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Citation: Singer M, Ashimatey BS, Zhou X, Chu Z, Wang R, Kashani AH (2020) Impaired layer specific retinal vascular reactivity among diabetic subjects. PLoS ONE 15(9): e0233871. https://doi.org/ 10.1371/journal.pone.0233871

Editor: Ji Yi, Boston Medical Center, Boston University School of Medicine, UNITED STATES

Received: May 12, 2020

Accepted: August 29, 2020

Published: September 11, 2020

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Data Availability Statement: Retinal images may contain identifiable data (Fukuta, Keisuke, Toshiaki Nakagawa, Yoshinori Hayashi, Yuji Hatanaka, Takeshi Hara, and Hiroshi Fujita. "Personal Identification Based on Blood Vessels of Retinal Fundus Images." In Medical Imaging 2008: Image Processing, 6914:69141V. International Society for Optics and Photonics, 2008. https://doi.org/10. <u>1117/12.769330</u>). We therefore cannot upload our data for unrestricted use based on the US Department of Health and Human Services guidelines (https://www.hhs.gov./hipaa/forprofessionals/privacy/laws-regulations/index.html) RESEARCH ARTICLE

Impaired layer specific retinal vascular reactivity among diabetic subjects

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Abstract

Purpose

To investigate layer specific retinal vascular reactivity (RVR) in capillaries of diabetic subjects without DR or with only mild non-proliferative diabetic retinopathy (NPDR).

Methods

A previously described nonrebreathing apparatus was used to deliver room air, 5% CO₂, or 100% O₂ to 41 controls and 22 diabetic subjects (with mild or no NPDR) while simultaneously acquiring fovea-centered 3x3mm² Swept-Source Optical Coherence Tomography Angiography (SS-OCTA) images. Vessel skeleton density (VSD) and vessel diameter index (VDI) were calculated for each gas condition for the superficial retinal layer (SRL) and deep retinal layer (DRL). The superficial layer analysis excluded arterioles and venules. Data analysis was performed using mixed factorial analysis of covariance stratified by diabetic status. All models were adjusted for age, gender, and hypertension, and statistical significance for multiple comparisons from posthoc comparisons were defined at p<0.017.

Results

Among controls, there was a significant difference in capillary VSD between all gas conditions (p<0.001). This difference was present in both the SRL and DRL. Among diabetics, there was no significant difference in response to CO_2 conditions in the SRL (p = 0.072), and a blunted response to both CO_2 (p = 0.9) and O_2 in the DRL (p = 0.019). A significant gas effect was detected in the capillary VDI in the SRL of controls (p = 0.001), which was driven by higher VDI in the oxygen condition compared to that of carbon dioxide.

Conclusions

Impairment in RVR in diabetic subjects is characterized by a paradoxical response to CO_2 in both the SRL and DRL as well as an attenuated response to O_2 in the DRL. These layer and

and our own IRB policy. Per our protocol for this study approved by the IRB, all data and images will only be accessible to the principal investigator and research staff that are part of the study. However, we are happy to make the data available to those with legitimate interest who have completed the requisite human subjects training (e.g. CITI training) and obtained the appropriate IRB approval. Interested parties may reach out to the University of Southern California IRB (irb@usc. edu) and the contact author (Amir Kashani at ahkashan@usc.edu).

Funding: This study was supported by the Tai Family Research Scholars Fund (MS), an unrestricted grant from Research to Prevent Blindness, https://www.rpbusa.org/rpb (AK) to the USC Roski Eye Institute, the National Eye Institute (R01EY030564, K08EY027006) (AK), and by Carl Zeiss Meditec, Inc., Dublin, CA, 94568 (AK). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing interests: Kashani - Carl Zeiss Meditec (Financial Support), Carl Zeiss Meditec (Recipient); Wang - Carl Zeiss Meditec (Patents US8180134B2, US9013555B2, US9759544B2 and US10354378B2), Carl Zeiss Meditec (Consultant), Kowa Inc (Patent), Insight Phototonic Solutions (Consultant). This does not alter our adherence to PLOS ONE policies on sharing data and materials. None of the commercial entities had any role in the design of the study, analysis of the data or preparation of the manuscript. gas specific impairments in diabetics seem to occur early in the disease and to be driven primarily at the capillary level.

Introduction

Blood flow through the retinal vasculature changes according to the metabolic needs of local tissue and the presence or absence of metabolites in the local environment [1]. One example of this spatial and temporal heterogeneity in blood flow is the reactivity of retinal vessels to blood gas perturbations. Retinal vessels change in caliber, dilating in hypercapnic conditions and constricting in hyperoxic conditions [2-7]. Such changes are mediated by regulatory mechanisms that allow for decentralized regulation of blood flow within the larger vessels of the retina [8]. For example, retinal arteries and arterioles can change in diameter in response to changes in metabolic demand or supply [9-11]. This retinal vascular reactivity (RVR) has been shown to be blunted in diabetics [12-16] even in the absence of clinically apparent diabetic retinopathy (DR) [2]. Most studies have demonstrated reactivity changes in arterioles or venules, but few have demonstrated impairment at the capillary level [1, 6]. Unlike many other imaging modalities that have been used to assess retinal vascular reactivity at the capillary level, Optical Coherence Tomography Angiography (OCTA) is an FDA approved technology that is commercially available [17, 18]. Our group has recently used Swept-Source OCTA (SS-OCTA) to investigate the impact of room air, hypercapnia and hyperoxia conditions on retinal capillaries in full thickness retina between controls and subjects with DR. That study concluded that subjects with DR preferentially responded to hyperoxia but not hypercapnia, while the healthy controls responded to both [19].

There is currently growing evidence that the deep retinal vascular plexus is more severely damaged by diabetes than the superficial plexus. Even in the absence of clinically detectible retinopathy, the deep vascular plexus shows lower capillary density than in controls [20-24]. Given that vascular damage may occur early in disease and vary by retinal layer, we expand our previous studies by investigating layer specific retinal vascular reactivity between healthy controls and subjects with mild non-proliferative diabetic retinopathy (NPDR) or no DR.

Methods

This was a prospective, observational study of diabetic patients and healthy controls. This study was approved by the institutional review board of the University of Southern California (USC) and adhered to the tenets of the Declaration of Helsinki.

Subjects

Subjects were recruited at the USC Roski Eye Institute and informed consents were obtained from each subject. Subjects were excluded if they had any past ocular history of glaucoma or retinal surgery, or past medical history of vascular diseases including heart disease, surgery, or vasculitis. Only those diabetic subjects without clinically apparent DR or with only mild NPDR were included. All subjects with significant media opacity such as cataracts or any other obscuration of the fundus were excluded. Subjects who reported a history of syncope, shortness of breath, lung disease, congestive heart failure, or recent hospitalization of greater than 1-day duration within the last 12 months were also excluded to minimize risk of adverse events during gas breathing. All eyes were imaged at the USC Roski Eye Institute between October

2017 and February 2020. Approximately, 46% of controls and 36% of diabetics overlapped with those from our previous study by Ashimatey et al. [19]

Gas delivery system

In order to create conditions of physiological hyperoxia and hypercarbia, a nonrebreathing apparatus was used to deliver gas mixtures. This gas delivery system has been described by Ashimatey et al., [19] and a step by step visual-illustration of the procedure is demonstrated in Kushner-Lenhoff et al. [25] In brief, three gas mixtures were delivered through a non-rebreathing gas apparatus to the subjects in the following order during the same visit: room air, a mixture of 5% carbon dioxide, atmospheric oxygen and balanced nitrogen, and finally 100% oxygen. There was a 10-minute refractory period between the hypercapnic and hyperoxic conditions to allow washout of the previous gas condition. The order of gas administration was established to maximize testing efficiency and to mitigate the impact of a persistent effect of a prior testing condition on the next condition. Room air was performed first because that was the baseline measure and acquiring images under room air condition after any gas breathing introduces the possibility of persistent gas effect from the prior gas condition. Yezhuvath et al. found that end tidal CO₂ and cerebrovascular reactivity returned to baseline 60 seconds after 5% CO₂ breathing and Tayyari et al suggested that an after-gas effect from O₂ could persist longer that of CO₂ [3, 26]. A 10-mins wash out time between CO₂ and O₂ was deemed adequate based on previous studies [3]. Therefore the order of CO_2 and O_2 gas administration was fixed for all subjects. During delivery of the gases, a fingertip pulse oximeter was used to monitor oxygen saturation. Testing was stopped if the oxygen saturation fell below 94% for any subject at any time.

SS-OCTA imaging protocol

During gas breathing, images were acquired using a 100kHz SS-OCTA system (PLEX Elite 9000, software version 1.7; Carl Zeiss Meditec, Dublin, CA, USA). The light source has a central wavelength of 1060 nm and a bandwidth of 100 nm, providing an axial resolution of ~ 6μ m and lateral resolution of ~ 20μ m (estimated at the surface of retina). The device uses an optical microangiopathy (OMAG) algorithm [27, 28] to determine the motion signal, which is interpreted as representing the flow of erythrocytes. Three OCTA volumes (3x3mm field-of-view) centered on the fovea were acquired starting 60 seconds after the onset of each gas breathing condition. Before OCTA image acquisitions were started, the subjects were asked to keep their eyes closed immediately after the imaging set-up was completed and the retinal region to be imaged was focused and aligned. Subjects were asked to blink two times before any OCTA image acquisitions and at the beginning of the imaging session to maximize image quality and signal strength.

Each OCTA volume is formed by 300 horizontal scans (B-scans) with each B-scan formed by 300 A-scans of 1536 pixels. One eye per patient was imaged, based on fixation ability, absence of co-morbid disease and media opacity. If both eyes were suitable, the imaged eye was chosen randomly. Images with poor decentration of the foveal avascular zone (FAZ), signal strength less than seven, significant motion artifacts or vitreous floaters were excluded. One representative image, the image with the highest signal strength and fewest artifacts, was selected per condition. The volumes were then automatically segmented into *en face* superficial retinal layer (SRL) and deep retinal layer (DRL). The segmentation software used is commercially available on the OCTA device and has been previously described by our group in the past [29, 30]. The SRL extends from the inner limiting membrane (ILM) to the inner plexiform layer (IPL), which is approximated as 70% of the thickness between the ILM and outer plexiform layer (OPL). The DRL extends from the IPL to the OPL, which is estimated to be 110 μ m above the retinal pigment epithelium (RPE). The *en face* DRL for the best images were projection-resolved using the commercially available projection artifact removal feature on the device [31, 32].

Quantitative vessel morphometric measures

Quantitative analysis with custom MATLAB software was performed on SRL and DRL images to calculate measures that have been described previously [33]. Vessel area density (VAD) represents the total area of the image occupied by vessels and is calculated by dividing the area occupied by vessel pixels by the dimensions of the image after image binarization. Vessel skeleton density (VSD) represents the total length of the blood vessels. VSD is calculated by skeletonizing the diameter of each vessel to one pixel and then dividing the pixels that represent all the skeletonized vessels by the total area of the image. Vessel diameter index represents the caliber of the blood vessels and is calculated by dividing VAD by VSD [25, 29, 34].

The central 3x3mm foveal centered region of the macula contains small arterioles which may preferentially drive the RVR response. In order to examine the isolated effect of gas conditions on capillaries only, a custom semi-automated MATLAB software was developed specifically to exclude non-capillary vessels that occur in the SRL (Fig 1). A manual threshold was empirically determined to exclude arterioles and venules without excluding precapillary arteriolar segments. VSD is then calculated by dividing the remaining skeletonized capillaries by all the pixels of the image that were not occupied by large vessels. In addition, VSD, VDI, and VAD of the SRL with the large vessels included was also quantified for comparison. Unless otherwise specified, results presented are those with large vessels excluded.

Statistical analysis

A 2 (layers: DRL, SRL) x 3 (gas conditions: room air, oxygen, carbon dioxide) mixed factorial analysis of covariance (ANCOVA) model with repeated effects of layer and breathing condition was used to assess differences in the OCTA metrics (VSD, VAD and VDI) by layer, gas breathing condition, and their interaction. Due to limitations in sample size and power to detect a three-way layer-gas-diabetes interaction, separate models were run for diabetic versus normal patients and for VSD measurements obtained for the images. All models were adjusted for age, gender, and hypertension. Differences of the differences in VSD by gas breathing condition between layers were estimated from the models. All statistical tests were two-sided with

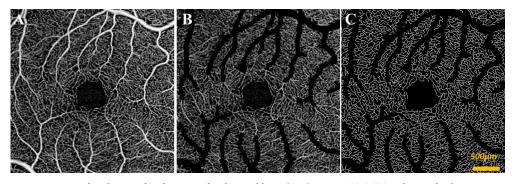


Fig 1. Large vessel exclusion of en face superficial retinal layer (SRL) 3x3mm SS-OCTA of control subject acquired during room air condition. (A) original image (B) original image with large vessels removed (C) image after large vessels excluded and remaining capillaries skeletonized.

https://doi.org/10.1371/journal.pone.0233871.g001

an alpha level set at 0.05 for statistical significance. The assumption of sphericity was assessed using the Mauchly's assessment of sphericity, and when the assumption of sphericity was violated, the p-value associated with the degree of freedom adjustment using the Greenhouse-Geisser correction was reported. The Bonferroni method was applied for posthoc multiple comparisons at p = 0.017. All statistical analyses were performed using SPSS version 25 (IBM SPSS Statistics for Windows, Version 25.0. Armonk, NY).

Results

Our study included 41 control (mean age 53.0 ± 18.9) and 22 diabetic subjects (mean age 53.7 ± 16.7). Table 1 summarizes the subject demographics. Fig 2 shows qualitative representations of the decorrelation signal intensity and capillary density under different gas breathing conditions in the SRL and DRL for a control subject. The same representations are shown for a diabetic subject in Fig 3. Subtle signal intensity changes and the apparent density of capillaries in the control subject were qualitatively increased in the hypercapnic condition compared to the other two conditions. Quantitative analysis confirmed this finding by showing that significant effects on VSD of gas breathing condition, layer, and the gas-layer interactions were present for both controls and diabetics, after controlling for age, gender, and hypertension (all p<0.001). Below we elaborate on these findings in more detail.

Retinal vascular reactivity to CO₂ is absent in SRL and DRL of diabetic subjects

There was no significant difference in mean VSD of diabetic subjects for the pairwise comparisons between room air and CO₂ gas conditions (SRL_{RA} 0.148 ± 0.002 vs SRL_{CO2} 0.146 ± 0.002, p = 0.072). As Fig 4 shows, the reactivity of retinal capillaries to CO₂ was not only absent in the SRL of diabetic subjects but the mean VSD under CO₂ was paradoxically lower than RA. In contrast, in the SRL of control subjects, there was a robust difference in all pairwise comparisons of the mean VSD among the three gas conditions in the expected directions (SRL_{RA} 0.154 ± 0.001 vs SRL_{CO2} 0.156 ± 0.002 vs SRL_{O2} 0.150 ± 0.002, p<0.017). Results for VAD were consistent with those of VSD, and the VSD results were unchanged when large vessels were not excluded from the SRL (see S1 Fig and S2 Fig). The omnibus findings of the mixed factorial analysis confirmed a significant gas condition (p<0.001), layer (p<0.001) and gaslayer interaction (p = 0.001) in the controls. In diabetic subjects, the mixed factorial analysis

Disease Classification	
Controls	Diabetics
41	22
53.0 ± 18.9	53.7 ± 16.7
20 (48.8%)	9 (40.9%)
10 (24.4%)	6 (27.3%)
N/A	15 no DR, 7 mild NPDR
OCTA signal strength (mean, [Min, Max]) under gas non-rebreathing	
conditions	
9.8 [9,10]	9.8 [9,10]
9.7 [8,10]	9.9 [9,10]
9.6 [8,10]	9.6 [9,10]
	Controls 41 53.0 ± 18.9 20 (48.8%) 10 (24.4%) N/A OCTA signal strength (mean 9.8 [9,10] 9.7 [8,10]

Table 1. Subject demographics.

The number, age, gender, hypertension status, and ocular diagnosis of subjects is summarized. RA means Room air.

https://doi.org/10.1371/journal.pone.0233871.t001

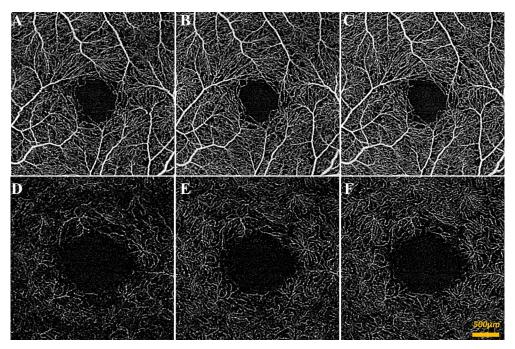


Fig 2. SS-OCTA of control subject during different gas conditions. (A-C) 3x3mm en face superficial retinal layer image of blood flow around central macula during oxygen condition (A), room air condition (B), and carbon dioxide condition (C). (D-E) 3x3mm en face deep retinal layer image of blood flow around central macula during oxygen condition (D), room air condition (E), and carbon dioxide condition (F). A qualitative increase in signal was apparent in the hypercapnic condition when compared to the other two conditions.

https://doi.org/10.1371/journal.pone.0233871.g002

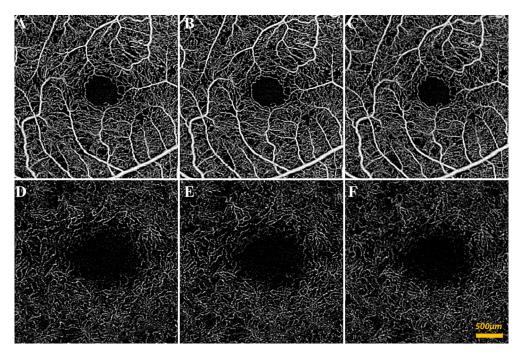


Fig 3. SS-OCTA of diabetic subject without diabetic retinopathy during different gas conditions. (A-C) 3x3mm en face superficial retinal layer image of blood flow around central macula during oxygen condition (A), room air condition (B), and carbon dioxide condition (C). (D-E) 3x3mm en face deep retinal layer image of blood flow around central macula during oxygen condition (F).

https://doi.org/10.1371/journal.pone.0233871.g003

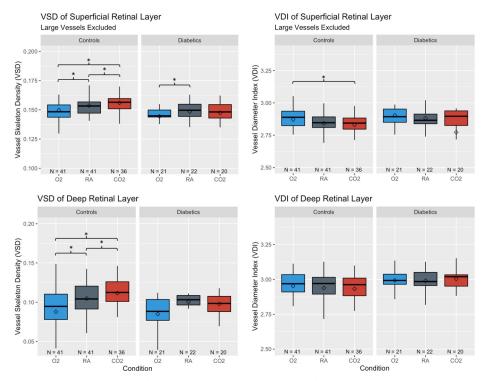


Fig 4. Vessel skeleton density (VSD) and vessel diameter index (VDI) of control and diabetic subjects under gas conditions. *Whiskers* indicate highest or lowest point within 1.5 times the interquartile range from the upper or lower quartile. *Diamonds* indicate the mean. *Stars* indicate significance for pairwise comparison between conditions from the ANCOVA model based on Bonferroni adjusted p-value of 0.017.

https://doi.org/10.1371/journal.pone.0233871.g004

confirmed that there was a significant effect of gas condition (p = 0.005), mainly driven by the response to O₂, layer (p < 0.001) as well as a significant gas-layer interaction (p = 0.02).

For the VDI metrics, there was a significant gas effect in the SRL of control subjects (SRL_{RA} 2.84 ± 0.01 vs SRL_{CO2} 2.83 ± 0.01 vs SRL_{O2} 2.87 ± 0.02, p = 0.001) which was driven primarily by the difference in VDI between O₂ and CO₂ (p = 0.002). Gas effect on the VDI metric in the SRL was not present in diabetic subjects (SRL_{RA} 2.88 ± 0.03 vs SRL_{CO2} 2.77 ± 0.03 vs SRL_{O2} 2.90 ± 0.02, p = 0.44). In the controls, the mixed factorial analysis confirmed that there was a significant effect of both gas conditions (p = 0.006) and layer (p<0.001) on VDI but no significant gas-layer interaction (p = 0.29). In diabetic subjects, the mixed factorial analysis neither confirmed a significant effect of gas condition (p = 0.42) nor a significant gas-layer effect (p = 0.34) but did show a significant difference between layers (p = 0.009). The VDI results were similar when large vessels were not excluded from the SRL (see S2 Fig).

Unlike the controls with significant pairwise comparisons between gas conditions (p<0.017), all pairwise comparisons of VSD between gas conditions in the DRL of diabetic subjects were non-significant. The mixed factorial analysis of the gas effect in the diabetics however showed significance (p = 0.005) and appeared to be driven by the response to hyperoxia from room air in the SRL (p = 0.005). The VSD response to hyperoxia in the DRL was trending towards significance (p = 0.019) but did not reach significance with the Bonferroni correction. Notably, as in the SRL, the difference in the VSD of the DRL under hypercapnic conditions was in a paradoxical direction and may reflect an aberrant physiological response in the diabetic subjects. In the controls the mixed factorial analysis confirmed significant gas effects on VSD in the DRL (DRL_{RA} 0.105 ± 0.003, DRL_{CO2} 0.112 ± 0.003, DRL_{O2} 0.087 ± 0.006, p<0.001). Results for VAD in the DRL were consistent with those of VSD (see S1 Fig).

Neither the controls nor subjects with diabetes had a significant gas effect on the VDI within the DRL.

Discussion

We investigated layer specific RVR during physiologically salient changes in inhaled oxygen and carbon dioxide in diabetic subjects and non-diabetic controls using SS-OCTA. Consistent with our previous findings [19] we found retinal vascular reactivity impairment in diabetic subjects can be assessed with OCTA and show that even in the early stages of the disease the impairment to hypercapnia previously reported are present. Whereas our earlier study consisted entirely of DR subjects with the full range of disease severity, our current cohort of diabetic subjects included only those without DR (68%) or those with mild NPDR. We extend our previous findings by demonstrating that the impairment in RVR in diabetic subjects is present in both the SRL and DRL and is more profound in the DRL. Namely, there was no significant effect of any gas condition on apparent capillary density in the DRL among diabetic subjects by pairwise comparison. Lastly, we show that the RVR findings are present even when larger arterioles and venules are excluded from the images. This suggests that capillaries themselves mediate the RVR and that this physiological response is abnormal in DR.

We found that retinal vascular reactivity to hyperoxia was preserved in the SRL of diabetic subjects but not significant in the DRL, even in the early stages of the disease. This is consistent with the recent report by Meshi et. al. that among diabetic subjects without DR, capillary density in the deep capillary plexus but not the superficial capillary plexus was significantly lower than that of controls [24]. Various other studies have reported a similar result for type 1 diabetics with no or mild DR [21–23]. It may therefore be that capillary damage in the DRL, including the ability to autoregulate, is more extensive than damage to the SRL in early disease.

We primarily used a measure of capillary density, VSD, which represents the length of capillaries in the image. Changes to VSD with the gas conditions may reflect blood flow within previously unperfused capillaries or vice versa, but more likely reflects incremental changes in flow through capillary segments. Due to the detection sensitivity of OCTA imaging in which flow below a certain threshold cannot be resolved, loss of signal for a given capillary segment between gas conditions may be from decreased perfusion through that segment outside the detection limits of the device. Differences in the VSD measures under the different gas conditions are thus convincing evidence of the change in perfusion per unit area.

Since capillaries typically only pass individual erythrocytes, it is possible that they function in binary states (only have open and closed states). Although it is not clear if Poiseuille's relation between flow rate and vessel diameter applies to capillaries, the changes in perfusion measures captured in our VSD metric may also be due to changes of vessel caliber induced by the gas conditions. The VDI results in the SRL were lower with increased CO₂ content in the controls. As VDI is a ratio of total vessel area (VAD) divided by skeletonized vessel length (VSD), the low VDI observed in the carbon dioxide condition compared to the oxygen condition is likely driven by a relatively large increase in vessel length compared to vessel width. Hence, the changes in VAD under the gas perturbation conditions are largely driven by the absence or presence of flow segments, rather than changes to flow caliber at the capillary level. This may suggest that capillary blood flow changes to gas perturbation function in a more binary manner (open or closed). A similar VDI trend was not noticed in the DRL of the controls and could mean the vessel dilation was simply neutralized by an increase in appearance of new vessel segments. The lack of changing VDI in the DRL could also be due to the confounding effect of OCTA signal [35].

Capillary pericyte loss, endothelial cells depletion and basement membrane thickening are among the earliest vasobliterations associated with clinically detectible signs diabetic

retinopathy [35, 36]. The potential mechanisms underlying retinal vascular reactivity assessment at the capillary level, such as assessed by our study, provide a pathway for implementing a clinically feasible protocol that can be an early biomarker of vascular function. Future studies may explore retinal vascular reactivity measures at the capillary level as a potential clinical variable for investigating the wide variability in the duration between when a subject first develops diabetes to when retinopathy is diagnosed.

The findings of our study come with some limitations. Namely our sample size was small and therefore we were unable to control for all potential confounders of retinal vascular reactivity including smoking [37]. Even though our sample size was small, our data was representative of the general trends previously described by our group and others, namely, that the mean VSD for the diabetic subjects was lower than the controls and VSD in the SRL was consistently greater than in the DRL. In addition, the relationship of VSD among retinal layers was predictable among controls and agreed with previous findings. One other limitation of our study is that the gas conditions we used were relatively mild to ensure subject comfort and safety. Nevertheless, the methodology is very useful because it can be applied using commercially available devices and on living human subjects with reasonable and reliable effect sizes. We have provided a detailed description of the methodology and its strengths and limitations in our previous publication Ashimatey et al. [19] as well as in Kushner-Lenhoff et al. [25]

Supporting information

S1 Fig. Vessel area density (VAD) of control and diabetic subjects under gas conditions. *Whiskers* indicate highest or lowest point within 1.5 times the interquartile range from the upper or lower quartile. *Diamonds* indicate the mean. *Stars* indicate significance for pairwise comparison between conditions from the ANCOVA model based on Bonferroni adjusted p-value of 0.017.

(TIF)

S2 Fig. Vessel skeleton density (VSD) and vessel diameter index (VDI) of control and diabetic subjects under gas conditions using all vessel analysis in the superficial retinal layer. *Whiskers* indicate highest or lowest point within 1.5 times the interquartile range from the upper or lower quartile. *Diamonds* indicate the mean. *Stars* indicate significance for pairwise comparison between conditions from the ANCOVA model based on Bonferroni adjusted pvalue of 0.017.

(TIF)

Acknowledgments

The authors would like to thank Melissa Mert and Luis de Sisternes for providing statistical and OCTA-related technical advice respectively during the manuscript development.

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Visualization: Maxwell Singer.

Writing – original draft: Maxwell Singer.

Writing – review & editing: Maxwell Singer, Bright S. Ashimatey, Xiao Zhou, Zhongdi Chu, Ruikang Wang, Amir H. Kashani.

References

- 1. Yu DY, Cringle SJ, Yu PK, Balaratnasingam C, Mehnert A, Sarunic MV, et al. Retinal capillary perfusion: Spatial and temporal heterogeneity. Progress in retinal and eye research. 2019; 70:23–54.
- Gilmore ED, Hudson C, Nrusimhadevara RK, Harvey PT, Mandelcorn M, Lam WC, et al. Retinal arteriolar diameter, blood velocity, and blood flow response to an isocapnic hyperoxic provocation in early sight-threatening diabetic retinopathy. Investigative ophthalmology & visual science. 2007; 48(4):1744– 50.
- Tayyari F, Venkataraman ST, Gilmore ED, Wong T, Fisher J, Hudson C. The relationship between retinal vascular reactivity and arteriolar diameter in response to metabolic provocation. Investigative ophthalmology & visual science. 2009; 50(10):4814–21.
- Klefter ON, Lauritsen A, Larsen M. Retinal hemodynamic oxygen reactivity assessed by perfusion velocity, blood oximetry and vessel diameter measurements. Acta ophthalmologica. 2015; 93(3):232– 41.
- Palkovits S, Lasta M, Told R, Schmidl D, Boltz A, Napora KJ, et al. Retinal oxygen metabolism during normoxia and hyperoxia in healthy subjects. Investigative ophthalmology & visual science. 2014; 55 (8):4707–13.
- 6. Duan A, Bedggood PA, Metha AB, Bui BV. Reactivity in the human retinal microvasculature measured during acute gas breathing provocations. Scientific reports. 2017; 7(1):2113.
- Werkmeister RM, Palkovits S, Told R, Gröschl M, Leitgeb RA, Garhöfer G, et al. Response of retinal blood flow to systemic hyperoxia as measured with dual-beam bidirectional Doppler Fourier-domain optical coherence tomography. PLoS One. 2012; 7(9):e45876.
- 8. Puro DG. Retinovascular physiology and pathophysiology: new experimental approach/new insights. Progress in retinal and eye research. 2012; 31(3):258–70.
- Chapman N, Haimes G, Stanton AV, Thom SA, Hughes AD. Acute effects of oxygen and carbon dioxide on retinal vascular network geometry in hypertensive and normotensive subjects. Clinical science (London, England: 1979). 2000; 99(6):483–8.
- Venkataraman ST, Hudson C, Rachmiel R, Buys YM, Markowitz SN, Fisher JA, et al. Retinal arteriolar vascular reactivity in untreated and progressive primary open-angle glaucoma. Investigative ophthalmology & visual science. 2010; 51(4):2043–50.
- 11. Duan A, Bedggood PA, Bui BV, Metha AB. Evidence of Flicker-Induced Functional Hyperaemia in the Smallest Vessels of the Human Retinal Blood Supply. PLoS One. 2016; 11(9):e0162621.
- 12. Grunwald JE, Riva CE, Brucker AJ, Sinclair SH, Petrig BL. Altered retinal vascular response to 100% oxygen breathing in diabetes mellitus. Ophthalmology. 1984; 91(12):1447–52.
- Patel V, Rassam SM, Chen HC, Kohner EM. Oxygen reactivity in diabetes mellitus: effect of hypertension and hyperglycaemia. Clinical science (London, England: 1979). 1994; 86(6):689–95.
- 14. Trick GL, Edwards P, Desai U, Berkowitz BA. Early supernormal retinal oxygenation response in patients with diabetes. Investigative ophthalmology & visual science. 2006; 47(4):1612–9.
- Lorenzi M, Feke GT, Cagliero E, Pitler L, Schaumberg DA, Berisha F, et al. Retinal haemodynamics in individuals with well-controlled type 1 diabetes. Diabetologia. 2008; 51(2):361–4.
- Fondi K, Wozniak PA, Howorka K, Bata AM, Aschinger GC, Popa-Cherecheanu A, et al. Retinal oxygen extraction in individuals with type 1 diabetes with no or mild diabetic retinopathy. Diabetologia. 2017; 60 (8):1534–40.

- Kashani AH, Chen CL, Gahm JK, Zheng F, Richter GM, Rosenfeld PJ, et al. Optical coherence tomography angiography: A comprehensive review of current methods and clinical applications. Progress in retinal and eye research. 2017; 60:66–100.
- Spaide RF, Fujimoto JG, Waheed NK, Sadda SR, Staurenghi G. Optical coherence tomography angiography. Progress in retinal and eye research. 2018; 64:1–55.
- Ashimatey BS, Green KM, Chu Z, Wang RK, Kashani AH. Impaired Retinal Vascular Reactivity in Diabetic Retinopathy as Assessed by Optical Coherence Tomography Angiography. Investigative ophthalmology & visual science. 2019; 60(7):2468–73.
- Dimitrova G, Chihara E. Implication of Deep-Vascular-Layer Alteration Detected by Optical Coherence Tomography Angiography for the Pathogenesis of Diabetic Retinopathy. Ophthalmologica Journal international d'ophtalmologie International journal of ophthalmology Zeitschrift fur Augenheilkunde. 2019; 241(4):179–82.
- Scarinci F, Picconi F, Giorno P, Boccassini B, De Geronimo D, Varano M, et al. Deep capillary plexus impairment in patients with type 1 diabetes mellitus with no signs of diabetic retinopathy revealed using optical coherence tomography angiography. Acta ophthalmologica. 2018; 96(2):e264–e5.
- 22. Carnevali A, Sacconi R, Corbelli E, Tomasso L, Querques L, Zerbini G, et al. Optical coherence tomography angiography analysis of retinal vascular plexuses and choriocapillaris in patients with type 1 diabetes without diabetic retinopathy. Acta diabetologica. 2017; 54(7):695–702.
- Simonett JM, Scarinci F, Picconi F, Giorno P, De Geronimo D, Di Renzo A, et al. Early microvascular retinal changes in optical coherence tomography angiography in patients with type 1 diabetes mellitus. Acta ophthalmologica. 2017; 95(8):e751–e5.
- 24. Meshi A, Chen KC, You QS, Dans K, Lin T, Bartsch DU, et al. ANATOMICAL AND FUNCTIONAL TESTING IN DIABETIC PATIENTS WITHOUT RETINOPATHY: Results of Optical Coherence Tomography Angiography and Visual Acuity Under Varying Contrast and Luminance Conditions. Retina (Philadelphia, Pa). 2019; 39(10):2022–31.
- Kushner-Lenhoff S, Ashimatey BS, Kashani AH. Retinal Vascular Reactivity as Assessed by Optical Coherence Tomography Angiography. J Vis Exp. 2020(157).
- Yezhuvath US, Lewis-Amezcua K, Varghese R, Xiao G, Lu H. On the assessment of cerebrovascular reactivity using hypercapnia BOLD MRI. NMR in biomedicine. 2009; 22(7):779–86.
- Wang RK, An L, Francis P, Wilson DJ. Depth-resolved imaging of capillary networks in retina and choroid using ultrahigh sensitive optical microangiography. Optics letters. 2010; 35(9):1467–9.
- An L, Qin J, Wang RK. Ultrahigh sensitive optical microangiography for in vivo imaging of microcirculations within human skin tissue beds. Optics express. 2010; 18(8):8220–8.
- 29. Kim AY, Chu Z, Shahidzadeh A, Wang RK, Puliafito CA, Kashani AH. Quantifying Microvascular Density and Morphology in Diabetic Retinopathy Using Spectral-Domain Optical Coherence Tomography Angiography. Investigative ophthalmology & visual science. 2016; 57(9):Oct362–70.
- Koulisis N, Kim AY, Chu Z, Shahidzadeh A, Burkemper B, Olmos de Koo LC, et al. Quantitative microvascular analysis of retinal venous occlusions by spectral domain optical coherence tomography angiography. PLoS One. 2017; 12(4):e0176404.
- Zhang A, Zhang Q, Wang RK. Minimizing projection artifacts for accurate presentation of choroidal neovascularization in OCT micro-angiography. Biomedical optics express. 2015; 6(10):4130–43.
- Zhang Q, Zhang A, Lee CS, Lee AY, Rezaei KA, Roisman L, et al. Projection artifact removal improves visualization and quantitation of macular neovascularization imaged by optical coherence tomography angiography. Ophthalmology Retina. 2017; 1(2):124–36.
- Chu Z, Lin J, Gao C, Xin C, Zhang Q, Chen CL, et al. Quantitative assessment of the retinal microvasculature using optical coherence tomography angiography. Journal of biomedical optics. 2016; 21 (6):66008.
- Reif R, Qin J, An L, Zhi Z, Dziennis S, Wang R. Quantifying optical microangiography images obtained from a spectral domain optical coherence tomography system. International journal of biomedical imaging. 2012; 2012:509783.
- Mizutani M, Kern TS, Lorenzi M. Accelerated death of retinal microvascular cells in human and experimental diabetic retinopathy. The Journal of clinical investigation. 1996; 97(12):2883–90.
- Cogan DG, Toussaint D, Kuwabara T. Retinal vascular patterns. IV. Diabetic retinopathy. Archives of ophthalmology (Chicago, III: 1960). 1961; 66:366–78.
- **37.** Rose K, Flanagan JG, Patel SR, Cheng R, Hudson C. Retinal blood flow and vascular reactivity in chronic smokers. Investigative ophthalmology & visual science. 2014; 55(7):4266–76.