxide induces regulated, death receptor-independent cell death through a Bcl-2-controlled pathway.** *Oncogene* 2005, **24**(47):7031-7042.
20. Bairey O, Vanichkin A, Shpilberg O: **Arsenic-trioxide-induced apoptosis of chronic lymphocytic leukemia cells.** *Int J Lab Hematol* 2010, **32**(1 Pt 1):e77-85.
21. Igaki T, Kanuka H, Inohara N, Sawamoto K, Nunez G, Okano H, Miura M: **Drob-1, a Drosophila member of the Bcl-2/CED-9 family that promotes cell death.** *Proc Natl Acad Sci USA* 2000, **97**(2):662-667.
22. Cory S, Huang DC, Adams JM: **The Bcl-2 family: roles in cell survival and oncogenesis.** *Oncogene* 2003, **22**(53):8590-8607.
23. Lin B, Kolluri SK, Lin F, Liu W, Han YH, Cao X, Dawson MI, Reed JC, Zhang XK: **Conversion of Bcl-2 from protector to killer by interaction with nuclear orphan receptor Nur77/TR3.** *Cell* 2004, **116**(4):527-540.
24. Zhang XK: **Targeting Nur77 translocation.** *Expert Opin Ther Targets* 2007, **11**(1):69-79.
25. Dai J, Weinberg RS, Waxman S, Jing Y: **Malignant cells can be sensitized to undergo growth inhibition and apoptosis by arsenic trioxide through modulation of the glutathione redox system.** *Blood* 1999, **93**(1):268-277.
26. Wang TS, Kuo CF, Jan KY, Huang H: **Arsenite induces apoptosis in Chinese hamster ovary cells by generation of reactive oxygen species.** *J Cell Physiol* 1996, **169**(2):256-268.
27. Bornstein J, Sagi S, Haj A, Harroch J, Fares F: **Arsenic Trioxide inhibits the growth of human ovarian carcinoma cell line.** *Gynecol Oncol* 2005, **99**(3):726-729.
28. Zhou C, Boggess JF, Bae-Jump V, Gehrig PA: **Induction of apoptosis and inhibition of telomerase activity by arsenic trioxide (As<sub>2</sub>O<sub>3</sub>) in endometrial carcinoma cells.** *Gynecol Oncol* 2007, **105**(1):218-222.
29. Kang MH, Reynolds CP: **Bcl-2 inhibitors: targeting mitochondrial apoptotic pathways in cancer therapy.** *Clin Cancer Res* 2009, **15**(4):1126-1132.
30. Lin B, Kolluri SK, Lin F, Liu W, Han YH, Cao X, Dawson MI, Reed JC, Zhang XK: **Conversion of Bcl-2 from protector to killer by interaction with nuclear orphan receptor Nur77/TR3.** *Cell* 2004, **116**(4):527-540.
31. Ferlini C, Cicchillitti L, Raspaglio G, Bartollino S, Cimitan S, Bertucci C, Mozzetti S, Gallo D, Persico M, Fattorusso C, Campiani G, Scambia G: **Paclitaxel directly binds to Bcl-2 and functionally mimics activity of Nur77.** *Cancer Res* 2009, **69**(17):6906-6914.
32. Colie S, Van Veldhoven PP, Kedjouar B, Bedia C, Albinet V, Sorli SC, Garcia V, Djavaheri-Mergny M, Bauvy C, Codogno P, Levade T, Andrieu-Abadie N: **Disruption of sphingosine 1-phosphate lyase confers resistance to chemotherapy and promotes oncogenesis through Bcl-2/Bcl-xL upregulation.** *Cancer Res* 2009, **69**(24):9346-9353.
33. Kim JW, Kong IG, Lee CH, Kim DY, Rhee CS, Min YG, Kim CW, Chung JH: **Expression of Bcl-2 in olfactory neuroblastoma and its association with chemotherapy and survival.** *Otolaryngol Head Neck Surg* 2008, **139**(5):708-712.
34. Zhang M, Zhang P, Zhang C, Sun J, Wang L, Li J, Tian Z, Chen W: **Prognostic significance of Bcl-2 and Bax protein expression in the patients with oral squamous cell carcinoma.** *J Oral Pathol Med* 2009, **38**(3):307-313.
35. Martin B, Paesmans M, Berghmans T, Branle F, Ghisdal L, Mascaux C, Meert AP, Steels E, Vallot F, Verdebout JM, Lafitte JJ, Sculier JP: **Role of Bcl-2 as a prognostic factor for survival in lung cancer: a systematic review of the literature with meta-analysis.** *Br J Cancer* 2003, **89**(1):55-64.
36. Kondo E, Miyake T, Shibata M, Kimura T, Iwagaki H, Nakamura S, Tanaka T, Ohara N, Ichimura K, Oka T, Yanai H, Shibasaki F, Yoshino T: **Expression of phosphorylated Ser70 of Bcl-2 correlates with malignancy in human colorectal neoplasms.** *Clin Cancer Res* 2005, **11**(20):7255-7263.
37. Lo Muzio L, Falaschini S, Farina A, Rubini C, Pezzetti F, Campisi G, De Rosa G, Capogreco M, Carinci F: **Bcl-2 as prognostic factor in head and neck squamous cell carcinoma.** *Oncol Res* 2005, **15**(5):249-255.



doi: 10.1186/1477-7819-8-31

**Cite this article as:** Zheng *et al.*, The conformation change of Bcl-2 is involved in arsenic trioxide-induced apoptosis and inhibition of proliferation in SGC7901 human gastric cancer cells *World Journal of Surgical Oncology* 2010, **8**:31

**Submit your next manuscript to BioMed Central  
and take full advantage of:**

- Convenient online submission
- Thorough peer review
- No space constraints or color figure charges
- Immediate publication on acceptance
- Inclusion in PubMed, CAS, Scopus and Google Scholar
- Research which is freely available for redistribution

Submit your manuscript at  
[www.biomedcentral.com/submit](http://www.biomedcentral.com/submit)

