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Interleukin-6 for early diagnosis of neonatal sepsis with premature rupture of the membranes A meta-analysis

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Abstract

Background: Premature rupture of the membranes (PROM) is the principal risk factor for neonatal sepsis. Interleukin-6 (IL-6) has been investigated for early diagnosis of neonatal sepsis, but not for diagnosis of neonatal sepsis with PROM. The objective of this study is to investigate the early diagnostic value of IL-6 for neonatal sepsis with PROM.

Methods: The literature was searched using PubMed, Embase, Cochrane Library, Web of Science, CNKI, Wan Fang, VIP, and CBM databases until March 2018. Each study was evaluated using Quality Assessment of Diagnostic Accuracy Studies tool-2. We used a bivariate diagnostic random-effects model.

Results: The overall pooled sensitivity, specificity, positive likelihood rate, negative likelihood rate, diagnostic odds ratio, and area under the summary receiver operating characteristic curve were 0.85 (95% confidence interval [CI]: 0.81-0.91), 0.88 (95% CI: 0.86-0.91), 9.94 (95% CI: 4.27-23.15), 0.14 (95% CI: 0.06-0.32), 79.26 (95% CI: 23.42-268.26), and 0.9473, respectively, which showed high accuracy in diagnosing neonatal sepsis with PROM. The types of sepsis might be connected with the source of heterogeneity (P = .0351).

Conclusion: IL-6 is therefore a sensitive and specific diagnostic marker for the early diagnosis of neonatal sepsis with PROM.

Abbreviations: 95% CI = 95% confidence interval, DOR = diagnostic odds ratio, IL-6 = interleukin-6, NLR = negative likelihood ratio, PLR = positive likelihood ratio, PROM = premature rupture of the membranes, QUADAS-2 = Quality Assessment of Diagnostic Accuracy Studies tool-2, SROC = summary receiver operating characteristic.

Keywords: interleukin-6, meta-analysis, neonatal sepsis, premature rupture of the membranes

1. Introduction

Premature rupture of the membranes (PROM) is membrane rupture before the onset of uterine contractions; the incidence of PROM is 10 per 100 pregnancies.^[1] Preterm premature PROM (PPROM) is defined as PROM prior to 37 weeks of gestation, which occurs in 3 of 100 pregnancies.^[2] PROM is associated with several neonatal diseases, including neonatal infection, necrotizing enterocolitis, and intraventricular hemorrhage.^[3–5]

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XQ and LZ contributed equally to this study.

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Neonatal sepsis is a serious systemic disease affecting infants in the first month of life.^[6] PPROM is the principal risk factor for neonatal sepsis, which is associated with increased fetal infection morbidity and mortality in the neonatal period.^[7–13] In recent years, although the management of newborns has improved, the incidence of neonatal sepsis is 1 to 10 per 1000 live births.^[14] Neonatal sepsis in PROM is easily misdiagnosed because the early clinical signs are nonspecific and variable.^[15] A positive blood culture is the reference standard for detecting neonatal sepsis with PROM. However, the blood culture results require at least 24 to 72 hours, and reveals positivity in 19.2% of cases, which is lowly sensitive.^[16,17] It has various practical limitations. Routine laboratory tests, such as C-reactive protein (CRP) and procalcitonin (PCT), which mediated the host response to bacterial infection and released in the neonatal blood, have low sensitivity.^[18,19] In addition, evidence indicates that antibiotic prophylaxis applied in 42% of the deliveries showed no beneficial effect on the incidence of neonatal sepsis and even made early diagnosis of neonatal sepsis more difficult.^[20,21] Therefore, early detection of neonatal sepsis with PROM should be developed to reduce the inadvertent use of antibiotics, cost of treatment, and overtreatment.

Interleukin-6 (IL-6) is a pleiotropic cytokine expressed by different cells in response to infections.^[22] Recently, IL-6 has been investigated for its validity in diagnosing neonatal sepsis.^[23] However, there are no large-scale multicenter studies or metaanalyses on IL-6 for the diagnosis of neonatal sepsis with PROM. Therefore, we conducted a meta-analysis to systematically assess all published studies on the diagnostic performance of IL-6 for detecting neonatal sepsis with PROM.

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2. Methods

2.1. Literature search

The study was conducted according to the Preferred Reporting Items for Systematic Reviews and Meta-Analyses criteria.^[24] A computer-aided literature search was performed using PubMed, Embase, the Cochrane Library, Web of Science, CNKI, WanFang, VIP, and CBM databases for relevant citations up to March 2018 without language restrictions. Our search terms were "neonate," "newborn," "infant," "sepsis," "septicemia," "premature rupture of membranes," "PROM," "interleukin-6," and "IL-6." The search strategy was as follows: ("interleukin-6" OR "IL-6") AND ("neonate" OR "newborn" OR "infant") AND ("sepsis" OR "septicemia") AND ("premature rupture of membranes" OR "PROM"). We (XQ and LZ) also manually searched the references of the included studies and relevant reviews.

2.2. Literature selection

Obtained studies from the literature were reviewed independently by 2 investigators (XQ and LZ) to ensure high accuracy. Inclusion criteria for this study were as follows: assessment of the diagnostic accuracy of IL-6 for neonatal sepsis in PROM; reporting on neonatal sepsis with PROM (population); provision of IL-6 in umbilical cord blood and neonatal peripheral blood as the index test and gold standard for blood culture (index test and reference standard); inclusion of sensitivity, specificity, or sufficient information to construct the 2×2 tables (outcome). For studies that were published more than once, only the most recent and comprehensive report was included. Reviews, letters, case reports, animal experiments, systematic reviews, and metaanalyses were excluded from the analysis.

2.3. Data extraction

Two investigators (XQ and LZ) independently extracted data from selected articles. The following data were recorded: author, year of publication, regions, study population, gestational age (week), sample size, specimen, assay method, cut-off value, diagnostic gold standard, type of sepsis, sensitivity, specificity, true positive (TP), false positive (FP), false negative (FN), and true negative (TN) of IL-6. The analyses were based on previous published studies. Therefore, no patient consent, ethical approval, and institutional review board are required.

2.4. Quality assessment

The methodological quality of the included studies was assessed using Quality Assessment of Diagnostic Accuracy Studies tool-2 (QUADAS-2).^[25] Two reviewers (XQ and LZ) independently performed the quality assessment. Four domains (patient selection, index test, reference standard, and flow and timing) were evaluated for the risk of bias, and 3 domains (patient selection, index test, and reference standard) were evaluated based on applicability. Participant spectrum bias and selection bias were determined. Information bias existed for the index test. Partial verification bias, differential verification bias, and disease progression bias were related to the reference standard. Excluded data bias existed for flow and timing.^[25] Signaling questions were asked to help estimate the risk of bias.^[25] In case of unresolved disagreement, a third reviewer (YT) was consulted.

2.5. Statistical analysis

Meta-Disc software (version 1.4) was used to perform the statistical analysis.^[26] The Spearman correlation analysis was used to assess the threshold effect, and P < .05 was used to indicate a significant threshold effect. We further analyzed the influence of the cut-off used in each study by subgroup analysis. A bivariate random effects model was used to calculate the pooled sensitivity, specificity, positive likelihood ratio (PLR), negative likelihood ratio (NLR), and diagnostic odds ratio (DOR).^[27-28] Based on sensitivity and specificity, we further constructed the summary receiver operating characteristic (SROC) curve. The area under the curve (AUC) was also calculated to evaluate the diagnostic performance of IP-10; a value close to 1 indicated that IP-10 is a good diagnostic tool.^[29-31] Heterogeneity among the included studies was evaluated using the Cochrane Q test and I^2 statistic. ^[32] Additionally, we conducted a meta-regression analysis to explore possible sources of heterogeneity. We used Deeks' test to assess publication bias. Stata software (version 14.0) was used to draw funnel plots for analysis of publication bias.

3. Results

3.1. Literature research

As shown in Figure 1, 682 literature citations were identified from database searches (PubMed: 147; Embase: 109; Cochrane Library: 9; Web of Science: 278; CNKI: 25; WanFang: 56; VIP: 8; CBM: 50). First, 216 duplicate studies were removed. Then, by reviewing the titles and abstracts, 422 articles that did not meet the inclusion criteria were excluded. After screening the full texts of 36 articles, 14 articles included other infections (pneumonia, necrotizing enterocolitis, etc.) were excluded, 18 articles which did not report related data (sensitivity, specificity, etc.) were excluded, and 2 articles detecting vaginal secretion and 2 articles detecting maternal blood were excluded. Ultimately, 8 articles^[7–11,2,12–13] including 9 trials met the inclusion criteria and were included in this meta-analysis.

3.2. Characteristics of the included studies

The characteristics of 8 eligible articles totaling 9 clinical trials, including 9 trials, are listed in Table 1.^[7–11,2,12–13] Seven trials were published in English while 2 were in Chinese.^[7,8] The publication year ranged from 2005 to 2017. Three trials were in Europe, and 6 trials were in Asia. There were 694 participants including 752 samples involved in this meta-analysis. The sensitivity, specificity, TP, FP, FN, and TN of IL-6 in each trial are shown in Table 1. Eight (89%) of studies included neonatal sepsis with PPROM and healthy neonates with PPROM, and only one study included neonatal sepsis with PROM and healthy neonates with PROM.^[8] Only one study did not report patient's age.^[11] Patients' age, number of patients, assay method, specimen, cut-off value, type of sepsis sensitivity, specificity, TP, FP, FN, and TN of IL-6 in each trial are shown in Table 1.

3.3. Quality assessment

The QUADAS-2 was used to assess the methodological quality of the included articles (Fig. 2). Patient selection showed low bias in 7studies and unclear bias in one study. All studies had low bias in their index tests. Seven studies were allocated as having low bias in their reference standard, and one study showed unclear bias. Six studies had low bias in flow and timing, and 2 studies showed



unclear bias. Regarding applicability concerns, 5 studies showed low concerns in patient selection, and 3 studies showed unclear concerns. All studies had applicability concerns as low concerns in the index tests; 2 studies were allocated as low concern in the reference standard, one study showed high concerns, and 5 studies showed unclear concern.

3.4. Pooled analysis

The Spearman correlation coefficient was -0.613, and the *P*-value was .079, and there was no threshold effect; the DOR

also showed a nonthreshold effect (Cochran-Q = 26.69, $I^2 = 70\%$, P = .0008). We used a random-effects model to detect IL-6 for neonatal sepsis with PROM. The sensitivity ranged from 0.47 to 0.95 (pooled sensitivity: 0.87, 95% CI: 0.81–0.91), whereas specificity ranged from 0.63 to 1.00 (pooled specificity: 0.88, 95% CI: 0.86–0.91) (Figs. 3 and 4). The pooled PLR of IL-6 was 9.94 (95% CI: 4.27–23.15), and the pooled NLR was 0.14 (95% CI: 0.06–0.32). In addition, the pooled DOR was 79.26 (95% CI: 23.42–268.26). DOR, which could be used to assess the accuracy of the test, showed that the discriminate effect in our study was good. For all aforementioned effect sizes, significant hetero-

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Author				Gestational age,			Assay			Type	Sensitivity	Specificity				
	Year	Region	Study population	weeks	z	Specimen	method	Cut-off	Sepsis diagnosis	of sepsis	(%)	(%)	đ	£	FN	Z
Song J ^[7]	2014	China	Case: neonatal sepsis with PPROM; control:	31.3±2.1	58	Neonatal peripheral blood	ELISA	7.8 ng/L	Culture, clinical, I	Early/late-onset	93	96	28	-	2	27
1			healthy neonates with PPROM						aboratory evaluation							
rang CY ^{t8]}	2017	China	Case: neonatal sepsis with PROM; control:	39.6 ± 2.9	187	Umbilical cord blood	ELISA	62.62 pg/mL	Culture, clinical,	Early onset	76.2	91	16	15	2	151
			healthy neonates with PROM						laboratory evaluation							
Hatzidaki Ea ^[9]	2005	Greece	Case: neonatal sepsis with PPROM; control:	32.6 ± 2.9	58	Umbilical cord blood	ELISA	108.5 pg/mL	Culture, clinical,	Early/late-onset	95	100	19	0	-	88
			healthy neonates with PPROM						laboratory evaluation							
Hatzidaki Eb ^[9]	2005	Greece	Case: neonatal sepsis with PPROM; control:	32.6 ± 2.9	58	Neonatal peripheral blood	ELISA	55 pg/mL	Culture, clinical,	Early/late-onset	06	97.4	18	-	2	37
			healthy neonates with PPROM						laboratory evaluation							
Cetin O ^[10]	2014	Turkey	Case: neonatal sepsis with PPROM; control:	32.4 ± 3.32	40	Umbilical cord blood	ELISA	11 pg/mL	Culture, clinical,	Early onset	06	63.3	6	÷	-	19
			healthy neonates with PPROM						laboratory evaluation							
Cobo T ^[11]	2013	Spain	Case: neonatal sepsis with PPROM; control:	unclear	176	Umbilical cord blood	ELISA	38 pg/mL	Culture, clinical,	Early onset	83	82	10	30	2	134
			healthy neonates with PPROM						laboratory evaluation							
Canpolat FE ^[2]	2011	Turkey	Case: neonatal sepsis with PPROM; control:	31.8	74	Umbilical cord blood	ELISA	7.6 pg/mL	Culture, clinical,	Early onset	93	96.7	30		2	4
			healthy neonates with PPROM						laboratory evaluation							
Sharehbaghi	2008	Iran	Case: neonatal sepsis with PPROM; control:	30.37 ± 2.38	45	Umbilical cord blood	ELISA	20 pg/mL	Culture, clinical,	Early onset	46	85	œ	4	6	24
MM ^[12]			healthy neonates with PPROM						laboratory evaluation							
ruan SW ⁽¹³⁾	2012	China	Case: neonatal sepsis with PPROM; control:	31.5 ± 2.2	56	Neonatal peripheral blood	ELISA	7.9 pg/mL	Culture, clinical,	Early/late-onset	94	97	28	-	2	25
			healthy neonates with PPROM						laboratory evaluation							
ELISA = enzyme-l	-linked im	Imunosorbe	ent assay, FN = false negative, FP = false positive,	TN = true negative,]	P = true	e positive.										

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geneities were observed ($I^2 > 50\%$). From the SROC in Figure 5, the AUC was 0.9473 with a standard error of 0.0314, which represented perfect discriminatory ability.

3.5. Sensitivity analysis

Excluding the study that showed high concerns in reference standard,^[9] the pooled sensitivity, specificity, and AUC of IL-6 for neonatal sepsis with PROM were 0.85 (95% CI: 0.78–0.90), 0.87 (95% CI: 0.84–0.90), and 0.9410, respectively, which showed high stability.

3.6. Meta-regression analysis

Meta-regression analysis was conducted to investigate potential sources of heterogeneity (Table 2). The specimen and cut-off were not significant sources of heterogeneity (P=.6932 and 0.9540, respectively). However, the types of sepsis might be connected with the source of heterogeneity. The diagnostic accuracy of IL-6 in early onset neonatal sepsis was 16.6 times higher than that in early/late-onset neonatal sepsis (RDOR=16.6, 95% CI: 1.31–209.61; P=.0351).

3.7. Subgroup analysis

Regarding the different specimens, a total of 580 samples in umbilical cord blood and 172 samples in neonatal peripheral blood were detected. The sensitivity and specificity in neonatal peripheral blood was higher than in umbilical cord blood (93% vs 83%, 97% vs 87%). Besides, the PLR of IL-6 in neonatal peripheral blood was higher than that in umbilical cord blood (27.86 vs 6.27). The NLR, DOR, and AUC are shown in Table 3.

When the cut-off of IL-6 was more than 30 pg/mL, the specificity was the same with IL-6 < 30 pg/mL (87%), and the sensitivity was comparable (89% and 88%). The PLR, NLR, DOR, and AUC are shown in Table 3.

Regarding types of sepsis, a total of 522 samples with early onset neonatal sepsis and 230 samples with early/late-onset neonatal sepsis were identified. The early/late-onset neonatal sepsis had a much higher sensitivity (93% vs 80%) and specificity (98% vs 86%) than early onset neonatal sepsis. Besides, the PLR of IL-6 in early/late-onset neonatal sepsis was higher than in early onset neonatal sepsis (31.84 vs 5.12). The NLR of IL-6 in early/ late-onset neonatal sepsis was lower than in early-onset neonatal sepsis (0.08 vs 0.22). The DOR and AUC are shown in Table 3.

3.8. Publication bias

We used Deek's funnel plot asymmetry test to assess publication bias of the diagnostic tests. The results showed that the *t*-value obtained from the funnel plot was -2.18, and the *P*-value was .07, indicating no striking publication bias in this meta-analysis.

4. Discussion

Neonatal sepsis with PROM is a serious systemic disease. The reference standard for the diagnosis of neonatal sepsis with PROM is a positive blood culture, which is time-consuming, less sensitive, and has a high false-negative rate.^[6,16,17] Besides, neonatal sepsis does not occur in every PROM case; it is thus important to accurately predict neonatal sepsis with PROM, so that active management instead of expectant management can be undertaken to reduce the likelihood of the inadvertent use of antibiotics, cost of treatment, and overtreatment.^[7,33]



Figure 2. Risk of bias and applicability concerns of included studies.



Figure 3. The forest plots of the pooled sensitivity of IL-6 to diagnose neonatal sepsis in PROM. CI = confidence interval, IL-6=interleukin-6, PROM = premature rupture of the membranes.



Figure 4. The forest plots of the pooled specificity of IL-6 to diagnose neonatal sepsis in PROM. CI = confidence interval, IL-6=interleukin-6, PROM = premature rupture of the membranes.



Figure 5. The summary receiver operating characteristic curve for assessment of IL-6 to diagnose neonatal sepsis in PROM. AUC=area under curve, SROC=summary receiver operating characteristic, IL-6=interleukin-6, PROM = premature rupture of the membranes, SE=standard error.

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Meta-regression analysis of the effects of specimen, cut-off, and type of sepsis.

C	ovariates	Coefficient	Standard error	P value RDOR		(95%CI)	
3							
	Cut-off	-0.077	1.2549	.954	0.93	(0.03; 30.17)	
	Specimen	-1.101	2.5965	.6932	0.33	(0.00; 449.12)	
	Type of sepsis	3.638	2.4823	.2166	38.03	(0.04; 37423.83)	
2							
	Specimen	-1.077	2.2803	.6565	0.34	(0.00; 119.61)	
	Type of sepsis	3.672	2.2033	.1565	39.32	(0.14; 11334.04)	
1							
	Type of sepsis	2.809	1.0363	.0351	16.6	(1.31; 209.61)	

Laboratory tests, such as CRP and PCT, are not sufficient for the early diagnosis of sepsis with PROM. The sensitivity and specificity of the CRP for the early diagnosis of neonatal sepsis with PROM were <60% and 80%, respectively.^[34,35] CRP often begins increasing after 12 to 24 hours of neonatal infection and peaks later, which is not conducive for the early diagnosis of neonatal sepsis with PROM. The sensitivity and specificity of PCT for the early diagnosis of neonatal sepsis with PROM are 53% and 45%, respectively. Besides, physiological PCT can increase from 1 to 10 ng/mL in normal neonates within 48 hours after delivery.^[35,36] IL-6 is a pleiotropic cytokine that is expressed by different cells in response to infections.^[22] Furthermore, IL-6 can be detected rapidly by flow cytometry with minimal blood volumes (0.05 mL). In recent years, IL-6 has been investigated for its validity in the early diagnosis of neonatal sepsis with PROM.^[2,7–13]

A meta-analysis by Shahkar et al^[23] showed that the sensitivity and specificity of IL-6 for neonatal sepsis were 79% and 84% in neonates without PROM. However, in our study, we first conducted a systematic review and meta-analysis to investigate the early diagnostic value of IL-6 as a potential biomarker for neonatal sepsis with PROM. The pooled sensitivity of IL-6 for the diagnosis of neonatal sepsis with PROM was 0.87, the pooled specificity was 0.88, the missed diagnosis rate was 0.13, and the misdiagnosis rate was 0.12, which showed that the diagnostic efficiency was high. A PLR of 9.94 suggests that neonates with sepsis and PROM have a 9.94-fold higher chance of being IL-6positive than neonates with PROM. This ratio suggests a potential role for IL-6 for neonatal sepsis with PROM. The DOR is a comprehensive evaluation of the diagnostic value of the index test; the pooled DOR was 61.81, which showed a good discriminating effect. The overall accuracy of IL-6 for the diagnosis of neonatal sepsis with PROM was favorable. These results indicate that IL-6 is a helpful biomarker for the early diagnosis of neonatal sepsis with PROM.

Different specimens may have different performance to IL-6. Generally speaking, umbilical cord blood and neonatal peripheral blood are neonatal blood. Although the specimens were not significant sources of heterogeneity (P = .6565), we found that the sensitivity and specificity in neonatal peripheral blood was higher than in umbilical cord blood. The small number of included studies might lead to this result. Therefore, further meta-analysis including more studies is needed to explain the reason.

With cut-off \geq 30 pg/mL, the specificity was the same with cut-off <30 pg/mL. Although the cut-off range was 7.6 to 108.5 pg/mL, it was not a significant source of heterogeneity (P = .954) and showed similar diagnostic value with the overall diagnostic accuracy.

In this meta-analysis, meta-regression proved that the diagnostic accuracy of IL-6 test for predicting neonatal sepsis with PROM was affected by the types of sepsis (P=.0351). Furthermore, the results of the subgroup analysis showed that the AUC of early/late-onset neonatal sepsis and early-onset neonatal sepsis was 0.9817 and 0.9098, respectively. The type of sepsis was one cause of the heterogeneity, because of different pathogens and degrees in inflammatory response between early/late-onset neonatal sepsis and early-onset neonatal sepsis.

Our meta-analysis had several limitations. First, IL-6 tests are usually detected along with other cytokines, but we did not address reliability with respect to combined analyses of IL-6 and

Table 3

Subgrou	p analysis of study	/ specim	en, cut-off, and t	ypes of sepsis.				
Subgroup		Studies	Sensitivity (95%)	Specificity (95%)	PLR (95%)	NLR (95%)	DOR	AUC
Specimen	Umbilical cord blood	6	0.82 (0.74, 0.89)	0.87 (0.84, 0.90)	6.27 (2.83, 13.88)	0.19 (0.07, 0.51)	40.40 (10.28, 158.69)	0.9227
	Neonatal blood	3	0.93 (0.84, 0.97)	0.97 (0.91, 0.99)	27.86 (9.14, 84.91)	0.08 (0.04, 0.17)	353.23 (85.29, 1462.87)	0.9871
Cut-off	<30 pg/mL	5	0.87 (0.79, 0.92)	0.88 (0.82, 0.93)	10.09 (1.92, 53.05)	0.13 (0.03, 0.61)	81.13 (9.87, 666.66)	0.9562
	\geq 30 pg/mL	4	0.87 (0.79, 0.94)	0.89 (0.85, 0.91)	11.50 (3.40, 38.85)	0.17 (0.10, 0.31)	76.02 (16.80, 343.88)	0.8871
Type of sepsis	Early onset	5	0.80 (0.70, 0.87)	0.86 (0.82, 0.89)	5.12 (2.61, 10.03)	0.22 (0.08, 0.61)	26.80 (7.51, 95.71)	0.9098
	Early/late-onset	4	0.93 (0.84, 0.97)	0.98 (0.93, 1.00)	31.86 (11.34, 89.53)	0.08 (0.04, 0.15)	417.63 (113.61, 1535.16)	0.9817

AUC = area under the curve, DOR = diagnostic odds ratio, NLR = negative likelihood ratio, PLR = positive likelihood ratio.

other cytokines. Second, some studies included individuals with neonatal sepsis and PROM after antibiotic treatment, while some were without antibiotic treatment. This may influence the diagnosis. Third, the heterogeneity was relatively high. Although different specimens and cut-off in meta-regression analysis are not significant sources of heterogeneity (P > .05), they can also increase the heterogeneity and reduce the generalizability of the overall performance. Third, some studies were rated as high risk, and this may have affected the pooled effects. Finally, publication bias was a concern. Because of the linguistic abilities of our team, only studies written in English or Chinese were included. The true accuracy of IL-6 tests for neonatal sepsis and PROM may be lower than our report.

5. Conclusions

The reference standard for the diagnosis of sepsis with PROM is a positive blood culture, which has various practical limitations. This meta-analysis shows that IL-6 might be a sensitive and specific diagnostic marker for the early diagnosis of neonatal sepsis with PROM. The diagnostic accuracy of IL-6 is not influenced by types of neonatal sepsis. To provide optimum diagnostic value, IL-6 should detect early onset and late-onset neonatal sepsis separately. Furthermore, large, multicentre, and prospective studies are warranted to support our findings.

Author contributions

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- Validation: Xia Qiu, Li Zhang, Dezhi Mu.
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References

- [1] Kayem G, Batteux F, Girard N, et al. Predictive value of vaginal IL-6 and TNF a bedside test repeated until delivery for the prediction of maternalfetal infection in cases of premature rupture of membranes. Eur J Obst Gvnecol Reprod Biol 2017;211:8–14.
- [2] Canpolat FE, Yiğit S, Korkmaz A, et al. Procalcitonin versus CRP as an early indicator of fetal infection in preterm premature rupture of membranes. Turk J Pediatr 2011;53:180–6.
- [3] Gulati S, Agrawal S, Raghunandan C, et al. Maternal serum interleukin-6 and its association with clinicopathological infectious morbidity in preterm premature rupture of membranes: a prospective cohort study. J Matern Fetal Neonatal Med 2011;25:1428–32.
- [4] Bigelow CA, Factor SH, Miller M, et al. Pilot randomized controlled trial to evaluate the impact of bed rest on maternal and fetal outcomes in women with preterm premature rupture of the membranes. Am J Perinatol 2016;33:356–63.
- [5] Pasquier JC, Picaud JC, Rabilloud M, et al. Neonatal outcomes after elective delivery management of preterm premature rupture of the membranes before 34 weeks' gestation (DOMINOS study). Eur J Obstet Gynecol Reprod Biol 2009;143:18–23.
- [6] Ganesan P, Shanmugam P, Sattar SB, et al. Evaluation of IL-6, CRP and hs-CRP as early markers of neonatal sepsis. J Clin Diagn Res 2016;10: DC13–7.

- [7] Song J. Application values of CRP, PCT, and IL-6 in early diagnosis of sepsis among neonates with premature rupture of membrane. Matern Child Health Care China 2014;29:1045–7.
- [8] Yang CY, Zhang BQ, Chen HQ. The relation between early-onset neonatal sepsis and the level of interleukin 6, C-reactive protein in cord blood of newborns with premature rupture of membrane. Chin J Neonatol 2017;32:110–4.
- [9] Hatzidaki E, Gourgiotis D, Manoura A, et al. Interleukin-6 in preterm premature rupture of membranes as an indicator of neonatal outcome. Acta Obstet Gynecol Scand 2005;84:632–8.
- [10] Cetin O, Dokurel Cetin I, Uludag S, et al. Serial ultrasonographic examination of the fetal thymus in the prediction of early neonatal sepsis in preterm premature rupture of membranes. Gynecol Obstet Invest 2014;78:201–7.
- [11] Cobo T, Kacerovsky M, Andrys C, et al. Umbilical cord blood IL-6 as predictor of early-onset neonatal sepsis in women with preterm prelabour rupture of membranes. PLoS One 2013;8:e69341.
- [12] Gharehbaghi MM, Peirovifar A, Gharehbaghi PM. Comparison of umbilical cord interleukin-6 in preterm infants with premature rupture of membranes and intact membranes. Saudi Med J 2008;29:224–8.
- [13] Yuan SW, Zhang YJ, Zheng WD, et al. Diagnostic value of procalcitonin, CRP and IL-6 in fetal sepsis with preterm premature rupture of membranes. Turk J Pediatr 2012;9:114–6.
- [14] Andaluz-Ojeda D, Iglesias V, Bobillo F, et al. Early natural killer cell counts in blood predict mortality in severe sepsis. Crit Care 2011;15: R243.
- [15] Shi J, Tang J, Chen D. Meta-analysis of diagnostic accuracy of neutrophil CD64 for neonatal sepsis. Ital J Pediatr 2016;42:57.
- [16] Jyothi P, Basavaraj MC, Basavaraj PV. Bacteriological profile of neonatal septicemia and antibiotic susceptibility pattern of the isolates. J Nat Sci Biol Med 2013;4:306–9.
- [17] Benitz WE. Adjunct laboratory tests in the diagnosis of early-onset neonatal sepsis. Clin Perinatol 2010;37:421–38.
- [18] Su H, Chang SS, Han CM, et al. Inflammatory markers in cord blood or maternal serum for early detection of neonatal sepsis-a systemic review and meta-analysis. J Perinatol 2014;34:268–74.
- [19] Zhou M, Cheng S, Yu J, et al. Interleukin-8 for diagnosis of neonatal sepsis: a meta-analysis. PLoS One 2015;10:e0127170.
- [20] Wójkowska-Mach J, Borszewska-Kornacka M, Domańska J, et al. Early onset infections of very-low-birth-weight infants in Polish neonatal intensive care units. Pediatr Infect Dis J 2012;31:691–5.
- [21] Ganesan P, Shanmugam P, Sattar SB, et al. Evaluation of IL-6, CRP and hs-CRP as early markers of neonatal sepsis. J Clin Diagn Res 2016;10:13–7.
- [22] Mihara M, Hashizume M, Yoshida H, et al. IL-6/IL-6 receptor system and its role in physiological and pathological conditions. Clin Sci 2012;122:143–59.
- [23] Shahkar L, Keshtkar A, Mirfazeli A, et al. The role of IL-6 for predicting neonatal sepsis: a systematic review and meta-analysis. Iran J Pediatr 2011;21:411–7.
- [24] Moher D, Liberati A, Tetzlaff J, et al. Preferred reporting items for systematic reviews and meta-analyses: the PRISMA statement. PLoS Med 2009;6:e1000097.
- [25] Whiting PF, Rutjes AW, Westwood ME, et al. QUADAS-2 Group-QUADAS-2: a revised tool for the quality assessment of diagnostic accuracy studies. Ann Intern Med 2011;155:529–36.
- [26] Zamora J, Abraira V, Muriel A, et al. Meta-DiSc: a software for metaanalysis of test accuracy data. BMC Med Res Methodol 2006;6:31.
- [27] Higgins JP, Thompson SG, Deeks JJ, et al. Measuring inconsistency in meta-analyses. Br Med J 2003;327:557.
- [28] Devillé WL, Buntinx F, Bouter LM, et al. Conducting systematic reviews of diagnostic studies: didactic guidelines. BMC Med Res Methodol 2002;2:9.
- [29] Moses LE, Shapiro D, Littenberg B. Combining independent studies of a diagnostic test into a summary ROC curve: data-analytic approaches and some additional considerations. Stat Med 1993;12:1293–316.
- [30] Vamvakas EC. Meta-analyses of studies of the diagnostic accuracy of laboratory tests: a review of the concepts and methods. Arch Pathol Lab Med 1998;122:675–86.
- [31] Arends LR, Hamza TH, van Houwelingen JC, et al. Bivariate random effects meta-analysis of ROC curves. Med Decis Making 2008;28:621–38.
- [32] Chappell FM, Raab GM, Wardlaw JM. When are summary ROC curves appropriate for diagnostic meta-analyses? Stat Med 2009;28:2653–68.
- [33] van Herk W, Stocker M, van Rossum AM. Recognising early onset neonatal sepsis: an essential step in appropriate antimicrobial use. J Infect 2016;72:77–82.

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- [34] Popowski T, Goffinet F, Maillard F, et al. Maternal markers for detecting early-onset neonatal infection and chorioamnionitis in cases of premature rupture of membranes at or after 34 weeks of gestation: a two-center prospective study. BMC Preg Childbirth 2011; 11:26.
- [35] Torbe A. Maternal plasma procalcitonin concentrations in pregnancy complicated by preterm premature rupture of membranes. Mediators Inflamm 2007;2007:35782.
- [36] Hu J, DU PF, Bei DD. Diagnostic value of interleukin 6 for neonatal sepsis: a meta-analysis. Chin J Contemp Pediatr 2015;17:1176–82.