

Case Report

MPGN Type 3 Associated with Pemphigus Herpetiformis Mimicking PGNMID and Dermatitis Herpetiformis

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Keywords

MPGN type 3 · Proliferative glomerulonephritis with monoclonal IgG deposits · Pemphigus herpetiformis · Dermatitis herpetiformis

Abstract

A 45-year-old man suffering from dermal blistering disease with proteinuria and hematuria underwent renal biopsy. The renal biopsy specimen suggested proliferative glomerulonephritis with monoclonal IgG deposits under routine light, immunofluorescence and electron microscopy. The staining for IgG subclasses (IgG1 and IgG2) and κ/λ light chain indicated secondary

immune complex type MPGN type 3. The patient had been diagnosed as having dermatitis herpetiformis (DH), a phenotype of gluten hypersensitivity prior to the appearance of the renal abnormality. Although common autoantibodies might be related to the pathogenesis of disorders in the skin and kidney, DH is mainly driven by IgA autoantibody, while MPGN is induced by IgG immune complexes. IgA was not observed in the glomeruli by immunofluorescence. Neither the examination for DH specific autoantibodies nor HLA-DQB1 genotype supported the diagnosis of DH. Reassessment of the skin biopsy record revealed that the blister was localized in the epidermis, suggesting pemphigus herpetiformis by IgG class anti-epidermal autoantibody, which also affected the renal disorder.

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Introduction

While immunoglobulins play a main role in humoral immunity, they sometimes induce various disorders involving many organs. Kidney and skin are also targets of immunoglobulin-associated disorders [1]. We present a case of a 45-year-old male with MPGN type 3 with blistering skin disease. He was previously diagnosed as having dermatitis herpetiformis (DH) by skin biopsy. It was speculated that the skin and renal disorders were due to a common autoantibody, but there was a discrepancy in the immunoglobulin class of the pathogenic autoantibodies between DH and MPGN. Reassessment of the histopathological findings in skin biopsy corrected the patient's diagnosis to pemphigus herpetiformis. Thus, this patient showed complex phenotypes in the kidney and skin that made it difficult to provide differential diagnoses.

Case Report

A 45-year-old Japanese male was found to have microscopic hematuria and proteinuria at the annual medical checkup. He was referred to the Juntendo University Shizuoka Hospital for further examination, by a local medical doctor. He had been working in general good health. Initial examination was normal but skin erythema on the trunk and extremities was treated in the dermatology clinic.

The patient was well-developed and well-nourished. His blood pressure in the sitting position was 138/76 mm Hg and pulse rate 66 beats per minute. His skin tanned well and was free of vesicles. Urinalysis showed a 2+ dipstick test for protein, 1+ for occult blood and was negative for sugar. Urinary sediments showed 10–19 red blood cells/HPF in the urinary sediments. The urinary protein-to-creatinine ratio was 1.2 g/gCr. Laboratory tests revealed serum urea nitrogen 12.1 mg/dL (normal values: 9–21 mg/dL), creatinine 0.68 mg/dL (0.4–0.9 mg/dL), uric acid 7.5 mg/dL (3.8–7.5 mg/dL) and an estimated glomerular filtration rate (eGFR) of 99.2 mL/min/1.73 m² (based upon the serum creatinine). The hematologic evaluations, liver function, serum cholesterol and serological tests were within normal ranges. Serological markers for syphilis, hepatitis B virus and hepatitis C virus were negative. Serum immunoelectrophoresis showed normal IgG, IgA and IgM precipitin arcs. Tests for anti-nuclear antibodies, anti-DNA antibodies, PR3-ANCA, MPO-ANCA and anti-glomerular basement membrane (anti-GBM) antibodies were negative. The complement 3 (C3) and C4 levels were within

normal ranges (Table 1). There were no findings suggestive of malignancy in the chest and abdominal X-ray and CT.

During renal biopsy, three cores were obtained by ultrasound guided needle biopsy. The renal cortex was approximately 70%. The LM showed 30 glomeruli. Three glomeruli were globally sclerosed and focal tubular atrophy with interstitial fibrosis and inflammatory cell infiltration were observed around them (Fig. 1a). The glomeruli showed moderate mesangial widening and thickening of the capillary walls. Lobular accentuation with segmental nodular formation due to an increase in cells and matrix were also observed (Fig. 1b). The majority of glomerular capillary loops showed double contours (Fig. 1c). In immunofluorescence, IgG, C3 and C1q were strongly positive in the granular deposits along the glomerular capillary walls and mesangial areas (Fig. 1d, g, h). IgA, IgM and fibrinogen revealed weak or negative signals (Fig. 1e, f). In electron microscopy, foot processes of glomerular epithelial cells were effaced (Fig. 2a). Massive electron-dense deposits (EDDs) were observed in the glomerular mesangial areas, basement membranes, subendothelial areas and subepithelial areas (Fig. 2b). Mesangial matrices and the numbers of cells were increased moderately. In the capillary loops, the GBM showed segmental doubling with EDDs located between the GBM. No inflammatory cells were observed in the glomerular capillary loops (Fig. 2c). The EDDs exhibited immune complex-like granular structures (Fig. 2d).

The findings are summarized as the Burkholder variant of MPGN type 3 [2] and considered in the differential diagnosis of proliferative glomerulonephritis with monoclonal IgG deposits (PGNMID). Immunofluorescence for anti-IgG1 and IgG2 antibodies were positive in both the glomerular capillary loops and the mesangial areas (Fig. 3a, b). Staining for anti-IgG3 and IgG4 antibodies was negative (data not shown). Immunohistochemistry revealed that the IgG deposited within the glomeruli were both κ and λ positive, suggesting that polymeric IgG was deposited in the glomeruli (Fig. 3c, d).

Since adult idiopathic MPGN is rare and the current pathological findings were atypical for primary MPGN, the patient's skin lesion, which had been treated for a long time, was examined in detail [3]. Twenty years ago, the patient had suffered from many blisters accompanied by severe itching. He was diagnosed as having DH by skin biopsy and treated with dapsons (diaphenylsulfone) 75 mg/day for 2 years. Seven years later, the DH recurred, and the patient was again treated with the same dose of dapsons. Since he responded well to the treatment, the dose of the drug was decreased to 50 mg/day and the treatment continued. During our clinical observation, the patient accidentally stopped taking dapsons. He noticed the onset of skin blisters and took a photo. The skin lesions quickly diminished after reinitiation of the dapsons treatment.

Since DH is recognized as a phenotype of gluten hypersensitivity, including celiac disease [4], and has a close relationship with specific HLA-DQB1 genotype covering serotype DQ2 and DQ8 in Europe and the United States [5], HLA-DQB1 typing was performed for the patient. His allele (1) was 06: 02 and allele (2) was 06: 04, corresponding to serotype DQ6(1)/DQ6(1), which is commonly observed among the Japanese community and is distinct from the gluten hypersensitivity-related allele [6]. Neither anti-tissue transglutaminase IgA antibody (anti-tTG IgA) nor anti-endomysial antibody (anti-EMA), autoantibodies specific to celiac disease, were detected in the patient's serum [7]. While IgG immune complex was predominantly observed in the renal biopsy specimen, there was little evidence indicating that the patient was affected with DH. In his past medical records, skin biopsy specimens were not examined by

immunofluorescence but only by light microscopy. The light microscopy specimen revealed that the blister was localized to the epidermis with accompanying edema. These findings did not suggest DH, but rather herpetiformis pemphigoid related to IgG autoantibodies against dermal antigens [8]. Histologically, DH exhibits subepidermal papillary blisters with infiltration of numerous neutrophils and variable eosinophils [4].

The patient was treated with the angiotensin II receptor blocker (ARB), losartan 50 mg, daily and dapsone, which is commonly used to treat pemphigus herpetiformis [9]. The treatments decreased the urinary protein excretion to below 0.5 g/gCr and did not impair renal function. The blood pressure remained below 130/80 mm Hg. The skin problems were also well-controlled.

Discussion

Prior to performing renal biopsy, we anticipated mild glomerulonephritis including IgA nephropathy (IgAN), based upon the clinical findings. In IgAN patients, recurrent episodes of visible hematuria or asymptomatic invisible hematuria, with or without proteinuria, are commonly observed [10]. IgAN, the most common form of glomerulonephritis and the major cause of end-stage kidney disease worldwide, is characterized by the presence of IgA1 deposits, often accompanied by C3 and IgG in glomerular mesangial areas [10].

Light microscopic findings suggested MPGN, and immunofluorescence revealed the dominant presence of IgG, C3 and C1q and subordinate IgA and IgM deposits in the glomeruli, supporting immunoglobulin-positive MPGN [11]. IgAN and C3 glomerulopathy were excluded from the potential candidates. Electron microscopic findings showed MPGN type 3 and the structure of the EDDs were granular, suggesting an immune complex type [12].

The pathologist suggested MPGN type 3 or PGNMID as the differential diagnosis and recommended assessment of the clonality of the deposited immunoglobulins in the glomeruli. PGNMID was reported by Nasr et al. [13] in 2009 as a proliferative glomerulonephritis with monoclonal IgG deposits, which is a form of renal involvement by monoclonal gammopathy, mimicking immune complex type glomerulonephritis. The histologic patterns are predominantly MPGN (57%), endocapillary proliferative (35%) with membranous features. Electron microscopy revealed granular, non-organized EDDs, and immunofluorescence microscopy demonstrated both single light-chain and heavy-chain subtypes [13]. While monoclonal IgG deposits in the glomeruli, M-proteins are not always found in the sera (29.7%) [13]. In our patient, PGNMID was excluded since the immunoglobulins were polyclonal (Fig. 3).

Based upon the current findings, infection, malignancy or drug related immunoglobulin positive MPGN was not suggestive. Since most cases of MPGN are secondary, for which an underlying cause can be found, the patient's skin disease, DH, was highlighted [12]. In the western community, gluten hypersensitivity is a well-known allergic disease [7]. Multiple phenotypes, including DH, celiac disease, mild depression, attention deficit/hyperactivity disorder (ADHD)-like behavior, abdominal pain, diarrhea, constipation, headache, bone and joint pain and chronic fatigue, are recognized as the gluten hypersensitivity family [7, 14].

DH is an IgA-mediated cutaneous disease, with IgA deposits appearing in a granular pattern at the top of the dermal papilla in the sublamina densa area of the basement membrane [15]. The patients with celiac disease associated with MPGN have been reported [16, 17]. It is

well known that celiac disease is tightly related to the specific human leukocyte antigen (HLA) class II genes, DQ2 and DQ8 [7]. Approximately 95% of celiac disease patients express HLA-DQ2, and the remaining patients have DQ8 among Caucasian patients [18]. Among the Japanese community, these HLA-loci are not common and are also rare among patients with celiac disease [19]. Nutritional disturbance due to malabsorption syndrome is not severe, and these patients exhibit good responses to treatment [20]. The reports of the characteristics of Japanese DH patients show that very few patients have HLA-DQ2 or 8 [21]. Our patient expressed HLA-DQ6(1)/DQ6(1) which is commonly observed among the Japanese population. Gluten is not a trigger for the onset of DH in two-thirds of such patients [21]. Although it is known that anti-tTG IgA and anti-EMA antibody are specific serological markers for the diagnosis of DH and celiac disease, most Japanese patients with DH are negative for these antibodies [21, 22]. It has also been shown that the patients exhibit a good response to dapsone and do not need to avoid dietary gluten [21].

In consideration of the characteristics of the Japanese DH patients, it was difficult to relate the DH and MPGN in our patient, since the predominant renal deposits consisted of IgG class immunoglobulin, while DH is considered IgA-mediated disease. It would have been simpler to understand the etiology of this patient if the renal and dermal manifestations had derived from IgG-class autoantibodies. Re-assessment of the skin biopsy record revealed that the blister was localized in the epidermis, strongly suggesting pemphigus herpetiformis, which is an IgG-related skin disease [8, 9].

Our patient showed a favorable clinical course for the skin and renal problems following treatment with dapsone and ARB, i.e. losartan, despite the presence of massive EDDs in the glomeruli. This could support that the MPGN type 3 in this patient was not primary but a secondary derived disease. If we had not performed staining for IgG subclasses or κ/λ light chains, we would have failed to diagnose the patient correctly and might have harmed the patient by massive immunosuppression, as PGNMID.

Conclusion

Herein we report the case of a patient with MPGN type 3 associated with pemphigus herpetiformis, which mimics PGNMID and DH. It was suggested that both the renal and dermal manifestations resulted from common IgG class autoantibodies.

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Statement of Ethics

The authors have no ethical conflicts to declare.

Disclosure Statement

The authors declare that they have no conflicts of interest to disclose.

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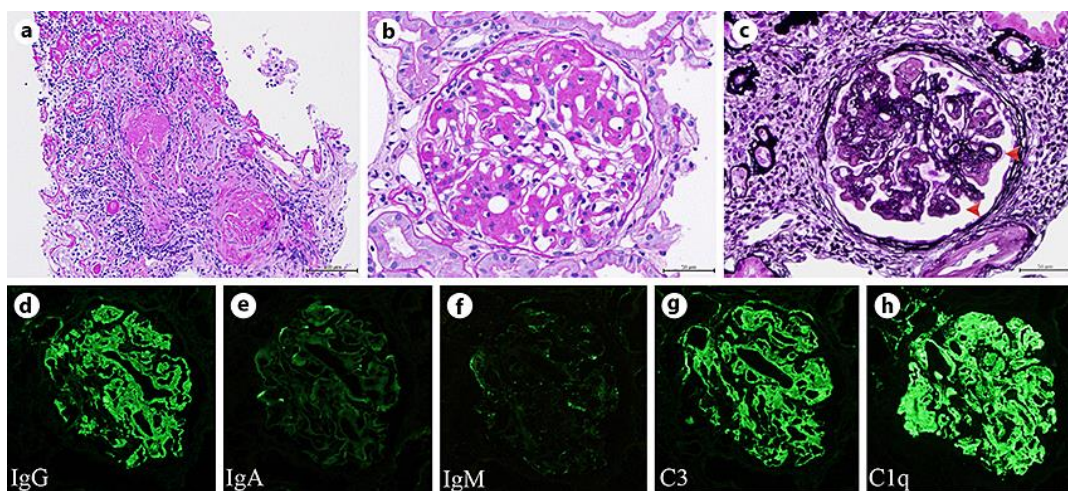


Fig. 1. LM findings of renal biopsy specimens. **a** Glomerular global sclerosis and focal tubular atrophy with interstitial fibrosis and cellular infiltration (PAS stain, ×200). **b** Lobular accentuation with segmental nodular formation in the glomerulus (PAS stain, ×200). **c** Mesangial enlargement and thickening capillary walls showing double contour (arrowheads) (PASM stain, ×400). **d–h** IF findings. Extensive granular deposits of IgG, C3 and C1q in the glomerular mesangial areas and along the glomerular capillary walls (×200).

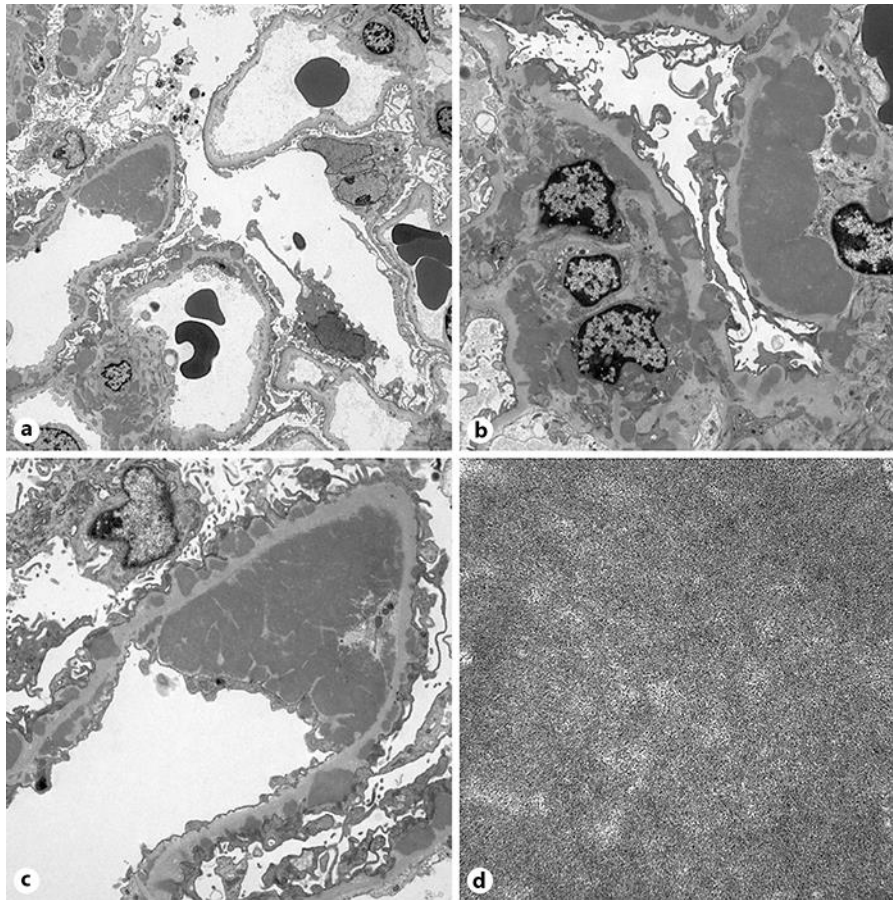


Fig. 2. Electron microscopy findings. **a** Part of a glomerulus showing epithelial foot process effacement, subendothelial, subepithelial and paramesangial EDDs. EDDs in the glomerular basement membrane (GBM) are observed ($\times 4,600$). **b** Massive EDDs in subendothelial, subepithelial and GBM. Hump-like subepithelial deposits are observed ($\times 6,400$). **c** A magnified glomerular capillary loop in Figure 3a. A large amount of EDDs in the subendothelial area ($\times 6,400$). **d** Unorganized and granular structure of EDDs ($\times 10,000$).

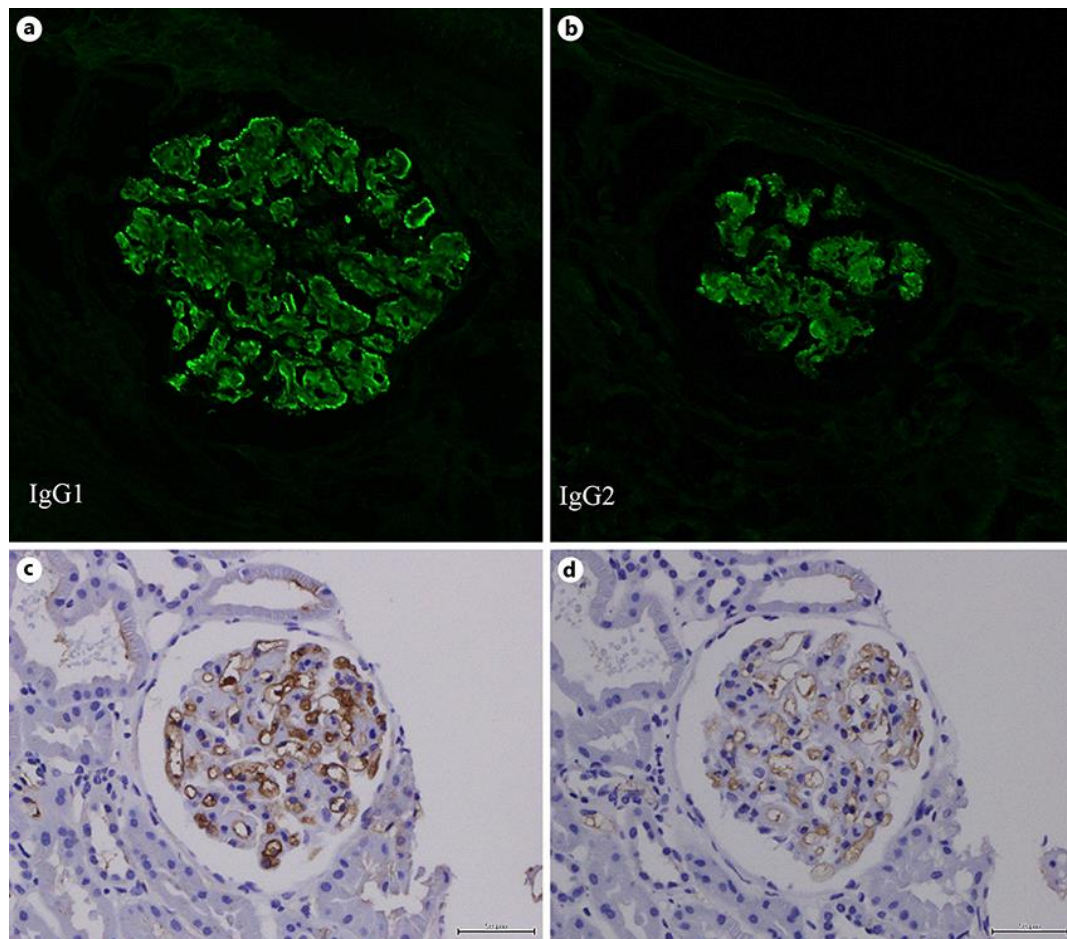


Fig. 3. **a, b** IF for IgG subclasses. Granular IgG1 and IgG2 deposits are observed in the glomerular mesangial areas and along the glomerular capillary walls ($\times 200$). **c, d** Immunohistochemistry for κ (**c**) and λ (**d**) light chain. Positive staining for both κ and λ light chains in the glomerular mesangial areas and along the glomerular capillary walls ($\times 200$).

Table 1. Laboratory findings

Urinalysis	Patient		Chemistry	Patient		Serology	Patient			
		Normal range			Normal range			Normal range		
SG	1.018	1.015–1.025	T.P	(g/dL)	6	6.3–7.8	ANA	(–)	(–)	
pH	7	5.0–7.8	Alb	(g/dL)	3.3	3.7–4.9	ADNA	(–)	(–)	
Protein	(2+)	(–)	AST	(IU/L)	20	11–40	IgG	(mg/dL)	1,624	739–1,649
Occult blood	(+)	(–)	ALT	(IU/L)	22	6–43	IgA	(mg/dL)	693.3	107–363
Glucose	(–)	(–)	LDH	(IU/L)	180	200–400	IgM	(mg/dL)	69.5	46–260
RBC	/HPF	10–19	CPK	(IU/L)	68	57–197	Cryoglobulin		(–)	(–)
WBC	/HPF	1–4	BUN	(mg/dL)	12.1	9–21	MPO-ANCA	(EU)	<10	<10
Dysmorphic RBC		(–)	Cr	(mg/dL)	0.68	0.4–0.9	PR3-ANCA	(EU)	<10	<10
Hyaline casts	/WF	(–)	UA	(mg/dL)	7.5	3.5–6.9	ant-GBM Ab	(U/mL)	<7	<7
Granular casts		(–)	Na	(mEq/L)	143	135–145	C3	(mg/dL)	80	70–130
Protein	g/day	1.2	K	(mEq/L)	4.2	3.5–4.9	C4	(mg/dL)	21.1	14–36
			Cl	(mEq/L)	106	96–108	TPHA		(–)	(–)
Peripheral blood			Ca	(mg/dL)	8.2	8.5–10.5	HBsAg		(–)	(–)
WBC	(/μL)	6,100	P	(mg/dL)	3.3	2.5–4.5	HCV Ab		(–)	(–)
RBC	(×10 ⁴ /μL)	470	CRP	(mg/dL)	0.3	<0.5	ASO	(IU/mL)	23	<240
Hb	(g/dL)	14.6	eGFR	(mL/min/1.73 m ²)	99.2					
Ht	(%)	41.1								
PLT	(×10 ⁴ /μL)	22.1								