



## Data in Brief

## Gene expression changes of single skeletal muscle fibers in response to modulation of the mitochondrial calcium uniporter (MCU)



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## ABSTRACT

The mitochondrial calcium uniporter (MCU) gene codifies for the inner mitochondrial membrane (IMM) channel responsible for mitochondrial  $\text{Ca}^{2+}$  uptake. Cytosolic  $\text{Ca}^{2+}$  transients are involved in sarcomere contraction through cycles of release and storage in the sarcoplasmic reticulum. In addition cytosolic  $\text{Ca}^{2+}$  regulates various signaling cascades that eventually lead to gene expression reprogramming. Mitochondria are strategically placed in close contact with the ER/SR, thus cytosolic  $\text{Ca}^{2+}$  transients elicit large increases in the  $[\text{Ca}^{2+}]$  of the mitochondrial matrix ( $[\text{Ca}^{2+}]_{\text{mt}}$ ). Mitochondrial  $\text{Ca}^{2+}$  uptake regulates energy production and cell survival. In addition, we recently showed that MCU-dependent mitochondrial  $\text{Ca}^{2+}$  uptake controls skeletal muscle trophism. In the same report, we dissected the effects of MCU-dependent mitochondrial  $\text{Ca}^{2+}$  uptake on gene expression through microarray gene expression analysis upon modulation of MCU expression by *in vivo* AAV infection. Analyses were performed on single skeletal muscle fibers at two time points (7 and 14 days post-AAV injection). Raw and normalized data are available on the GEO database (<http://www.ncbi.nlm.nih.gov/geo/>) (GSE60931).

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Specifications	
Organism/cell line/tissue	Single skeletal muscle fibers from mouse EDL
Sex	Male
Sequencer or array type	Agilent-028005 SurePrint G3 Mouse GE 8 × 60K Microarray type
Data format	Raw and normalized
Experimental factors	Two time points (7 days and 14 days) for each experimental factor: MCU up-regulation, MCU silencing, wild type mice, expression of control shRNA (shluc).
Experimental features	Two time points were considered for each condition (MCU up-regulation, MCU silencing, wild type, expression of control shRNA (shluc)). Each condition was analyzed with at least three biological replicates. AAV-MCU, AAV-shMCU and AAV-shluc were used for mice <i>in-vivo</i> infection. After mice sacrifice EDL skeletal muscles were excised and single myofibers were recovered and characterized. Only type 2b myofiber types were used in gene expression experiments.
Consent	<i>In vivo</i> experiments were performed in accordance with the Italian law D. L.vo n.26/2014.

(continued)

Specifications	
Sample source location	CD1 mice from University of Padova, Italy

## 1. Direct link to deposited data

Deposited data can be found here: <http://www.ncbi.nlm.nih.gov/geo/query/acc.cgi?acc=GSE60931>.

## 2. Experimental design, materials and methods

## 2.1. Experimental design

To dissect the transcriptional effects of mitochondrial calcium uniporter (MCU) gene modulation in *in-vivo* single skeletal muscle fiber we performed a time series gene expression analysis after MCU up- or down-regulation [1]. The MCU gene encodes the channel of the inner mitochondrial membrane (IMM) responsible for mitochondrial  $\text{Ca}^{2+}$  uptake which controls the aerobic metabolism, cell death, and

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**Table 1**  
Primers used for qRT-PCR.

Primer name	Sequence	Description
MCU_endo_FOR	AAAGGAGCCAAAAGTCACG	Endogenous MCU
MCU_endo_REV	AACGGCGTGAGTTACAAACA	Endogenous MCU
MCU_eso_FOR	AATTGCTCAGGCAGAAATGGA	Exogenous MCU from AAV-MCU
MCU_eso_REV	CTTATCGTCGTCATCCTTGAATC	Exogenous MCU from AAV-MCU
Txn1_FOR	TCCAATGTGGTTCCTTGA	Reference gene
Txn1_REV	GGCTTCAAGCTTTTCCTTGT	Reference gene

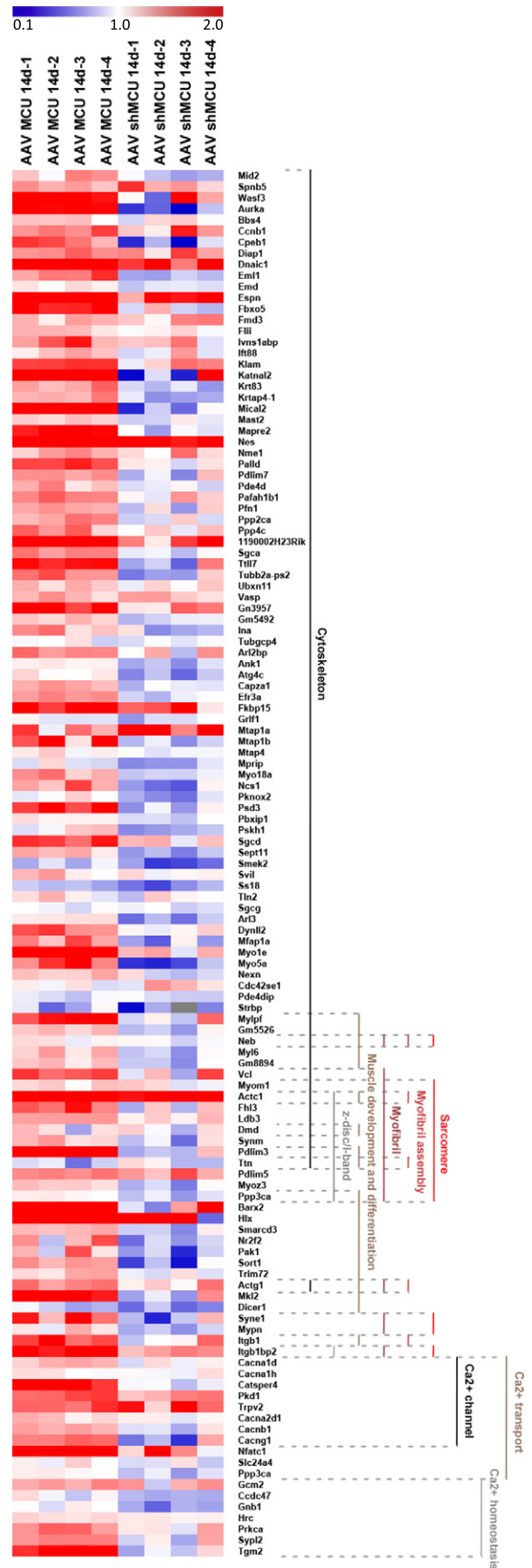
survival pathways [2–4]. Muscle activity leads to major swings in mitochondrial  $[Ca^{2+}]$ , which, in turn controls muscle trophism [1].

## 2.2. Materials and methods

Adult 2 month old mice were used for the experiments. Untreated and adeno-associated virus (AAV) injected EDL muscles were collected. Muscles were digested in type I collagenase (10 mg/ml in DMEM). Single myofibers were dissociated. Intact, not contracted single myofibers were picked under a stereomicroscope and washed in PBS. For AAV-shluc and AAV-shMCU infected muscles, ZsGreen-expressing myofibers were selected using an inverted microscope (DMI4000, Leica). Each single isolated myofiber was lysed in 250  $\mu$ l of TRIzol Reagent (Life Technologies) and RNA was extracted in the aqueous phase following the manufacturer's instructions. To purify RNA, spin-columns of the RNeasy Micro Kit (Qiagen) were used (Fig. 1). Before performing microarray experiments about 1/5 of the purified RNA was used for qRT-PCR measurements of MCU expression levels (Fig. 2) and for fiber characterization (Fig. 1). RNA was retrotranscribed using the SuperScript III Reverse Transcriptase (Life Technologies) according to the manufacturer's specifications. Gene-specific primers for exogenous MCU (AAV-MCU), and endogenous MCU were selected with Primer3Plus (<http://www.bioinformatics.nl/cgi-bin/primer3plus/primer3plus.cgi>) and the specificity of each primer set was monitored by dissociation curve analysis. Thioredoxin 1 was chosen as the reference gene according to the results in [5]. The selected primers are described in Table 1.

The qRT-PCR experiments were performed in a 7500 Real-time PCR System (Life Technologies) using the SYBR Green technology of GoTaq qPCR Master Mix (Promega). qRT-PCR confirmed the induction and silencing of MCU in selected fibers for microarray analysis (Fig. 2). Moreover, MCU induction is time dependent evidencing a smaller MCU up-regulation 7 days after AAV injection (~8 fold) than that 14 days after AAV injection (~360 fold).

The remaining purified RNA of the selected fibers was exponentially amplified using the TransPlex Whole Transcriptome Amplification 2 Kit (Sigma-Aldrich) to obtain a sufficient amount of cDNA for the microarray experiments. Briefly, RNA was reverse transcribed in a cDNA library, and then the library was exponentially amplified for 18 cycles, a few cycles below the amplification "plateau" observed in a PCR test reaction. To remove the residual primers and nucleotides, the amplification product was purified with the GenElute PCR Clean-up columns (Sigma-Aldrich). The resulting cDNA was quantified with a Nanodrop ND-1000 spectrophotometer (Thermo Scientific). 2  $\mu$ g of amplified-purified cDNA was directly labeled using the Genomic DNA Enzymatic Labeling Kit (Agilent Technologies). The kit uses random primers and the exo-Klenow fragment to directly label cDNA samples with Cy3-dUTP nucleotides. Labeled cDNA was purified using Amicon 30 kDa filters (Millipore) and quantified using the NanoDrop ND-1000 spectrophotometer (Thermo Scientific). On average, cDNA yield was about 4  $\mu$ g and the specific activity of 30 pmol Cy3 per  $\mu$ g of cDNA (Fig. 1).



**Fig. 3.** Heat map of differentially expressed genes between AAV-MCU and AAV-shMCU after 14 days. Selected genes are those enriched for the cytoskeleton, myofibrils, sarcomere organization and calcium ion homeostasis functions. Gene expression values are relative to the average expression in the control condition: the blue color means low expression, the red color high expression.

The microarray experiments were performed using SurePrint G3 Mouse Gene Expression 8 × 60K microarrays (Agilent Technologies) (GEO platform: [GPL13912](#)). 800 ng of labeled cDNA target was mixed with 5 µl of 10× Blocking Agent (Agilent Technologies) and water to a final volume of 25 µl. Samples were denatured at 95 °C for 2 min and added to 25 µl of 2× GEx Hybridization Buffer HI-RPM (Agilent Technologies). A 40 µl mix was dispensed onto the array. Slides were loaded into the Agilent SureHyb chambers and hybridization was performed in a hybridization oven at 65 °C for 17 h with a 10 rpm rotation. After hybridization, slides were washed using a Wash Buffer Kit (Agilent Technologies) and dried at RT.

Microarray slides were scanned using a G2505C scanner (Agilent Technologies) at a 3 µm resolution. Probe features were extracted using the Feature Extraction Software v. 10.7.3.1 with GE\_1\_Sep09 protocol (Agilent Technologies). The Feature Extraction Software directly performed intra-array normalizations. Raw data are available in the GEO database (accession number [GSE60931](#)). The genomic results described in [1] were produced as following described. Inter-array normalization of expression levels was performed with a quantile method [6] and the values for within-array replicate spots were then averaged. Feature Extraction Software, which provided spot quality measures, was used to evaluate the quality and reliability of the hybridization. In particular, the flag “glsFound” (set to 1 if the spot had an intensity value significantly different from that of the local background and to 0 when otherwise) was used to filter out unreliable probes: the flag equals to 0 was to be noted as “not available” (NA). Probes with a high proportion of NA values were removed from the dataset in order to carry out more solid and unbiased statistical analyses. 45% of NA was used as the threshold in the filtering process, and a total of 30,073 of 39,570 probes were obtained. The comparison of MCU expression obtained with qRT-PCR and microarray evidenced a correlation of 98.8% (Fig. 2). To identify the differentially expressed probes in at least one condition, one way ANOVA analysis was performed using a threshold  $p$ -value  $\leq 0.01$ . Significant differentially expressed probes were used to search specific expression clusters according to the Self Organizing Tree Algorithm (SOTA) [7] as implemented in MultiExperiment Viewer

version 4.8.1 (tMev) of the TM4 Microarray Software Suite [8]. Gene ontology analysis was performed using DAVID web tool [9], while pathway analysis was performed applying Gene Set Enrichment Analysis (GSEA) as implemented in Graphite web tool [10]. The KEGG pathway database was used and only pathways that presented at least 10 mapped genes in common with our gene expression matrix were considered in the analysis. Interestingly, the analysis evidenced that, after MCU up-regulation, genes involved in Ca<sup>2+</sup> homeostasis are activated, while, but when MCU was down-regulated, the same genes were lesser activated or down-regulated (Fig. 3).

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